

# Can an inhibitor of a multidrug pump become a substrate?

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Thesis submitted to the University of Nottingham for the  
degree of Master of Research



## **COVID-19 impact statement**

The research for this thesis was performed October 2019-March 2020 followed by a 7-month interruption to studies and then finished November 2020-May-2021. This was due to closure of the university labs and the break also had the impact of additional work for lab shutting down and lab set up (freezing cell lines, thawing and reviving cells etc, all adding time and further disruption).

After returning in November, safety measures limited lab occupancy to 5 people and only permitted coming into the lab when absolutely necessary for lab work. This was challenging because working from home for all non-laboratory activities required me to pre-plan all lab work from home which was difficult without access to all the lab resources e.g., checking stock of reagents. This also resulted in reduced flexibility for working in general. Additional cleaning requirements and social distancing measures both made day to day work more challenging and time consuming.

As an early stage career researcher social distancing was particularly impactful on my learning and productivity as training and supervision were both limited. This required me to work more independently and change my plans, for example I was planning to use confocal microscopy but the machine I was trained on was no longer in use due to COVID-19 restrictions.



## Abstract

ABCG2 is an ATP binding cassette (ABC) transporter that is involved in multidrug resistance, particularly anti-cancer drugs such as methotrexate and mitoxantrone. Distinguishing between substrates and inhibitors is important for rational design of drugs that won't be transported by ABCG2 and inhibitors that prevent transport of existing ABCG2 substrates. In this project, the hypothesis being proposed is that ligands with a higher affinity for ABCG2 act as inhibitors and the transient conformational changes required for transport do not occur. Whereas transported substrates have a lower affinity for ABCG2 and the conformational changes can occur. Therefore, if the affinity for a widely used inhibitor, Ko143, was reduced, would it become a substrate that is transported by ABCG2? A series of mutants (T435A, N436A, F439A, S440W, M549E, A397S/V401A/L539A and L405A/I543A/V546A) were designed using cryo-EM structures of ABCG2 bound to MZ29 (a Ko143 analogue) and mutational studies of substrate transport. The aim was to reduce affinity of ABCG2 for Ko143. All mutant proteins, except M549E, were successfully expressed in HEK293T cells and trafficked to the cell membrane. Using flow cytometry, cellular accumulation of fluorescent Ko143 derivatives (Ko143-Cy5 and Ko143-X-BY630) was measured which indicates whether they are exported by ABCG2 or not. Ko143-Cy5 fluorescence was significantly higher in WT-ABCG2 expressing cells compared with the untransfected control and the other ABCG2 mutants, except for A397S/V401A/L539A. Ko143-X-BY630 showed a similar pattern but without significance. The reduction in cellular accumulation of the fluorescent Ko143 derivatives in the mutants could be caused by the reduced affinity leading to ABCG2-mediated transport or increased diffusion out of the cell. Transport of Ko143-Cy5 or Ko143-X-BY630 cannot be ruled out but it is not detectable in this

experiment. Consideration of the data in this thesis alongside emerging structural and functional data from other laboratories will continue to shed light on the interaction of substrates and inhibitors with ABCG2.

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Also, a huge thank you to my boyfriend, Joe Davies, for being a constant source of motivation and for encouraging me to do a Master's in the first place.



## **Declaration**

This thesis, “Can an inhibitor of a multidrug pump become a substrate?”, is the result of my own work undertaken during my period of registration at the University of Nottingham under the supervision of Dr Ian Kerr. Technical assistance, and collaborations where relevant, has been acknowledged.

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## Abbreviations

<b>ABC</b>	ATP-binding cassette
<b>ABCG2</b>	ATP-binding cassette subfamily G member 2
<b>ABCG5/G8</b>	ABCG5 heterodimerised with ABCG8
<b>ADP</b>	Adenosine diphosphate
<b>AMP-PNP</b>	Adenylyl-imidodiphosphate
<b>ANOVA</b>	Analysis of variance
<b>APS</b>	Ammonium persulfate
<b>ATP</b>	Adenosine triphosphate
<b>BLAST</b>	Basic local alignment search tool
<b>bp</b>	Base pair
<b>BSA</b>	Bovine serum albumin
<b>CFTR</b>	Cystic fibrosis transmembrane conductance regulator
<b>CMV</b>	Cytomegalovirus
<b>Cryo-EM</b>	Cryogenic electron microscopy
<b>DHEAS</b>	Dehydroepiandrosterone sulfate
<b>DMEM</b>	Dulbecco's modified eagle medium
<b>DMSO</b>	Dimethyl sulfoxide
<b>DNA</b>	Deoxyribose nucleic acid
<b>dNTP</b>	Deoxynucleotides
<b>E<sub>1</sub>S</b>	Estrone 3-sulfate
<b>E3040S</b>	E3040 sulfate
<b>EC<sub>50</sub></b>	Concentration at which a drug evokes half maximal response
<b>EDTA</b>	Ethylenediaminetetraacetic acid
<b>FACS</b>	Fluorescence-activated cell sorting
<b>FBS</b>	Foetal bovine serum
<b>FSC</b>	Forward scatter
<b>FTC</b>	Fumitremorgin C
<b>GFP</b>	Green fluorescent protein
<b>GPCR</b>	G protein-coupled receptor
<b>HBSS</b>	Hank's balanced salt solution
<b>HDL</b>	High density lipoprotein
<b>HEK293S</b>	Human embryonic kidney 293S cell line

<b>HEK293T</b>	Human embryonic kidney 293S cell line
<b>IC<sub>50</sub></b>	Half-maximal inhibitory concentration
<b>IgG</b>	Immunoglobulin G
<b>kb</b>	Kilo bases (1000 base pairs)
<b>kDa</b>	Kilo Daltons
<b>LB</b>	Luria-Bertani
<b>NBD</b>	Nucleotide binding domain
<b>NCBI</b>	National Center for Biotechnology Information
<b>PBS</b>	Phosphate buffered saline
<b>PBS-T</b>	Phosphate buffered saline with 0.1% (v/v) Tween-20
<b>PCR</b>	Polymerase chain reaction
<b>PEI</b>	Polyethyleneimine
<b>P<sub>i</sub></b>	Inorganic phosphate
<b>SDS</b>	Sodium dodecyl sulfate
<b>SDS-PAGE</b>	Sodium dodecyl sulfate polyacrylamide gel electrophoresis
<b>SEM</b>	Standard error of the mean
<b>SSC</b>	Side scatter
<b>SV40</b>	Simian vacuolating virus 40
<b>TBE</b>	Tris/borate/EDTA
<b>TEMED</b>	N,N,N',N'-tetramethylethane-1,2-diamine
<b>TLC-S</b>	Taurolithocholate sulfate
<b>TM</b>	Transmembrane helix
<b>TMD</b>	Transmembrane domain
<b>UV</b>	Ultraviolet
<b>WT</b>	Wild type
<b>4-MUS</b>	4-methylumbelliferone sulfate
<b>ΔMFI</b>	Change in median fluorescence intensity

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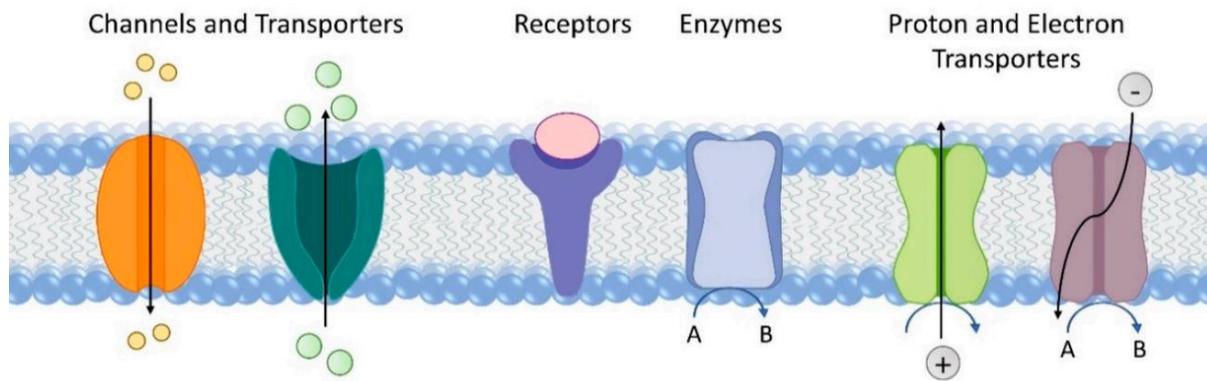
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# Chapter 1 Introduction

## 1.1 Membrane transporters

All forms of life have a lipid membrane of some sort to contain their molecular components to allow biochemical reactions to take place. Without membranes, the compartmentalisation necessary for the vast majority of cellular processes cannot occur. Archaea, bacteria and eukaryotes have lipid bilayers which act as a barrier so that what enters and exits the cell can be controlled. In eukaryotic organisms, different cellular processes are facilitated and regulated by their spatial separation in organelles.

Integral membrane proteins have sections that are inserted into the cell membrane and can perform a variety of roles (Figure 1.1). For example, flippases and floppases can alter the shape and lipid composition of the cell membrane by transferring lipids from the inner layer to the outer layer or vice versa. Receptors such as G-protein coupled receptors (GPCRs) can aid in the communication between cells, for example by triggering signalling cascades when a signal molecule binds. Transporter and channel proteins control what enters and exits the cell or organelle. This can include transport of charged ions, for example cystic fibrosis transmembrane conductance regulator (CFTR) exports  $\text{Cl}^-$  from endothelial cells to control mucus consistency and in neurons  $\text{Na}^+$  influx can trigger an electrical impulse. Larger molecules such as peptides, amino acids, nucleotides and phospholipids can also be transported. Transporters can also have a protective effect by exporting potentially harmful xenobiotics back into the lumen of the gastrointestinal tract or away from the foetus when expressed in the placenta (Horsey et al., 2016). This is the role of the membrane transporter, ABCG2, studied in this thesis.



**Figure 1.1 Schematic of the cell membrane.** The lipid bilayer (blue spheres and tails) contain different types of membrane proteins involved in signalling, enzymology and substrate flux. Image retrieved from (Zhang et al., 2020)

### 1.1.1 Facilitated diffusion vs active transport

Some molecules can cross the cell membrane unaided because they are small and hydrophobic so can easily diffuse through the hydrophobic lipid bilayer. Movement of these molecules occurs down the concentration gradient, from a more concentrated cytosol to the less concentrated extracellular space, or vice versa. More polar compounds (e.g. water) or ions (e.g.  $\text{Na}^+$  or  $\text{H}^+$ ) cannot diffuse across the cell membrane without the help of channels or carrier proteins by facilitated diffusion. These proteins selectively facilitate the diffusion of chemical entities through a more suitable chemical environment. Flux can be controlled, for example voltage gated  $\text{Na}^+$  channels undergo conformational changes to open the channel when membrane depolarisation occurs (de Lera Ruiz and Kraus, 2015).

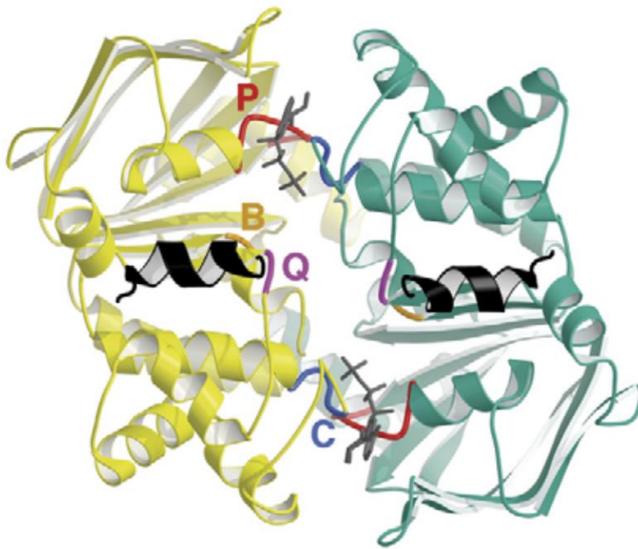
Active transport can be against a concentration gradient and the transported compound is “pumped” in or out of the cell. An important step in the transport cycle of primary active transporters is ATP hydrolysis, which provides the energy for movement against the concentration gradient. An example of proteins that perform active transport are ATP-binding cassette (ABC) transporters which are described in

more detail below. The subject of this project, ABCG2, is part of this family of proteins. The true substrate of active transport proteins is ATP because its hydrolysis to produce ADP and  $P_i$  is catalysed by the transporter protein, however, in this thesis, the substrate will refer to the transported compound as opposed to an inhibitor of the pump.

### **1.1.2 ATP-binding cassette transporters**

The ABC family of proteins are mostly membrane proteins: in humans ABCA, ABCB, ABCC, ABCD and ABCG subfamilies are membrane associated but ABCE and ABCF are not (Kerr et al., 2011). The basic structure of an ABC transporter is two nucleotide binding domains (NBDs) and two transmembrane domains (TMDs), however some ABC transporters, named “half-transporters”, must dimerize in order to achieve this formation. The NBDs are highly conserved, even between eukaryotic and prokaryotic transporters of otherwise unrelated function. They contain several motifs that are essential for ATP-hydrolysis, which is what drives the active transport. Figure 1.2 shows that the P loop (or Walker-A motif) from one NBD and the LSGGQ motif (or C-loop) from the other NBD contact the nucleotide, resulting in two ATP molecules bound at two separate points along the NBD:NBD interface (Jones and George, 1999, Hollenstein et al., 2007). The Walker-B motif contains a catalytic glutamate which is capable of performing a nucleophilic attack on ATP via a water molecule (Hollenstein et al., 2007). The TMDs, which span the cell membrane, are less conserved and relate more to the specificity for the transport substrate and perform conformational changes required for substrate transport. TMDs contain a coupling helix (black helices, Figure 1.2) that contact the NBD (Q loops) to couple ATP hydrolysis to the conformational changes required for substrate transport

(Hollenstein et al., 2007). ATP binding and hydrolysis is coupled to inward and outward facing conformations of the transporter TMDs which in the case of exporters, allows binding of substrate on the intracellular side and release on the extracellular side (Hollenstein et al., 2007, Manolaridis et al., 2018).



**Figure 1.2 The NBDs of the ABC transporter Sav1866.** The NBDs of Sav1866 crystallised with bound AMP-PNP, a non-hydrolysable ATP analogue, shown in stick representation. There are 2 bound AMP-PNP molecules per NBD dimer. The NBDs are shown as if looking down from the membrane. The 2 NBDs are in green and yellow and mechanistically important sequence motifs are shown with single letter: red P (P-loop), yellow B (Walker-B motif), purple Q (Q-loop), blue C (LSGGQ or C-loop). Bound AMP-PNP is shown as grey sticks and coupling helices from the TMDs are the short black helices. Figure adapted from Hollenstein et al. (2007)

## 1.2 ABCG subfamily

### 1.2.1 Overview

The ABCG subfamily is part of the ABC family of proteins and the 5 members of this subfamily all share a common ancestral gene. Besides the conserved ABC motifs, there is very little protein sequence homology between the ABCG proteins, except G1 and G4 which have 72% homology (Kerr et al., 2011). This plays down to the

TMDs being largely non-conserved, for substrate selectivity (Hollenstein et al., 2007). The ABCG family all transport lipids and, with the exception of ABCG2, have a narrower functionality than other ABC subfamilies (Kerr et al., 2011, Kerr et al., 2021). All ABCG proteins are half-transporters because they only consist of one TMD and one NBD which requires them to at least dimerize to function. This is so that the ATP binding site can form at the interface between two NBDs (section 1.1.2). ABCG5 and ABCG8 form an obligate heterodimer to complete this binding site whereas ABCG2 homodimerises but higher order oligomeric structures have been observed (Wong et al., 2016, Kerr et al., 2011). ABCG1 and ABCG4 can homodimerise but have also been shown to heterodimerise with each other *in vitro* (Hegyí and Homolya, 2016).

ABCG1 has widespread expression and is located in the brain alongside ABCG4 and both contribute to cholesterol and desmosterol efflux to high-density lipoprotein (HDL) from astrocytes (Wang et al., 2008). ABCG5/G8 is involved in limiting absorption of dietary sterols by localising in the apical membranes of hepatocyte canaliculi and gall bladder epithelial cells. ABCG2 has a broad specificity, having a wide range of substrates structurally unrelated to each other. This is useful in its protective role against xenobiotics (Kerr et al., 2011). Its function, structure and mechanism are described in more detail below.

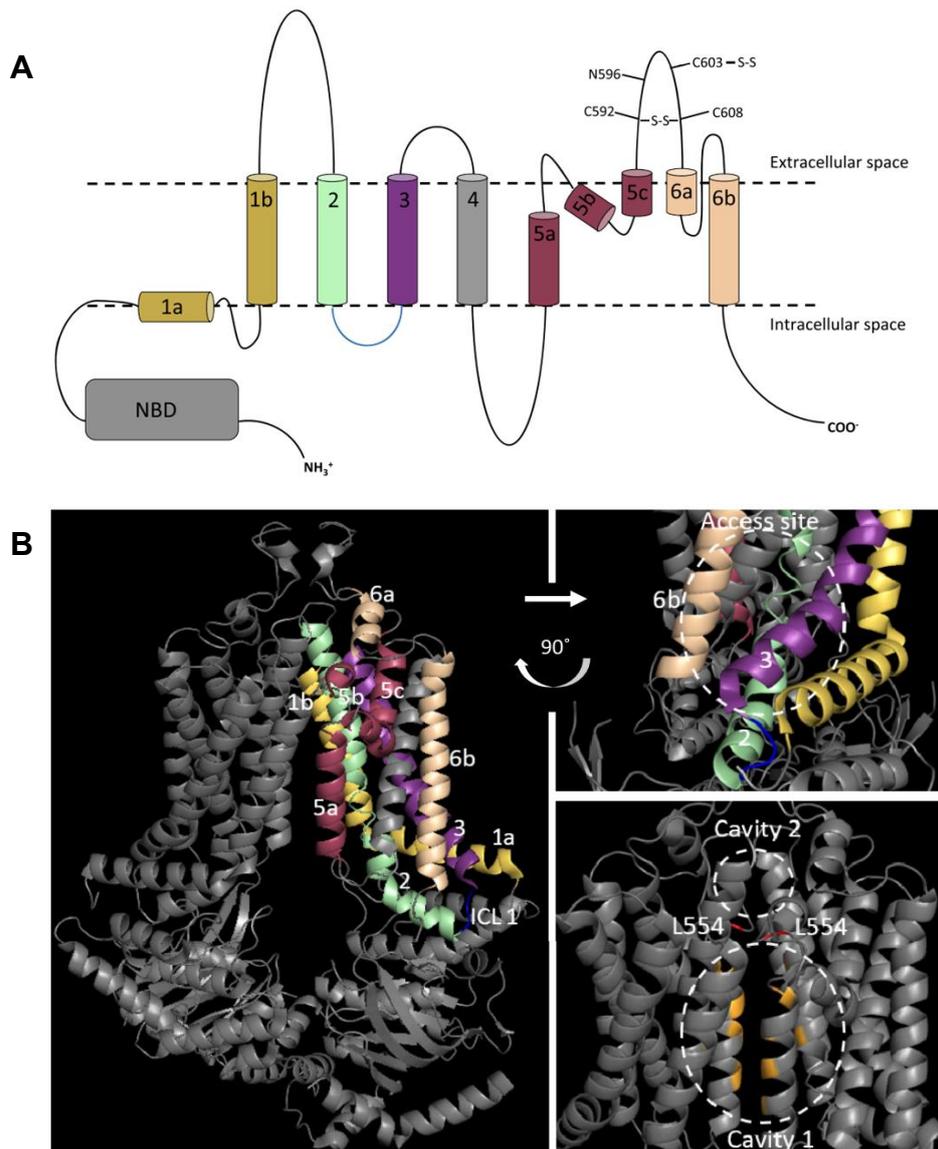
### **1.2.2 ABCG2 function and clinical implications**

ABCG2 is an important protein to study because of its role in multidrug resistance. Its natural role as a defence against environmental toxins means that some drugs fall victim to this multidrug pump. Expression of ABCG2 on apical membranes of epithelial cells of the gastrointestinal tract, liver canalicular membranes and the apical

membrane of proximal tubular cells in the kidneys, contributes to limiting absorption and promoting excretion of xenobiotics (Horsey et al., 2016). Expression in placental syncytiotrophoblasts and in mammary glands have contradictory roles, with ABCG2 pumping xenobiotics away from the foetus and into milk respectively. Along with reduced uptake and increased excretion, ABCG2 contributes to the failure of some anti-cancer drugs by its overexpression in cancer cells. ABCG2 expression has been linked to poor outcome in acute myeloid leukaemia, diffuse large B-cell lymphoma, and lung and oesophageal cancer (Horsey et al., 2016). Overexpression of ABCG2 has also been found in other cancer cells exhibiting a multidrug resistance phenotype, including topotecan-selected ovarian tumour cell line T8 and gefitinib-resistant non-small cell lung cancer (NSCLC) cells (Mo and Zhang, 2012). This is why it is so important to study ABCG2 further, to be able to design drugs that won't be exported by this multidrug transporter. Drugs that have been shown to be transported by ABCG2 include: methotrexate, mitoxantrone, pheophorbide A, topotecans, flavopiridol, imatinib, gefitinib, nilotinib and others (Homolya et al., 2011, Volk and Schneider, 2003, Kapoor et al., 2018, Robey et al., 2005). Multidrug resistance is caused because ABCG2 limits absorption, increases excretion and lowers cellular concentration of these drugs. An endogenous substrate is urate and successful transport by ABCG2 leads to urate excretion. A naturally occurring single nucleotide polymorphism, rs2231142, which results in Q141K mutant ABCG2, is a loss of function mutation causing an increased level of serum urate which is linked to hyperuricemia and gout (Woodward et al., 2009).

### 1.2.3 ABCG2 structure and mechanism

ABCG2 contains one transmembrane domain (TMD), which consists of six transmembrane helices (TM1-TM6), and one nucleotide binding domain (NBD), which is cytoplasmic (Kapoor et al., 2018). Since two NBDs are required for ATP hydrolysis to occur, dimerisation is required for the protein to be functional. Higher forms of oligomerisation, such as tetramers, have been observed, although the physiological relevance is unclear (Wong et al., 2016). In the TM5-TM6 extracellular loop region (Figure 1.3) there is an intramolecular disulfide bond (C592 and C608), an intermolecular disulfide bond (C603 from each monomer) and a glycosylation site (N596) (Diop and Hrycyna, 2005, Henriksen et al., 2005, Kapoor et al., 2018). The mutation of C603 does not prevent surface expression of the protein or affect its function so presumably the C603 inter-molecular disulfide bond just has a small stabilising effect. However, mutation of C592 or C608 did impact stability and trafficking of ABCG2 to the cell membrane (Henriksen et al., 2005). Mutation of N596 prevents glycosylation which leads to increased ubiquitin-mediated proteasomal proteolysis (Nakagawa et al., 2009).



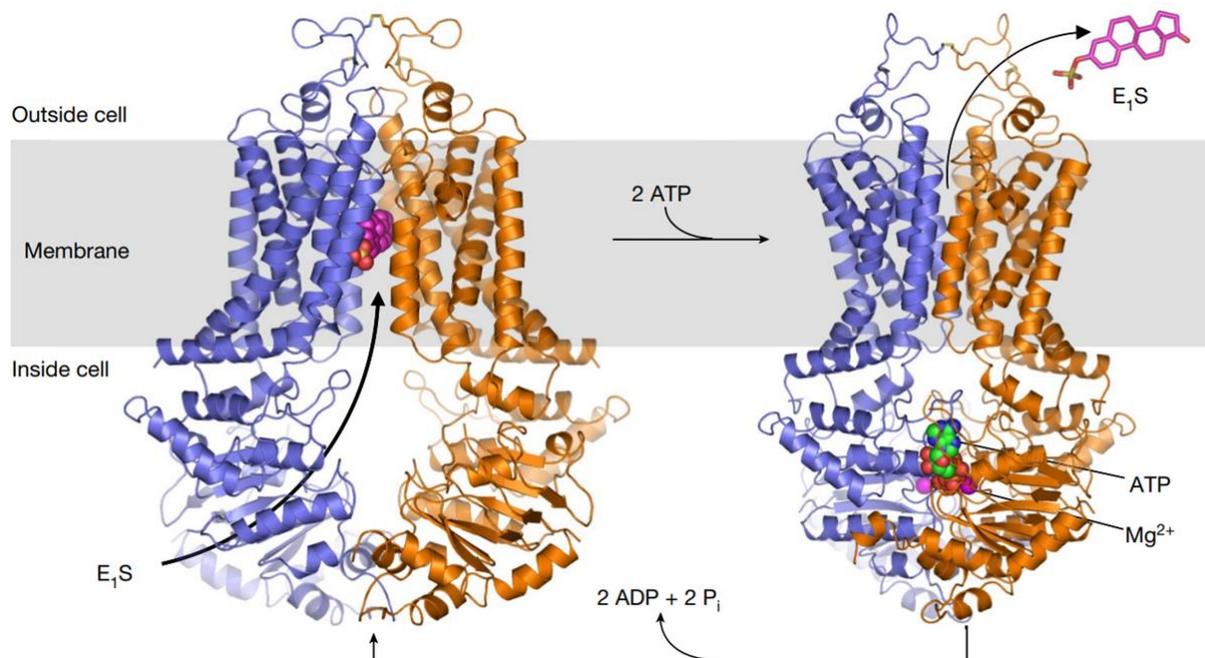
**Figure 1.3 ABCG2 topology and structure. (A)** Schematic representation of the topology of ABCG2 within the cell membrane. The transmembrane helices 1-6 (TM1-TM6) are shown in different colours. Key residues for intramolecular (C592 and C608) and intermolecular (C603) disulfide bonds and glycosylation (N596) are shown. **(B)** Cartoon representation of the ABCG2 dimer from PDB 6ETI (Jackson et al., 2018). TM1-6 are the same colours as in A. Rotation by 90° gives a clearer view of the “access site” which is surrounded by TM2, TM3 and TM6b. The bottom right panel shows the “leucine plug” (L554, red), cavity 1 and cavity 2. Figure taken from (Kapoor et al., 2018).

There are multiple binding sites for substrates, shown in Figure 1.3. The main site that the substrate binds to before being transported is cavity 1, which is located between the two ABCG2 monomers and surrounded by TM2 and TM5 (László et al.,

2016). Cavity 2 is on the extracellular side of ABCG2 and is the final binding site before the substrate is released into the extracellular space. Cavity 1 and cavity 2 are separated by a “leucine plug” which consists of L554 from each ABCG2 monomer and blocks movement of the substrate into cavity 2 before conformational changes occur (Taylor et al., 2017). Both these cavities are described by structural biology data (see below). A third binding site, referred to as the “access site” and predicted by mutagenesis studies and molecular modelling, is located between TM2, TM3 and TM6 and includes the residue R482 which has been linked to substrate specificity (Kapoor et al., 2018). The R482G mutation leads to broader substrate specificity, with drugs such as doxorubicin, daunorubicin and rhodamine 123 becoming transported when they otherwise would not (Ozvegy-Laczka et al., 2005, Tamura et al., 2007). Another substrate binding site (not shown) has also been predicted by molecular docking (László et al., 2016).

In recent years, cryo-EM structures have become available which has helped elucidate the structure and mechanism of ABCG2. The first was published by Taylor et al. (2017), which showed ABCG2 locked in an inward facing conformation by an inhibitory anti-ABCG2 antibody, 5D3, on the extracellular side. The NBDs did not interact to form the “ATP sandwich dimer” as no nucleotide was bound (as described in Figure 1.2) but were still in contact through a novel ABCG-family specific NBD:NBD interaction surface, consistent with other inward facing structures (Manolaridis et al., 2018, Jackson et al., 2018). Manolaridis et al. (2018) solved two further structures of ABCG2: the E211Q mutant (ABCG2<sub>E211Q</sub>) bound to either the transport substrate estrone 3-sulfate (E<sub>1</sub>S) or to the catalytic substrate ATP. This helped shed light on the mechanism of transport by ABCG2. As seen in Figure 1.4, the E<sub>1</sub>S-bound ABCG2<sub>E211Q</sub> adopts an open, inward facing conformation with the

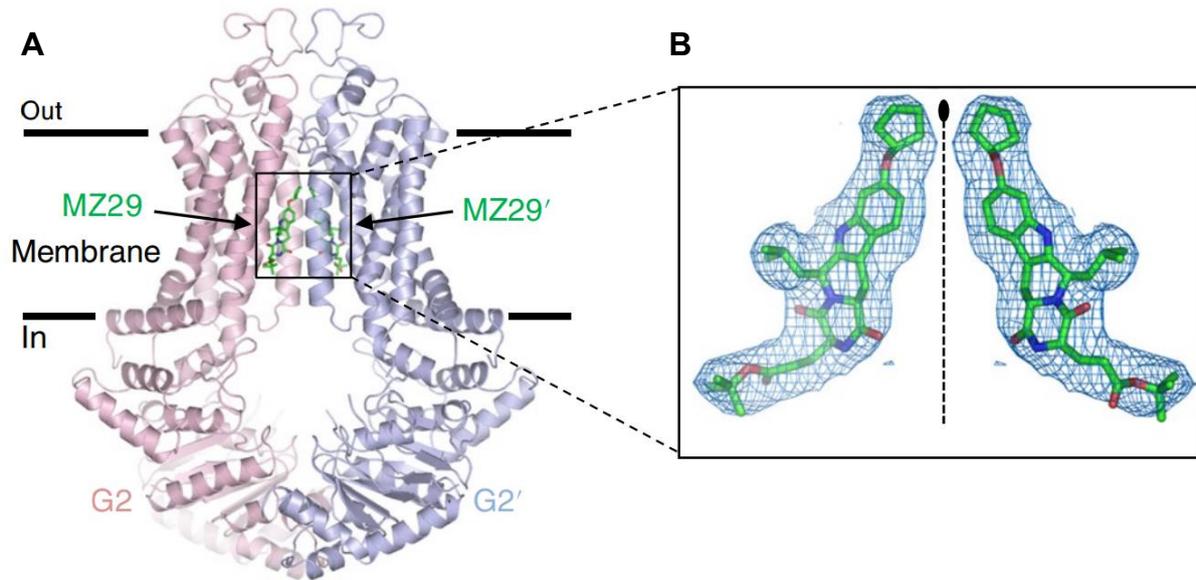
substrate located in cavity 1. The ATP-bound ABCG2<sub>E211Q</sub> shows a closed, outward facing conformation, where cavity 1 is completely closed, forcing the substrate towards cavity 2 and then the substrate is released on the extracellular side. The NBDs in this structure do dimerise in the classical fashion (Figure 1.2) with ATP-bound at the interface (Manolaridis et al., 2018). This structure is consistent with the idea that conformational change and transport of substrate occurs on ATP binding and not on ATP hydrolysis, as previously predicted by radioligand binding studies (McDevitt et al., 2008). ATP hydrolysis likely resets the inward facing conformation (Manolaridis et al., 2018).



**Figure 1.4 Mechanism of substrate transport by ABCG2.** Cartoon representation of E<sub>1</sub>S (substrate)-bound (left, PDB 6HCO) and ATP-bound (right, PDB 6HBU) ABCG2<sub>E211Q</sub>. ABCG2<sub>E211Q</sub> monomers are displayed in blue and orange. Figure taken from Manolaridis et al. (2018).

### 1.2.4 Inhibitors

Inhibitors of ABCG2 prevent transport of substrates and therefore increase their cellular accumulation. This would also apply to drug molecules which are substrates, helping prevent multidrug resistance. There are currently no clinically available ABCG2 inhibitors but Ko143 is widely used in research. Ko143 is derived from a fungal toxin, fumitremorgin C (FTC), which is highly neurotoxic. Ko143 is more potent and less toxic, although still not clinically utilisable (Allen et al., 2002, Toyoda et al., 2019). Jackson et al. (2018) published the only current structures of ABCG2 bound to inhibitors: MZ29 (PDB 6ETI) and MB136 (PDB 6FEQ). MZ29 is a Ko143 derivative and binds to cavity 1, just as substrates do (Figure 1.5). Two MZ29 molecules bind between TM1b and TM2 of one monomer and TM5a of the other and vice versa. This was concordant with data indicating that a 2:1 inhibitor:ABCG2 molar ratio was required for full inhibition of ATPase activity (Jackson et al., 2018). The larger MB136 binds in the same location but only one molecule fits in cavity 1 and a 1:1 ratio is sufficient for maximum inhibition (Jackson et al., 2018). Since stable electron densities were found for these inhibitors in absence of the stabilizing antibody, 5D3, transport of these molecules is minimal. Binding of other inhibitors, such as febuxostat and elacridar, has not been structurally determined so it is unclear whether they inhibit allosterically (i.e. at a site distinct from cavity 1) or orthosterically (i.e. at cavity 1)(Toyoda et al., 2019).



**Figure 1.5 Structure of inhibitor-bound ABCG2.** (A) Cartoon representation of the ABCG2 homodimer from PDB 6ETI. MZ29 (green sticks), a Ko143 derivative, is bound between the two ABCG2 monomers (pink and blue) in cavity 1. (B) Rotated (45°) view of the bound MZ29 molecules (green sticks) with their electron microscopy density (blue). The dotted line represents the two-fold symmetry axis. Both A and B were retrieved from Jackson et al. (2018).

### 1.3 Project aims

Distinguishing the features of ABCG2 inhibitor binding from features of ABCG2 substrate binding would be an important step in tackling multidrug resistance. The aim of this project is to use the cryo-EM structures of inhibitor-bound ABCG2 to identify key residues involved in inhibitor binding. Then relevant mutations of ABCG2 will be made and assessed for whether the mutations prevent inhibition or even transforms the inhibitor into a substrate.

## Chapter 2 Materials and Methods

### 2.1 Materials and reagents

All molecular biology reagents were purchased from New England Biolabs (NEB, Hitchin, UK) and Promega (Southampton, UK), unless stated otherwise. The primers for site directed mutagenesis were ordered from Sigma-Aldrich (Poole, UK). All other cell culture materials and reagents, except Zeocin™ (Invitrogen) and polyethyleneimine (PEI, Polyscience Inc.), were obtained from Sigma-Aldrich or Thermo Fisher Scientific unless otherwise mentioned.

### 2.2 Molecular biology

#### 2.2.1 Site-directed mutagenesis

Mutations were introduced into a WT-ABCG2 expression vector by performing mutagenic polymerase chain reactions (PCR). These were set up using mutagenic primers shown in Table 2.1 (0.5 µM), dNTPs (0.2 mM), pcDNA™3.1/Zeo(+)\_SNAP\_ABCG2<sup>1</sup> template (50ng), Pfu polymerase (1.0-1.5 U), and polymerase buffer made up to 50 µL. Denaturation, annealing and elongation steps were performed using a SensoQuest Thermocycler and cycled 18 times at the temperatures shown in Table 2.2 (with the annealing temperature varied until successful amplification was observed). Presence of amplified PCR product was confirmed by agarose gel electrophoresis. PCR products were mixed with Gel

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<sup>1</sup> Zeo(+) refers to presence of a Zeocin™ resistance gene (BleoR). SNAP refers to the SNAP-tag® which is a modified alkylguanine DNA alkyltransferase which can specifically and covalently bind to benzyl guanine fluorophores as a way of tagging the protein of interest (Tirat et al., 2006). However, labelling of ABCG2 was not used in this project.

Loading Dye and loaded onto a 1% (w/v) agarose gel which was prepared in TBE buffer (10.8% (w/v) Tris, 5.5% (w/v) boric acid, 20 mM Na<sub>2</sub>EDTA) and contained ~1 µg/mL ethidium bromide. The resolved gels were then viewed under UV light (using a Syngene GeneGenius gel imaging system).

**Table 2.1 Forward primers used in the generation of the mutant ABCG2 constructs.** Codon changes that result in an amino acid change are highlighted in yellow, uppercase letters indicate bases that are different from the template and lowercase letters identical to the template. The complementary reverse primers are not shown but have identical length, T<sub>m</sub> and GC.

Name	Sequence (5'-3')	Length (bp)	T <sub>m</sub> (°C)	GC (%)
T435A	cttcttctgacgGcTaaaccagtggttc	28	68.95	50.0
N436A	cttctgacgaccGCcagtggttc	25	71.82	60.0
F439A	gaccaaccagtgGccagcagtggttc	27	71.78	55.56
S440W	caaccagtggttTgGagtggttcagcc	28	70.13	46.43
M549E	ctgttttgtggttCatgGAgattttttcaggtctg	34	72.81	38.24
A397S/V401A	gcctctataTctcagatcattgCcacagtcgtactg	36	74.46	47.22
L539A	ctgtagcaacaGctctTatgaccatctg	28	65.17	46.43
L405A	gtcacagtcgtaGCgggactggttatag	28	67.79	53.56
I543A/V546A	cttctcatgaccGCctgttttgCCtttatgatgatt	36	77.65	41.67

**Table 2.2 PCR thermocycling parameters.** Steps 2-4 were cycled 18 times before proceeding to step 5, as demonstrated by the arrow. Annealing temperatures were varied (50-65 °C) if these standard parameters were unsuccessful.

Step	Temperature (°C)	Time (min)	Number of cycles
1. Heat	95	1	1
2. Denature	95	1	18
3. Anneal	55	1	18
4. Extend	72	12	18
5. Extend	72	10	1
6. Cool	10	∞	1

### 2.2.2 *DpnI* digest and transformation

To remove template DNA, PCR products were digested with *DpnI* (20 units) for 90 minutes at 37 °C. The enzyme was then deactivated by incubating at 80 °C for 20 min. *DpnI* digested-PCR product was transformed into DH5α competent *E. coli*. First 100 µL of DH5α cells were thawed on ice and 5 µL of the *DpnI* digested PCR product was added. The competent cell mixture was then incubated on ice for 1 hour, heat

shocked at 42 °C for 1 min, and then placed back on ice for 2 min. The cells were then supplemented with 250 µL Luria-Bertani (LB) medium (1% (w/v) tryptone, 1% (w/v) NaCl, 0.5% (w/v) yeast extract) and shaken at 37 °C for 1 hour. All 350 µL was subsequently spread onto LB-agar plates (1.5% (w/v) agar, 100 µg/mL ampicillin) and incubated overnight at 37 °C.

### **2.2.3 Plasmid preparation**

Single colonies from LB-agar plates were grown overnight at 37 °C with shaking, after being picked and inoculated into 5 mL of ampicillin-supplemented (100 µg/mL) LB medium. For long-term storage, the resultant bacterial cultures were stored at -80 °C as glycerol stocks (500 µL of bacterial culture, 500 µL of 30% (v/v) glycerol). The remainder of the cultures were centrifuged for 5 mins at ~3000 x g and the supernatants were discarded. Plasmid DNA was then extracted from the resulting bacterial pellet using a NucleoSpin® Plasmid kit (Macherey-Nagel). The manufacturer's protocol was followed except instead of centrifuging at 11,000 x g, the samples were centrifuged at 13,000 x g.

DNA plasmid concentration and purity were determined using the Nanodrop 2000® (Thermo Fisher Scientific). The purity was confirmed by the A260/A280 ratio, where a value of >1.7 was deemed acceptable for future transfection.

### **2.2.4 Sequencing**

To confirm presence of the desired experimental mutations and absence of other mutations, the purified DNA underwent Sanger sequencing. The mutant constructs were sent to Source Bioscience (Nottingham, UK) along with the primers shown in Table 2.3. The quality of the chromatograms was analysed in Chromas

(Technelysium Pty Ltd) and the sequences were aligned with the template sequence using BLAST (NCBI) to ensure only the desired mutation was incorporated plasmid DNA.

**Table 2.3 Sequencing primers.** A series of primers were used to sequence the full SNAP-ABCG2 coding region (bases 1026-3543 of the pcDNA™3.1/Zeo(+)\_SNAP\_ABCG2 plasmid). “Region sequenced” refers to the minimum region where sequencing data was of a high quality for all mutants.

Primer name	Sequence (5'-3')	Region sequenced (bp)
T7 promoter (F)	TAATACGACTCACTATAGGG	978-1992
SeqF1	CACAGGTGGAGGCAAATCTT	1935-2852
SeqF2	GCAGGGACGAACAATCATCT	2361-3310
Seq482	AACTCTTTGTGGTAGA	3028-3749

## 2.3 Cell culture

### 2.3.1 Maintenance of cell cultures

HEK293T cells were grown in T25 (25 cm<sup>2</sup>) flasks at 37 °C, 5% CO<sub>2</sub> in Dulbecco's modified eagle medium (DMEM, 4500 mg/L glucose, L-glutamine, sodium pyruvate, and sodium bicarbonate) supplemented with 10% (v/v) foetal bovine serum (FBS) and 1% (v/v) penicillin-streptomycin (P/S, 100 U/mL penicillin, 100 µg/mL streptomycin). Once cells reached 70-90% confluency based on visual inspection, the cultures were passaged (typically twice a week). Media was removed, cells were washed once with pre-warmed phosphate buffered saline (PBS) and incubated with 0.5 mL trypsin/EDTA for 1-3 min. The trypsin/EDTA was then quenched with 4.5 mL of DMEM and cells were then detached by repeated pipetting, before being centrifuged 225-250 x g for 5 min. The cell pellets were then resuspended in DMEM and reseeded at a typical dilution of 1/10.

### 2.3.2 Transfection

Cells were seeded at  $1.25 \times 10^5$  cell/mL (determined using a haemocytometer) into a 6-well plate, ~24 hours prior to transfection. Two hours before transfecting, the media was replaced with 5% serum media (DMEM, 5% FBS, 0.5% P/S). Cells were transfected using linear polyethylenimine (PEI) at a molar PEI nitrogen:DNA phosphorus ratio of 15:1. Transfection mixtures were made by the addition of 9  $\mu$ L of PEI (from a 10 mM working stock solution) to 2  $\mu$ g DNA, as described by Cox et al. (2018), and were left no longer than 5 minutes before adding 100  $\mu$ L of media and dropwise addition to the HEK293T cells. 24 hours later media was replaced with 10% FBS-supplemented media.

### 2.3.3 Zeocin™ selection

Transfected cells were transferred to T25 flasks by trypsinization (described in 2.2.1 and scaled down appropriately). Flasks contained 5 mL DMEM (10% FBS, 1% P/S) and approximately 5 hours later, Zeocin™ was added to total concentration of 200  $\mu$ g/mL. Media was changed every 2-3 days and Zeocin™ concentration was maintained at 200  $\mu$ g/mL for several weeks until growth of Zeocin™ resistant colonies was observed. Transfected cells expressing sfGFP-ABCG2 were employed as a control for Zeocin™ selection. sfGFP tagged proteins were chosen because protein expression (fluorescence) could be monitored under an epifluorescence microscope without addition of fluorescent reagents. This helped confirm when the other cells lines were stably transfected. At this point, Zeocin™ concentration was dropped to 40  $\mu$ g/mL.

### **2.3.4 Epifluorescence microscopy**

Expression of GFP-ABCG2 in pcDNA™3.1/Zeo(+)\_sfGFP\_ABCG2 transfected HEK293T cells was monitored with epifluorescence microscopy. Images were obtained using the Zeiss Axiovert S100 microscope and Zeiss AxioCam MRm monochrome digital camera and processed with AxioVision version 4.8.2 SP3.

### **2.3.5 Long-term storage**

For long-term storage, 80% of cells in a T25 flask were resuspended in 4 mL ice cold freezing medium (10% (v/v) DMSO in FBS) and aliquoted into 1 mL cryovials. The other 20% were maintained as described in 2.2.1. The cryovials were then frozen slowly (-1 °C/min) by storage in a precooled (4 °C) Mr Frosty™ (containing isopropanol) in a -80 °C freezer. Frozen cryovials were transferred to liquid nitrogen for longer term storage. When needed, cells were rapidly thawed by pipetting up and down with warm media, centrifuged at 225-250 x g, resuspended in 5 mL DMEM (10% FBS, 1 %P/S) and maintained in a T25 flask as described in 2.2.1.

## **2.4 SDS-PAGE and western blotting**

### **2.4.1 Cell harvesting and lysis**

Cell culture monolayers were washed once with PBS, detached with Trypsin/EDTA and centrifuged at 235 x g for 5 minutes (as described in 2.2.1). The pellet was then washed once with PBS and stored at -80 °C if not required immediately. Pellets were resuspended in 250 µL ice cold lysis buffer consisting PBS supplemented with 10% (v/v) glycerol and EDTA-free protease inhibitor cocktail III (Calbiochem, 1:200 dilution). Cells were then sonicated at 40% output for 4 x 5 seconds, being stored on ice for at least 2 minutes between bursts.

### 2.4.2 Protein concentration determination

To ensure equal protein loading in sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and western blot analysis, a modified Lowry assay was performed. Using a Bio-Rad DC protein assay kit and a standard curve of 0-10 µg bovine serum albumin (BSA), total protein concentration of the cell lysates was determined. All samples and the standard curve were analysed in duplicate.

### 2.4.3 Sample preparation and SDS-PAGE

SDS-PAGE was performed as described by Laemmli (1970). Equal quantities of cell lysate (typically 20-100 µg) were incubated with protein loading buffer (50 mM Tris-HCl pH 6.8, 2% (w/v) SDS, 10% (v/v) glycerol, 0.1% (w/v) bromophenol blue, 100 mM 2-mercaptoethanol) at 37 °C for 30 minutes. Resolving gels (10% (w/v) polyacrylamide, 0.175% (w/v) SDS, 0.15% (w/v) ammonium persulfate (APS), 0.05% (v/v) N,N,N',N'-tetramethylethane-1,2-diamine (TEMED), 375 mM Tris base pH 8.8) and stacking gels (4% (w/v) polyacrylamide, 0.175% (w/v) SDS, 0.15% (w/v) APS, 0.06% (v/v) TEMED, 125 mM Tris base pH 6.8) were prepared before being placed in an electrophoresis tank filled with protein running buffer (25 mM Tris Base, 192 mM glycine, 3.5 mM SDS). Samples were loaded alongside a molecular weight marker (Invitrogen™ SeeBlue™ Plus2 Pre-stained Protein Standard) and electrophoresed at constant current (30 mA) until loading dye had fully eluted. Gels were then either used in a western blot (section 2.4.4) or stained with InstantBlue™ by rocking at room temperature for 2 hours. An ABCG2-positive sample was run on every gel as a control for the western blots.

#### **2.4.4 Western blotting**

Proteins were transferred from the polyacrylamide gel (section 2.4.3) to a nitrocellulose membrane by electrophoresis (200 mA, 2 hours, 4 °C) in transfer buffer (25 mM Tris base, 192 mM glycine, 20% (v/v) methanol), first described by Towbin et al. (1979). Transient staining of the nitrocellulose with Ponceau S solution (0.1% (w/v) Ponceau S, 1% (v/v) acetic acid) allowed confirmation of effective transfer of proteins. Membranes were washed with PBS-T (0.1% (v/v) Tween-20 in PBS) for 5 minutes, then incubated with non-fat milk (5% (w/v) in PBS-T) at room temperature for 1 hour to prevent antibody from binding to non-specific sites. The blots were subsequently incubated with the primary antibody (anti-ABCG2 antibody BXP-21, 1:2000 dilution in non-fat milk) overnight at 4 °C. Then the blots were washed with PBS-T several times over a period of 15-20 minutes to remove any unbound primary antibody and then incubated with the secondary antibody (rabbit anti-mouse horseradish peroxidase, 1:5000 dilution in non-fat milk) at room temperature for 1 hour. The 15-20-minute washes were then repeated before incubating with the chemiluminescence substrate (SuperSignal™ West Pico PLUS, Thermo Fisher Scientific) for 1 minute and imaging with the LAS-3000 mini (Fujifilm).

### **2.5 Flow cytometry**

#### **2.5.1 Cell surface expression**

ABCG2-transfected and untransfected HEK293T cells were seeded at  $1 \times 10^6$  cells/mL in FACS buffer (0.2-1.0% fatty acid-free BSA in PBS or Hank's balanced salt solution, HBSS) and incubated for 30 mins on ice with 5D3 antibody (1.82-5.00 µg/mL, Millipore), isotype control (1 µg/mL anti-TRP1, Santa Cruz Biotechnology or 3 µg/mL mouse IgG isotype control, Invitrogen) or nothing (negative control). Cells

were then centrifuged at 350 x g for 5 min at 4 °C and the supernatant was discarded. Cell pellets were washed twice by resuspending in FACS buffer and spun down at 350 x g for 5 min at 4 °C, twice. Then, the 5D3 and isotype control cells were incubated with AlexaFluor647 (5-10 µg/mL goat anti-Mouse IgG (H+L) Alexa Fluor 647, Invitrogen) in 1 mL FACS buffer for 1 hour on ice. Cells were then centrifuged at 350 x g for 5 min at 4 °C, the supernatant was discarded and washed twice. Finally, cells were resuspended in FACS buffer and fluorescence was measured by flow cytometry using the Beckman Coulter Astrios EQ Cell Sorter (channel 640-671/30).

Data was initially analysed using Kaluza 2.1. Firstly, side scatter height (channel 488-SSC) was plotted against forward scatter height (channel 488-FSC1) and was gated to exclude debris from live cells. Then side scatter height was plotted against side scatter area and gated to allow monodispersed cells to be separated from doublets. Then a histogram of AlexaFluor647 fluorescence vs number of cells was plotted and the gate placed at the edge of the isotype control peak was used to determine percentage of positive cells i.e. percentage of cells with more fluorescence with 5D3 than with the isotype control.

### **2.5.2 Transport assay**

ABCG2-transfected and untransfected HEK293T cells were seeded at  $1 \times 10^6$  cells/mL in phenol red-free DMEM and incubated at 37 °C, 5% CO<sub>2</sub> for 1 hour with either DMSO (0.1% (v/v)) or fluorescent derivatives of Ko143 (2 µM, Ko143-Cy5 or Ko143-X-BY630; synthesized by Sarah Mistry, School of Pharmacy, University of Nottingham). After the incubation, cells were kept on ice. Cells were then centrifuged at 350 x g for 5 min at 4 °C and the supernatant was discarded. Cell pellets were then resuspended in phenol red-free DMEM and spun down again at 350 x g for 5

min at 4 °C. Finally, cells were resuspended in phenol red-free DMEM and fluorescence was measured by flow cytometry using the Beckman Coulter Astrios EQ Cell Sorter (channel 640-671/30). Flow cytometry is capable of simultaneously measuring ABCG2 function and expression for each cell in suspension. This allows the gating out of lower ABCG2-expressing cells when looking at the transport data, however, it was not possible in this project.

Data was gated for monodispersity as in 2.5.1. Then, the median fluorescence of the DMSO control was subtracted from the median fluorescence of the fluorescent Ko143-treated cells. This removes background fluorescence giving a value for  $\Delta MFI$  (change in median fluorescence intensity).

$$\Delta MFI = MFI_{Fluorescent\ Ko143} - MFI_{DMSO}$$

## 2.6 Monolayer transport assay

A 96-well plate was pre-treated with poly-L-lysine (10 µg/mL, 100 µL in each well) for 1 hour, then aspirated and left to dry. Subsequently,  $3 \times 10^4$  HEK293S cells (HEK293 cells adapted for growth in suspension (Lin et al., 2014)), expressing WT-ABCG2 or not, were seeded. 48 hours later, media was removed and cells were incubated at 37 °C with either DMSO; substrate (Hoechst 33342) in the presence or absence of inhibitor (Ko143 or a fluorescent Ko143 derivative); or a fluorescent Ko143 derivative in the presence or absence of Ko143. Concentrations are shown in Table 2.4. After 45 min, media was removed and cells were incubated with either DMSO (1:1000), fluorescent Ko143 derivative (1 µM), or Ko143 (1 µM) as shown in Table 2.4. Then, after another 45 min, media was replaced with HBSS. Hoechst 33342 fluorescence (excitation  $350 \pm 10$  nm, emission  $460 \pm 20$  nm) and Ko143-X-BY630 fluorescence (excitation  $620 \pm 10$  nm, emission  $660 \pm 20$  nm) or Ko143-Cy5 fluorescence

(excitation  $610 \pm 30$  nm, emission  $675 \pm 50$  nm) was measured with the CLARIOstar microplate reader (BMG Labtech). The CLARIOstar microplate reader measures average fluorescence of each well so it is not possible to separate low ABCG2-expressing cells from higher expressing cells. All dilutions were made with HBSS and DMSO concentration was the same in each well. Fluorescent derivatives used above were Ko143-X-BY630 or Ko143-Cy5. Data was collected in collaboration with James Mitchell-White (Kerr Lab, School of Life Sciences, University of Nottingham). Background fluorescence was considered by subtracting the average DMSO fluorescence value from each datapoint.

$$Fluorescence = Fluorescence_{Raw} - Average\ Fluorescence_{DMSO}$$

Where  $Fluorescence_{Raw}$  is the Hoechst 33342 or fluorescent Ko143 derivative fluorescence for an individual datapoint and  $Average\ Fluorescence_{DMSO}$  is the mean fluorescence value of DMSO treated cells for each plate and cell line.

**Table 2.4 Monolayer transport assay incubation steps** First incubation step involves substrate, inhibitors or DMSO control diluted with HBSS. The corresponding second incubation step is also shown. For Ko143-Cy5 experiments, Ko143-Cy5 was used in place of Ko143-X-BY630.

First incubation	Second Incubation
DMSO	DMSO
Hoechst 33342 (3 $\mu$ M)	DMSO
Hoechst 33342 (3 $\mu$ M) + Ko143-X-BY630 (1 $\mu$ M)	Ko143-X-BY630 (1 $\mu$ M)
Hoechst 33342 (3 $\mu$ M) + Ko143 (1 $\mu$ M)	Ko143 (1 $\mu$ M)
Ko143-X-BY630 (1 $\mu$ M)	DMSO
Ko143-X-BY630 (1 $\mu$ M) + Ko143 (1 $\mu$ M)	Ko143 (1 $\mu$ M)

## 2.7 Data analysis

All data were analysed using GraphPad Prism 9.1.0. Multiple data sets were compared with WT and untransfected HEK293T cells using a one-way ANOVA with a Dunnett's multiple comparisons test. All experiments were repeated a minimum of 3 independent occasions for statistical analysis and figure legends confirm the number of technical repeats (n). The monolayer transport assay data was analysed with a two-way ANOVA with Tukey's multiple comparisons test.

## Chapter 3 Results

### 3.1 Hypothesis formation

The hypothesis studied in this project was based around the question, “what makes an inhibitor an inhibitor and not a substrate that is transported by ABCG2?”. This could potentially be explained by differences in affinity, with inhibitors having much higher affinity or potency than substrates, whether that be measured by dissociation constants or by  $IC_{50}$  values, as suggested by Jackson et al. (2018). Further investigation of the literature, provided some evidence for this, with inhibitors having between 3-fold and ~2000-fold higher affinity than substrates (Table 3.1) The only apparent exceptions to this are TLC-S and sulfasalazine. Sulfasalazine has been used as a substrate and an inhibitor in various research (Miyata et al., 2016, Karlsson et al., 2010). TLC-S (tauroolithocholate sulfate) is a bile acid which at certain concentrations can disrupt the cell membrane which could explain its slight inhibitory effect but low affinity for ABCG2 (Chiang, 2003). The apparent affinity values, shown in Table 3.1, are the concentration at which either half maximal transport occurred or half maximal inhibition was achieved ( $K_m$  or  $IC_{50}$ ). These values cannot be directly compared and  $IC_{50}$  values will vary depending on the specific conditions used in the assay but can act as a guide to how strongly these compounds bind to ABCG2. Jackson et al. (2018) also found a ~3,000-fold difference in affinity between their fluorescent Ko143 derivative (inhibitor) and the substrate E<sub>1</sub>S.

**Table 3.1 Apparent affinities of substrates and inhibitors**

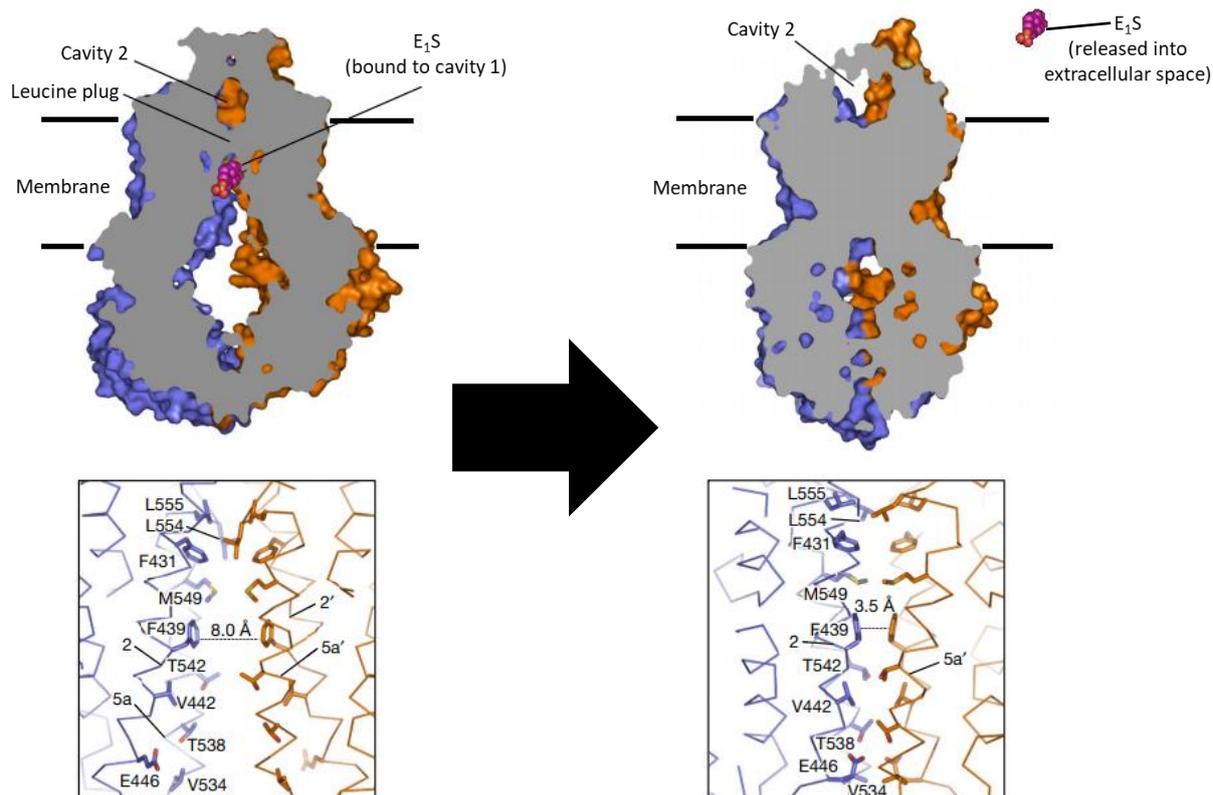
Compound name	Substrate or Inhibitor	Apparent affinity ( $\mu\text{M}$ )	Reference
Fumitremorgin C	Inhibitor	$0.731 \pm 0.092$	(Ochoa-Puentes et al., 2013)
Ko143	Inhibitor	$0.128 \pm 0.017$	(Kohler and Wiese, 2015)
Benzbromarone	Inhibitor	0.20	(Miyata et al., 2016)
Topiroxostat	Inhibitor	0.18	(Miyata et al., 2016)
Febuxostat	Inhibitor	0.027	(Miyata et al., 2016)
Sulfasalazine	Substrate/Inhibitor	0.6	(Karlsson et al., 2010)
TLC-S	Inhibitor	37	(Suzuki et al., 2003)
Rosuvastatin	Substrate	2.3	(Miyata et al., 2016)
E <sub>1</sub> S	Substrate	$16.6 \pm 3.4$	(Suzuki et al., 2003)
Mitoxantrone	Substrate	61	(Suzuki et al., 2003)
DHEAS	Substrate	55	(Suzuki et al., 2003)
4-MUS	Substrate	$12.9 \pm 2.1$	(Suzuki et al., 2003)
E3040S	Substrate	$26.9 \pm 4.0$	(Suzuki et al., 2003)
SN-38	Substrate	4.0	(Nakatomi et al., 2001)
SN-38-glucuronide	Substrate	26	(Nakatomi et al., 2001)

The difference in affinity can be rationalised by the cryo-EM structures determined by Manolaridis et al (2018). The substrate-bound (E<sub>1</sub>S), inward facing ABCG2 structure (PDB 6HCO) showed that F439 from opposite monomers are 8 Å apart and interacting with the bound substrate. In ATP-bound, outward facing structure (PDB 6HBU) these residues are only 3.5 Å apart (Figure 3.1). This means the substrate must be released from cavity 1 before it completely collapses, suggesting transient conformational changes which push out the substrate, resembling a peristaltic motion (Manolaridis et al., 2018). The resulting hypothesis is that inhibitors have a higher affinity for cavity 1 which means they are less likely to be released to allow the

transient conformational changes to occur. *In other words, high affinity inhibitors are proposed to lock ABCG2 in the inward facing conformation.*

One extension of this theory is that there would be an inverse relationship between binding affinity and maximal transport, suggested by Manolaridis et al. (2018). This group removed the hydrogen bond potential in a T435A mutant causing an increase in E<sub>1</sub>S transport. This could indicate that reducing affinity for a compound increases substrate character (i.e. increased transport) and increasing affinity increases inhibitor character (less transport, more inhibition).

If inhibitors have high affinity and substrates have lower affinity, will reducing the affinity of an inhibitor cause it to become a substrate which is transported? To test this hypothesis a series of experimental mutants were made with the goal of reducing affinity for the most commonly used inhibitor Ko143.

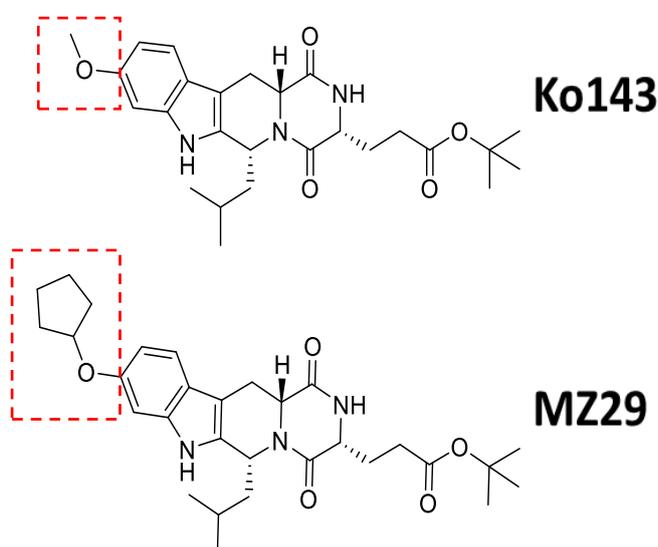


**Figure 3.1 Cavity 1 completely collapses to promote substrate transport.** When the substrate  $E_1S$  is bound in cavity 1 of human ABCG2 (top left, PDB 6HCO), the F439 residues from each monomer are 8.0 Å apart (bottom left). ATP binding triggers conformational changes which lead to  $E_1S$  being extruded into the extracellular space (top right, PDB 6HBU) and cavity 1 collapses so that F439 residues are only 3.5 Å apart (bottom right). Cross sections (top left and top right) and stick models (bottom left and bottom right) of each monomer are shown in orange and blue. The leucine plug residue is L554. Figure adapted from Manolaridis et al. (2018).

### 3.2 Mutant design

In order to decide which experimental mutations to make, the structure of ABCG2 was visualised in PyMOL. This allowed determination of residues that are involved in the binding of Ko143 and provided information for the design of mutants that would have a reduced affinity for Ko143. The structure used was the cryo-EM structure published by Jackson et al. (2018) of ABCG2 bound to MZ29, a derivative of Ko143. MZ29 and Ko143 only differ in one place: the methoxy group of Ko143 is replaced by an O-cyclopentyl group (Figure 3.2). First, all residues with atoms within 4.0 Å of

MZ29 were identified; a total of fifteen were found. Three of these residues (F431, F432 and L555, shown in white in Figure 3.3 B and C) were excluded since they only interacted with the O-cyclopentyl group of MZ29 and would be too far away (4.7-5.9 Å) to interact with the carbon of the methoxy group in Ko143. Also, Manolaridis et al. (2018) found that no functional protein was expressed with L555A because it is likely to have structural importance, so mutations to different residues could have the same issue. The remaining residues are shown in Figure 3.3 A, where their interactions with specific parts of Ko143 are also shown.



**Figure 3.2 Comparison of the structures of Ko143 and MZ29.** The red dashed squares highlight the difference between Ko143 and MZ29: a methoxy group and O-cyclopentyl group respectively. Structures are from Jackson et al. (2018).

In an attempt to narrow down the important residues further, the human ABCG2 protein sequence was aligned with ABCG2 sequences from other mammals (rabbit, cow and mouse) where Ko143 has been shown to be an inhibitor (Weidner et al., 2015, Manzini et al., 2017, Wei et al., 2012, Halwachs et al., 2016). All residues with atoms within 4.0 Å of MZ29 were fully conserved (Figure 3.3 D-F), in fact most residues in ABCG2 are conserved (81.61-86.28% sequence identity compared with

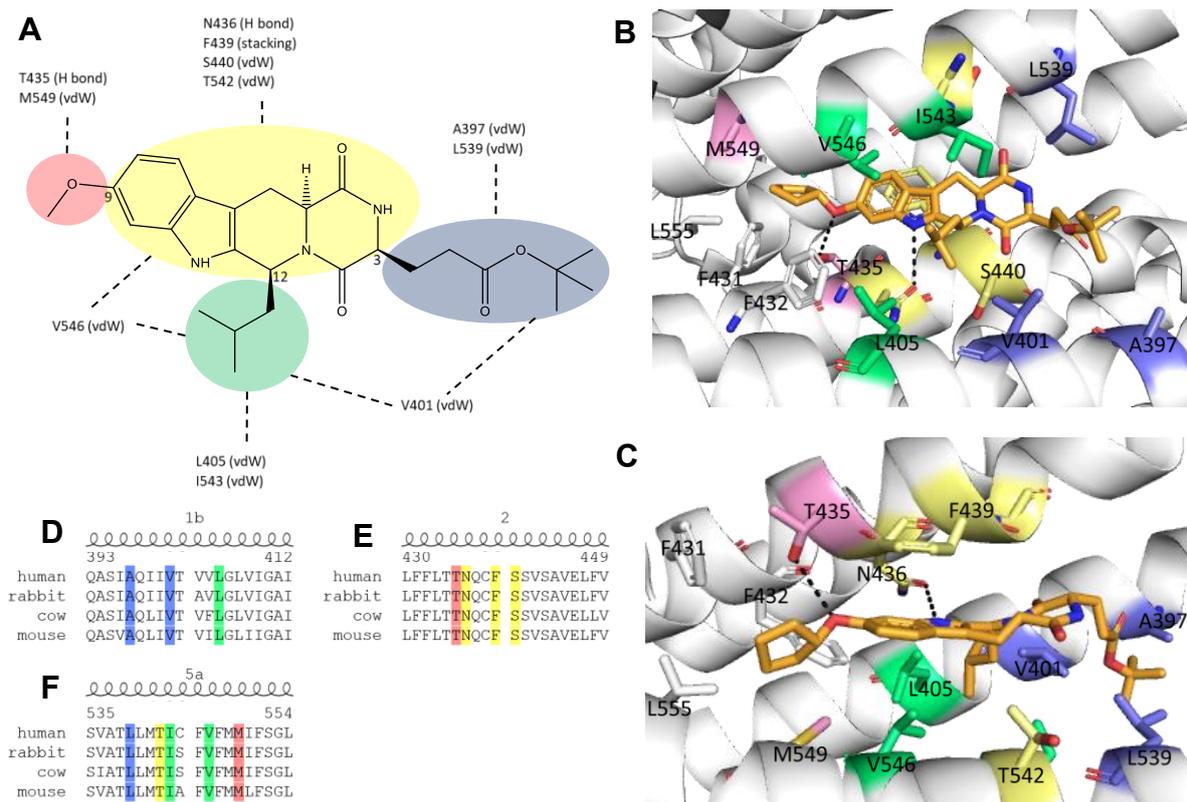
human ABCG2). Therefore, all highlighted residues could play a role in binding, and none can be ruled out at this stage based off the sequence alignments alone.

### 3.2.1 A397S/V401A/L539A – Mut1

A397, V401 and L539 interact with the *tert*-Butyl ester (blue in Figure 3.3 A) via van der Waals forces. Mutating these residues to reduce hydrophobicity could reduce the affinity of ABCG2 for Ko143. This is supported by the research performed by Weidner et al. (2015) where the acid metabolite of Ko143, lacking the *tert*-Butyl ester, has no inhibitory effect on mitoxantrone transport by ABCG2. V401A and L539A would lower affinity for Ko143 since mutation to alanine would introduce a smaller hydrophobic group which would form fewer or weaker van der Waals interactions. However, this approach could not be applied to A397 which cannot be mutated to anything smaller, besides glycine which would add flexibility to  $\alpha$ -helix 1b (Figure 3.3 B, C and D), potentially causing unwanted effects to the overall protein structure (Högel et al., 2018). For this reason and to further decrease affinity, the mutation to the polar residue serine was introduced in this triple mutant. Of the three residues interacting with the *tert*-Butyl ester, A397S is the furthest away from the oxygens and is therefore the least likely to hydrogen bond but could still cause a hydrophobic/hydrophilic repulsion that would result in a decrease in affinity. Making the triple mutant, A397S/V401A/L539A, will ensure the hydrophobicity is sufficiently reduced to have an impact on the affinity for Ko143's *tert*-Butyl ester. This mutant will be referred to as Mut1 throughout this thesis.

### 3.2.2 L405A/I543A/V546A – Mut2

The isobutyl group, highlighted by the green circle in Figure 3.3 A, forms van der Waals interactions with V401, L405, I543 and V546. Another triple mutant was designed to target these hydrophobic interactions: L405A/I543A/V546A (Mut2). Cox et al. (2018) made the single mutants L405A and I543A, both of which showed a significant reduction in mitoxantrone and pheophorbide A transport. It is, therefore, feasible these residues will also play a role in the binding of inhibitors. V546A, however, compared to WT had no change in E<sub>1</sub>S transport but did double the EC<sub>50</sub> of E<sub>1</sub>S-induced ATPase activity (Manolaridis et al., 2018). This means the V546A mutant reduces the affinity for the substrate E<sub>1</sub>S but perhaps not enough for reduced transport. By combining V546A with L405A and I543A the hydrophobicity will be lowered to a greater extent which will have a larger effect on reducing the affinity, when applied to Ko143.



**Figure 3.3 ABCG2 residues interacting with Ko143.** Schematic showing ABCG2 residues interacting with Ko143. Four chemical features of Ko143 are involved in interactions: methoxy (pink), polycyclic core (yellow), *tert*-Butyl ester (blue), isobutyl (green). Carbons 3, 9 and 12 are labelled (Jackson et al., 2018) (**B and C**) MZ29-bound ABCG2 with interacting residues shown as sticks (within 4.0 Å). Polar interactions (hydrogen bonds) shown as black dotted lines. Residues interacting with one feature of MZ29 are coloured as in A. V401 (blue) is coloured to match the other residues interacting with the *tert*-Butyl ester (triple mutant) but it also interacts with the isobutyl group. V546 (green) is coloured in the same manner but also interacts with the polycyclic core. F431, F432 and L555 (white) interact with O-cyclopentyl group of MZ29 but not Ko143. Some residues from TM1b and TM5a (opposite monomers) are hidden for clarity. PDB 6ETI was used (Jackson et al., 2018) (**D-F**) Partial sequences from the alignment of ABCG2 from human, rabbit, cow and mouse. Shows conservation of residues interacting with Ko143 in TM1b (D), TM2 (E) and TM5a (F) (highlighted with same colours as in B and C).

### 3.2.3 T435A and M549E

Single mutants were made to target the affinity for the methoxy group of Ko143 (pink in Figure 3.3). This is because the methoxy group has been shown to be a key component of potent inhibitors, with Ko143 being four times more potent than its demethoxy analogue Ko134 when measuring inhibition of ABCG2-mediated mitoxantrone efflux (Allen et al., 2002). Demethoxy fumitremorgin C is also 10 times less potent than native fumitremorgin C (He et al., 1999, as cited in Allen et al., 2002) so small changes to the interactions with the methoxy will potentially have a greater effect than with other groups. This is supported by Jackson et al. (2018). They found that changing the substituent at C9 (the methoxy group, Figure 3.3 A) had a large influence on binding affinity and suggested that the removal of the T435 hydrogen bond causes a reduction in affinity. For these reasons, the mutation T435A was made which removes the hydrogen bond.

M549 also interacts with the methoxy group via van der Waals forces and has been shown to be important in substrate transport. In previous studies, M549A decreased mitoxantrone and pheophorbide A transport but not E<sub>1</sub>S transport (Haider et al., 2015, Manolaridis et al., 2018). Jackson et al. (2018) found that the addition of hydrophilic groups to C9 of the polycyclic core of Ko143 caused the inhibitor to become inactive. They suggest this is due to the hydrophobicity at the bottom of cavity 1, nevertheless, it would be interesting to see the effect of mutating M549 to a glutamic acid which is a similar sized but hydrophilic residue. This mutation would remove the hydrophobic interactions without adding a hydrogen bond between the carboxyl and the methoxy (4.7 Å between 2 closest oxygen atoms). In addition, since

M549A had no effect on E<sub>1</sub>S transport, a more drastic change (M549E) is more likely to have an effect on affinity for Ko143.

### 3.2.4 N436A, F439A and S440W

The polycyclic core of Ko143 (yellow in Figure 3.3 A) has 5 amino acids interacting with it: N436, F439, S440, T542 and V546. N436 hydrogen bonds via the carbonyl oxygen of the side chain with the NH of the indole ring in Ko143 (Figure 3.3 C).

Mutating to an alanine removes this hydrogen bond and minimizes advantageous hydrophobic interactions with the polycyclic core. Previous mutation of N436A led to depleted E<sub>1</sub>S transport activity and there was also no E<sub>1</sub>S-induced ATPase activity suggesting it is important for substrate binding, therefore, the mutation N436A could reduce affinity for Ko143 (Manolaridis et al., 2018).

F439 forms stacking interactions with the benzene ring in the polycyclic core of Ko143, this is a stronger bond than van der Waals but weaker than hydrogen bonds so would be interesting to mutate. Removing the aromaticity of phenylalanine would eliminate this interaction. Mutating to any other amino acid, besides tryptophan or tyrosine, would do this but introduction of other interactions must also be avoided. This is why mutation to alanine was made: no hydrogen bonds are introduced and hydrophobic interactions are minimized. Previous data on the F439A mutant has shown that it had an effect on E<sub>1</sub>S transport and E<sub>1</sub>S-induced ATPase activity so could also have an effect on Ko143 affinity (Manolaridis et al., 2018).

S440 has been shown to be involved in mitoxantrone and pheophorbide A transport, with mutation to alanine causing a significant reduction in transport compared with the WT (Cox et al., 2018). However, the hydroxyl group of S440 is pointing away from MZ29 in the 6ETI structure (Figure 3.3 C) so would not form any polar

interactions with MZ29 nor Ko143, based on the assumption that they bind in the same manner. Perhaps the orientation of S440 is different when binding to substrates (although S440 also points away in the E<sub>1</sub>S-bound structure, 6HCO) or the hydrophilicity of this residue contributes to creating an environment suitable for binding. Since S440A will not necessarily have a clear effect on the affinity for Ko143, a tryptophan mutation was made. The idea was to mutate to the largest amino acid to cause steric hindrance so that Ko143 can no longer sit in the pocket in a way that optimises the other interactions. This could have major ramifications on binding by causing too much steric hindrance or removing too many other interactions. On the other hand, interactions could be introduced including  $\pi$ - $\pi$  stacking and hydrogen bonds with the NH of tryptophan.

T542 forms van der Waals interactions with the polycyclic core of Ko143, however, this residue was not mutated. Removing one van der Waals interaction would likely have very little effect on the total affinity especially since there are 4 other amino acids interacting with the polycyclic core, including a stronger hydrogen bond and stacking interaction. Furthermore, there was no literature to support its involvement in Ko143 or substrate binding. Since the aim of this project is to assess the importance of affinity for inhibitor function, there is no need to mutate this residue unnecessarily.

### 3.2.5 Summary

In summary, the mutants designed are: Mut1 (A397S/V401A/L539A), Mut2 (L405A/I543A/V546A), T435A, M549E, N436A, F439A and S440W. These mutations target different functional groups of Ko143 and are proposed to affect the affinity in different ways (Table 3.2). For example, Mut1 and Mut2 remove multiple weaker interactions, whereas T435A and N436A remove stronger hydrogen bonds. M549E

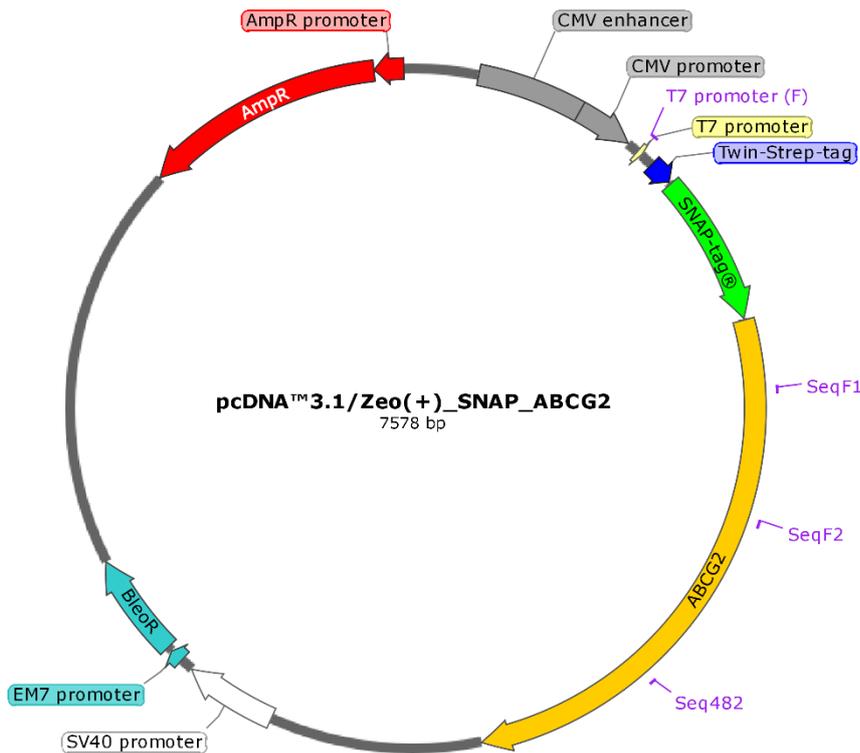
and S440W are mutating residues that are potentially less important for affinity, to more extreme mutations designed to disrupt binding. In future experiments, if the triple mutations had a functional impact, there would be value in assessing the contribution of each residue to Ko143 binding and transport. However, for this project the compounded effect of three minor changes in both Mut1 and Mut2 is more likely to noticeably impact Ko143 binding.

**Table 3.2 ABCG2 mutants and their desired effect on Ko143 binding.** Mut1 and Mut2 are A397S/V401A/L539A and L405/I543A/V546A triple mutants respectively.

Mutant	Ko143 functional group	Desired effect
T435A	Methoxy group	Removes hydrogen bond
M549E	Methoxy group	Removes hydrophobic interaction; Introduces hydrophilicity
N436A	Polycyclic core	Removes hydrogen bond
F439A	Polycyclic core	Removes stacking interactions
S440W	Polycyclic core	Increases steric hindrance
Mut1	<i>tert</i> -Butyl ester	Reduces hydrophobic interactions
Mut2	Isobutyl group	Reduces hydrophobic interactions

### 3.3 Construct generation

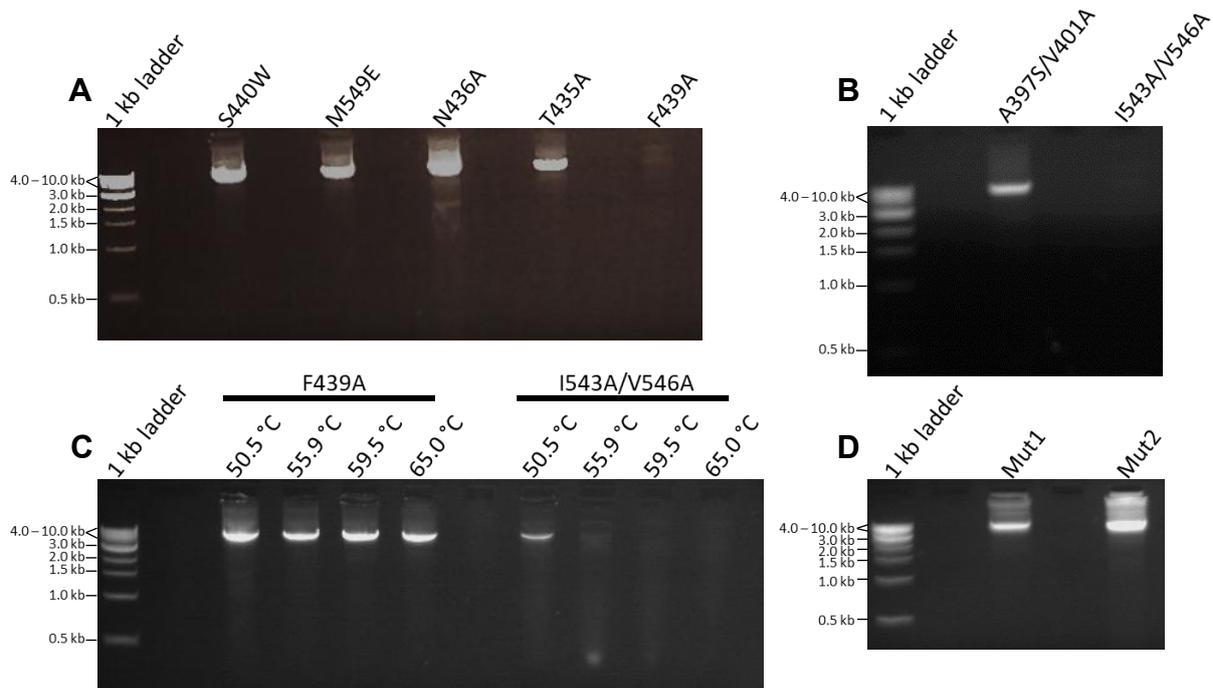
Mutant ABCG2 constructs were created by performing mutagenic PCR with pcDNA™3.1/Zeo(+)\_SNAP\_ABCG2 plasmid as the template DNA which is shown in Figure 3.4. Key features of this plasmid include an ampicillin resistance gene (AmpR) for bacterial selection; a Zeocin™ resistance gene (BleoR) for mammalian cell selection; and the cytomegalovirus (CMV) promoter for enhanced expression of the protein of interest, SNAP-tagged ABCG2.



**Figure 3.4 Schematic of the template construct used for mutagenic PCR.** The pcDNA™3.1/Zeo(+) backbone contains the coding region for twin-strep and SNAP-tagged ABCG2. Key features are shown including AmpR and BleoR to confer ampicillin and Zeocin™ resistance respectively and a CMV promoter for enhanced expression. Primers used in sequencing are shown in purple.

In order to make the mutant constructs, mutagenic PCR was performed using primers containing the mutant codon. The primers were designed to optimise the following factors:  $\Delta G$  of self-dimer formation greater than -10 kcal/mol;  $\Delta G$  of hairpin formation greater than -5 kcal/mol; GC content between 40% and 60%; and a score higher than 75% (all found using Premier Biosoft's Netprimer). For the triple mutants Mut1 and Mut2, two sets of primers were designed. The first set made two mutations relatively close in sequence to each other (A397S and V401A for Mut1, I543A and V546A for Mut2). The second set included the final mutation (L539A for Mut1, L405A for Mut2).

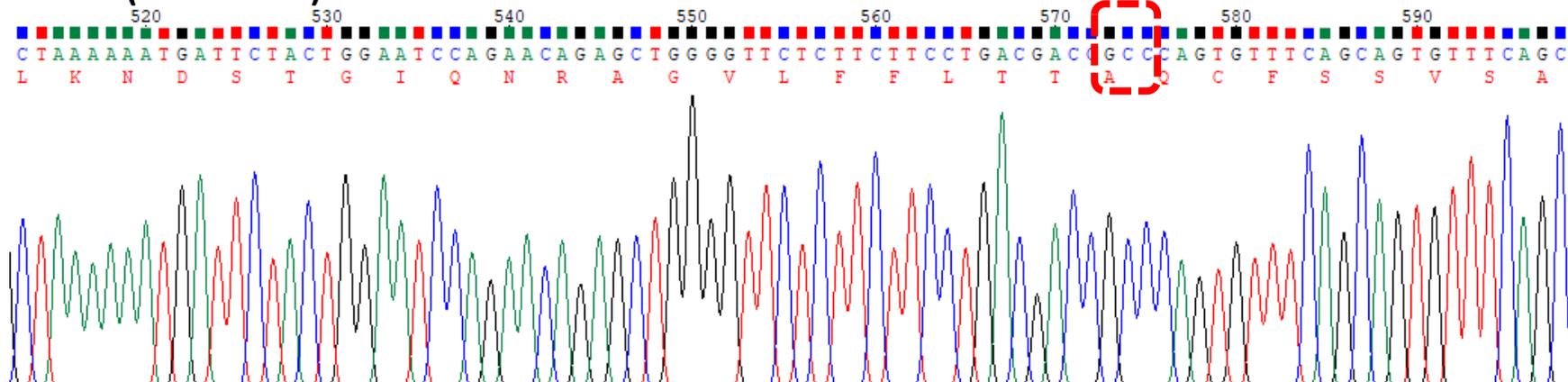
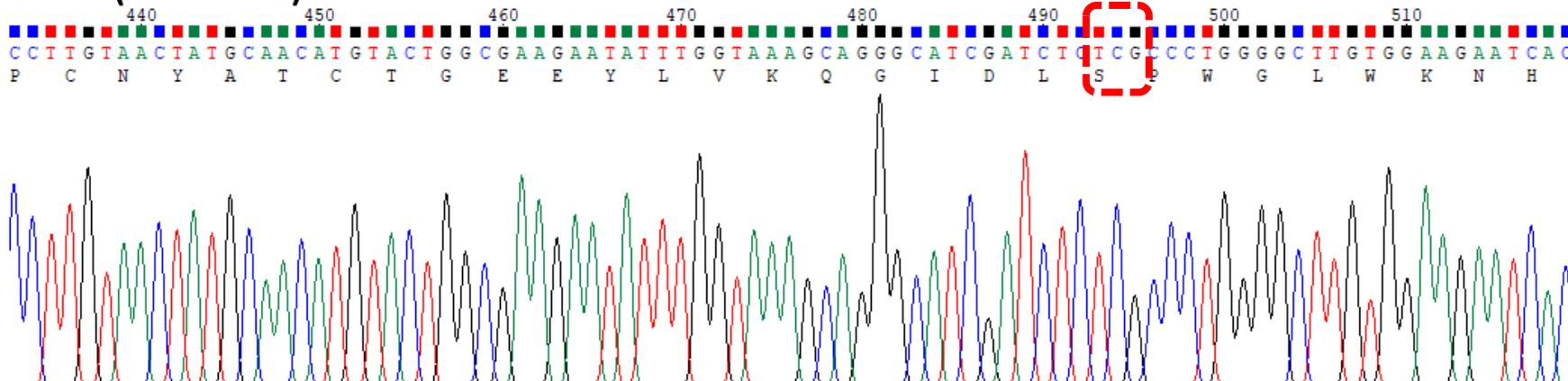
PCRs were performed as described in 2.2.1 and resulting DNA was confirmed on agarose gel electrophoresis, shown in Figure 3.5. For the single mutants, S440W, M549E, N436A and T435A have clear bands between 4.0 kb and 10.0 kb which correlates with the construct being 7.6 kb. There is no band, however, for F439A so a gradient PCR was conducted at a range of annealing temperatures (4 temperatures 50.5-65.0 °C), all of which were successful (Figure 3.5 C). Two rounds of PCR were performed for the triple mutants, Mut1 and Mut2. First, the double mutant primers were used (A397S/V401A and I543A/V546A, Table 2.1) on the WT template shown in Figure 3.4. A397S/V401A was successful at the original annealing temperature (55.0 °C, Figure 3.5 B) and I543A/V546A was successful at 50.5 °C (Figure 3.5 C). Once successful PCR was confirmed (Figure 3.5 B and C) and sequences were checked by Sanger sequencing (see below), the PCR products were used as the template in the second round of PCR with the second set of primers (L539A for Mut1 and L405A for Mut2). The resulting PCR products were confirmed by agarose gel electrophoresis shown in Figure 3.5 D.



**Figure 3.5 Confirmation of successful PCR by gel electrophoresis.** PCR was performed using the pcDNA™3.1/Zeo(+)\_SNAP\_ABCG2 plasmid as the template DNA and the mutagenic primers shown in Table 2.1. **(A)** For the single mutants, all but F439A have undergone successful PCR **(B)** First round of PCR for Mut1 and Mut2, using the double mutant primers to create A397S/V401A and I543A/V546A respectively. A397S/V401A was successful but I543A/V546A was not. **(C)** PCR was performed at a range of annealing temperatures (50.5-65.0 °C) for F439A and I543A/V546A. F439A was successful at all temperatures and I543A/V546A at 50.5 °C **(D)** Second round of PCR for Mut1 and Mut2, using the A397S/V401A and the I543A/V546A (50.5 °C) PCR products respectively as the templates, was successful.

The PCR products were then treated with *DpnI* to break down the methylated template DNA (WT), leaving only the mutated DNA intact. The PCR products were then transformed into DH5 $\alpha$  competent *E. coli* and following plasmid isolation, the DNA was sequenced by Source BioScience (Nottingham). Sanger sequencing was performed using a series of primers that covered the entire region of interest (SNAP-labelled ABCG2). Primers used, as well as the region they cover, are shown in Table 2.3 and the position they bind on the plasmid is shown in Figure 3.4. Sequence chromatograms were first viewed in Chromas to confirm quality of the data and then

sequences were compared with the WT construct by aligning in BLAST to see if the mutation was successfully incorporated (Appendix, section 6.2). Successful mutagenesis can be observed for N436A (Figure 3.6, top panel) and similar data for all other single and triple experimental mutants was obtained (Appendix, section 6.1). For N436A there was one unexpected difference between WT and observed sequence (Figure 3.6, bottom panel): an adenine to guanine which changed the TCA codon to TCG. Both these codons code for serine so the construct was still able to be used. Strictly speaking this mutant is N436A/S622S but will be referred to as N436A throughout this thesis. All the other mutant constructs were exactly as expected.

**N436A (AAC→GCC)****S622S (TCA→TCG)**

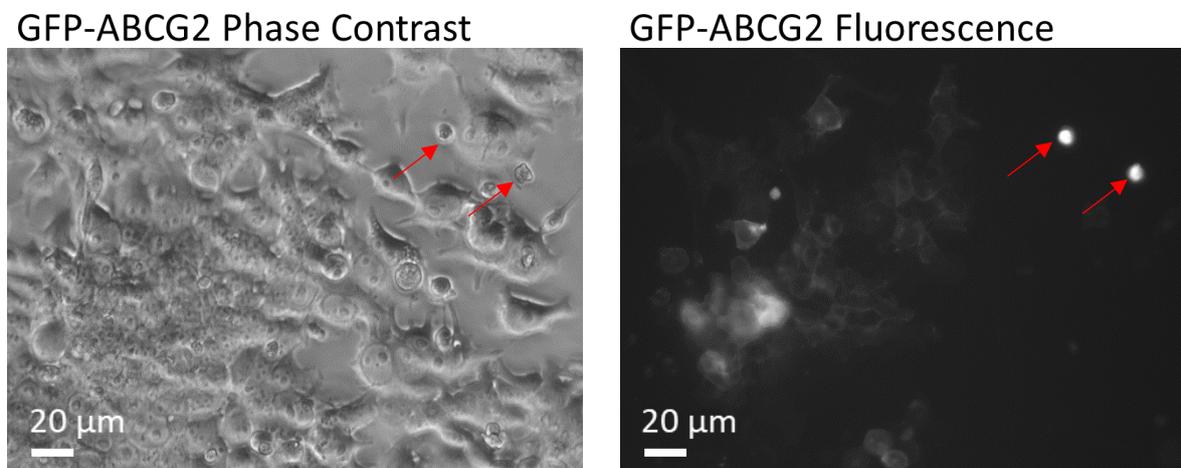
**Figure 3.6 Sequencing chromatograms to confirm the desired mutation within the mutant ABCG2 constructs.** The top panel shows a portion of the N436A sequencing data obtained using the SeqF2 primer, with the desired mutation highlighted by the red box. The bottom panel shows the additional mutation (red box) in the N436A construct (data collected using the Seq482 primer). The other mutant constructs showed similar results to the top panel, with no additional mutations.

### 3.4 Protein expression

The mammalian expression system used in this study was HEK293T cells. They are derived from HEK293 cells which were created from normal human embryonic kidney (HEK) cells exposed to sheared fragments of human adenovirus type 5 DNA, making them more infectious. They also contain the temperature-sensitive SV40 T-antigen tsA1609 which activates replication of vectors with the SV40 origin of replication (Figure 3.4). This allows high levels of expression of the protein of interest, SNAP-tagged ABCG2 (Graham et al., 1977, DuBridge et al., 1987, Rio et al., 1985).

HEK293T cells, which have no endogenous ABCG2 expression, were transiently transfected with WT and mutant pcDNA<sup>TM</sup>3.1/Zeo(+)\_SNAP\_ABCG2 plasmids using the PEI transfection method described by Boussif et al. (1995). The molar PEI nitrogen: DNA phosphorus ratio was 15:1 (section 2.3.2). Stable cell lines were generated by Zeocin<sup>TM</sup>-mediated selection. Zeocin<sup>TM</sup> concentration in the media was maintained at 200 µg/mL for 36-43 days before reducing to 40 µg/mL. This kills the cells that have not taken up the plasmid, while successfully transfected cells are resistant due to the newly acquired Zeocin<sup>TM</sup>-resistance gene, BleoR (Figure 3.4). The SNAP-labelled ABCG2 is not particularly conducive to rapid observations of expression in live cells; therefore, as a guide for zeocin selection efficacy, the pcDNA<sup>TM</sup>3.1/Zeo(+)\_sfGFP\_ABCG2 plasmid, coding for GFP-tagged WT-ABCG2, was transfected alongside the WT and mutant plasmids described above. This allowed the expression of ABCG2 to be monitored by epifluorescence microscopy. Figure 3.7 shows that, even after 5 days of Zeocin<sup>TM</sup> selection, most of the GFP-ABCG2 cells are fluorescent meaning that they are expressing GFP-ABCG2. Observation of untransfected HEK293T cells confirmed that cell death was not

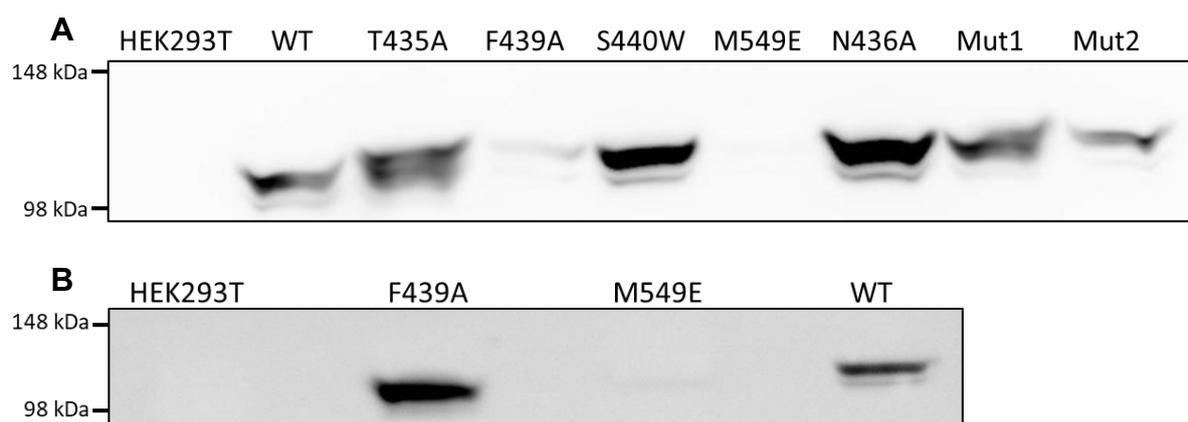
complete until several weeks of exposure. Once stable cell lines were generated, aliquots were frozen in liquid nitrogen for long term storage.



**Figure 3.7 Confirmation of GFP fluorescence as an indicator of transfection efficacy.** pcDNA™3.1/Zeo(+)\_sfGFP\_ABCG2 plasmid was transfected into HEK293T cells alongside the WT and mutant constructs described in section 3.2. After 5 days of Zeocin™-selection, epifluorescence microscopy was used to view the GFP-ABCG2 fluorescence of the cells. Fluorescence of these cells indicates that transfection was successful. Cells that are rounded and intensely fluorescent (red arrowheads) are dying/dead cells, presumably reflecting cells that have not taken up the plasmid and so now suffering the cytotoxic effects of zeocin. Scale bars represent approximately 20 µm.

To determine expression levels of ABCG2 in the WT and mutant cell lines, western blotting was performed (Figure 3.8). First cells were harvested, then lysed in ice cold buffer by sonication. 35 µg of cell lysate was resolved electrophoretically on 10% polyacrylamide gel and transferred to nitrocellulose or stained with InstantBlue™. Blots were then probed against the anti-ABCG2 antibody, BXP-21. Figure 3.8 A shows bands of varying intensity between 98 kDa and 148 kDa, which is a larger apparent molecular weight than the expected size of ABCG2 (72.3 kDa) tagged with SNAP-tag® (19.4 kDa) and twin strep tag (3.4 kDa). However, many membrane proteins have mobility differences on SDS-PAGE due to increased interactions with SDS molecules, increasing their apparent molecular weight compared with the

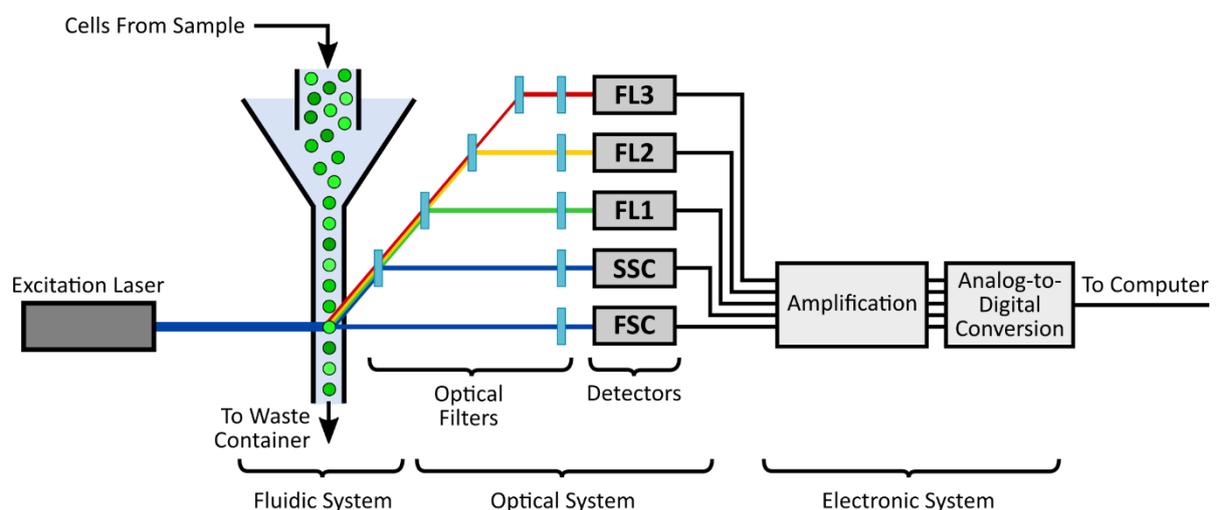
globular protein standards (Rath et al., 2009). F439A and M549E have weak BXP-21 reactive bands alongside lower protein loading demonstrated by the gels stained with InstantBlue™. Increasing the protein loading to 70 µg (Figure 3.8 B) confirmed that F439A was expressed but showed that M549E still had minimal expression of ABCG2. As a result, this mutant was discounted from further experiments. Very low/absent expression of M549E might result from the extreme nature of this mutation, with a hydrophobic methionine being replaced by a hydrophilic glutamic acid, since M549A was successfully expressed in other work (Haider et al., 2015, Manolaridis et al., 2018).



**Figure 3.8 Western blots showing mutant and WT-ABCG2 expression in stable cell lines.** 35 µg (A) or 70 µg (B) of whole cell lysates underwent SDS-PAGE and immunoblotted against the anti-ABCG2 antibody, BXP-21. Untransfected HEK293T cells were used as a negative control. Change in relative expression of F439A compared to WT can be explained by lower protein loading of F439A and M549E in A and lower protein loading of WT in B. The molecular weights of pre-stained standard proteins were identified from the nitrocellulose.

### 3.5 Confirmation of protein localisation by flow cytometry

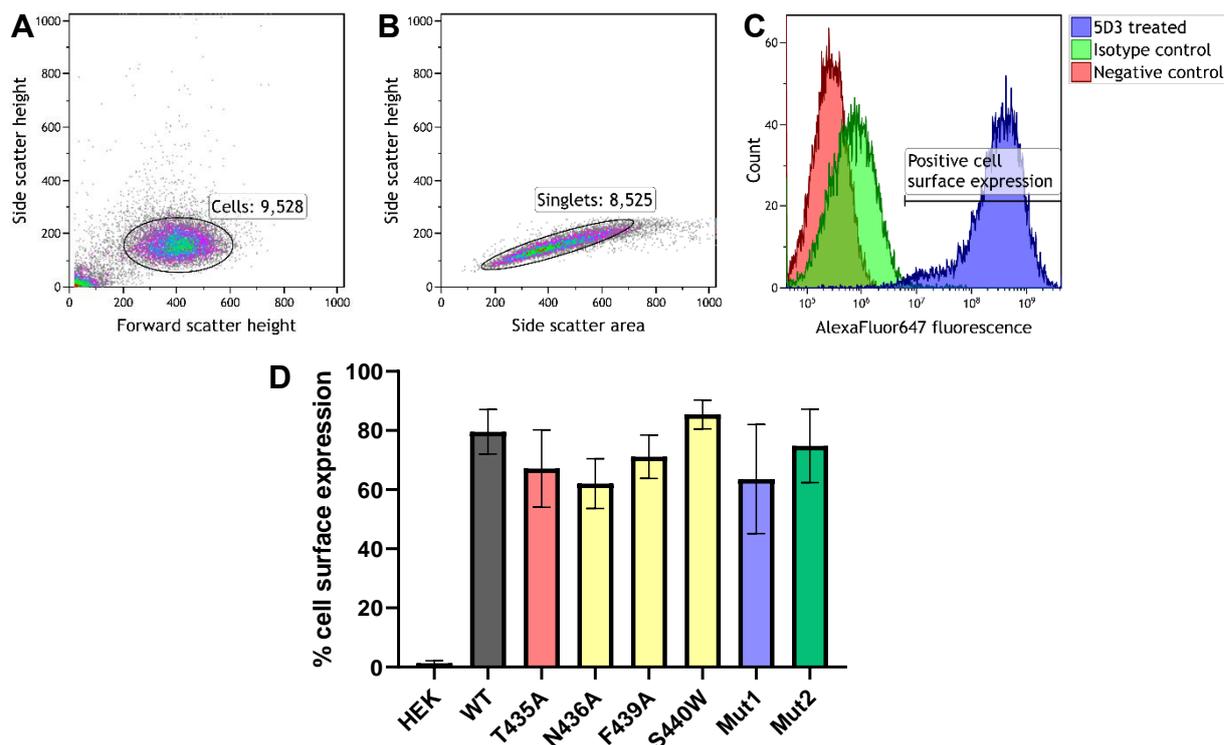
Western blots can confirm total expression of a protein, however, exclusively using this technique for membrane proteins does not provide any information on localisation with the cell. Activity of membrane proteins is influenced by its lipid bilayer surroundings (Lee, 2004, Szilagyi et al., 2017) so localisation on the cell membrane must be confirmed. This is especially true for transporters such as ABCG2, where substrates can only be exported if the protein is correctly localised. In this thesis, flow cytometry was used to quantify the proportion of cells with significant cell surface expression of ABCG2. The monoclonal antibody 5D3 recognises a cell surface epitope within the extracellular loop 3 of ABCG2 and can therefore be used to measure cell surface localisation of ABCG2 (Ozvegy-Laczka et al., 2008). In this project, cells were incubated with 5D3 antibody or isotype control and then the Alexafluor647 secondary antibody and fluorescence was measured by flow cytometry.



**Figure 3.9 Schematic representation of flow cytometry.** The basic flow cytometry setup consists of a single stream of cells intercepted by a laser. Light scattered by the cells pass through or are reflected by filters and are detected by side scatter (SSC), forward scatter (SSC) or fluorescent (FL1, FL2, FL3) detectors. Figure acquired from AAT Bioquest (2019).

During flow cytometry, a single stream of cells is produced which allows a laser to illuminate each cell as it flows past (Figure 3.9). The light from the laser is scattered by the cell in all directions but is only detected in the forward direction (same direction as the laser) and at 90° angle from the laser beam (Jaroszeski and Radcliff, 1999). This is referred to as forward scatter (FSC) and side scatter (SSC) respectively. If the cells are fluorescent, either through fluorescent antibodies or proteins (e.g. GFP), the laser will excite the fluorophore and the emitted light will be detected in one of the fluorescent detectors (FL1, FL2, FL3), 90° from the laser. Different wavelengths of emitted light will be either reflected or passed through optical filters so that they are detected by the correct detector.

Side scatter values relate to granularity/structure and forward scatter is proportional to size (Jaroszeski and Radcliff, 1999). This means that different types of cell will appear in different positions when SSC and FSC are plotted against each other and data points can be gated to separate data from each type. This is shown in Figure 3.10 A, where the clear cluster is HEK293T cells and datapoints outside this gate are likely to represent damaged cells and cell debris. Side scatter height plotted against side scatter area can separate monodispersed cells from doublets. Figure 3.10 B shows the gating for singlets, for the untransfected HEK293T cells. This particular sample shows very few doublets but they would appear in a cluster shifted to the right of the one gated. It is important that doublets are gated out because they could give falsely high fluorescence values.



**Figure 3.10 Cell surface ABCG2 expression was confirmed by flow cytometry.** Cells were treated with either 5D3 (extracellular binding anti-ABCG2 antibody) or isotype control and then AlexaFluor647 secondary antibody. Negative control cells were not treated with any antibody. **(A and B)** Data was first gated to exclude debris from live cells by plotting side scatter height against forward scatter height (A). Side scatter height was then plotted against side scatter area to separate singlets from doublets (B). Number of cells in each gate is shown. Untransfected HEK293T (negative control) is shown and is representative of all data. Density plots are shown, with grey being the sparsest datapoints and red/green being the densest. **(C)** After gating, histograms of cell count vs AlexaFluor647 fluorescence were plotted. C shows the overlay of WT histograms (5D3, isotype control and negative control) for one repeat. It is representative of the remaining data. The right-hand edge of the isotype control peak was used as a threshold for positive cell surface expression for each cell line and repeat. **(D)** Mean percentage of cell surface ABCG2 expression ( $n=4$ ) of untransfected HEK293T cells (HEK), WT and mutants, derived from 5D3 histograms and isotype control set thresholds. % cell surface expression is the percentage of cells with fluorescence over the threshold set in C. Error bars depict standard error of the mean (SEM). Colours of the bars represent which functional group of Ko143 the mutants interact with (as in Figure 3.3): methoxy (pink), polycyclic core (yellow), *tert*-Butyl ester (blue), isobutyl (green). Control cell lines (HEK and WT) are shown in grey.

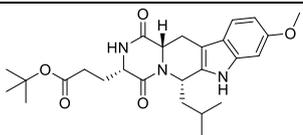
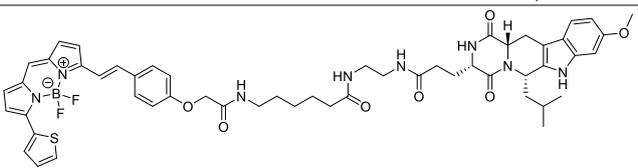
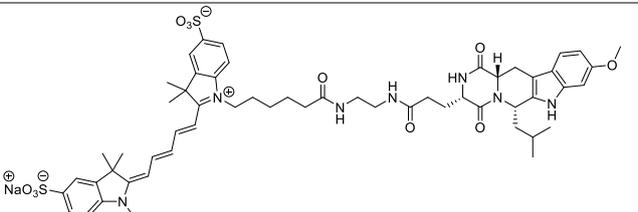
Once data was gated for cells and singlets, histograms for Alexafluor647 fluorescence for each cell line were plotted (Figure 3.10 C). Thresholds for positive cell surface expression were set using the isotype controls for each cell line with any cell displaying greater AlexaFluor647 fluorescence than the 99th centile of the isotype control considered as ABCG2-expressing (Figure 3.10 C). Figure 3.10 D shows the mean percentage of cells with fluorescence over the threshold (% cell surface expression) for all cell lines over 4 repeats. ABCG2 is localised to the plasma membrane to varying degrees but all cell lines (except for untransfected HEK293T cells) have at least 60% cell surface expression and are not significantly different from each other ( $p > 0.05$ ). This is sufficient for use in the transport assays.

### 3.6 Transport assays

Transport assays have often been performed using flow cytometry and take advantage of many ABCG2 substrates being fluorescent e.g. mitoxantrone, pheophorbide A and Hoechst 33342 (Cox et al., 2018, Kapoor et al., 2020). When cells expressing WT-ABCG2 are incubated with fluorescent substrates, ABCG2 will pump out the substrate leaving a low level of fluorescence in the cell. When treated with inhibitors or using ABCG2 mutants that do not transport the substrate, the substrate will accumulate in the cell giving a higher fluorescence. Ko143, however, is not fluorescent. In order to measure its transport, fluorescent derivatives of Ko143 were used (kindly synthesized by Sarah Mistry, School of Pharmacy, University of Nottingham). Two derivatives, Ko143-X-BY630 and Ko143-Cy5, are shown in Table 3.3 and consist of Ko143 tagged with a fluorophore at the *tert*-Butyl ester end. These will likely bind to ABCG2 in a similar manner to Ko143 with the Ko143-like regions of the fluorescent derivatives binding to cavity 1. All the experimental mutants (except

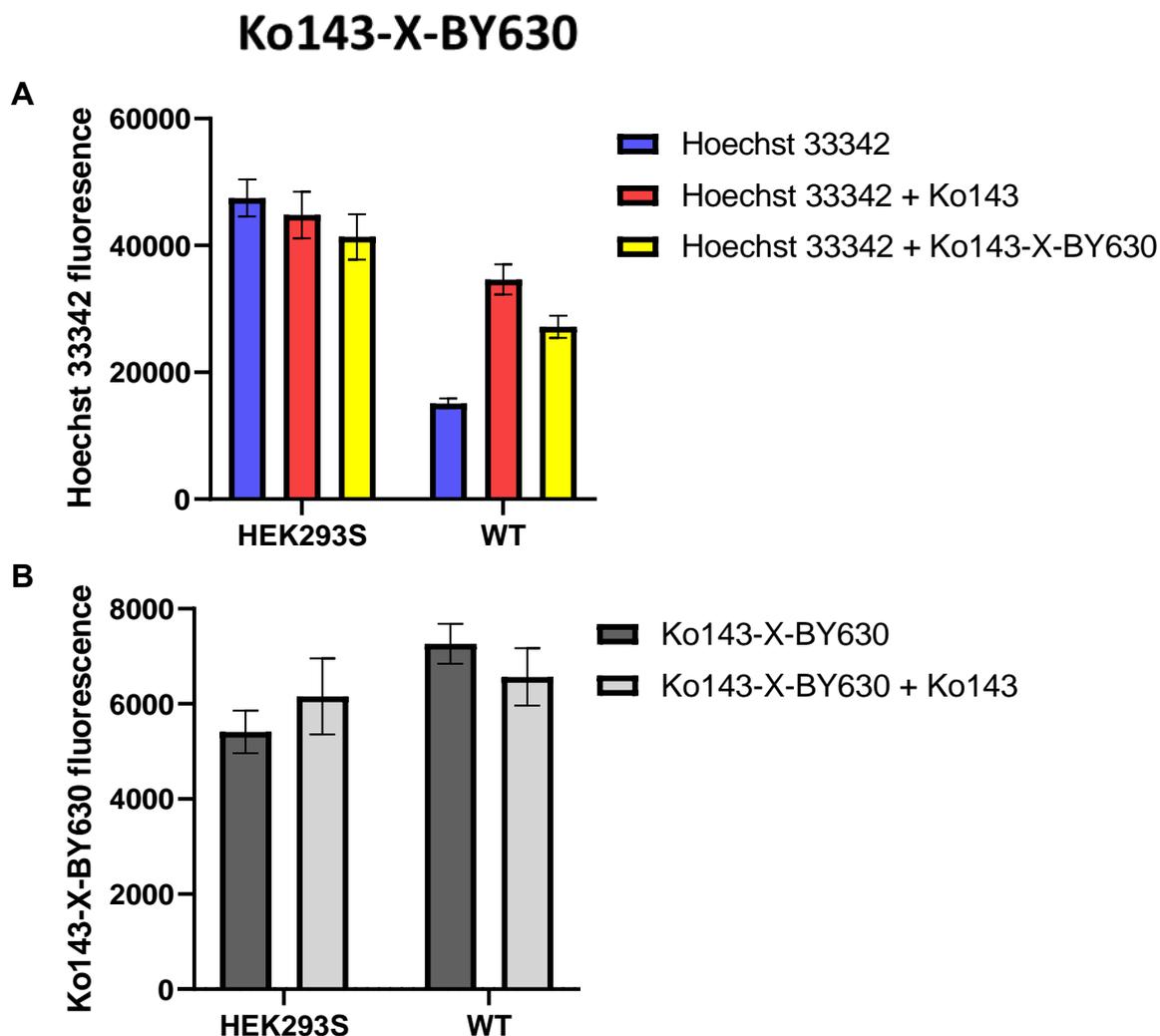
for Mut1) are designed to reduce affinity for functional groups that are common between Ko143, Ko143-X-BY630 and Ko143-Cy5. Therefore, the effect on binding/transport of the fluorescent derivatives compared with untagged Ko143 should be minimal. However, Mut1 has 3 mutations (A397S/V401A/L539A) all focused around reducing the affinity for the *tert*-Butyl ester of Ko143 (Figure 3.3) but for both fluorescent derivatives the *tert*-Butyl ester is replaced by a fluorescent tag (Table 3.3). Ko143-X-BY630 and Ko143-Cy5 are less hydrophobic than Ko143 in the equivalent region, meaning the experimental mutations would have less of an effect on the affinity than originally intended. In addition, any reduction in affinity caused by a decrease in hydrophobicity, could be cancelled out by the possible formation of a hydrogen bond between A397S and the amides present in the fluorescent Ko143 derivatives.

**Table 3.3 Details of Ko143 and its fluorescent derivatives.**

Name	Structure	Excitation wavelength (nm)	Emission wavelength (nm)
Ko143		N/A	N/A
Ko143-X-BY630		636	651
Ko143-Cy5		649	666

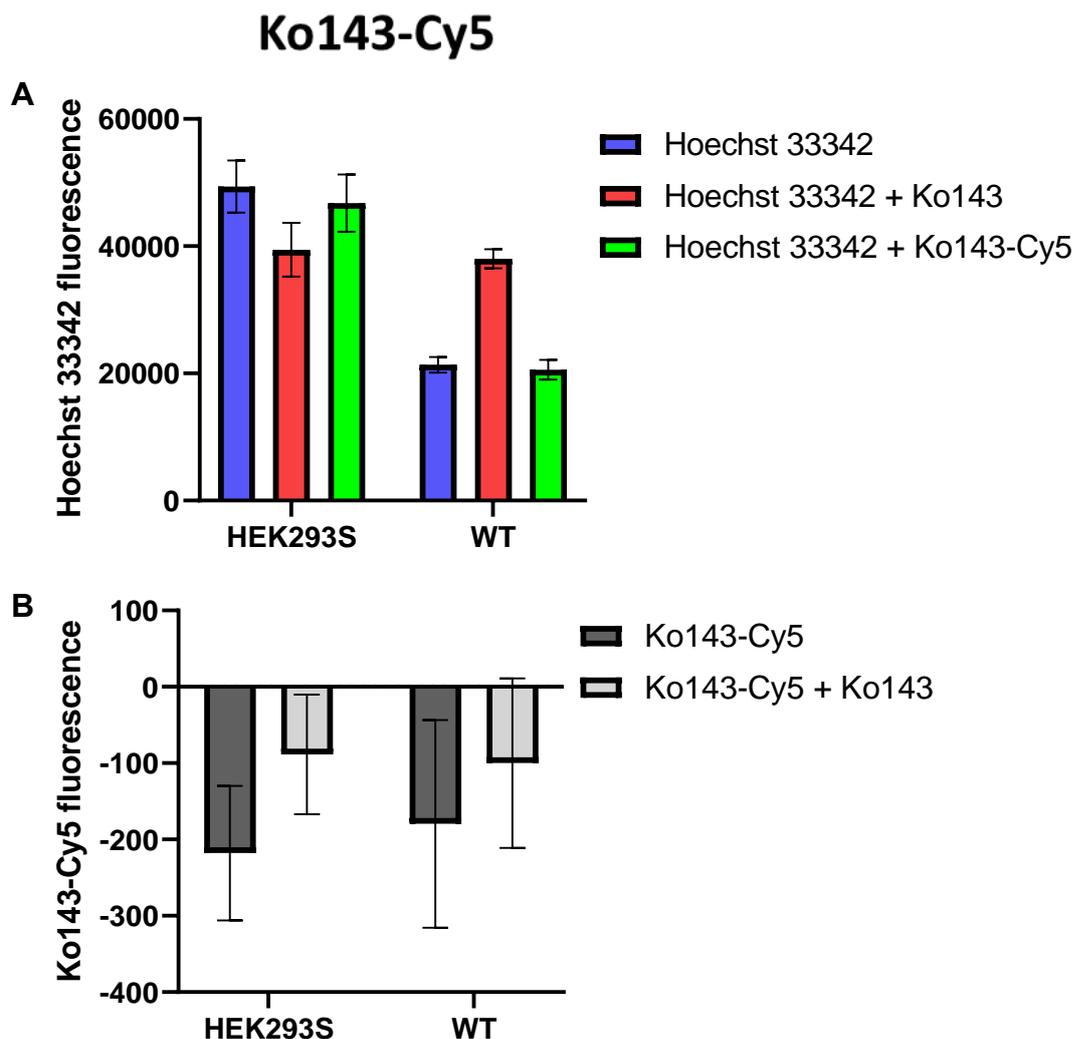
### 3.6.1 Monolayer transport assay

Before analysing whether the fluorescent Ko143 derivatives are transported by the mutant ABCG2 proteins, their inhibitory activity compared to Ko143 must be determined. To accomplish this, a monolayer transport assay was performed in collaboration with James Mitchell-White (Kerr Lab, School of Life Sciences, University of Nottingham) using suspension growth-adapted HEK293 (HEK293S) cells which were untransfected or expressing WT-ABCG2 (Lin et al., 2014). Similar to Haider et al. (2015), HEK293S cells were treated with either Hoechst 33342 in the presence or absence of inhibitor (Ko143 or fluorescent derivative). Hoechst 33342 is a fluorescent substrate of ABCG2 whose excitation and emission wavelength (361/497 nm) does not overlap with that of the fluorescent Ko143 derivatives (Table 3.3). The HEK293S and WT cells were also treated with a fluorescent Ko143 derivative (Ko143-X-BY630 or Ko143-Cy5) in the presence or absence of Ko143 (also in absence of Hoechst 33342). The purpose of this is to confirm that the fluorescent derivatives are able to enter HEK293 cells.



**Figure 3.11 Ko143-X-BY630 inhibits Hoechst 33342 transport by ABCG2. (A)** Hoechst 33342 fluorescence was measured in untransfected HEK293S and WT expressing HEK293S cells treated with Hoechst 33342 in the presence or absence of an inhibitor (Ko143 or Ko143-BY630). Hoechst 33342 treated WT-ABCG2 expressing cells have significantly less Hoechst 33342 fluorescence than untransfected HEK293S cells. Ko143 and Ko143-X-BY630 inhibit ABCG2 causing a significant increase in Hoechst 33342 accumulation in WT-ABCG2 expressing cells. **(B)** Ko143-X-BY630 fluorescence was measured in untransfected HEK293S and WT expressing HEK293S cells treated with Ko143-X-BY630 in presence or absence of Ko143. All conditions were significantly different from zero showing that Ko143-X-BY630 enters HEK293S and WT-ABCG2 expressing cells. Data also shows no significant Ko143-inhibitable transport of Ko143-BY630. Data was collected on 3 separate occasions. Background fluorescence was accounted for by subtracting the average DMSO fluorescence from each data point (section 2.6). Error bars depict standard error of the mean (SEM).

Figure 3.11 shows the results of the monolayer transport assay for Ko143-X-BY630. When measuring Hoechst 33342 fluorescence (Figure 3.11 A), WT-ABCG2 expressing cells have a significantly different fluorescence ( $p < 0.0001$ , comparing the two blue bars) to the HEK293S cells because ABCG2 pumps Hoechst 33342 out of the cell. Treatment with Ko143 restores cellular accumulation of Hoechst 33342 giving a higher fluorescence that is not significantly different from the equivalent HEK293S cells. This is because Ko143 inhibits ABCG2 so Hoechst 33342 was not exported. Figure 3.11 A shows that Ko143-X-BY630 acts as inhibitor to almost the same extent as untagged Ko143. Hoechst 33342 transport by WT-ABCG2 expressing cells treated with Ko143-X-BY630 was not significantly different from those treated with Ko143 (compare red and yellow bars, right hand side, Figure 3.11 A) but the magnitude of inhibition appears slightly lower with Ko143-X-BY630. As seen in Figure 3.11 B, Ko143-X-BY630 successfully enters the cell since the Ko143-X-BY630 fluorescence in HEK293S and WT cells is significantly different from zero ( $p < 0.0001$ ). Also, in this case, the addition of the fluorescent tag to Ko143 does not cause it to be transported by WT-ABCG2. This is demonstrated by no significant difference in Ko143-X-BY630 fluorescence between HEK293S and WT cells. In addition, there is no significant difference between Ko143-BY630 treated WT cells in absence or presence of Ko143. This means that, unlike with Hoechst 33342 transport, Ko143 does not affect Ko143-X-BY630 accumulation.



**Figure 3.12 Ko143-Cy5 does not inhibit Hoechst 33342 transport by ABCG2. (A)** Hoechst 33342 fluorescence was measured in untransfected HEK293S and WT-ABCG2 expressing HEK293S cells treated with Hoechst 33342 in the presence or absence of an inhibitor (Ko143 or Ko143-Cy5). Hoechst 33342 treated WT-ABCG2 expressing cells have significantly less Hoechst 33342 fluorescence than untransfected HEK293S cells. Ko143 inhibits ABCG2 causing a significant increase in Hoechst 33342 accumulation in WT-ABCG2 expressing cells. Ko143-Cy5 does not inhibit Hoechst 33342 transport because fluorescence is not significantly different from WT cells treated with just Hoechst 33342. **(B)** Ko143-Cy5 fluorescence was measured in untransfected HEK293S and WT-ABCG2 expressing HEK293S cells treated with Ko143-Cy5 in presence or absence of Ko143. All conditions were not significantly different from zero showing that Ko143-Cy5 did not enter HEK293S and WT cells. Data was collected on 3 separate occasions. Background fluorescence was accounted for by subtracting the average DMSO from each data point (section 2.6). Error bars depict standard error of the mean (SEM).

The monolayer transport assay was repeated with Ko143-Cy5 instead of Ko143-X-BY630 (Figure 3.12). In contrast to the data for Ko143-X-BY630, Ko143-Cy5 did not act as an inhibitor in the monolayer transport assay. As shown in Figure 3.12 A, there is low cellular accumulation of Hoechst 33342 in WT-ABCG2 expressing cells treated with Ko143-Cy5, which is not significantly different from WT cells treated with Hoechst 33342 alone. This is explained by Figure 3.12 B, where Ko143-Cy5 fluorescence was not significantly different from zero in WT and HEK293S cells. This indicates that Ko143-Cy5 may not successfully enter the cell. Perhaps this is due to the more hydrophilic nature of Ko143-Cy5 compared with Ko143-BY630, having more charged atoms (Table 3.3). This would make it more difficult to cross the largely hydrophobic lipid bilayer of the cell membrane. From this data, it is not possible to determine whether Ko143-Cy5 acts as an inhibitor or not because Ko143-Cy5 did not successfully enter the cell. A membrane-based activity assay, such as the one described in Kapoor et al. (2020) would be able to confirm this ambiguous result.

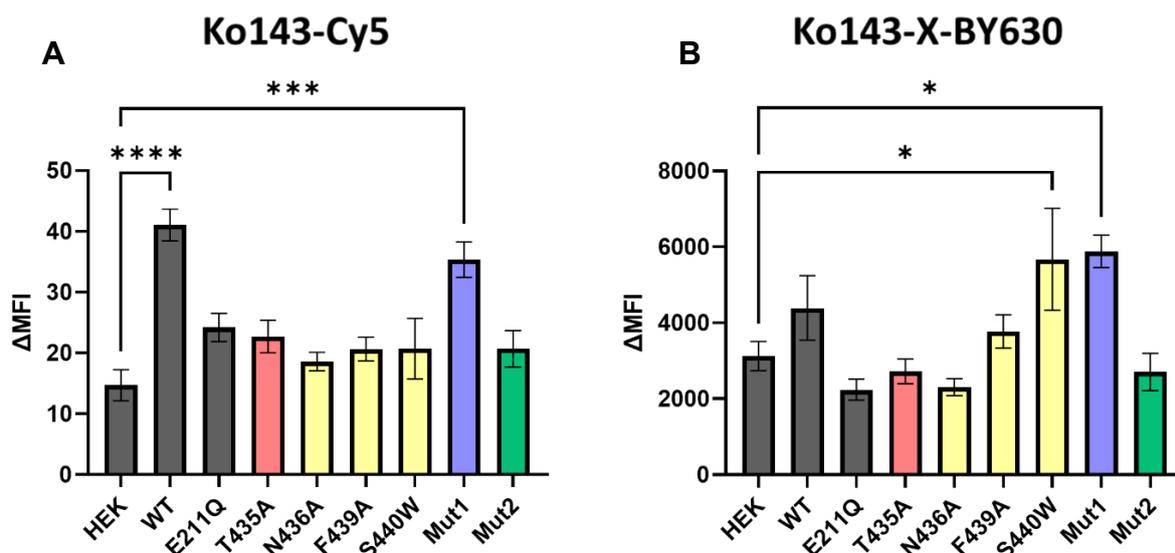
In summary, the monolayer transport assay measured the cellular accumulation of fluorescent Hoechst 33342 (ABCG2 substrate) and the fluorescent Ko143 derivatives. Ko143-X-BY630 successfully enters the HEK293S cells; inhibits Hoechst 33342 transport by ABCG2 to the same extent as untagged Ko143; and is not transported by ABCG2. Ko143-Cy5, however, did not successfully enter the HEK293S cells and therefore, it is not possible to assess the impact the fluorescent tag has on ABCG2 inhibition.

### 3.6.2 Flow cytometry

Using the same principle of cellular accumulation as in section 3.6.1, flow cytometry was used to measure whether the mutant cell lines transported the fluorescent Ko143 derivatives. The overarching hypothesis is that if the mutant ABCG2 proteins now recognise Ko143 as a substrate rather than an inhibitor then the accumulation of the fluorescent Ko143 derivatives in those cell lines will be lower. Although the monolayer transport assay indicated that Ko143-Cy5 did not enter cells, the preliminary data suggested that the flow cytometry assay could detect intracellular Ko143-Cy5. Therefore, both fluorescent Ko143 derivatives were used in flow cytometry. The underlying technical differences between the two assays are discussed in Chapter 4.

For the flow cytometry transport assay, mutant cell lines were incubated for 1 hour with 2  $\mu$ M Ko143-Cy5 or Ko143-X-BY630 and then fluorescence was measured by flow cytometry. WT-ABCG2 expressing cells and untransfected HEK293T (HEK) cells were used as controls as well cells expressing the inactive mutant, E211Q, which has the catalytic glutamic acid in the Walker-B sequence of the NBD mutated to glutamine (Haider et al., 2015), a kind gift of Joseph Morris (Kerr Lab, School of Life Sciences, University of Nottingham). To account for autofluorescence/background fluorescence, control cells for each cell line were treated with DMSO for 1 hour. After gating out debris and doublets (as described in section 3.5), the median fluorescence for each DMSO sample was subtracted from the median fluorescence of the fluorescent Ko143 derivatives. This gave a value for change in median fluorescence intensity ( $\Delta$ MF<sub>I</sub>) which were averaged over at least 3 repeats and shown in Figure 3.13.

For Ko143-Cy5 (Figure 3.13 A and Table 3.4), WT-ABCG2 expressing cells have a significantly higher ( $p < 0.0001$ )  $\Delta$ MFI than HEK293T cells. This difference is not due to transport of Ko143-Cy5 because HEK293T does not express functionally detectable levels of ABCG2. However, the higher fluorescence of WT cells could be explained by Ko143-Cy5 binding to cavity 1 of ABCG2 leading to cellular retention of the fluorescent inhibitor. All of the cavity 1 mutant cell lines (described in section 3.2), except for Mut1, show significantly different Ko143-Cy5 fluorescence compared to WT-ABCG2 expressing cells ( $p < 0.001$ ) but the same Ko143-Cy5 fluorescence as HEK293T cells. This is possibly due to reduced binding to ABCG2 caused by the designed/purposeful reduction in affinity for Ko143. With this explanation, Ko143-Cy5 would diffuse out of the mutant ABCG2-expressing cells more easily than with the WT-ABCG2 expressing cells, giving a similar fluorescence to HEK293T cells. Altered expression levels of ABCG2 in the mutant cell lines does not appear to be an explanation as Ko143-Cy5 fluorescence does not correlate with expression level of ABCG2 (Figure 3.8). For example, N436A expression was higher than WT-ABCG2 so cannot explain why  $\Delta$ MFI of N436A expressing cells is lower than WT-ABCG2 expressing cells. The triple mutant Mut1 has a  $\Delta$ MFI significantly higher than HEK ( $p < 0.001$ ) but not significantly different from WT. As mentioned in section 3.6, the experimental mutations made in Mut1 are focused around reducing the affinity of the *tert*-Butyl ester of Ko143, which is not present in fluorescent Ko143 derivatives. This explains why Mut1 has similar fluorescence to WT cells and supports the idea that the other cavity 1 mutants have reduced binding.



**Figure 3.13 Cellular accumulation of fluorescent Ko143 derivatives in mutant cell lines.** After gating for monodispersity (see Figure 3.10 A and B), median fluorescence intensity (MFI) was measured for mutant, WT and untransfected cell lines, treated with a fluorescent Ko143 derivative, Ko143-Cy5 (A) or Ko143-X-BY630 (B). Background fluorescence (DMSO control) was subtracted to give  $\Delta$ MFI. Data was collected on 3 separate occasions for A and 4 separate occasions for B (except F439A where  $n=3$ ). Colours of the bars represent which functional group of Ko143 the mutants interact with (as in Figure 3.3): methoxy (pink), polycyclic core (yellow), *tert*-Butyl ester (blue), isobutyl (green). Control cell lines (HEK, WT and E211Q) are shown in grey. The lines labelled with \* ( $p < 0.05$ ), \*\*\* ( $p < 0.001$ ) or \*\*\*\* ( $p < 0.0001$ ) represent mutant cell lines that have a significantly different  $\Delta$ MFI from HEK293T cells (HEK). Error bars depict standard error of the mean (SEM).

For the Ko143-X-BY630 transport assay (Figure 3.13 B and Table 3.4), the fluorescence level for each cell line shows a similar pattern to the Ko143-Cy5 data. However, S440W is an exception to this and the  $\Delta$ MFI for all mutant cell lines and HEK293T cells are not significantly different from WT-ABCG2 expressing cells. WT-ABCG2 expressing cells have a  $\Delta$ MFI that is slightly higher than HEK293T cells along with T435A, N436A, F439A and Mut2 expressing cells having a similar fluorescence to HEK293T cells. As before, Mut1 has a significantly higher  $\Delta$ MFI than HEK293T cells ( $p < 0.05$ ) but this time the fluorescence is higher (but not significantly) than WT cells. As with Ko143-Cy5, this can be explained by the addition of the fluorescent tag which could diminish the effect of the hydrophobicity-

reducing experimental mutations. S440W has significantly more Ko143-X-BY630 accumulation than HEK cells ( $p < 0.05$ , despite the greater variability of this data set), with a fluorescence more similar to Mut1.

**Table 3.4 Summary of flow cytometry transport assay results.** Significant differences are shown in red (lower) and green (higher) and p values are also shown. Where differences were not significant the phrase “trend to lower/higher” is used.

Cell Line	Ko143-Cy5		Ko143-X-BY630	
	Compared to WT	Compared to HEK	Compared to WT	Compared to HEK
HEK293T	Lower ( $p < 0.0001$ )	N/A	Trend to lower	N/A
WT	N/A	Higher ( $p < 0.0001$ )	N/A	Trend to higher
E211Q	Lower ( $p < 0.01$ )	Trend to higher	Trend to lower	Trend to lower
T435A	Lower ( $p < 0.001$ )	Trend to higher	Trend to lower	Trend to lower
N436A	Lower ( $p < 0.0001$ )	Trend to higher	Trend to lower	Trend to lower
F439A	Lower ( $p < 0.001$ )	Trend to higher	Trend to lower	Trend to lower
S440W	Lower ( $p < 0.001$ )	Trend to higher	Trend to higher	Higher ( $p < 0.05$ )
Mut1	Trend to lower	Higher ( $p < 0.001$ )	Trend to higher	Higher ( $p < 0.05$ )
Mut2	Lower ( $p < 0.001$ )	Trend to higher	Trend to lower	Trend to higher

In summary, Ko143-Cy5 has significantly higher accumulation in WT-ABCG2 expressing cells compared to HEK293T cells and the other ABCG2 mutants, excluding Mut1. Ko143-X-BY630 shows similar results but without a significant difference (Table 3.4). The exception is S440W which, along with Mut1, has a significantly higher  $\Delta$ MFI than HEK293T cells. Both Ko143-Cy5 and Ko143-X-BY630 data support the idea that reduced affinity of the mutants for fluorescent Ko143 derivatives causes a decrease in “stickiness” that leads to a lower  $\Delta$ MFI. Transport of Ko143-Cy5 or Ko143-X-BY630 cannot be ruled out but it is not detectable in this experiment.

## Chapter 4 Discussion

At the beginning of this project, a hypothesis was proposed that inhibitors of ABCG2 have a higher affinity for cavity 1 than substrates. It was speculated that reducing the affinity of ABCG2 for Ko143, a well-known inhibitor, would cause it to be transported instead of inhibiting. Data collected was inconclusive with regards to the hypothesis, however, it makes way for additional experiments which could further interrogate the hypothesis. Distinguishing between substrates and inhibitors is important from an academic perspective as well as a clinical one. Not only does it allow better characterisation of substrates which allows drugs to be rationally designed to avoid transport by ABCG2, but also aids development of selective ABCG2 inhibitors. Multidrug resistance is large problem and there are currently no clinically available inhibitors of ABCG2. Known inhibitors such as fumitremorgin C, which is extremely neurotoxic, and Ko143, which is less toxic, are not suitable *in vivo* (Toyoda et al., 2019). If this hypothesis is further validated, ABCG2 inhibitors could be designed by altering non-toxic, selective substrates to have a stronger binding affinity to the transporter. This provides a more guided approach to inhibitor design but also increases the chances of it being clinically suitable since the structurally related substrates have already been approved.

### 4.1 Summary of results

On a whole, the Ko143-X-BY630 and Ko143-Cy5 flow cytometry data show similar patterns which could be rationalised by the cavity 1 mutants having lower fluorescent Ko143 derivative binding than the WT-ABCG2. However, the flow cytometry data does not rule out the possibility that Ko143-Cy5 or Ko143-X-BY640 are transported by the cavity 1 mutants (except Mut1). If  $\Delta\text{MFI}$  was significantly lower than in

HEK293T cells, this would indicate that the fluorescence Ko143 derivatives were pumped out. Therefore, a reduction in  $\Delta$ MFI to a value significantly lower than in WT-ABCG2 expressing cells, as observed with most mutants for Ko143-Cy5, could mean that it is transported or that a reduced level of binding to ABCG2 occurs due to the mutations made to cavity 1.

Work by Gose et al. (2020), published after the cavity 1 mutants were designed (section 3.2), confirms a reduction in affinity for Ko143 in ABCG2 mutants F439A and N436A. An aromatic residue 439 was found to be essential for substrate binding and transport, with the authors calling the residue an “aromatic clamp” (Gose et al., 2020). In their work, F439A lost Hoechst 33342 and pheophorbide A transport ability in contrast to their F439W and F439Y mutants. In addition, their thermal stabilisation assay indicated that F439A-ABCG2 reduced binding of multiple substrates and inhibitors including Ko143. Similarly, Manolaridis et al. (2018) found that F439 could be essential for substrate transport because the two F439 residues from both monomers come together which causes cavity 1 to completely collapse. Mutating to a smaller residue could in fact stop substrates being forced out of cavity 1. So, even if it is possible to make Ko143 into a substrate by reducing affinity, this mutant might not be able to transport it regardless. This confirms that in the case of F439A, the most likely scenario is that the fluorescent Ko143 derivatives do not bind ABCG2. This explains why cellular accumulation of Ko143-Cy5 and Ko143-X-BY630 was comparable to untransfected HEK293T cells.

Mutation of N436A by Gose et al. (2020) showed less impact than the F439A mutation in their thermostability assay. This indicated that this residue contributed less to the binding of Ko143, which might be comparable to the lower fluorescence

for Ko143-X-BY630 in the N436A mutant compared to the F439A mutant in the flow cytometry assay (Figure 3.13), although this difference did not reach significance. Therefore, Ko143-X-BY630 could be transported by N436A-ABCG2. Transport by this mutant is possible for some substrates: Hoechst 33342 and pheophorbide A transport activity of N436A is similar to WT but E<sub>1</sub>S transport is strongly reduced (Gose et al., 2020, Manolaridis et al., 2018). This supports the idea that difference in fluorescent Ko143 accumulation is due to differences in binding affinity as opposed to increased transport. Gose et al. (2020) suggest that N436A binding selectively to ligands could be due to altered cholesterol modulation of activity or a conformational change which may affect interactions with certain ligands. This could explain why the difference in fluorescence between N436A and F439A is not seen for Ko143-Cy5 but is seen for Ko143-X-BY630.

When treated with Ko143-Cy5, S440W expressing cells showed significantly lower fluorescence than WT-ABCG2 expressing cells and a more comparable fluorescence to untransfected HEK293T cells. In contrast, when treated with Ko143-X-BY630, S440W expressing cells had significantly higher fluorescence than HEK293T cells. This is the exception to the pattern, which on whole matches that of the Ko143-Cy5 flow cytometry data. It is unlikely that there are differences in affinity between the two fluorescent Ko143 derivatives that are not noticed in the other mutants. This is because residue 440 is located in cavity 1 and the structures of Ko143 and both fluorescent Ko143 derivatives would be the same in this location. However, the variability of the data for this particular mutant is high and it is not significantly different from WT-ABCG2 expressing cells, as are the other mutants. The E211Q mutant is not expected to transport anything since the catalytic residue required for ATP hydrolysis has been mutated (Haider et al., 2015, Hollenstein et al., 2007). Its

lower  $\Delta$ MFI may be partially explained by this mutant consistently expressing at a lower level than WT, meaning fewer ABCG2 protein molecules for Ko143-Cy5 to bind to (Cox, 2019, Kapoor, 2020).

There was a discrepancy between the monolayer transport assay (section 3.6.1) and the flow cytometry transport assay (section 3.6.2) with regards to Ko143-Cy5 fluorescence. The monolayer transport assay had no significant accumulation of Ko143-Cy5 which suggests that Ko143-Cy5 did not enter the cell. This result alone could be rationalised by the altered hydrophobicity of Ko143 by adding the fluorescent tag (Table 3.3). However, Ko143-Cy5 *did* successfully enter the cell in flow cytometry, suggesting cell permeability is merely reduced and not abolished. The assays differ in terms of Ko143-Cy5 concentration (2  $\mu$ M in flow cytometry vs 1  $\mu$ M in monolayer transport assay), total duration of Ko143-Cy5 incubation and subsequent washes and second incubations. Additionally, flow cytometry is more sensitive than the monolayer transport assay so it is a better technique for distinguishing low fluorescence levels from the background compared with the microplate reader (Basiji et al., 2007). This could mean that for both techniques, Ko143-Cy5 entered the cells equally well but flow cytometry was just more capable of measuring it. Future work will include performing a Hoechst 33342 transport assay with flow cytometry to assess the inhibitory capability of Ko143-Cy5 and is explained in more detail in section 4.3.

## **4.2 Alternate ideas and support of the hypothesis**

As mentioned in section 3.1, E<sub>1</sub>S transport was increased when T435 was mutated to alanine, which suggests that there is an inverse relationship between affinity and maximal transport (Manolaridis et al., 2018). The same mutant was made in this

project but instead of E<sub>1</sub>S transport, the potential transport of fluorescent Ko143 derivatives was studied. The accumulation of Ko143-X-BY630 and Ko143-Cy5 in T435A expressing cells in the flow cytometry experiment (Figure 3.13) was lower than in WT-ABCG2 expressing cells and as stated before, this suggests less binding to ABCG2 but does not rule out the possibility of transport. Since T435A has already shown to increase transport of a substrate, it seems plausible that transport of Ko143 is also possible, especially in conjunction with the flow cytometry data. Gose et al. (2020) found that there was a strong correlation between binding affinity of kinase inhibitors for ABCG2 and their IC<sub>50</sub> for transport inhibition. This supports the idea that there is a spectrum of inhibitors to substrates, where the higher the affinity of a compound, the more inhibitor character it has (Table 3.1). This suggests that perhaps Ko143 can be turned into a substrate and that the lower fluorescence of the cavity 1 mutants (except Mut1) compared with the WT-ABCG2 expressing cells in Figure 3.13 could be due to transport of the fluorescent Ko143 derivatives.

However, there are some other ideas. Kapoor et al. (2018) hypothesised that substrates enter cavity 1 via a surface “access site”, binding to which triggers conformational changes required for ATP binding and hydrolysis. In contrast, they propose that inhibitors bind directly to cavity 1 so ATP hydrolysis does not occur and the inhibitor is not transported. If this is true then reducing the affinity for an inhibitor will only reduce IC<sub>50</sub> (Gose et al., 2020) and won't make it a substrate. However, with the hypothesis in this project, ATP hydrolysis would also be inhibited because without the release of the inhibitor from cavity 1, the transport cycle cannot reset. However, phenolic indenoindole inhibitors of ABCG2 stimulate ATPase activity while also inhibiting mitoxantrone transport (Gozzi et al., 2015). Gozzi et al. (2015) suggest that this is due to multiple binding sites but perhaps they enter cavity 1 via the access site

which allows the conformational changes to occur. By combining these ideas, it is possible that some inhibitors can enter via the access site, activating ATPase activity, and others cannot. In relation to the hypothesis in this project, maybe it is only possible for an inhibitor to become a substrate (by decreasing affinity), if it enters via the access site.

Cholesterol has a unique relationship between structure and function when it comes to binding to and modulating ABCG2. At least 20% (w/w) cholesterol in the cell membrane is required for function of ABCG2 (Storch et al., 2007) and Telbisz et al. (2013) found that depletion of cholesterol in Madin Darby canine kidney cells inhibits pheophorbide A transport. In their cryo-EM structures, Jackson et al. (2018) found ordered cholesterol molecules peripheral to ABCG2, which are potentially involved in modulation. Two cholesterol molecules are observed in cavity 1 in the cryo-EM structures by Taylor et al. (2017) and bind in the same location as MZ29, the Ko143 derivative (Jackson et al., 2018, Kerr et al., 2021). The outcome of this binding is unknown since cholesterol is not considered to be a substrate for transport. Perhaps it can act as an inhibitor in cavity 1, preventing the protein from hydrolysing ATP in a so-called “futile cycle” and is also an allosteric modulator at the sites on the protein periphery (Kerr et al., 2021).

New cryo-EM structures of ABCG2 show that part of the transmembrane helix 2 (TM2, residues 434-438) is unwound in the apo-state compared with the substrate/inhibitor-bound state where this region is fully helical (Orlando and Liao, 2020, Jackson et al., 2018, Manolaridis et al., 2018). This unravelling positions some residues important for ligand binding (e.g. F439) away from cavity 1 which is sealed off in this conformation. Since R482, which is situated in the access site and is known

for affecting substrate specificity, interacts with the unravelled portion of TM2, it has been suggested that control over the conformation of TM2 could be a factor in distinguishing between inhibitors and substrates (Orlando and Liao, 2020, Kapoor et al., 2018).

Egido et al. (2015) found that compounds that activate ABCG2 ATPase activity tend to be hydrophilic, non-amphiphilic and highly charged. On the other hand, inhibitors tend to be hydrophobic, amphiphilic and moderately charged to non-charged. In contrast to the hypothesis suggested in this thesis, they propose that the molecular properties relating to amphiphilicity, hydrophobicity, ionisation indicate whether a compound is an inhibitor or substrate (Egido et al., 2015). So, you would be able to predict where the compound falls on the substrate-inhibitor spectrum based on its properties. It would be interesting to examine how these properties correlate to affinity for ABCG2.

### **4.3 Future experiments**

Some experiments are already underway in the Kerr lab with the aim to provide more data to support or disprove the hypothesis. Since flow cytometry had more success with Ko143-Cy5 cell permeability than the monolayer transport assay, a Hoechst 33342 transport assay will be performed using a combined approach of both assays (section 2.5.2 and 2.6). This will include WT and untransfected HEK cells treated with Hoechst 33342 in absence or presence of Ko143 or Ko143-Cy5 as described in Table 2.4 and in Cox et al. (2018) and in Kapoor et al. (2020). If this shows a different result to the monolayer transport assay, a dose response curve will be produced with flow cytometry by measuring inhibition by Ko143, Ko143-Cy5 and Ko143-X-BY630 at different concentrations. This produces a value for half maximal inhibition

concentration ( $IC_{50}$ ). Comparison of this value for the original Ko143 with the modified derivatives would indicate whether they were more or less potent inhibitor of ABCG2. For the cavity 1 mutants,  $IC_{50}$  of untagged Ko143 will be calculated by performing a plate reader accumulation assay as described by Horsey et al. (2020) and measuring mitoxantrone transport at a range of Ko143 concentrations. This would enable quantification of the impact of any one mutant on the interaction of ABCG2 with Ko143.

As for the flow cytometry transport assay (section 2.5.2 and 3.6.2), perhaps adding fluorescent Ko143 in excess will negate the effect of WT having increased binding to fluorescent Ko143. The overall fluorescence of the cell would be higher in non-transporting cell lines so the impact of the “stickiness” of fluorescent Ko143 derivatives to ABCG2 would be decreased. This could potentially decrease the difference between HEK293T cells and the WT-ABCG2 expressing cells and could help clarify whether the decrease in fluorescence from mutant to WT-ABCG2 expressing cells is due to transport of fluorescent Ko143 derivatives or not.

Another method distinguishing between transport and decreased binding would be to prepare inside-out vesicles from cells overexpressing WT and mutant ABCG2 (Karlsson et al., 2010, Toyoda et al., 2019). ABCG2 would then pump substrates and potentially the fluorescent Ko143 derivatives into the vesicle and the fluorescence of these vesicles could be measured to find the accumulation of the fluorescent Ko143 derivatives inside the vesicle. The benefit of this approach is that the cell permeability of Ko143-Cy5 would not be an issue. Ko143-Cy5 in the media would be able interact with cavity 1 of ABCG2 without having to cross the cell membrane first. Also, the medium can be treated with reagents for ATP regeneration or with a non-

hydrolysable ATP analogue AMP-PNP, which allows the measurement of ATP-dependent transport (Karlsson et al., 2010, Toyoda et al., 2019). The difference between vesicle fluorescence in the presence of ATP or AMP-PNP would show how much the fluorescent Ko143 derivatives are transported.

In an ideal world, affinity measurements of the mutant ABCG2 proteins for Ko143 could be compared to WT and a range of substrates, perhaps by measuring thermal shift of ligand binding as described by Gose et al. (2020). This is because there will be a minimum affinity threshold at which transport is no longer possible. It could be possible that the mutations made in this project are too strong and resultant affinity of the Ko143 derivatives is too low to bind, let alone be transported. So even if it is possible to make a substrate out of an inhibitor, we do not know if these experimental mutations are too extreme.

Ko143-X-BY630 inhibits Hoechst 33342 transport to almost the same extent as unmodified Ko143 which means in terms of affinity Ko143-X-BY630 is a good model for how Ko143 would behave in the same circumstances. Yet, the fluorescent tag might still have an impact on transport, potentially being too large or even restricting the conformational changes required for transport. Ko143-Cy5 behaves similarly to Ko143-X-BY630 in the flow cytometry but even after the inhibitory capability is confirmed the tag could prevent transport. Perhaps some inhibitors can never be substrates for the same reason and the hypothesis in this project might only apply to some inhibitors. Radiolabelling Ko143 with a radioactive atom, such as  $^3\text{H}$ , would be a suitable method to overcome the effect of the large fluorescent tags.

If the hypothesis in this project is ever supported by more conclusive data, the next step would be to make chemical modifications to existing substrates to increase

affinity for ABCG2. Not only does this have the potential for creating non-toxic inhibitors from substrates such as E<sub>1</sub>S, but perhaps altering drugs that are affected by multidrug resistance (e.g. mitoxantrone or methotrexate, (Doyle and Ross, 2003)) might also prevent their own transport.

#### **4.4 Conclusion**

Without further experiments there is currently ambiguity about whether the mutants made lead to Ko143 transport. However, it is likely that lower fluorescence of the cavity 1 mutants is due to reduced binding to the fluorescent Ko143 derivatives or perhaps a small level of transport. Data from Gose et al. (2020) compliments the hypothesis in this thesis, showing more potent inhibitors have a higher affinity than weaker inhibitors. Mutations to introduce a smaller reduction in affinity or measuring the affinity for Ko143 in the current mutants will help clarify if the experimental mutations are too strong to transport. It is potentially possible for inhibitors to become substrates but other factors, such as access site binding, could also influence substrate/inhibitor quality which prevents some inhibitors being substrates.

## Chapter 5      References

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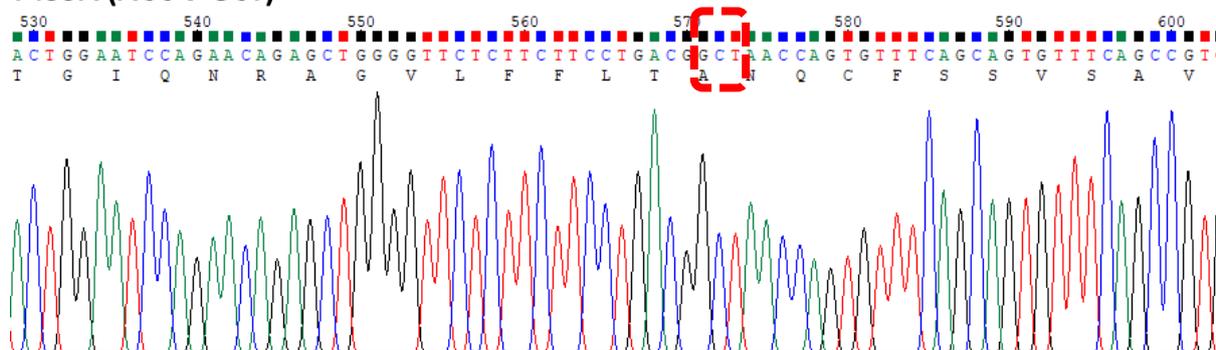


## Chapter 6 Appendix

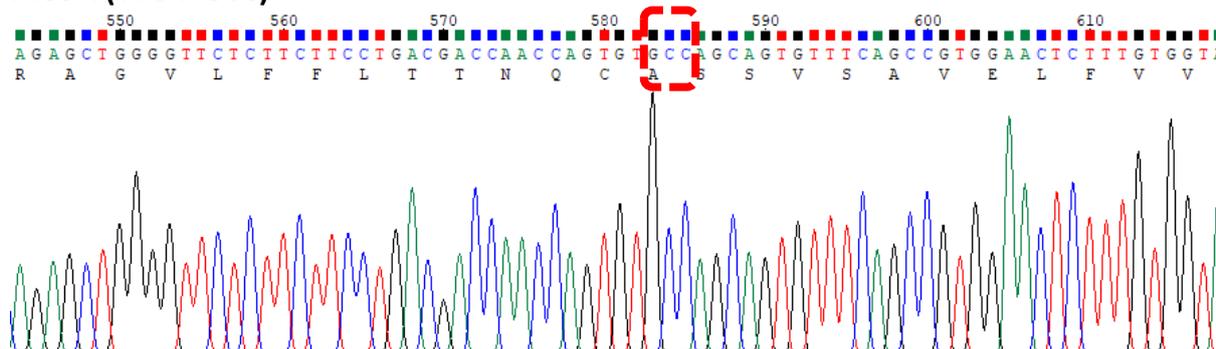
### 6.1 Sequencing chromatograms

Portions of the sequencing chromatograms of the remaining mutant ABCG2 constructs (N436A is shown in Figure 3.6) are shown below. The SeqF2 primer was used for T435A, F439A, S440W, A397S/V401A (Mut1) and L405A (Mut2). The Seq482 primer was used for M549E, L539A (Mut1) and I543A/V546A (Mut2). The red box highlights the desired mutation and no other mutations were found.

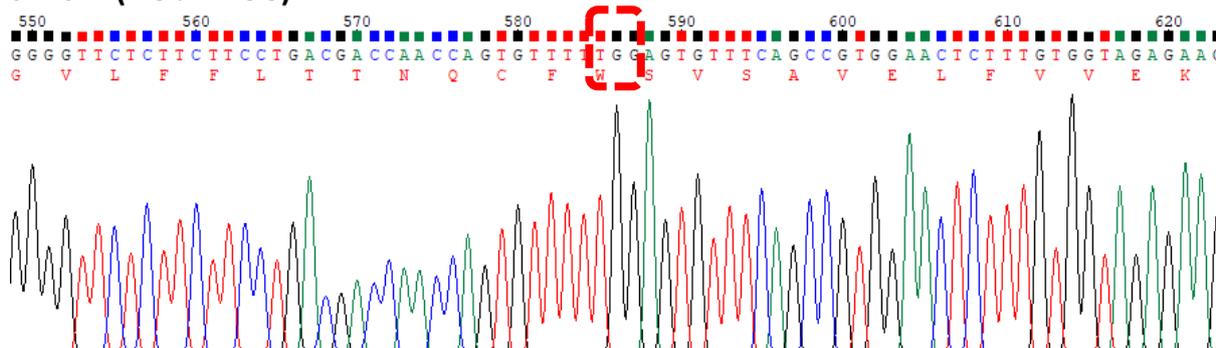
#### T435A (ACC→GCT)



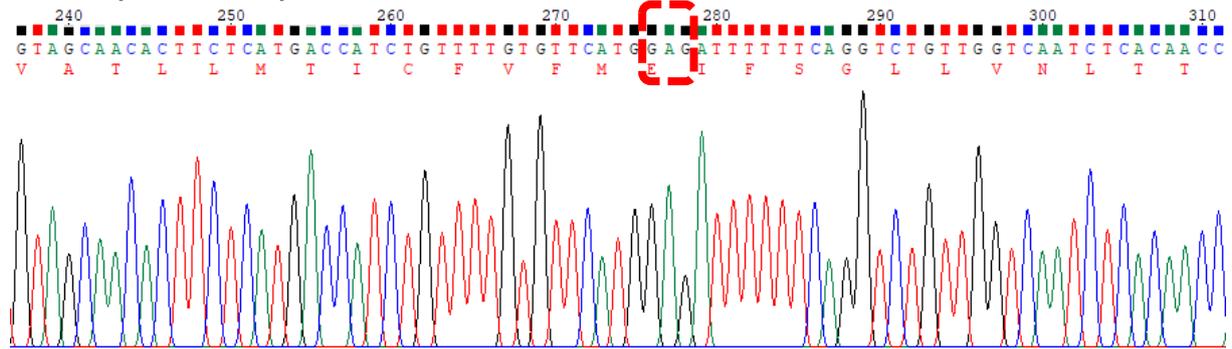
#### F439A (TTC→GCC)



#### S440W (AGC→TGG)

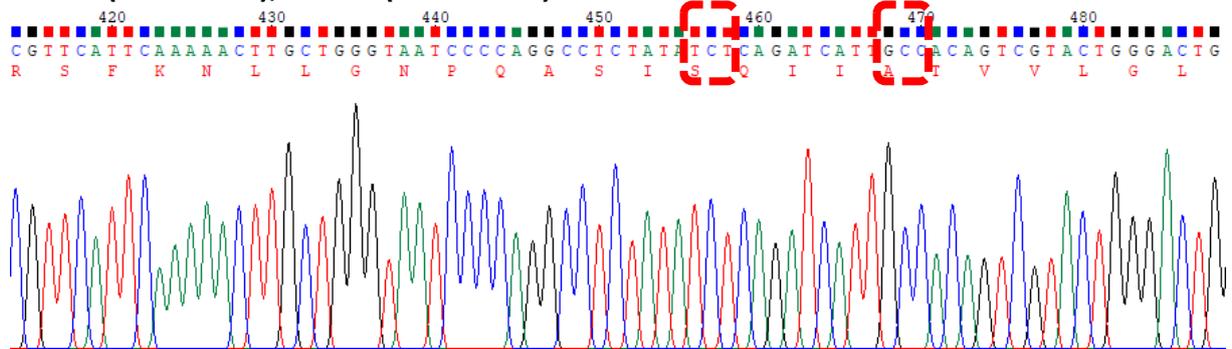


**M549E (ATG→GAG)**



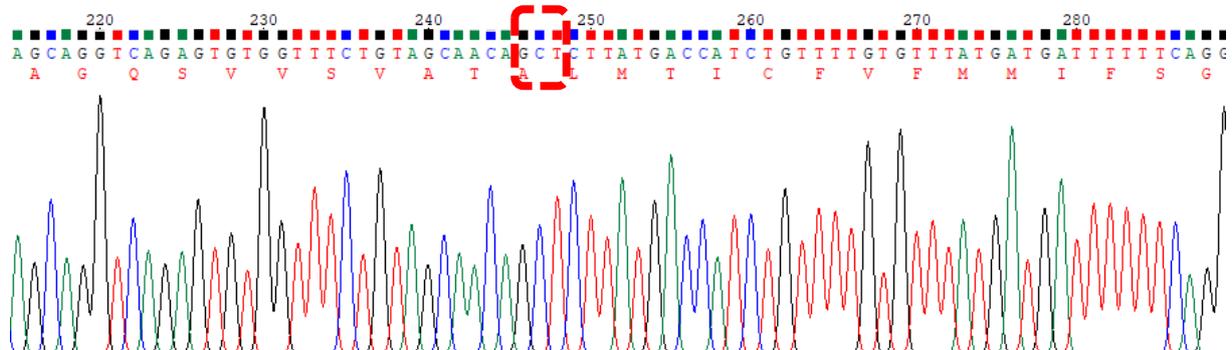
**Mut1**

**A397S (GCT→TCT), V401A (GTC→GCC)**



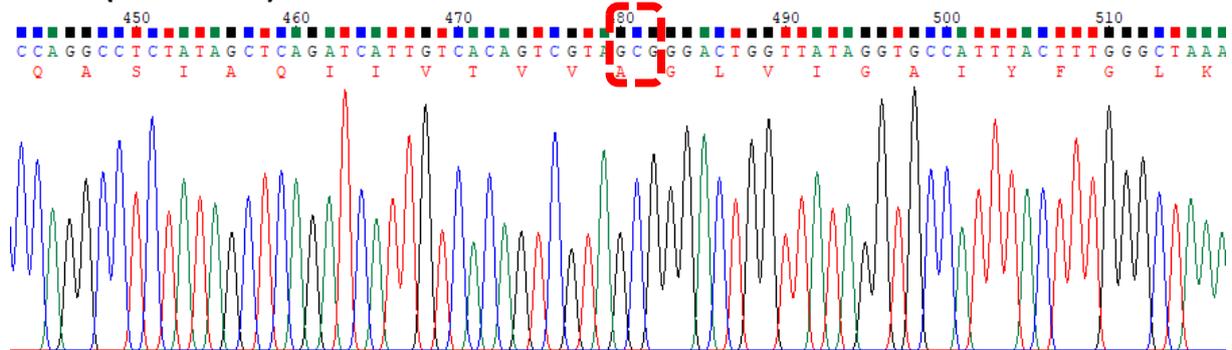
**Mut1**

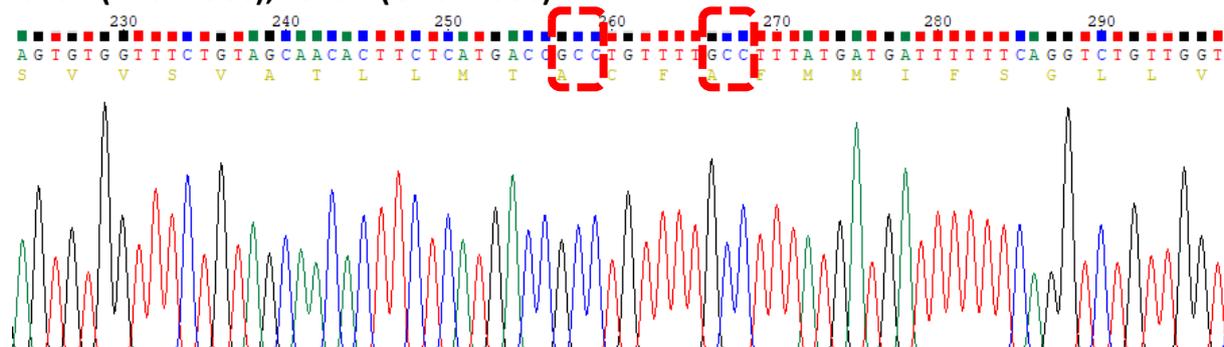
**L539A (CTT→GCT)**



**Mut2**

**L405A (CTG→GCG)**



**Mut2****I543A (ATC→GCC), V546A (GTG→GCC)****6.2 Sequence alignments**

The sequence alignments of the WT construct (Query) aligned with the sequence determined by Sanger sequencing (Sbjct) is shown below for each primer.

**6.2.1 T435A**

## T7 promoter (F)

Score	Expect	Identities	Gaps	Strand
2508 bits(1358)	0.0	1373/1383(99%)	2/1383(0%)	Plus/Plus
Query 912	AGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTTCGAGAAGGGAGGTGGAAGCGG			971
Sbjct 16	AGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTTCGAGAAGGGAGGTGGAAGCGG			75
Query 972	TGGAGGCTCAGGAGGCAGCGCATGGTCCCACCCCAAGTTTGAAAAGCTTGCCACCATGGA			1031
Sbjct 76	TGGAGGCTCAGGAGGCAGCGCATGGTCCCACCCCAAGTTTGAAAAGCTTGCCACCATGGA			135
Query 1032	CAAAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCTGGGCAAGCTGGAACGTGC			1091
Sbjct 136	CAAAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCTGGGCAAGCTGGAACGTGC			195
Query 1092	TGGGTGCGAACAGGGCCTGCACGAGATCAAGCTGCTGGGCAAGGAACATCTGCCCGCGA			1151
Sbjct 196	TGGGTGCGAACAGGGCCTGCACGAGATCAAGCTGCTGGGCAAGGAACATCTGCCCGCGA			255
Query 1152	CGCCGTGGAAAGTGCC TGCCCGAGCCCGCTGCTGGGCGGACAGAGCCACTGATGCAAGC			1211
Sbjct 256	CGCCGTGGAAAGTGCC TGCCCGAGCCCGCTGCTGGGCGGACAGAGCCACTGATGCAAGC			315
Query 1212	CACCGCTGGCTCAACGCCTACTTTACCAGCTGAGGCCATCGAGGAGTTCCCTGTGCC			1271
Sbjct 316	CACCGCTGGCTCAACGCCTACTTTACCAGCTGAGGCCATCGAGGAGTTCCCTGTGCC			375
Query 1272	AGCCCTGCACCACCCAGTGTTCACGAGGAGAGCTTTACCCGCCAGGTGCTGTGGAAACT			1331
Sbjct 376	AGCCCTGCACCACCCAGTGTTCACGAGGAGAGCTTTACCCGCCAGGTGCTGTGGAAACT			435
Query 1332	GCTGAAAGTGGTGAAGTTCGGAGAGGT CATCAGCTACCAGCAGCTGGCCGCGCTGGCCGG			1391
Sbjct 436	GCTGAAAGTGGTGAAGTTCGGAGAGGT CATCAGCTACCAGCAGCTGGCCGCGCTGGCCGG			495
Query 1392	CAATCCCGCCGCGCACCGCCGCTGAAAACCGCCCTGAGCGGAAATCCCGTGCCCATTTCT			1451
Sbjct 496	CAATCCCGCCGCGCACCGCCGCTGAAAACCGCCCTGAGCGGAAATCCCGTGCCCATTTCT			555
Query 1452	GATCCCCTGCCACCGGGTGGTGTCTAGCTCTGGCGCCGTGGGGGGCTACGAGGGCGGGCT			1511
Sbjct 556	GATCCCCTGCCACCGGGTGGTGTCTAGCTCTGGCGCCGTGGGGGGCTACGAGGGCGGGCT			615
Query 1512	CGCCGTGAAAGAGTGGCTGCTGGCCACGAGGGCCACAGACTGGGCAAGCTGGGCTGGG			1571
Sbjct 616	CGCCGTGAAAGAGTGGCTGCTGGCCACGAGGGCCACAGACTGGGCAAGCTGGGCTGGG			675
Query 1572	CGAATTCATGCTTCCAGTAATGTCGAAGTTTTTATCCAGTGTCAACAAGGAACACCAA			1631
Sbjct 676	CGAATTCATGCTTCCAGTAATGTCGAAGTTTTTATCCAGTGTCAACAAGGAACACCAA			735
Query 1632	TGGCTTCCCGCGACAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTGTTAAG			1691
Sbjct 736	TGGCTTCCCGCGACAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTGTTAAG			795
Query 1692	TTTTATAACATCTGCTATCGAGTAAAAGTGAAGAGTGGCTTTCTACCTTGTGCGAAAACC			1751
Sbjct 796	TTTTATAACATCTGCTATCGAGTAAAAGTGAAGAGTGGCTTTCTACCTTGTGCGAAAACC			855
Query 1752	AGT TGAGAAAGAAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGCCAT			1811
Sbjct 856	AGT TGAGAAAGAAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGCCAT			915
Query 1812	CCTGGGACCCACAGGTGGAGGCAAAATCTTCGTTATTAGATGTCTTAGCTGCAAGGAAAGA			1871
Sbjct 916	CCTGGGACCCACAGGTGGAGGCAAAATCTTCGTTATTAGATGTCTTAGCTGCAAGGAAAGA			975
Query 1872	TCCAAGTGGATATCTGGAGATGTTCTGATAAATGGAGCACACGACCTGCCAACTTCAA			1931
Sbjct 976	TCCAAGTGGATATCTGGAGATGTTCTGATAAATGGAGCACACGACCTGCCAACTTCAA			1035
Query 1932	ATGTAATTCAGGTTACGTTGATCAAGATGATGTTGTGATGGGCACCTGACGGTGAGAGA			1991
Sbjct 1036	ATGTAATTCAGGTTACGTTGATCAAGATGATGTTGTGATGGGCACCTGACGGTGAGAGA			1095
Query 1992	AAACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGaaaaaaa			2051
Sbjct 1096	AAACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAAAAAAA			1155
Query 2052	CGAACGGATTAAACAGGGTCATTCAAGAGTTAGGCTTGGATAAAGTGGCAGACTCCAAGGT			2111
Sbjct 1156	CGAACGGATTAAACAGGGTCATTCAAGAGTTAGGCTTGGATAAAGTGGCANACTCCAAGGT			1215
Query 2112	TGGAACCTCAGTTTATCCGTGGTGTCTTGGAGGAGAAAAG-AAAAAGGACTAGTATAGGAA			2170
Sbjct 1216	TGGAACCTCAGTTTATCCGTGGTGTCTTGGAGGAGAAAANAAAAAGGACTAGTNTAGGAA			1275
Query 2171	TGGAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTTAGACT			2230
Sbjct 1276	TGGAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGNACCTACAACCTGGCTTAGACT			1335
Query 2231	CAAGCACAGCAAATGCTGTCTTTTGCCTTGAAAAGGATGCTAAGCAGGG-ACGAAACA			2289
Sbjct 1336	CAAGCACAGCAAATGCTGTCTTTTGCCTTGAAAAGGATGCTAAGCCGGGGANNAACA			1395
Query 2290	ATC 2292			
Sbjct 1396	ATC 1398			

## SeqF1

Score	Expect	Identities	Gaps	Strand
2362 bits(1279)	0.0	1334/1365(98%)	9/1365(0%)	Plus/Plus
Query 1877	GTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTCAAATGTA			1936
Sbjct 24	GTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTCAAATGTA			83
Query 1937	ATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACCTGACGGTGAGAGAAAAC			1996
Sbjct 84	ATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACCTGACGGTGAGAGAAAAC			143
Query 1997	TACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGaaaaaaCGAAC			2056
Sbjct 144	TACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAAAAAAAACGAAC			203
Query 2057	GGATTAACAGGGTCATTCAAGAGTAGGTCGGATAAAGTGGCAGACTCCAAGGTTGGAA			2116
Sbjct 204	GGATTAACAGGGTCATTCAAGAGTAGGTCGGATAAAGTGGCAGACTCCAAGGTTGGAA			263
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Sbjct 264	CTCAGTTTATCCCGTGGTGTCTGGAGGAGAAAAGAAAAGGACTAGTATAGGAATGGAC			323
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Sbjct 324	TTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTTAGACTCAAGCA			383
Query 2237	CAGCAAAATGCTGTCC TTTTGTCTGAAAAGGATGCTAAGCAGGGACGAACAATCATCT			2296
Sbjct 384	CAGCAAAATGCTGTCC TTTTGTCTGAAAAGGATGCTAAGCAGGGACGAACAATCATCT			443
Query 2297	TCTCCATTATCAGCCTCGATATCCATCTTCAAGTTGTTTGTAGCCTCACCTTATGG			2356
Sbjct 444	TCTCCATTATCAGCCTCGATATCCATCTTCAAGTTGTTTGTAGCCTCACCTTATGG			503
Query 2357	CCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGACTTTGAATCAG			2416
Sbjct 504	CCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGACTTTGAATCAG			563
Query 2417	CTGGTTATCACTGTGAGGCTATAATAACCTGCAGACTTCTTCTTGGACATCATTAATG			2476
Sbjct 564	CTGGTTATCACTGTGAGGCTATAATAACCTGCAGACTTCTTCTTGGACATCATTAATG			623
Query 2477	GAGATTCCACTGCTGTGGCATTAAACAGAGAAGAAGACTTAAAGCCACAGAGATCATAG			2536
Sbjct 624	GAGATTCCACTGCTGTGGCATTAAACAGAGAAGAAGACTTAAAGCCACAGAGATCATAG			683
Query 2537	AGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTTATGTCAACTCCT			2596
Sbjct 684	AGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTTATGTCAACTCCT			743
Query 2597	CCTTCTACAAGAGACAAAAGCTGAATTACATCAACTTTCCGGGGTGAGAAGAAGAAGA			2656
Sbjct 744	CCTTCTACAAGAGACAAAAGCTGAATTACATCAACTTTCCGGGGTGAGAAGAAGAAGA			803
Query 2657	AGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTCACTCAACTCAGATGGG			2716
Sbjct 804	AGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTCACTCAACTCAGATGGG			863
Query 2717	TTTCTAAGCGTTCATTCAAAAACTTGTGGGTAATCCCAGGCTCTATAGCTCAGATCA			2776
Sbjct 864	TTTCTAAGCGTTCATTCAAAAACTTGTGGGTAATCCCAGGCTCTATAGCTCAGATCA			923
Query 2777	TTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAAATGATT			2836
Sbjct 924	TTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAAATGATT			983
Query 2837	CTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTTCTGACGACCAACCAAGTGTTC			2896
Sbjct 984	CTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTTCTGACGCTAACCAAGTGTTC			1043
Query 2897	GCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACATGAATACA			2956
Sbjct 1044	GCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACATGAATACA			1103
Query 2957	TCAGCGGATACTACAGAGTGTCACTTATTTCTTGGAAAACTGTTATCTGATTTATTAC			3016
Sbjct 1104	TCAGCGGATACTACAGAGTGTCACTTATTTCTTGGAAAACTGTTATCTGATTTATTAC			1163
Query 3017	CCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTACTTCATGTTAGGATTGA			3076
Sbjct 1164	CCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTACTTCATGTTAGGATTGA			1223
Query 3077	AGCCAAAAGGCAGATGCCCTTCTCGTTATGAT-GTTTACCCCTTATGATGGTGGCTTATTCA			3135
Sbjct 1224	AGCCAAAAGGCAGATGCCCTTCTCGTTATGAAAGTTTACCCCTTATGANGGTGGCTTATTCA			1283
Query 3136	GCCAGTTCATGG-CACTGG-CCATAGCAGC-AGGTGAGAGT-GTGGTTCTGTAGCAAC			3191
Sbjct 1284	GCCAGTTCATGGNCNCTGGNCCATAGCANCCAGGTCANAANNGTGGTTNNNGTAACAAC			1343
Query 3192	ACTT-CTCATGACCATCTG-TTTTGTG-TTTATGAT-GATTTTTT			3232
Sbjct 1344	NNTTNCNNGGACCATCTGGTTTTGGGGTTAAGAANGAATTTTT			1388

## SeqF2

Score	Expect	Identities	Gaps	Strand
2457 bits(1330)	0.0	1363/1384(98%)	1/1384(0%)	Plus/Plus
Query 2325	CTTCAAGTTGTTT	GATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGCC		2384
Sbjct 16	CTTC-AGTTGTTT	GATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGCC		74
Query 2385	TGCTCAGGAGGCCTT	GGGATACTTTGAATCAGCTGGTTATCACTGTGAGGCCTATAATAA		2444
Sbjct 75	TGCTCAGGAGGCCTT	GGGATACTTTGAATCAGCTGGTTATCACTGTGAGGCCTATAATAA		134
Query 2445	CCCTGCAGACTTCTT	CTGGACATCATTAAATGGAGATTCACCTGCTGTGGCATTAAACAG		2504
Sbjct 135	CCCTGCAGACTTCTT	CTGGACATCATTAAATGGAGATTCACCTGCTGTGGCATTAAACAG		194
Query 2505	AGAAAGAGACTTTAA	AGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCAT		2564
Sbjct 195	AGAAAGAGACTTTAA	AGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCAT		254
Query 2565	AGAAAAATTAGCGG	AGATTTATGTCAACTCCTCCTTCTACAAAGAGACAAAAGCTGAATT		2624
Sbjct 255	AGAAAAATTAGCGG	AGATTTATGTCAACTCCTCCTTCTACAAAGAGACAAAAGCTGAATT		314
Query 2625	ACATCAACTTTCCG	GGGTGAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTA		2684
Sbjct 315	ACATCAACTTTCCG	GGGTGAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTA		374
Query 2685	CACCACCTCCTTCT	GTCACTCAACTCAGATGGGTTTCTAAGCGTTCAATCAAAAACCTTGCT		2744
Sbjct 375	CACCACCTCCTTCT	GTCACTCAACTCAGATGGGTTTCTAAGCGTTCAATCAAAAACCTTGCT		434
Query 2745	GGGTAATCCCCAG	GCCTCTATAGCTCAGATCATTGTACAGTCTGACTGGGACTGGTTAT		2804
Sbjct 435	GGGTAATCCCCAG	GCCTCTATAGCTCAGATCATTGTACAGTCTGACTGGGACTGGTTAT		494
Query 2805	AGGTGCCATTTACT	TTGGGCTAAAAATGATTC TACTGGAAATCCAGAACAGAGCTGGGGT		2864
Sbjct 495	AGGTGCCATTTACT	TTGGGCTAAAAATGATTC TACTGGAAATCCAGAACAGAGCTGGGGT		554
Query 2865	TCTCTTCTCCTG	ACGACCAACAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTGT		2924
Sbjct 555	TCTCTTCTCCTG	ACGACCAACAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTGT		614
Query 2925	GGTAGAGAAGAAG	CTTCATACATGAATACATCAGCGGATACTACAGAGTGCATCTTA		2984
Sbjct 615	GGTAGAGAAGAAG	CTTCATACATGAATACATCAGCGGATACTACAGAGTGCATCTTA		674
Query 2985	TTTCCTTGGAAA	ACTGTTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATATT		3044
Sbjct 675	TTTCCTTGGAAA	ACTGTTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATATT		734
Query 3045	TACCTGTATAGT	GACTTCATGTTAGGATTGAAGCCAAAGGCAGATGCCCTTCTCGTTAT		3104
Sbjct 735	TACCTGTATAGT	GACTTCATGTTAGGATTGAAGCCAAAGGCAGATGCCCTTCTCGTTAT		794
Query 3105	GATGTTTACCCT	TATGATGGTGGCTTATTCAGCCAGTTCATGGCACTGGCCATAGCAGC		3164
Sbjct 795	GATGTTTACCCT	TATGATGGTGGCTTATTCAGCCAGTTCATGGCACTGGCCATAGCAGC		854
Query 3165	AGGTGAGAGTGG	TTCTGTAGCAACACTTCTCATGACCATCTGTTTGTGTTTATGAT		3224
Sbjct 855	AGGTGAGAGTGG	TTCTGTAGCAACACTTCTCATGACCATCTGTTTGTGTTTATGAT		914
Query 3225	GATTTTTTCAGG	TCTGTTGGTCAATCTCACAACCATTGCATCTTGGCTGTCATGGCTTCA		3284
Sbjct 915	GATTTTTTCAGG	TCTGTTGGTCAATCTCACAACCATTGCATCTTGGCTGTCATGGCTTCA		974
Query 3285	GTACTTCAGCAT	TCCACGATATGGATTACGGCTTTCAGCATAATGAATTTTGGGACA		3344
Sbjct 975	GTACTTCAGCAT	TCCACGATATGGATTACGGCTTTCAGCATAATGAATTTTGGGACA		1034
Query 3345	AAACTTCGCCAG	GACTCAATGCAACAGGAAACAATCCTTGTAACTATGCAACATGTAC		3404
Sbjct 1035	AAACTTCGCCAG	GACTCAATGCAACAGGAAACAATCCTTGTAACTATGCAACATGTAC		1094
Query 3405	TGGCGAAGAATA	TTTGGTAAAGCAGGGCATCGATCTCTACCCTGGGGCTTGTGGAAGAA		3464
Sbjct 1095	TGGCGAAGAATA	TTTGGTAAAGCAGGGCATCGATCTCTACCCTGGGGCTTGTGGAANAA		1154
Query 3465	TCACGTGGCCTT	GGCTTGTATGATTGTATTTTCCCTCACAATTGCC TACCTGAAATTGTT		3524
Sbjct 1155	TCACGTGGCCTT	GGCTTGTATGATTGTATTTTCCCTCACAATTGCC TACCTGAAATTGTT		1214
Query 3525	ATTTCTTAAAAA	TATTTCTTAAATTTGGATTCTAGAGGGCCCGTTTAAACCCGCTGATCAG		3584
Sbjct 1215	ATTTCTTAAAAA	TATTTCTTAAATTTGGATTCTAGAGGGCCCGTTTAAACCCGCTGATCAG		1274
Query 3585	CCTCGACTGTGC	CTTCTAGTTGCCAGCCATCTGTTGTTTCCCTCCCCCTCCCCCGTCCCT		3644
Sbjct 1275	CCTCGACTGTGC	CTTCTAGTTGCCAGCCATCTGTTGTTTCCCTCCCCCTCCCCCGTCCCT		1334
Query 3645	TGACCTTGGAA	GGTGCCACTCCACTGCTTTTCTAATAAAATGAGGAAATTCATCGC		3704
Sbjct 1335	TGACCTTGGAA	GGTGCCACTCCACTGCTTTTCTAATAAAATGAGGAAATTCATCGC		1394
Query 3705	ATTG 3708			
Sbjct 1395	ATTG 1398			

## Seq482

Score	Expect	Identities	Gaps	Strand
2283 bits(1236)	0.0	1285/1310(98%)	12/1310(0%)	Plus/Plus
Query 2960	GCGGATACTACAGAGTGCATCTTATTTCCTTGGAAAACGTGTTATCTGATTATTACCCA			3019
Sbjct 13	GCGGNNACTACAGAGTGCATCTTATTTCCTTGGAAAACGTGTTATCTGATTATTACCCA			72
Query 3020	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			3079
Sbjct 73	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			132
Query 3080	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCCTATGATGGTGGCTTATTCAGCCA			3139
Sbjct 133	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCCTATGATGGTGGCTTATTCAGCCA			192
Query 3140	GTTCCATGGCACTGGCCATAGCAGCAGGTGAGTGTGGTTTCTGTAGCAACACTTCTCA			3199
Sbjct 193	GTTCCATGGCACTGGCCATAGCAGCAGGTGAGTGTGGTTTCTGTAGCAACACTTCTCA			252
Query 3200	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGTCGTGGTCAATCTCACAAACA			3259
Sbjct 253	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGTCGTGGTCAATCTCACAAACA			312
Query 3260	TTGCATCTTGGCTGTGATGGCTTCACTTCCAGCATTCCACGATATGGATTTACGGCTT			3319
Sbjct 313	TTGCATCTTGGCTGTGATGGCTTCACTTCCAGCATTCCACGATATGGATTTACGGCTT			372
Query 3320	TGCAGCATAATGAATTTTTGGGACAAAACCTTCCAGGACTCAATGCAACAGGAAACA			3379
Sbjct 373	TGCAGCATAATGAATTTTTGGGACAAAACCTTCCAGGACTCAATGCAACAGGAAACA			432
Query 3380	ATCCTTGTAACTATGCAACATGACTGGCGAAGAATATTGGTAAAGCAGGGCATCGATC			3439
Sbjct 433	ATCCTTGTAACTATGCAACATGACTGGCGAAGAATATTGGTAAAGCAGGGCATCGATC			492
Query 3440	TCTCACCTGGGGCTTGTGGAAGAATCACGTGGCCTTGGCTTGTATGATTGTTATTTCC			3499
Sbjct 493	TCTCACCTGGGGCTTGTGGAAGAATCACGTGGCCTTGGCTTGTATGATTGTTATTTCC			552
Query 3500	TCACAAATGCCTACCTGAAATTTGTTATTTCTTAAAAAATATTCTTAAATTGGATTCTAGA			3559
Sbjct 553	TCACAAATGCCTACCTGAAATTTGTTATTTCTTAAAAAATATTCTTAAATTGGATTCTAGA			612
Query 3560	GGGCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCTTCTAGTTGCCAGCCATCTGTT			3619
Sbjct 613	GGGCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCTTCTAGTTGCCAGCCATCTGTT			672
Query 3620	GTTTGGCCCCCCCCGCTTCCCTTGACCCCTGGAAGGTGCCACTCCCCTGTCCTTTCC			3679
Sbjct 673	GTTTGGCCCCCCCCGCTTCCCTTGACCCCTGGAAGGTGCCACTCCCCTGTCCTTTCC			732
Query 3680	TAATAAAATGAGGAAATTCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTgggggt			3739
Sbjct 733	TAATAAAATGAGGAAATTCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTgggggt			792
Query 3740	gggggtggggCAGGACAGCAAGGGGGAGGATTGGGAAGCAATAGCAGGCATGCTGGGGAT			3799
Sbjct 793	gggggtggggCAGGACAGCAAGGGGGAGGATTGGGAAGCAATAGCAGGCATGCTGGGGAT			852
Query 3800	GCGGTGGGCTCTATGGCTTCTGAGGCGAAAGAACCAGCTGGGGCTCTAGGGGGATCCC			3859
Sbjct 853	GCGGTGGGCTCTATGGCTTCTGAGGCGAAAGAACCAGCTGGGGCTCTAGGGGGATCCC			912
Query 3860	CACGCGCCCTGTAGCGGCGCATTAAAGCGCGCGGGGTGGTGGTTACGCGCAGCGTGACC			3919
Sbjct 913	CACGCGCCCTGTAGCGGCGCATTAAAGCGCGCGGGGTGGTGGTTACGCGCAGCGTGACC			972
Query 3920	GCTACACTTGCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCTTTCTCGCC			3979
Sbjct 973	GCTACACTTGCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCTTTCTCGCC			1032
Query 3980	ACGTTTCGCGGGCTTCCCCGTCAAGCTCTAAATCGGGGCATCCCTTTAGGGTCCGATTT			4039
Sbjct 1033	ACGTTTCGCGGGCTTCCCCGTCAAGCTCTAAATCGGGGCATCCCTTTAGGGTCCGATTT			1092
Query 4040	AGTGCITTTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGTGGTTACGTTAGTGGG			4099
Sbjct 1093	AGTGCITTTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGTGGTTACGTTAGTGGG			1152
Query 4100	CCATCGCCCTGATAGACGG- TTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTAAATAG			4158
Sbjct 1153	CCATCGCCCTGATAGACGG- TTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTAAATAG			1212
Query 4159	TGG- ACTCTTGTCCAACTGG- AACAACT- CAACCCTATCT- -CGGTCTATT- TTT			4212
Sbjct 1213	TGGAACCTTGGTCCAACTGGNAACAACACTTCAACCNNNTTNNCGGCTATTCTNTT			1272
Query 4213	TGA- TTTATAAGGGA- TTTTGGGGA- TTTTCGGCC- TATTGG- TTAATAA			4257
Sbjct 1273	TGAATTTATAAGGGAATTTGCCNAATTTTCGGCCCTATTGGGTTAAAAA			1322

## 6.2.2 N436A/S622S

## T7 promoter (F)

Score	Expect	Identities	Gaps	Strand
2300 bits(1245)	0.0	1293/1323(98%)	9/1323(0%)	Plus/Plus
Query 912	AGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGAAGCGG			971
Sbjct 16	AGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGAAGCGG			75
Query 972	TGGAGGCTCAGGAGGCAGCGCATGGTCCACCCCCAGTTTGAAAAGCTTGCCACCATGGA			1831
Sbjct 76	TGGAGGCTCAGGAGGCAGCGCATGGTCCACCCCCAGTTTGAAAAGCTTGCCACCATGGA			135
Query 1032	CAAAGACTGCGAAAATGAAGCGCACCCCTGGATAGCCCTCTGGGCAAGCTGGAACCTGTC			1091
Sbjct 136	CAAAGACTGCGAAAATGAAGCGCACCCCTGGATAGCCCTCTGGGCAAGCTGGAACCTGTC			195
Query 1092	TGGGTGCGAACAGGGCCTGCACGAGATCAAGCTGCTGGGCAAAGGAACATCTGCCGCCGA			1151
Sbjct 196	TGGGTGCGAACAGGGCCTGCACGAGATCAAGCTGCTGGGCAAAGGAACATCTGCCGCCGA			255
Query 1152	CGCCGTGGAAGTGCC TGCCCCAGCCCGCTGCTGGGCGGACAGAGCCACTGATGCAGGC			1211
Sbjct 256	CGCCGTGGAAGTGCC TGCCCCAGCCCGCTGCTGGGCGGACAGAGCCACTGATGCAGGC			315
Query 1212	CACCGCTGGCTCAACGCCTACTTTCACAGCCTGAGGCCATCGAGGAGTTCCTGTGCC			1271
Sbjct 316	CACCGCTGGCTCAACGCCTACTTTCACAGCCTGAGGCCATCGAGGAGTTCCTGTGCC			375
Query 1272	AGCCCTGCACCACCCAGTGTTCAGCAGGAGAGCTTTACCCGCCAGGTGCTGTGAAACT			1331
Sbjct 376	AGCCCTGCACCACCCAGTGTTCAGCAGGAGAGCTTTACCCGCCAGGTGCTGTGAAACT			435
Query 1332	GCTGAAGTGGTGAAGTTCGGAGAGGT CATCAGTACCAGCAGCTGGCCGCCCTGGCCGG			1391
Sbjct 436	GCTGAAGTGGTGAAGTTCGGAGAGGT CATCAGTACCAGCAGCTGGCCGCCCTGGCCGG			495
Query 1392	CAATCCCGCCGCCACCGCCCGGTGAAAACCGCCCTGAGCGGAAATCCCGTGCCATTCT			1451
Sbjct 496	CAATCCCGCCGCCACCGCCCGGTGAAAACCGCCCTGAGCGGAAATCCCGTGCCATTCT			555
Query 1452	GATCCCCTGCCACCGGGTGGTGTCTAGCTTGCGCCGTGGGGGGCTACGAGGGCGGGCT			1511
Sbjct 556	GATCCCCTGCCACCGGGTGGTGTCTAGCTTGCGCCGTGGGGGGCTACGAGGGCGGGCT			615
Query 1512	CGCCGTGAAAGAGTGGCTGCTGGCCACGAGGGCCACAGACTGGGCAAGCC TGGGCTGGG			1571
Sbjct 616	CGCCGTGAAAGAGTGGCTGCTGGCCACGAGGGCCACAGACTGGGCAAGCC TGGGCTGGG			675
Query 1572	CGAATTCATGCTTCCAGTAATGTCGAAGTTTTTATCCAGTGTCAACAAGGAAACACAA			1631
Sbjct 676	CGAATTCATGCTTCCAGTAATGTCGAAGTTTTTATCCAGTGTCAACAAGGAAACACAA			735
Query 1632	TGGCTTCCCGCGACAGCTTCCAATGACCTGAAGGCATT TACTGAAGGAGCTGTGTTAAG			1691
Sbjct 736	TGGCTTCCCGCGACAGCTTCCAATGACCTGAAGGCATT TACTGAAGGAGCTGTGTTAAG			795
Query 1692	TTTTATAACATCTGCTATCGAGTAAACTGAAGAGTGGCTTCTACCTTGTGCAAAACC			1751
Sbjct 796	TTTTATAACATCTGCTATCGAGTAAACTGAAGAGTGGCTTCTACCTTGTGCAAAACC			855
Query 1752	AGTTGAGAAAGAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGCCAT			1811
Sbjct 856	AGTTGAGAAAGAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGCCAT			915
Query 1812	CCTGGGACCCACAGGTGGAGGCAAATCTTCGTTATTAGATGCTTAGCTGCAAGGAAAGA			1871
Sbjct 916	CCTGGGACCCACAGGTGGAGGCAAATCTTCGTTATTAGATGCTTAGCTGCAAGGAAAGA			975
Query 1872	TCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCAGCTGCCAACTTCAA			1931
Sbjct 976	TCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCAGCTGCCAACTTCAA			1035
Query 1932	ATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGA			1991
Sbjct 1036	ATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGA			1095
Query 1992	AAACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAAC TATGACGAATCATGaaaaaaa			2051
Sbjct 1096	AAACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAAC TATGACGAATCATG-AAAAAA			1153
Query 2052	CGAACGGATTAAACAGGGTCATTCAAGAGTAGGCTGGATAAAGTGGCAGACTCCAAGGT			2111
Sbjct 1154	CGAACGGATTANNAGGGTCATT CAGAANTTAGGCTGGAT -AAGTGGCAGACTCCAAGGT			1212
Query 2112	TGGAACCTCAGTTTATCCGTGGTGTGCTGGAGGAGAAAGAAAAAGGACTAGTATAGGAAT			2171
Sbjct 1213	-GGAACTCN-TTTATCCGNGNN-TNTCTGGAGGAGAAAGAAAAAGGACTAGTNNAGGAAT			1269
Query 2172	GGAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTTAGACTC			2231
Sbjct 1270	GGAGCTTATNNCTGATCCTTCCNNTTGTCTT-GGATGACCNTNCA-CTGGCTTACA-TC			1326
Query 2232	AAG 2234			
Sbjct 1327	AAG 1329			

## SeqF1

Score	Expect	Identities	Gaps	Strand
2362 bits(1279)	0.0	1318/1340(98%)	6/1340(0%)	Plus/Plus
Query 1873	CCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTCAAA			1932
Sbjct 20	CCNAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTCAAA			79
Query 1933	TGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGACTCTGACGGTGAGAGAA			1992
Sbjct 80	TGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGACTCTGACGGTGAGAGAA			139
Query 1993	AACTTACAGTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGaaaaaaaaC			2052
Sbjct 140	AACTTACAGTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAAAAAAAAAC			199
Query 2053	GAACGGATTAAACAGGGTCATTCAAGAGTTAGGCTGGATAAAGTGGCAGACTCCAAGGTT			2112
Sbjct 200	GAACGGATTAAACAGGGTCATTCAAGAGTTAGGCTGGATAAAGTGGCAGACTCCAAGGTT			259
Query 2113	GGAACTCAGTTTATCCGTGGTGTGCTGGAGGAGAAAAAGAAAGGACTAGTATAGGAATG			2172
Sbjct 260	GGAACTCAGTTTATCCGTGGTGTGCTGGAGGAGAAAAAGAAAGGACTAGTATAGGAATG			319
Query 2173	GAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGACTTACACTCA			2232
Sbjct 320	GAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGACTTACACTCA			379
Query 2233	AGCACAGCAAAATGCTGTCCTTTGCTCCTGAAAAGGATGCTAAGCAGGGACGAAACAATC			2292
Sbjct 380	AGCACAGCAAAATGCTGTCCTTTGCTCCTGAAAAGGATGCTAAGCAGGGACGAAACAATC			439
Query 2293	ATCTTCTCCATTTCATCAGCCTCGATATCCATCTTCAAGTTGTTGATAGCCTCACCTTA			2352
Sbjct 440	ATCTTCTCCATTTCATCAGCCTCGATATCCATCTTCAAGTTGTTGATAGCCTCACCTTA			499
Query 2353	TTGGCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGATACTTTGAA			2412
Sbjct 500	TTGGCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGATACTTTGAA			559
Query 2413	TCAGCTGGTTATCACGTGAGGCCTATAATAACCTGCAGACTTCTTCTGGACATCATT			2472
Sbjct 560	TCAGCTGGTTATCACGTGAGGCCTATAATAACCTGCAGACTTCTTCTGGACATCATT			619
Query 2473	AATGGAGATTCCACTGCTGTGGCATTAAACAGAGAAGAAAGACTTTAAAGCCACAGAGATC			2532
Sbjct 620	AATGGAGATTCCACTGCTGTGGCATTAAACAGAGAAGAAAGACTTTAAAGCCACAGAGATC			679
Query 2533	ATAGAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTATGTCAAC			2592
Sbjct 680	ATAGAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTATGTCAAC			739
Query 2593	TCCCTCTTACAAAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAAGAAG			2652
Sbjct 740	TCCCTCTTACAAAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAAGAAG			799
Query 2653	AAGAAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTTCATCAACTCAGA			2712
Sbjct 800	AAGAAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTTCATCAACTCAGA			859
Query 2713	TGGGTTTCTAAGCGTTCATTCAAAAACTTGCTGGGTAATCCCCAGGCCCTATAGCTCAG			2772
Sbjct 860	TGGGTTTCTAAGCGTTCATTCAAAAACTTGCTGGGTAATCCCCAGGCCCTATAGCTCAG			919
Query 2773	ATCATTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAAAT			2832
Sbjct 920	ATCATTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAAAT			979
Query 2833	GATTCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTGACGACCAACCAGTGT			2892
Sbjct 980	GATTCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTGACGACCAACCAGTGT			1039
Query 2893	TTCAGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACATGAA			2952
Sbjct 1040	TTCAGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACATGAA			1099
Query 2953	TACATCAGCGGATACACAGAGTGCATCTTATTTCTTGGAAAACTGTTATCTGATTTA			3012
Sbjct 1100	TACATCAGCGGATACACAGAGTGCATCTTATTTCTTGGAAAACTGTTATCTGATTTA			1159
Query 3013	TTACCCATGAGGATGTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGA			3072
Sbjct 1160	TTACCCATGAGGATGTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGA			1219
Query 3073	TTGAAGCCAAAGGCAGATGCCCTTCTCGTTATGATGTTTACCCTTATGATGGT-GGCTTA			3131
Sbjct 1220	TTGAAGCCAAAGGCAGATGCCCTTCTCGTTATGATGTTTACCCTTATGAAAGGNGGCTTA			1279
Query 3132	TTCAGCCAGTTCAT-GGCACTGGCCATAGCAGC-AGGTGAGAGTG-TGGTTTCTG-TAG			3187
Sbjct 1280	TTCAGCCAGTTCATNGGCCTGGCCATANNAACCAAGGTGAGANNGGTGGTTNCNGGTAA			1339
Query 3188	CAACACTT-CTCATGACCAT 3206			
Sbjct 1340	CAANCTTTCTCAGGACCAT 1359			

## SeqF2

Score	Expect	Identities	Gaps	Strand
2492 bits(1349)	0.0	1396/1427(98%)	5/1427(0%)	Plus/Plus
Query 2324	TCTTCAAGTGTGTTGATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGC			2383
Sbjct 14	TCTTC-AGTTGTTGATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGC			72
Query 2384	CTGCTCAGGAGGCTTTGGGATACTTTGAATCAGCTGGTTATCACTGTGAGGCCTATAATA			2443
Sbjct 73	CTGCTCAGGAGGCTTTGGGATACTTTGAATCAGCTGGTTATCACTGTGAGGCCTATAATA			132
Query 2444	ACCCTGCAGACTTCTCTTGGACATCATTAAATGGAGATTCCACTGCTGTGGCATTAAACA			2583
Sbjct 133	ACCCTGCAGACTTCTCTTGGACATCATTAAATGGAGATTCCACTGCTGTGGCATTAAACA			192
Query 2584	GAGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCA			2563
Sbjct 193	GAGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCA			252
Query 2564	TAGAAAAATAGCGGAGATTTATGTCAACTCCTCCTTCTACAAGAGACAAAAGCTGAAT			2623
Sbjct 253	TAGAAAAATAGCGGAGATTTATGTCAACTCCTCCTTCTACAAGAGACAAAAGCTGAAT			312
Query 2624	TACATCAACTTTCCGGGGTGAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCT			2683
Sbjct 313	TACATCAACTTTCCGGGGTGAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCT			372
Query 2684	ACACCACCTCCTTCTGTCAACTCAGATGGGTTCTAAGCGTTTCAAAAACTTGC			2743
Sbjct 373	ACACCACCTCCTTCTGTCAACTCAGATGGGTTCTAAGCGTTTCAAAAACTTGC			432
Query 2744	TGGGTAATCCCAGGCCCTATAGCTCAGATCATTGTACAGTCTGACTGGGACTGGTTA			2883
Sbjct 433	TGGGTAATCCCAGGCCCTATAGCTCAGATCATTGTACAGTCTGACTGGGACTGGTTA			492
Query 2884	TAGGTGCCATTTACTTTGGGCTAAAAAATGATTCTACTGGAATCCAGAACAGAGCTGGGG			2863
Sbjct 493	TAGGTGCCATTTACTTTGGGCTAAAAAATGATTCTACTGGAATCCAGAACAGAGCTGGGG			552
Query 2864	TTCTCTTCTCCTGACGACCAACCAAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTG			2923
Sbjct 553	TTCTCTTCTCCTGACGACCCCAAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTG			612
Query 2924	TGGTAGAGAAGAAGCTCTTCATACATGAATACATCAGCGGATACTACAGAGTGCATCTT			2983
Sbjct 613	TGGTAGAGAAGAAGCTCTTCATACATGAATACATCAGCGGATACTACAGAGTGCATCTT			672
Query 2984	ATTTCCCTGGAAAACTGTTATCTGATTATTACCCATGAGGATGTTACCAAGTATTATAT			3043
Sbjct 673	ATTTCCCTGGAAAACTGTTATCTGATTATTACCCATGAGGATGTTACCAAGTATTATAT			732
Query 3044	TTACCTGTATAGTGTACTTCATGTTAGGATTGAAGCCAAAGGCAGATGCCTTCTCGTTA			3183
Sbjct 733	TTACCTGTATAGTGTACTTCATGTTAGGATTGAAGCCAAAGGCAGATGCCTTCTCGTTA			792
Query 3184	TGATGTTTACCCTTATGATGGTGGCTTATTCAGCCAGTCCATGGCACTGGCCATAGCAG			3163
Sbjct 793	TGATGTTTACCCTTATGATGGTGGCTTATTCAGCCAGTCCATGGCACTGGCCATAGCAG			852
Query 3164	CAGGTCAGAGTGTGGTTCTGTAGCAACTTCTCATGACCATCTGTTTGTGTTTATGA			3223
Sbjct 853	CAGGTCAGAGTGTGGTTCTGTAGCAACTTCTCATGACCATCTGTTTGTGTTTATGA			912
Query 3224	TGATTTTTTACAGTCTGTGGTCAATCTCACAACTTGCATCTGGCTGTATGGCTTC			3283
Sbjct 913	TGATTTTTTACAGTCTGTGGTCAATCTCACAACTTGCATCTGGCTGTATGGCTTC			972
Query 3284	AGTACTTCAGCATTCCACGATATGGATTACGGCTTTCAGCATAATGAATTTTGGGAC			3343
Sbjct 973	AGTACTTCAGCATTCCACGATATGGATTACGGCTTTCAGCATAATGAATTTTGGGAC			1032
Query 3344	AAAACCTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGTAACTATGCAACATGTA			3403
Sbjct 1033	AAAACCTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGTAACTATGCAACATGTA			1092
Query 3484	CTGGCGAAGAATATTTGGTAAAGCAGGGCATCGATCTCTCACCTGGGGCTTGTGGAAGA			3463
Sbjct 1093	CTGGCGAAGAATATTTGGTAAAGCAGGGCATCGATCTCTCGCCCTGGGGCTTGTGGAANA			1152
Query 3464	ATCACGTGGCCTTGGCTGTATGATTGTTATTTCCCTACAATTGCCTACCTGAAATTTGT			3523
Sbjct 1153	ATCACGTGGCCTTGGCTGTATGATTGTTATTTCCCTACAATTGCCTACCTGAAATTTGT			1212
Query 3524	TATTTCTAAAAAATATTCTTAAATGGATTCTAGAGGGCCCGTTAAACCCGCTGATCA			3583
Sbjct 1213	TATTTCTAAAAAATATTCTTAAATGGATTCTAGAGGGCCCGTTAAACCCGCTGATCA			1272
Query 3584	GCCTCGACTGTGCTTCTAGTTGCCAGCCATCTGTTGTTGCCCTCCCCGTGCTTCC			3643
Sbjct 1273	GCCTCGACTGTGCTTNNAGTTGCCAGCCATCTGTTGTTGCCCTCCCCGTGCTTCC			1332
Query 3644	TTGACCCTGGAAGGTGCCACTCCCCTTCCCTTCCCTTCCCTTCCCTTCCCTTCCCTTCCCT			3702
Sbjct 1333	TTGACCCTGGAAGGTGCCNNCCNN-TNNCTTCC-AAATAAANGGAGGAAATGTCATC			1390
Query 3783	GCATTGTCTG-AGTAGGTGTACTTATTCTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT			3748
Sbjct 1391	GCATTGGCCGNANTAGGTNNCATNNNTTTCGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG			1437

## Seq482

Score	Expect	Identities	Gaps	Strand
2052 bits(1111)	0.0	1143/1166(98%)	2/1166(0%)	Plus/Plus
Query 2960	GCGGATACTACAGAGTGCATCTTATTCCCTTGGAAAACTGTTATCTGATTATTACCCA			3019
Sbjct 11	GCGGNNACTACAGAGTGCATCTTATTCCCTTGGAAAACTGTTATCTGATTATTACCCA			70
Query 3020	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTACTTCATGTTAGGATTGAAGC			3079
Sbjct 71	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTACTTCATGTTAGGATTGAAGC			130
Query 3080	CAAAGGCAGATGCCCTCTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTCAGCCA			3139
Sbjct 131	CAAAGGCAGATGCCCTCTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTCAGCCA			190
Query 3140	GTTCCATGGCACTGGCCATAGCAGCAGGTCAGAGTGTGGTTTCTGTAGCAACACTTCTCA			3199
Sbjct 191	GTTCCATGGCACTGGCCATAGCAGCAGGTCAGAGTGTGGTTTCTGTAGCAACACTTCTCA			250
Query 3200	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGCTGTGGTCAATCTCACAAACA			3259
Sbjct 251	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGCTGTGGTCAATCTCACAAACA			310
Query 3260	TTGCATCTTGGCTGTGATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCTT			3319
Sbjct 311	TTGCATCTTGGCTGTGATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCTT			370
Query 3320	TGCAGCATAATGAATTTTTGGGACAAAACCTTCCCGCAGGACTCAATGCAACAGGAAACA			3379
Sbjct 371	TGCAGCATAATGAATTTTTGGGACAAAACCTTCCCGCAGGACTCAATGCAACAGGAAACA			430
Query 3380	ATCCTTGAACATGCAACATGACTGGCGAAGAATATTGGTAAAGCAGGGCATCGATC			3439
Sbjct 431	ATCCTTGAACATGCAACATGACTGGCGAAGAATATTGGTAAAGCAGGGCATCGATC			490
Query 3440	TCTCACCTGGGGCTGTGGAAAGAAACACGTGGCCTTGGCTTGTATGATTGTTATTTTCC			3499
Sbjct 491	TCTCGCCCTGGGGCTGTGGAAAGAAACACGTGGCCTTGGCTTGTATGATTGTTATTTTCC			550
Query 3500	TCACAATTGCCACCTGAAATGTTATTTCTTAAAAAATATTCTTAAATTGGATTCTAGA			3559
Sbjct 551	TCACAATTGCCACCTGAAATGTTATTTCTTAAAAAATATTCTTAAATTGGATTCTAGA			610
Query 3560	GGGCCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCTTCTAGTTGCCAGCCATCTGTT			3619
Sbjct 611	GGGCCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCTTCTAGTTGCCAGCCATCTGTT			670
Query 3620	GTTTGGCCCTCCCGGCTGCTTCCCTTACCCTGGAAAGGTGCCACTCCCACTGTCCTTTCC			3679
Sbjct 671	GTTTGGCCCTCCCGGCTGCTTCCCTTACCCTGGAAAGGTGCCACTCCCACTGTCCTTTCC			730
Query 3680	TAATAAAATGAGGAAATGTCATCGCATGTCTGAGTAGGTGTCATTCTATTCTgggggt			3739
Sbjct 731	TAATAAAATGAGGAAATGTCATCGCATGTCTGAGTAGGTGTCATTCTATTCTgggggt			790
Query 3740	ggggggggCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGAT			3799
Sbjct 791	ggggggggCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGAT			850
Query 3800	GCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTTAGGGGTATCCC			3859
Sbjct 851	GCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGNGCTTAGNGGTATCCC			910
Query 3860	CACGCGCCCTGTAGCGGCGCATTAAAGCGCGGCGGGTGTGGTGGTTACGCGCAGCGTGACC			3919
Sbjct 911	CACGCGCCCTGTAGCGGCGCATTAAAGCGCGGCGGGTGTGGTGGTTACGCGCAGCGTGACC			970
Query 3920	GCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCCCTTCTCGCC			3979
Sbjct 971	GCTACACTTGCCAGCGCCCTANCGCCCGCTCCTTTCGCTTCTTCCCTTCCCTTCTCGCC			1030
Query 3980	ACGTTTCGCCGGCTTCCCGTCAAGCTCTAAATCGGGG-CATCCCTTATAGGGTTCAGGATT			4038
Sbjct 1031	ACGTTTCGCCGGCTTCCCGTCAAGCTCTAAATCGGGGNCCTCCCTTATAGGNTTCAGGATT			1090
Query 4039	TAGTGCTTTACGGCACCTCGACCCCAAAAA-ACCTTGATTAGGGTGATGGTTACAGTAGTG			4097
Sbjct 1091	TAGTGCTTTACGGCACCTCNACCCNAAAAANACTTGATTAGGGTGAAAGTTACAGTANTG			1150
Query 4098	GGCCATCGCCCTGATAGACGGTTTTTT 4123			
Sbjct 1151	GGCCATCNCCTGAANNACGGCTTTT 1176			

## 6.2.3 F439A

## T7 promoter (F)

Score	Expect	Identities	Gaps	Strand
2558 bits(1385)	0.0	1414/1436(98%)	3/1436(0%)	Plus/Plus
Query 907	ACTTAAAGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGA			966
Sbjct 15	ACTTNNNGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGA			74
Query 967	AGCGGTGGAGGCTCAGGAGGCAGCGCATGGTCCACCCCAAGTTTGAAAAGCTTGCCACC			1026
Sbjct 75	AGCGGTGGAGGCTCAGGAGGCAGCGCATGGTCCACCCCAAGTTTGAAAAGCTTGCCACC			134
Query 1027	ATGGACAAAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCTGGGCAAGCTGGAA			1086
Sbjct 135	ATGGACAAAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCTGGGCAAGCTGGAA			194
Query 1087	CTGTCTGGGTGCGAACAGGGCTGCACGAGATCAAGCTGCTGGGCAAGGAAACATCTGCC			1146
Sbjct 195	CTGTCTGGGTGCGAACAGGGCTGCACGAGATCAAGCTGCTGGGCAAGGAAACATCTGCC			254
Query 1147	GCCGACGCCCTGGAAAGTGCCTGCCAGCCGCGCTGCTGGGCGGACCAGAGCCACTGATG			1206
Sbjct 255	GCCGACGCCCTGGAAAGTGCCTGCCAGCCGCGCTGCTGGGCGGACCAGAGCCACTGATG			314
Query 1207	CAGGCCACCCCTGGCTCAACGCCTACTTTACCAGCCTGAGGCCATCGAGGAGTCCCT			1266
Sbjct 315	CAGGCCACCCCTGGCTCAACGCCTACTTTACCAGCCTGAGGCCATCGAGGAGTCCCT			374
Query 1267	GTGCCAGCCCTGCACACCCAGTGTTCAGCAGGAGAGCTTTACCCTGAGGTGCTGTGG			1326
Sbjct 375	GTGCCAGCCCTGCACACCCAGTGTTCAGCAGGAGAGCTTTACCCTGAGGTGCTGTGG			434
Query 1327	AAACTGCTGAAAGTGGTGAAGTTCGGAGAGGTTCATCAGCTACCAGCAGCTGGCCGCCCTG			1386
Sbjct 435	AAACTGCTGAAAGTGGTGAAGTTCGGAGAGGTTCATCAGCTACCAGCAGCTGGCCGCCCTG			494
Query 1387	GCCGGCAATCCCGCCGCCACCGCCGCGTGAAAACCCGCTGAGCGGAAATCCCGTGCC			1446
Sbjct 495	GCCGGCAATCCCGCCGCCACCGCCGCGTGAAAACCCGCTGAGCGGAAATCCCGTGCC			554
Query 1447	ATTCTGATCCCTGCCACCGGGTGGTCTAGCTCTGGCGCGTGGGGGGTACGAGGGC			1506
Sbjct 555	ATTCTGATCCCTGCCACCGGGTGGTCTAGCTCTGGCGCGTGGGGGGTACGAGGGC			614
Query 1507	GGGCTCGCCGTGAAAGAGTGGTCTGGCCACGAGGGCCACAGACTGGGCAAGCTGGG			1566
Sbjct 615	GGGCTCGCCGTGAAAGAGTGGTCTGGCCACGAGGGCCACAGACTGGGCAAGCTGGG			674
Query 1567	CTGGGCGAATTCATGCTTCCAGTAATGTCGAAGTTTTATCCAGTGTACAAGGAAAC			1626
Sbjct 675	CTGGGCGAATTCATGCTTCCAGTAATGTCGAAGTTTTATCCAGTGTACAAGGAAAC			734
Query 1627	ACCAATGGCTTCCCGCCGACAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTG			1686
Sbjct 735	ACCAATGGCTTCCCGCCGACAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTG			794
Query 1687	TTAAGTTTTATAACATCTGCTATCGAGTAAAACGAAGAGTGGCTTTCTACCTTGTGGA			1746
Sbjct 795	TTAAGTTTTATAACATCTGCTATCGAGTAAAACGAAGAGTGGCTTTCTACCTTGTGGA			854
Query 1747	AAACCAGTTGAGAAAATAATATCGAATATCAATGGGATCATGAAACCTGGTCTCAAC			1806
Sbjct 855	AAACCAGTTGAGAAAATAATATCGAATATCAATGGGATCATGAAACCTGGTCTCAAC			914
Query 1807	GCCATCTGGGACCCACAGGTGGAGGCAAACTTCGTTATTAGATGCTTAGCTGCAAGG			1866
Sbjct 915	GCCATCTGGGACCCACAGGTGGAGGCAAACTTCGTTATTAGATGCTTAGCTGCAAGG			974
Query 1867	AAAGATCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAAC			1926
Sbjct 975	AAAGATCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAAC			1034
Query 1927	TTCAAATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTTGACGGTG			1986
Sbjct 1035	TTCAAATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTTGACGGTG			1094
Query 1987	AGAGAAAACCTACAGTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGaa			2046
Sbjct 1095	AGAGAAAACCTACAGTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAA			1154
Query 2047	aaaaCGAACGGATTAAACAGGGTCATTCAAGAGTTAGGTCTGGATAAAGTGGCAGACTCC			2106
Sbjct 1155	AAAAACGAAACGGATTAAACAGGGTCATTCAAGAGTTAGGTCTGGATAAAGTGGCAGACTCC			1214
Query 2107	AAGGTTGGAACCTCAGTTTATCCGTGGTGTGCTGGAGGAGAAAAGAAAAA-GGACTAGTAT			2166
Sbjct 1215	AAGGTTGGAACCTCAGTTTATCCGTGGTGTGCTGGAGGAGAAAAGAAAAAAGGACTAGTAT			1274
Query 2166	AGGAATGGAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTT			2226
Sbjct 1275	AGGAATGGAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAACTACAACCTGGCTT			1334
Query 2226	AGACTCAAGCACAGCAAAATGCTGTCTTTTGTCTGAAAAGGATGCTAAGCAGGGACG			2286
Sbjct 1335	AGACTCAAGCANNAGCAAAATGCTGTCTTTTGTCTGAAAAGGATGCTAAGCAGGGANN			1394
Query 2286	AACAATCATCTTCT-CCATTATCAGCCTCG-ATATTCATCTTCAAGTTGTTTGA			2339
Sbjct 1395	AACAATCANCCTNNCCATTATCANNCCNCCNANNTTCCATCTTCAAGTTNNTTGA			1450

## SeqF1

Score	Expect	Identities	Gaps	Strand
2386 bits(1292)	0.0	1333/1353(99%)	8/1353(0%)	Plus/Plus
Query 1870	GATCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTC			1929
Sbjct 17	GATCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTC			76
Query 1930	AAATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGA			1989
Sbjct 77	AAATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGA			136
Query 1990	GAAAACCTACAGTTCCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGaaaa			2049
Sbjct 137	GAAAACCTACAGTTCCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAAAA			196
Query 2050	aaCGAACGGATTAACAGGGTCATTCAAGAGTTAGGTCTGGATAAAGTGGCAGACTCCAAG			2109
Sbjct 197	AACGAAACGGATTAACAGGGTCATTCAAGAGTTAGGTCTGGATAAAGTGGCAGACTCCAAG			256
Query 2110	GTTGGAACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAAGGACTAGTATAGGA			2169
Sbjct 257	GTTGGAACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAAGGACTAGTATAGGA			316
Query 2170	ATGGAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACGGCTTAGAC			2229
Sbjct 317	ATGGAGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACGGCTTAGAC			376
Query 2230	TCAAGCACAGCAAATGCTGTCTTTTGTCTCTGAAAAGGATGTCTAAGCAGGGACGAACA			2289
Sbjct 377	TCAAGCACAGCAAATGCTGTCTTTTGTCTCTGAAAAGGATGTCTAAGCAGGGACGAACA			436
Query 2290	ATCATCTTCTCCATTATCAGCCTCGATATTCATCTTCAAGTTGTTTATAGCCTCACC			2349
Sbjct 437	ATCATCTTCTCCATTATCAGCCTCGATATTCATCTTCAAGTTGTTTATAGCCTCACC			496
Query 2350	TTATTGGCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGATACTTT			2409
Sbjct 497	TTATTGGCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGATACTTT			556
Query 2410	GAATCAGCTGGTTATCACTGTGAGGCCATAATAACCTGCAGACTTCTTCTTGGACATC			2469
Sbjct 557	GAATCAGCTGGTTATCACTGTGAGGCCATAATAACCTGCAGACTTCTTCTTGGACATC			616
Query 2470	ATTAATGGAGATTCCACTGCTGGCATTAAACAGAGAAGAGACTTTAAAGCCACAGAG			2529
Sbjct 617	ATTAATGGAGATTCCACTGCTGGCATTAAACAGAGAAGAGACTTTAAAGCCACAGAG			676
Query 2530	ATCATAGAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTTATGTC			2589
Sbjct 677	ATCATAGAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTTATGTC			736
Query 2590	AACCTCTCTTCTACAAAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAAG			2649
Sbjct 737	AACCTCTCTTCTACAAAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAAG			796
Query 2650	AAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCTTCTGTATCAACTC			2709
Sbjct 797	AAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCTTCTGTATCAACTC			856
Query 2710	AGATGGGTTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCAGGCCCTATAGCT			2769
Sbjct 857	AGATGGGTTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCAGGCCCTATAGCT			916
Query 2770	CAGATCATTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTACTTTGGGCTAAAA			2829
Sbjct 917	CAGATCATTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTACTTTGGGCTAAAA			976
Query 2830	AATGATTCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTGACGACCAACCAG			2889
Sbjct 977	AATGATTCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTGACGACCAACCAG			1036
Query 2890	TGTTTCAGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACAT			2949
Sbjct 1037	TGTTTCAGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACAT			1096
Query 2950	GAATACATCAGCGGATACTACAGAGTGTATCTTATTTCTTGGAAAACCTGTTATCTGAT			3009
Sbjct 1097	GAATACATCAGCGGATACTACAGAGTGTATCTTATTTCTTGGAAAACCTGTTATCTGAT			1156
Query 3010	TTATTACCCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTA			3069
Sbjct 1157	TTATTACCCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTA			1216
Query 3070	GGATTGAAGCCAAAGGCGAGATGCCTTCTCGTTATGATGTTTACCCTTATGAT-GGTGGC			3128
Sbjct 1217	GGATTGAAGCCAAAGGCGAGATGCCTTCTCGTTATGATGTTTACCCTTATGAGGGTGGC			1276
Query 3129	TTATTCAGCCAGTTCATGG-CACTGG-CCATAGC-AGCAGGTCAG-AGT-GTGGTTTCT			3183
Sbjct 1277	TTATTCAGCCAGTTCATGGCACTGGNCCATAGNACCAGGTCNGNAANNGTGGTTTCT			1336
Query 3184	GTAGCAACACTTCTC-ATGACCATCTG-TTTTG 3214			
Sbjct 1337	GTAACAACACTTNCNAGGACCATCCGGTTTTG 1369			

## SeqF2

Score	Expect	Identities	Gaps	Strand
2396 bits(1297)	0.0	1323/1338(99%)	2/1338(0%)	Plus/Plus
Query 2325	CTTCAAGTTGTTGATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGCC			2384
Sbjct 16	CTTC-AGTTGTTGATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGCC			74
Query 2385	TGCTCAGGAGGCCTTGGGATACTTTGAATCAGCTGGTTATCACTGTGAGGCCTATAATAA			2444
Sbjct 75	TGCTCAGGAGGCCTTGGGATACTTTGAATCAGCTGGTTATCACTGTGAGGCCTATAATAA			134
Query 2445	CCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATTCCACTGCTGTGGCATTAAACAG			2504
Sbjct 135	CCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATTCCACTGCTGTGGCATTAAACAG			194
Query 2505	AGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCAT			2564
Sbjct 195	AGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCAT			254
Query 2565	AGAAAAATTAGCGGAGATTTATGTCAACTCCTCCTTCTACAAAGAGACAAAAGCTGAATT			2624
Sbjct 255	AGAAAAATTAGCGGAGATTTATGTCAACTCCTCCTTCTACAAAGAGACAAAAGCTGAATT			314
Query 2625	ACATCAACTTCCGGGGGTGAGAAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTA			2684
Sbjct 315	ACATCAACTTCCGGGGGTGAGAAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTA			374
Query 2685	CACCACCTCCTTCTGTCATCAACTCAGATGGGTTTCTAAGCGTTCAATCAAAAACCTTGCT			2744
Sbjct 375	CACCACCTCCTTCTGTCATCAACTCAGATGGGTTTCTAAGCGTTCAATCAAAAACCTTGCT			434
Query 2745	GGGTAATCCCCAGGCCTCTATAGCTCAGATCATTGTACAGTCTGACTGGGACTGGTTAT			2804
Sbjct 435	GGGTAATCCCCAGGCCTCTATAGCTCAGATCATTGTACAGTCTGACTGGGACTGGTTAT			494
Query 2805	AGGTGCCATTTACTTTGGGCTAAAAAATGATTTACTGGAATCCAGAACAGAGCTGGGGT			2864
Sbjct 495	AGGTGCCATTTACTTTGGGCTAAAAAATGATTTACTGGAATCCAGAACAGAGCTGGGGT			554
Query 2865	TCTCTTCTCCTGACGACCAACCAAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTGT			2924
Sbjct 555	TCTCTTCTCCTGACGACCAACCAAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTGT			614
Query 2925	GGTAGAGAAGAAGCTCTTCATACATGAATACATCAGCGGATACTACAGAGTGCATCTTA			2984
Sbjct 615	GGTAGAGAAGAAGCTCTTCATACATGAATACATCAGCGGATACTACAGAGTGCATCTTA			674
Query 2985	TTTCCTTGGAAAAGTGTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATATT			3044
Sbjct 675	TTTCCTTGGAAAAGTGTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATATT			734
Query 3045	TACCTGTATAGTACTTTCATGTTAGGATTGAAGCCAAAGGCAGATGCCCTTCTCGTTAT			3104
Sbjct 735	TACCTGTATAGTACTTTCATGTTAGGATTGAAGCCAAAGGCAGATGCCCTTCTCGTTAT			794
Query 3105	GATGTTTACCCCTTATGATGGTGGCTTATTCAGCCAGTTCATGGCACTGGCCATAGCAGC			3164
Sbjct 795	GATGTTTACCCCTTATGATGGTGGCTTATTCAGCCAGTTCATGGCACTGGCCATAGCAGC			854
Query 3165	AGGTCAGAGTGGGTTTCTGTAGCAACACTTCTCATGACCATCTGTTTTGTGTTTATGAT			3224
Sbjct 855	AGGTCAGAGTGGGTTTCTGTAGCAACACTTCTCATGACCATCTGTTTTGTGTTTATGAT			914
Query 3225	GATTTTTTCAGGCTCTGTTGGTCAATCTCACAACCATTGCATCTTGGCTGTCATGGCTTCA			3284
Sbjct 915	GATTTTTTCAGGCTCTGTTGGTCAATCTCACAACCATTGCATCTTGGCTGTCATGGCTTCA			974
Query 3285	GTACTTCAGCATTCCACGATATGGATTACGGCTTTCAGCATAATGAATTTTGGGACA			3344
Sbjct 975	GTACTTCAGCATTCCACGATATGGATTACGGCTTTCAGCATAATGAATTTTGGGACA			1034
Query 3345	AAACTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGTAACTATGCAACATGTAC			3404
Sbjct 1035	AAACTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGTAACTATGCAACATGTAC			1094
Query 3405	TGGCGAAGAATATTTGGTAAAGCAGGGCATCGATCTCTACCCCTGGGGCTTGTGGAAGAA			3464
Sbjct 1095	TGGCGAANAATATTTGGTAAAGCAGGGCATCGATCTCTACCCCTGGGGCTTGTGGAANAA			1154
Query 3465	TCACGTGGCCTTGGCTTGTATGATTGTATTTTCTCACAAATGCCCTACCTGAAATGTT			3524
Sbjct 1155	TCACGTGGCCTTGGCTTGGATGATTGTATTTTCTCACAAATGCCCTACCTGAAATGTT			1214
Query 3525	ATTTCTTAAAAAATATCTTAAATGGATTCTAGAGGGCCCGTTAAACCCGCTGATCAG			3584
Sbjct 1215	ATTTCTTAAAAAATATCTTAAATGGATTCTAGAGGGCCCGTTAAACCCGCTGATCAG			1274
Query 3585	CCTCGACTGTGCCTCTAGTTGCCAGCCATCTGTTGTTGCCCC-TCCCCGTGCCTTCC			3643
Sbjct 1275	CTCCAACNTGCTTCAAGTTGCCAGCCATCTGTTGTTGCCCCNTCCCCGGGCTTCC			1334
Query 3644	TTGACCTTGGAAAGGTGCC 3661			
Sbjct 1335	TTGACCTTGGAAAGGGCC 1352			

## Seq482

Score	Expect	Identities	Gaps	Strand
2300 bits(1245)	0.0	1301/1330(98%)	16/1330(1%)	Plus/Plus
Query 2966	ACTACAGAGTGCATCTTATTTCCCTGGAAAACGTGTTATCTGATTTATTACCCATGAGGA			3025
Sbjct 20	ACTACAGAGTGCATCTTATTTCCCTGGAAAACGTGTTATCTGATTTATTACCCATGAGGA			79
Query 3026	TGTTACCAAGTATTATATTACCTGTATAGTGTACTTCATGTTAGGATTGAAGCCAAAGG			3085
Sbjct 80	TGTTACCAAGTATTATATTACCTGTATAGTGTACTTCATGTTAGGATTGAAGCCAAAGG			139
Query 3086	CAGATGCCTTCTCGTTATGATGTTTACCCCTATGATGGTGGCTTATTCAGCCAGTTCCA			3145
Sbjct 140	CAGATGCCTTCTCGTTATGATGTTTACCCCTATGATGGTGGCTTATTCAGCCAGTTCCA			199
Query 3146	TGGCACTGGCCATAGCAGCAGGTGAGAGTGGTTTCTGTAGCAACACTTCTCATGACCA			3205
Sbjct 200	TGGCACTGGCCATAGCAGCAGGTGAGAGTGGTTTCTGTAGCAACACTTCTCATGACCA			259
Query 3206	TCTGTTTTGTGTTTATGATGATTTTTTTCAGGCTGTGGTCAATCTCACAAACCATTCAT			3265
Sbjct 260	TCTGTTTTGTGTTTATGATGATTTTTTTCAGGCTGTGGTCAATCTCACAAACCATTCAT			319
Query 3266	CTTGGCTGTGCATGGCTTCAGTACTTCAGCATTCCACGATATGGATTACGGCTTTCAGC			3325
Sbjct 320	CTTGGCTGTGCATGGCTTCAGTACTTCAGCATTCCACGATATGGATTACGGCTTTCAGC			379
Query 3326	ATAATGAATTTTTGGGACAAAACCTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTT			3385
Sbjct 380	ATAATGAATTTTTGGGACAAAACCTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTT			439
Query 3386	GTAACATGCAACATGTACTGGCGAAGAATATTTGGTAAAGCAGGGCATCGATCTCTCAC			3445
Sbjct 440	GTAACATGCAACATGTACTGGCGAAGAATATTTGGTAAAGCAGGGCATCGATCTCTCAC			499
Query 3446	CCTGGGGCTGTGGAAAGAATCACGTGGCCTTGGCTTGTATGATTGTTATTTCCACAAA			3505
Sbjct 500	CCTGGGGCTGTGGAAAGAATCACGTGGCCTTGGCTTGTATGATTGTTATTTCCACAAA			559
Query 3506	TTGCCACCTGAAATGTTATTTCTAAAAAATATCTTAAATTGGATTCTAGAGGGCCC			3565
Sbjct 560	TTGCCACCTGAAATGTTATTTCTAAAAAATATCTTAAATTGGATTCTAGAGGGCCC			619
Query 3566	GTTTAAACCCGCTGATCAGCCTCGACTGTGCCTCTAGTTGCCAGCCATCTGTTGTTGC			3625
Sbjct 620	GTTTAAACCCGCTGATCAGCCTCGACTGTGCCTCTAGTTGCCAGCCATCTGTTGTTGC			679
Query 3626	CCCTCCCCCGTGCCTTCCCTTGACCTGGAAGGTGCCACTCCCCTGTCTTTCCATAATA			3685
Sbjct 680	CCCTCCCCCGTGCCTTCCCTTGACCTGGAAGGTGCCACTCCCCTGTCTTTCCATAATA			739
Query 3686	AATGAGGAAATGTCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTGGGGGTGGGGT			3745
Sbjct 740	AATGAGGAAATGTCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTGGGGGTGGGGT			799
Query 3746	GGGACAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGATGCGGTG			3805
Sbjct 800	GGGACAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGATGCGGTG			859
Query 3806	GGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTTAGGGGGTATCCCCACGCG			3865
Sbjct 860	GGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTTAGGGGGTATCCCCACGCG			919
Query 3866	CCCTGTAGCGCGCATTAAGCGCGCGGGGTGGTGGTTACGCGCAGCGTGACCGCTACA			3925
Sbjct 920	CCCTGTAGCGCGCATTAAGCGCGCGGGGTGGTGGTTACGCGCAGCGTGACCGCTACA			979
Query 3926	CTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCTTCTGCCACGTTT			3985
Sbjct 980	CTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCTTCTGCCACGTTT			1039
Query 3986	GCCGGCTTCCCGCTCAAGCTCTAAATCGGGGCATCCCTTTAGGGTCCGATTTAGTGCT			4045
Sbjct 1040	GCCGGCTTCCCGCTCAAGCTCTAAATCGGGGCATCCCTTTAGGGTCCGATTTAGTGCT			1099
Query 4046	TTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGTGGTTACAGTAGTGGCCATCG			4105
Sbjct 1100	TTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGTGGTTACAGTAGTGGCCATCG			1159
Query 4106	CCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTTAATAGTGGACT			4165
Sbjct 1160	CCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTTAATAGTGGACT			1219
Query 4166	TTGTTCCAAACTGG-AACAACAC-TCAACCTAT--CTCGG-TCTATTCTTTTGA-TTT			4218
Sbjct 1220	TTGTTCCAAACTGGNAANAACACCTCAACCTANNNTCTGGGTCTATTCTTTTGAATTT			1279
Query 4219	ATAAGGGA-TTTTGGGGA-TTTC-GGCCTATTGG-TTAAAAAAT-G-AG-CTGATTT-AA			4278
Sbjct 1280	ANAAGGGAATTTGNCNAATTTTCGGCTATTGGGTAAAAAANNNGNANNTGATTTTAA			1339
Query 4271	CAAAAA-TTT	4279		
Sbjct 1340	CAAAAAATTT	1349		

## 6.2.4 S440W

## T7 promoter (F)

Score	Expect	Identities	Gaps	Strand
2497 bits(1352)	0.0	1368/1380(99%)	1/1380(0%)	Plus/Plus
Query 914	CTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGAAGCGGTG			973
Sbjct 28	CTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGAAGCGGTG			79
Query 974	GAGGCTCAGGAGGAGCGCATGGTCCCACCCAGTTTGAAAAGCTTGCCACCATGGACA			1833
Sbjct 80	GAGGCTCAGGAGGAGCGCATGGTCCCACCCAGTTTGAAAAGCTTGCCACCATGGACA			139
Query 1834	AAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCGGGCAAGCTGGAAGTGTCTG			1893
Sbjct 140	AAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCGGGCAAGCTGGAAGTGTCTG			199
Query 1894	GGTGCGAACAGGGCTGCACGAGATCAAGCTGCTGGGCAAGGAACATCTGCCGCCGACG			1153
Sbjct 200	GGTGCGAACAGGGCTGCACGAGATCAAGCTGCTGGGCAAGGAACATCTGCCGCCGACG			259
Query 1154	CCGTGGAAGTGCCGCCCCAGCCGCGTGGGCGGACAGAGCCACTGATGCAGGCCA			1213
Sbjct 260	CCGTGGAAGTGCCGCCCCAGCCGCGTGGGCGGACAGAGCCACTGATGCAGGCCA			319
Query 1214	CCGCTGGCTCAACGCCTACTTTCACAGCCTGAGGCCATCGAGGAGTTCCTGTGCCAG			1273
Sbjct 320	CCGCTGGCTCAACGCCTACTTTCACAGCCTGAGGCCATCGAGGAGTTCCTGTGCCAG			379
Query 1274	CCCTGCACCACCCAGTGTTCAGCAGGAGAGCTTTACCCGCCAGGTGCTGTGAAAATGC			1333
Sbjct 380	CCCTGCACCACCCAGTGTTCAGCAGGAGAGCTTTACCCGCCAGGTGCTGTGAAAATGC			439
Query 1334	TGAAAGTGGTGAAGTTCGGAGAGGTCAACAGTACCAGCAGTGGCCGCCCTGGCCGGCA			1393
Sbjct 440	TGAAAGTGGTGAAGTTCGGAGAGGTCAACAGTACCAGCAGTGGCCGCCCTGGCCGGCA			499
Query 1394	ATCCCGCCGCCACCGCCGCGTGAACCCGCCCTGAGCGGAAATCCCGTGCCATTCTGA			1453
Sbjct 500	ATCCCGCCGCCACCGCCGCGTGAACCCGCCCTGAGCGGAAATCCCGTGCCATTCTGA			559
Query 1454	TCCCCTGCCACCCGGGTGGTGTAGCTCTGGCGCCGTGGGGGGCTACGAGGGCGGGCTCG			1513
Sbjct 560	TCCCCTGCCACCCGGGTGGTGTAGCTCTGGCGCCGTGGGGGGCTACGAGGGCGGGCTCG			619
Query 1514	CCGTGAAAAGAGTGGCTGCTGGCCACGAGGGCCACAGACTGGGCAAGCTGGGCTGGCG			1573
Sbjct 620	CCGTGAAAAGAGTGGCTGCTGGCCACGAGGGCCACAGACTGGGCAAGCTGGGCTGGCG			679
Query 1574	AATTCATGTCTCCAGTAATGTCGAAGTTTTATCCAGTGTCAAGGAAACACCAATG			1633
Sbjct 680	AATTCATGTCTCCAGTAATGTCGAAGTTTTATCCAGTGTCAAGGAAACACCAATG			739
Query 1634	GCTTCCCCGCGACAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTGTTAAGTT			1693
Sbjct 740	GCTTCCCCGCGACAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTGTTAAGTT			799
Query 1694	TTCATAACATCTGCTATCGAGTAAACTGAAGAGTGGCTTCTACCTTGTGAAAACAG			1753
Sbjct 800	TTCATAACATCTGCTATCGAGTAAACTGAAGAGTGGCTTCTACCTTGTGAAAACAG			859
Query 1754	TTGAGAAAAGAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGCCATCC			1813
Sbjct 860	TTGAGAAAAGAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGCCATCC			919
Query 1814	TGGGACCCACAGGTGGAGGCAAACTTCGTTATAGATGCTTAGCTGCAAGGAAAGATC			1873
Sbjct 920	TGGGACCCACAGGTGGAGGCAAACTTCGTTATAGATGCTTAGCTGCAAGGAAAGATC			979
Query 1874	CAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCAGCCTGCCAACTTCAAAT			1933
Sbjct 980	CAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCAGCCTGCCAACTTCAAAT			1039
Query 1934	GTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGAAA			1993
Sbjct 1040	GTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGAAA			1099
Query 1994	ACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGaaaaaaCG			2053
Sbjct 1100	ACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAAAAAACG			1159
Query 2054	AACGGATTAACAGGGTCATTCAAGAGTAGGCTGGATAAAGTGGCAGACTCCAAGGTTG			2113
Sbjct 1160	AACGGATTAACAGGGTCATTCAAGAGTAGGCTGGATAAAGTGGCAGACTCCAAGGTTG			1219
Query 2114	GAACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAAGGACTAGTATAGGAATGG			2173
Sbjct 1220	GAACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAAGGACTAGTATAGGAATGG			1279
Query 2174	AGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTTAG-ACTCA			2232
Sbjct 1280	AGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAACTACAACCTGGCTTATAACTCA			1339
Query 2233	AGCACAGCAAAATGCTGTCTTTTGTCTCTGAAAAGGATGCTAAGCAGGGACGAAACAATC			2292
Sbjct 1340	AGCNNAGCAAAATGCTGTCTTTTGTCTCTGAAAAGGATGCTAANCAGGGANNAANAATC			1399

## SeqF1

Score	Expect	Identities	Gaps	Strand
2392 bits(1295)	0.0	1358/1391(98%)	11/1391(0%)	Plus/Plus
Query 1869	AGATCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTT			1928
Sbjct 16	AGATCC-AGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTT			74
Query 1929	CAAAATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACCTCTGACGGTGAG			1988
Sbjct 75	CAAAATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACCTCTGACGGTGAG			134
Query 1989	AGAAAACTTACAGTTCACAGCAGCTCTTCGGCTTGCAACAATATGACGAATCATGaaaa			2048
Sbjct 135	AGAAAACTTACAGTTCACAGCAGCTCTTCGGCTTGCAACAATATGACGAATCATGAAAA			194
Query 2049	aaaCGAACGGATTAACAGGGTCATTCAAGAGTTAGGCTCGGATAAAGTGGCAGACTCCAA			2108
Sbjct 195	AAACGAACGGATTAACAGGGTCATTCAAGAGTTAGGCTCGGATAAAGTGGCAGACTCCAA			254
Query 2189	GGTGGAACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAGGACTAGTATAGG			2168
Sbjct 255	GGTGGAACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAGGACTAGTATAGG			314
Query 2169	AATGGAGCTTATCAGTATCCTTCCATCTTGTTCTTGGATGAGCCTACAACGGCTTAGA			2228
Sbjct 315	AATGGAGCTTATCAGTATCCTTCCATCTTGTTCTTGGATGAGCCTACAACGGCTTAGA			374
Query 2229	CTCAAGCACAGCAAAATGCTGTCTTTTGTCTGAAAAGGATGCTAAGCAGGGACGAAAC			2288
Sbjct 375	CTCAAGCACAGCAAAATGCTGTCTTTTGTCTGAAAAGGATGCTAAGCAGGGACGAAAC			434
Query 2289	AATCATCTTCTCCATTTCATCAGCCTCGATATCCATCTTCAAGTTGTTGATAGCCTCAC			2348
Sbjct 435	AATCATCTTCTCCATTTCATCAGCCTCGATATCCATCTTCAAGTTGTTGATAGCCTCAC			494
Query 2349	CTTATTGGCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCCTTGGGATACTT			2408
Sbjct 495	CTTATTGGCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCCTTGGGATACTT			554
Query 2489	TGAATCAGCTGGTTATCACTGTGAGGCTATAATAACCTGCAGACTTCTTCTGGACAT			2468
Sbjct 555	TGAATCAGCTGGTTATCACTGTGAGGCTATAATAACCTGCAGACTTCTTCTGGACAT			614
Query 2469	CATTAATGGAGATTCACCTGCTGTGGCATTAAACAGAGAAGAAGACTTAAAGCCACAGA			2528
Sbjct 615	CATTAATGGAGATTCACCTGCTGTGGCATTAAACAGAGAAGAAGACTTAAAGCCACAGA			674
Query 2529	GATCATAGAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATAGCGGAGATTTATGT			2588
Sbjct 675	GATCATAGAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATAGCGGAGATTTATGT			734
Query 2589	CAACTCTCTCTTACAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAA			2648
Sbjct 735	CAACTCTCTCTTACAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAA			794
Query 2649	GAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTATCAACT			2708
Sbjct 795	GAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTATCAACT			854
Query 2789	CAGATGGGTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCAGGCTCTATAGC			2768
Sbjct 855	CAGATGGGTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCAGGCTCTATAGC			914
Query 2769	TCAGATCATGTGACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAA			2828
Sbjct 915	TCAGATCATGTGACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAA			974
Query 2829	AAATGATTC TACTGGAATCCAGAACAGAGCTGGGGTTCTTCTTCTTCTGACGACCAACCA			2888
Sbjct 975	AAATGATTC TACTGGAATCCAGAACAGAGCTGGGGTTCTTCTTCTTCTGACGACCAACCA			1034
Query 2889	GTGTTTCAGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTCTTCATACA			2948
Sbjct 1035	GTGTTTCAGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTCTTCATACA			1094
Query 2949	TGAATACATCAGCGGATACTACAGAGTGTATCTTATTTCTTGGAAAACCTGTTATCTGA			3008
Sbjct 1095	TGAATACATCAGCGGATACTACAGAGTGTATCTTATTTCTTGGAAAACCTGTTATCTGA			1154
Query 3089	TTTATTACCCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTACTTCATGTT			3068
Sbjct 1155	TTTATTACCCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTACTTCATGTT			1214
Query 3069	AGGATTGAAGCCAAAGGCAGATGCCCTTCTCGTATGATGTTACCCCTATGATGGT-GG			3127
Sbjct 1215	AGGATTGAAGCCAAAGGCAGATGCCCTTCTCGTATGATGTTACCCCTATGAAAGGGNGG			1274
Query 3128	CTTATTCAGCCAGTTCATGGCACTGG-CCATAGCAGCAGGTGAGT-GTGGTTCTGT			3185
Sbjct 1275	CTTATTCAGCCAGTTCATGGCACTGGNCCATAGCANAGGTGAGANNNGTGGTTCTGG			1334
Query 3186	AGCAACACTTCTC-ATGACCATCTG-TTTTGTGTTT-ATG-ATGA-TTTTTTCAGG-TCT			3239
Sbjct 1335	ANCAANCTTNNNATGACCATCTGGTTTGGGGTTNAANNAAGAAATTTTTTCAGGGTCG			1394
Query 3240	GTTGG-TCAAT 3249			
Sbjct 1395	GTTGGGTCAAT 1405			

## SeqF2

Score	Expect	Identities	Gaps	Strand
2466 bits(1335)	0.0	1372/1397(98%)	4/1397(0%)	Plus/Plus
Query 2325	CTTCAAGTTGTTGATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGCC			2384
Sbjct 15	CTTC-AGTTGTTGATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGCC			73
Query 2385	TGCTCAGGAGGCCCTTGGGATACTTTGAATCAGCTGGTTACTCTGTGAGGCCATAATAA			2444
Sbjct 74	TGCTCAGGAGGCCCTTGGGATACTTTGAATCAGCTGGTTACTCTGTGAGGCCATAATAA			133
Query 2445	CCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATTCCACTGCTGTGGCATTAAACAG			2504
Sbjct 134	CCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATTCCACTGCTGTGGCATTAAACAG			193
Query 2505	AGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCAT			2564
Sbjct 194	AGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCAT			253
Query 2565	AGAAAAATTAGCGGAGATTTATGTCAACTCCTCCTTCTACAAAGAGACAAAAGCTGAATT			2624
Sbjct 254	AGAAAAATTAGCGGAGATTTATGTCAACTCCTCCTTCTACAAAGAGACAAAAGCTGAATT			313
Query 2625	ACATCAACTTTCGGGGGTGAGAAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTA			2684
Sbjct 314	ACATCAACTTTCGGGGGTGAGAAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCTA			373
Query 2685	CACCACCTCCTTCTGTATCAACTCAGATGGGTTTCTAAGCGTTCAATCAAAAACTTGCT			2744
Sbjct 374	CACCACCTCCTTCTGTATCAACTCAGATGGGTTTCTAAGCGTTCAATCAAAAACTTGCT			433
Query 2745	GGGTAATCCCGGCCCTCTATAGCTCAGATCATGTGACAGTCTGACTGGGACTGGTTAT			2804
Sbjct 434	GGGTAATCCCGGCCCTCTATAGCTCAGATCATGTGACAGTCTGACTGGGACTGGTTAT			493
Query 2805	AGGTGCCATTTACTTTGGGCTAAAAAATGATTTACTGGAATCCAGAACAGAGCTGGGGT			2864
Sbjct 494	AGGTGCCATTTACTTTGGGCTAAAAAATGATTTACTGGAATCCAGAACAGAGCTGGGGT			553
Query 2865	TCTCTTCTCTGACGACCAACAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTGT			2924
Sbjct 554	TCTCTTCTCTGACGACCAACAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTGT			613
Query 2925	GGTAGAAGAAGACTCTTCATACATGAATACATCAGCGGATACTACAGAGTGTATCTTA			2984
Sbjct 614	GGTAGAAGAAGACTCTTCATACATGAATACATCAGCGGATACTACAGAGTGTATCTTA			673
Query 2985	TTTCTTGGAAAACTGTTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATATT			3044
Sbjct 674	TTTCTTGGAAAACTGTTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATATT			733
Query 3045	TACCTGTATAGTGTACTTTCATGTTAGGATTGAAGCCAAAGGCAGATGCCTTCTCGTTAT			3104
Sbjct 734	TACCTGTATAGTGTACTTTCATGTTAGGATTGAAGCCAAAGGCAGATGCCTTCTCGTTAT			793
Query 3105	GATGTTTACCCTTATGATGGTGGCTTATTACGCCAGTTCATGGCACTGGCCATAGCAGC			3164
Sbjct 794	GATGTTTACCCTTATGATGGTGGCTTATTACGCCAGTTCATGGCACTGGCCATAGCAGC			853
Query 3165	AGGTGAGAGTGGTTCCTGTAGCAACTTCTCATGACCATCTGTTTGTGTTTATGAT			3224
Sbjct 854	AGGTGAGAGTGGTTCCTGTAGCAACTTCTCATGACCATCTGTTTGTGTTTATGAT			913
Query 3225	GATTTTTTCAGGCTGTTGGTCAATCTCACAACATTGCATCTTGGCTGTATGGCTTCA			3284
Sbjct 914	GATTTTTTCAGGCTGTTGGTCAATCTCACAACATTGCATCTTGGCTGTATGGCTTCA			973
Query 3285	GTACTTCAGCATTCCACGATATGGATTACGGCTTTCAGCATAATGAATTTTGGGACA			3344
Sbjct 974	GTACTTCAGCATTCCACGATATGGATTACGGCTTTCAGCATAATGAATTTTGGGACA			1033
Query 3345	AAACTTCTGCCAGGACTCAATGCAACAGGAACAATCCTTGTAACTATGCAACATGTAC			3404
Sbjct 1034	AAACTTCTGCCAGGACTCAATGCAACAGGAACAATCCTTGTAACTATGCAACATGTAC			1093
Query 3405	TGGCGAAGAATATTTGGTAAAGCAGGGCATCGATCTCTCACCTGGGGCTTGTGGAAGAA			3464
Sbjct 1094	TGGCGAAGAATATTTGGTAAAGCAGGGCATCGATCTCTCACCTGGGGCTTGTGGAAGAA			1153
Query 3465	TCACGTGGCCTTGGCTTGTATGATTGTTATTTTCTCACAAATGGCTACCTGAAATGTT			3524
Sbjct 1154	TCACGTGGCCTTGGCTTGGATGATTGTTATTTTCTCACAAATGGCTACCTGAAATGTT			1213
Query 3525	ATTTCTTAAAAAATATTTCTTAAATTGGATTCTAGAGGGCCCGTTTAAACCCGCTGATCAG			3584
Sbjct 1214	ATTTCTTAAAAAATATTTCTTAAATTGGATTCTAGAGGGCCCGTTTAAACCCGCTGATCAG			1273
Query 3585	CCTCGACTGTGCCCTTCTAGTTGCCAGCCATCTGTT-GTTTGGCCCTCCCCCGTCCCTCC			3643
Sbjct 1274	CCTCGACTGTGCCCTTCTAGTTGCCAGCCATCTGTT-GTTTGGCCCTCCCCCGTCCCTCC			1333
Query 3644	TTGACCCTGGAAAGGTGCCACTCCCAGTGT-CCTTTCTAATAAAAATGAGGAAATTCATC			3702
Sbjct 1334	TTGACCCTGGAAAGGTGCCACTCCCAGTGT-CCTTTCTAATAAAAATGAGGAAATTCATC			1392
Query 3703	GCATTGTCTGAGTAGGT 3719			
Sbjct 1393	GCATTGTCTGANTAGGT 1409			

## Seq482

Score	Expect	Identities	Gaps	Strand
2287 bits(1238)	0.0	1303/1337(97%)	17/1337(1%)	Plus/Plus
Query 2960	GCGGATACTACAGAGTGCATCTTATTTCCTTGGAAAACGTTATCTGATTATTACCCA			3019
Sbjct 12	GCGGNNACTACAGAGTGCATCTTATTTCCTTGGAAAACGTTATCTGATTATTACCCA			71
Query 3020	TGAGGATGTACCAAGTATTATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			3079
Sbjct 72	TGAGGATGTACCAAGTATTATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			131
Query 3080	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTCAGCCA			3139
Sbjct 132	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTCAGCCA			191
Query 3140	GTTCCATGGCACTGGCCATAGCAGCAGGTCAGAGTGTGGTTTCTGTAGCAACTTCTCA			3199
Sbjct 192	GTTCCATGGCACTGGCCATAGCAGCAGGTCAGAGTGTGGTTTCTGTAGCAACTTCTCA			251
Query 3200	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGTCGTGGTCAATCTCACAACCA			3259
Sbjct 252	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGTCGTGGTCAATCTCACAACCA			311
Query 3260	TTGCATCTTGGCTGTCATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCTT			3319
Sbjct 312	TTGCATCTTGGCTGTCATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCTT			371
Query 3320	TGCAGCATAATGAATTTTGGGACAAAACCTCTGCCAGGACTCAATGCAACAGGAAACA			3379
Sbjct 372	TGCAGCATAATGAATTTTGGGACAAAACCTCTGCCAGGACTCAATGCAACAGGAAACA			431
Query 3380	ATCCTTGTAACTATGCAACATGACTGGCGAAGAAATTTGGTAAAGCAGGGCATCGATC			3439
Sbjct 432	ATCCTTGTAACTATGCAACATGACTGGCGAAGAAATTTGGTAAAGCAGGGCATCGATC			491
Query 3440	TCTCACCTGGGGCTTGTGGAAAGAACACGTGGCCTTGGCTTGTATGATTGTTATTTCC			3499
Sbjct 492	TCTCACCTGGGGCTTGTGGAAAGAACACGTGGCCTTGGCTTGTATGATTGTTATTTCC			551
Query 3500	TCACAATGGCTACCTGAAATTTGTTATTTCTTAAAAAATATTCTTAAATGGATTCTAGA			3559
Sbjct 552	TCACAATGGCTACCTGAAATTTGTTATTTCTTAAAAAATATTCTTAAATGGATTCTAGA			611
Query 3560	GGGCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTT			3619
Sbjct 612	GGGCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTT			671
Query 3620	GTTTGGCCCTCCCGCTGCTTCTTTCAGCCCTGGAAAGGTGCCACTCCCACTGTCTTTCC			3679
Sbjct 672	GTTTGGCCCTCCCGCTGCTTCTTTCAGCCCTGGAAAGGTGCCACTCCCACTGTCTTTCC			731
Query 3680	TAATAAAATGAGGAAATGTCATCGCATGTCTGAGTAGGTGTCATTCTATTCTGGGGGT			3739
Sbjct 732	TAATAAAATGAGGAAATGTCATCGCATGTCTGAGTAGGTGTCATTCTATTCTGGGGGT			791
Query 3740	GGGGTGGGGCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGAT			3799
Sbjct 792	GGGGTGGGGCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGAT			851
Query 3800	GCGGTGGGCTCTATGGCTTCTGAGCGGAAAGAACCAGCTGGGGCTCTAGGGGGATCCC			3859
Sbjct 852	GCGGTGGGCTCTATGGCTTCTGAGCGGAAAGAACCAGCTGGGGCTCTAGGGGGATCCC			911
Query 3860	CACGCGCCCTGTAGCGGCATTAAGCGCGCGGGTGTGGTGGTTACGCGCAGCGTGACC			3919
Sbjct 912	CACGCGCCCTGTAGCGGCATTAAGCGCGCGGGTGTGGTGGTTACGCGCAGCGTGACC			971
Query 3920	GCTACACTTGCAGCGCCCTAGCGCCCGCTCTTTCGCTTCTTCCCTTCTTCTCGCC			3979
Sbjct 972	GCTACACTTGCAGCGCCCTAGCGCCCGCTCTTTCGCTTCTTCCCTTCTTCTCGCC			1031
Query 3980	ACGTTTCGCGGCTTTCCCGTCAAGCTCTAAATCGGGGCATCCCTTAGGGTCCGATTT			4039
Sbjct 1032	ACGTTTCGCGGCTTTCCCGTCAAGCTCTAAATCGGGGCATCCCTTAGGGTCCGATTT			1091
Query 4040	AGTGCTTACGGCACCTCGACCCCAAAAAAATTGATTAGGGTATGGTTACGTAGTGGG			4099
Sbjct 1092	AGTGCTTACGGCACCTCGACCCCAAAAAAATTGATTAGGGTATGGTTACGTAGTGGG			1151
Query 4100	CCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTTGG-AGTCCACGTTCTTAAATAG			4158
Sbjct 1152	CCATCGCCCTGATANACGGTTTTTCGCCCTTTGACGTTGGAAAGTCCACGTTCTTAAATAG			1211
Query 4159	TGG-ACTCTTGTTCACAACTGGAA-CAACACT-CAACCCATCT-CCG-TCATTCTTT			4213
Sbjct 1212	TGGAACCTCTGTTCACAACTGGAAACAANNNTTCAACCCATCCNCGGNTCTATTCTTT			1271
Query 4214	-GATTTATAA-GGGA-TTTTGGGGA-TTTC-GGCTATTGG-TTAAAAA-TGAGC-TG-			4264
Sbjct 1272	TGATTTNNAAGGGAAATTTGCCNAATTTCCGGCTATTGGTTAAAAAATGNNCTGG			1331
Query 4265	ATTT-AACAATAA-TTT		4279	
Sbjct 1332	ATTTTAAACAAAAATTT		1348	

## 6.2.5 M549E

## T7 promoter (F)

Score	Expect	Identities	Gaps	Strand
2499 bits(1353)	0.0	1382/1400(99%)	4/1400(0%)	Plus/Plus
Query 989	TTAAGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGAAG			968
Sbjct 14	TTAAGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGAAG			73
Query 969	CGGTGGAGGCTCAGGAGGCAGCGCATGGTCCCACCCCAAGTTGAAAAGCTTGCCACCAT			1028
Sbjct 74	CGGTGGAGGCTCAGGAGGCAGCGCATGGTCCCACCCCAAGTTGAAAAGCTTGCCACCAT			133
Query 1029	GGACAAAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCTGGGCAAGCTGGAACT			1088
Sbjct 134	GGACAAAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCTGGGCAAGCTGGAACT			193
Query 1089	GTCGGGTGCGAACAGGGCCTGCACGAGATCAAGCTGCTGGGCAAGGAACTCTGCCGC			1148
Sbjct 194	GTCGGGTGCGAACAGGGCCTGCACGAGATCAAGCTGCTGGGCAAGGAACTCTGCCGC			253
Query 1149	CGACGCCGTGGAAAGTGCTGCCCCAGCCGCGTCTGGGCGGACAGAGCCACTGATGCA			1208
Sbjct 254	CGACGCCGTGGAAAGTGCTGCCCCAGCCGCGTCTGGGCGGACAGAGCCACTGATGCA			313
Query 1209	GGCCACCGCTGGCTCAACGCCACTTTCACCAGCCTGAGGCCATCGAGGAGTTCCTGT			1268
Sbjct 314	GGCCACCGCTGGCTCAACGCCACTTTCACCAGCCTGAGGCCATCGAGGAGTTCCTGT			373
Query 1269	GCCAGCCCTGCACCACCCAGTGTTCAGCAGGAGAGCTTACCAGCCAGGTGCTGTGGAA			1328
Sbjct 374	GCCAGCCCTGCACCACCCAGTGTTCAGCAGGAGAGCTTACCAGCCAGGTGCTGTGGAA			433
Query 1329	ACTGCTGAAAGTGGTGAAGTTCGGAGAGGTGATCAGCTACCAGCAGCTGGCCGCCCTGGC			1388
Sbjct 434	ACTGCTGAAAGTGGTGAAGTTCGGAGAGGTGATCAGCTACCAGCAGCTGGCCGCCCTGGC			493
Query 1389	CGGCAATCCCGCCGCGCACCGCCCGTGAAAACCGCCCTGAGCGGAAATCCCGTGCCCAT			1448
Sbjct 494	CGGCAATCCCGCCGCGCACCGCCCGTGAAAACCGCCCTGAGCGGAAATCCCGTGCCCAT			553
Query 1449	TCTGATCCCCGCCACCGGGTGGTGTCTAGCTCTGGCGCCGTGGGGGGCTACGAGGGCGG			1508
Sbjct 554	TCTGATCCCCGCCACCGGGTGGTGTCTAGCTCTGGCGCCGTGGGGGGCTACGAGGGCGG			613
Query 1509	GCTCGCCGTGAAAGAGTGGCTGCTGGCCACGAGGGCCACAGACTGGGCAAGCTGGGCT			1568
Sbjct 614	GCTCGCCGTGAAAGAGTGGCTGCTGGCCACGAGGGCCACAGACTGGGCAAGCTGGGCT			673
Query 1569	GGGCGAATTTCATGCTTCCAGTAAATGTCGAAGTTTTTATCCAGTGTCAAGGAAACAC			1628
Sbjct 674	GGGCGAATTTCATGCTTCCAGTAAATGTCGAAGTTTTTATCCAGTGTCAAGGAAACAC			733
Query 1629	CAATGGCTTCCCGCGACAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTGTT			1688
Sbjct 734	CAATGGCTTCCCGCGACAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTGTT			793
Query 1689	AAGTTTTTCATAACATCTGCTATCGAGTAAAACGAAGAGTGGCTTCTACCTTGTGCGAA			1748
Sbjct 794	AAGTTTTTCATAACATCTGCTATCGAGTAAAACGAAGAGTGGCTTCTACCTTGTGCGAA			853
Query 1749	ACCAGTTGAGAAAGAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGC			1808
Sbjct 854	ACCAGTTGAGAAAGAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGC			913
Query 1809	CATCCTGGGACCCACAGGTGGAGGCAAACTTCGTTATTAGATGCTTACTGCAAGGAA			1868
Sbjct 914	CATCCTGGGACCCACAGGTGGAGGCAAACTTCGTTATTAGATGCTTACTGCAAGGAA			973
Query 1869	AGATCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCAGACTGCCAACTT			1928
Sbjct 974	AGATCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCAGACTGCCAACTT			1033
Query 1929	CAAAATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGA			1988
Sbjct 1034	CAAAATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGA			1093
Query 1989	AGAAAACTTACAGTCTCAGCAGCTCTCGGCTTGCAACAATATGACGAATCATGaaa			2048
Sbjct 1094	AGAAAACTTACAGTCTCAGCAGCTCTCGGCTTGCAACAATATGACGAATCATGAAAA			1153
Query 2049	aaaCGAACGGATTAACAGGGTCATTCAAGAGTTAGGCTGGATAAAGTGGCAGACTCCAA			2108
Sbjct 1154	AAAAGAACGGATTAACAGGGTCATTCAAGAGTTAGGCTGGATAAAGTGGCAGACTCCAA			1213
Query 2109	GGTTGGAACCTCAGTTTATCCGTTGGTGTCTGGAGGAGAAAGAAAAGGACTAGTATAGG			2168
Sbjct 1214	GGTTGGAACCTCAGTTTATCCGTTGGTGTCTGGAGGAGAAAGAAAAGGACTAGTATAGG			1273
Query 2169	AATGGAGCTTATCAGTATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTTAG-			2227
Sbjct 1274	AATGGAGCTTATCAGTATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTTAA			1333
Query 2228	ACTCAAGCACAGCAAAAT-GCTGTCTTTGCTCTGAAAAGGATGCTAAGCAGGGACGA			2286
Sbjct 1334	ACTCAAGCACAGCAAAANGGCTGTCTTTNGNCCCTGAAAAGGATGCTAACCNGGGACAA			1393
Query 2287	AC-AATCATC-TTCTCCATT 2304			
Sbjct 1394	ANNAATCANCNTTNTCCATT 1413			

## SeqF1

Score	Expect	Identities	Gaps	Strand
2348 bits(1271)	0.0	1303/1320(99%)	7/1320(0%)	Plus/Plus
Query 1872	TCCAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGC CAACTTCAA			1931
Sbjct 19	TCCNAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGC CAACTTCAA			78
Query 1932	ATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGA			1991
Sbjct 79	ATGTAATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGA			138
Query 1992	AAACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGaaaaaaaa			2051
Sbjct 139	AAACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAAAAAAAA			198
Query 2052	CGAACGGATTAAACAGGGTCATTCAAGAGTTAGGTCTGGATAAAGTGGCAGACTCCAAGGT			2111
Sbjct 199	CGAACGGATTAAACAGGGTCATTCAAGAGTTAGGTCTGGATAAAGTGGCAGACTCCAAGGT			258
Query 2112	TGGAACCTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAGGACTAGTATAGGAAT			2171
Sbjct 259	TGGAACCTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAGGACTAGTATAGGAAT			318
Query 2172	GGAGCTTATCACTGATCCTTCCATCTTGTTCTGGATGAGCCTACAACCTGGCTTAGACTC			2231
Sbjct 319	GGAGCTTATCACTGATCCTTCCATCTTGTTCTGGATGAGCCTACAACCTGGCTTAGACTC			378
Query 2232	AAGCACAGCAAAATGCTGTCTTTGCTCCTGAAAAGGATGTCTAAGCAGGGACGAACAAT			2291
Sbjct 379	AAGCACAGCAAAATGCTGTCTTTGCTCCTGAAAAGGATGTCTAAGCAGGGACGAACAAT			438
Query 2292	CATCTTCCATTATCAGCCTCGATATCCATCTTCAAGTTGTTTATAGCCTCACCTT			2351
Sbjct 439	CATCTTCCATTATCAGCCTCGATATCCATCTTCAAGTTGTTTATAGCCTCACCTT			498
Query 2352	ATTGGCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCCCTTGGGATACCTTGA			2411
Sbjct 499	ATTGGCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCCCTTGGGATACCTTGA			558
Query 2412	ATCAGCTGGTTATCACTGTGAGGCCATAATAACCTGCAGACTTCTTCTGGACATCAT			2471
Sbjct 559	ATCAGCTGGTTATCACTGTGAGGCCATAATAACCTGCAGACTTCTTCTGGACATCAT			618
Query 2472	TAATGGAGATTCACACTGCTGTGGCATTAAACAGAGAAGAAGACTTTAAAGCCACAGAGAT			2531
Sbjct 619	TAATGGAGATTCACACTGCTGTGGCATTAAACAGAGAAGAAGACTTTAAAGCCACAGAGAT			678
Query 2532	CATAGAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTTATGTCAA			2591
Sbjct 679	CATAGAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTTATGTCAA			738
Query 2592	CTCCTCCTTCTACAAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGTGAGAAAGAA			2651
Sbjct 739	CTCCTCCTTCTACAAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGTGAGAAAGAA			798
Query 2652	GAAGAAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTCACTCAACTCAG			2711
Sbjct 799	GAAGAAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTCACTCAACTCAG			858
Query 2712	ATGGGTTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCCAGGCCCTATAGCTCA			2771
Sbjct 859	ATGGGTTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCCAGGCCCTATAGCTCA			918
Query 2772	GATCATTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAAA			2831
Sbjct 919	GATCATTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAAA			978
Query 2832	TGATTCCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTTCTGACGACCAACCAGTG			2891
Sbjct 979	TGATTCCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTTCTGACGACCAACCAGTG			1038
Query 2892	TTTCAGCAGTGTTCAGCCGTGGAACCTTTTGTGGTAGAGAAGAAGCTCTTCATACATGA			2951
Sbjct 1039	TTTCAGCAGTGTTCAGCCGTGGAACCTTTTGTGGTAGAGAAGAAGCTCTTCATACATGA			1098
Query 2952	ATACATCAGCGGATACTACAGAGTGTATCTTATTTCTTGGAAAACCTGTTATCTGATTT			3011
Sbjct 1099	ATACATCAGCGGATACTACAGAGTGTATCTTATTTCTTGGAAAACCTGTTATCTGATTT			1158
Query 3012	ATTACCCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGG			3071
Sbjct 1159	ATTACCCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGG			1218
Query 3072	ATTGAAGCCAAAGGCAGATGCCTTCTCGTTATGAT-GTTTACCTTATGAT-GGTGGCT			3129
Sbjct 1219	ATTGAAGCCAAAGGCAGATGCCTTCTCGTTATGAAAGTTTACCCTTATGAAGGGTGGCT			1278
Query 3130	TATTCAGCCAGTTCATGGC-ACTGG-CCATAGCAGC-AGGTCAGAGT--GTGGTTTCTG			3184
Sbjct 1279	TATTCAGCCAGTTCATGGNACTGGCCATANCNNCCAGGTCAGAAANNNGTGGTTTCTG			1338

## SeqF2

Score	Expect	Identities	Gaps	Strand
2348 bits(1271)	0.0	1327/1363(97%)	7/1363(0%)	Plus/Plus
Query 2324	TCTTCAAGTTGTTTGTATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGC			2383
Sbjct 14	TCTTC-AGNTGTTTGTATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGC			72
Query 2384	CTGCTCAGGAGGCTTGGGATACTTTGAATCAGCTGGTTATCACTGTGAGGCTATAATA			2443
Sbjct 73	CTGCTCAGGAGGCTTGGGATACTTTGAATCAGCTGGTTATCACTGTGAGGCTATAATA			132
Query 2444	ACCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATCCACTGCTGTGGCATTAAACA			2503
Sbjct 133	ACCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATCCACTGCTGTGGCATTAAACA			192
Query 2504	GAGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCA			2563
Sbjct 193	GAGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCA			252
Query 2564	TAGAAAAATAGCGGAGATTTATGTCAACTCCTCTTACAAGAGACAAAAGCTGAAT			2623
Sbjct 253	TAGAAAAATAGCGGAGATTTATGTCAACTCCTCTTACAAGAGACAAAAGCTGAAT			312
Query 2624	TACATCAACTTTCCGGGGTGAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCT			2683
Sbjct 313	TACATCAACTTTCCGGGGTGAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCT			372
Query 2684	ACACCACCTCCTTCTGTCACTCAACTCAGATGGGTTTCTAAGGTTTATTCAAAAACTTGC			2743
Sbjct 373	ACACCACCTCCTTCTGTCACTCAACTCAGATGGGTTTCTAAGGTTTATTCAAAAACTTGC			432
Query 2744	TGGGTAATCCCAGGCTCTATAGCTCAGATCATTGTCCAGTCTGACTGGGACTGGTTA			2803
Sbjct 433	TGGGTAATCCCAGGCTCTATAGCTCAGATCATTGTCCAGTCTGACTGGGACTGGTTA			492
Query 2804	TAGGTGCCATTTACTTTGGGCTAAAAAATGATTCTACTGGAATCCAGAACAGAGCTGGG			2863
Sbjct 493	TAGGTGCCATTTACTTTGGGCTAAAAAATGATTCTACTGGAATCCAGAACAGAGCTGGG			552
Query 2864	TTCTCTTCTTCTGACGACCAACAGTGTTCAGCAGTGTTCAGCCGTGGAACTCTTTG			2923
Sbjct 553	TTCTCTTCTTCTGACGACCAACAGTGTTCAGCAGTGTTCAGCCGTGGAACTCTTTG			612
Query 2924	TGGTAGAGAAGAAGCTCTTCATACATGAATACATCAGCGGATACTACAGAGTGTATCTT			2983
Sbjct 613	TGGTAGAGAAGAAGCTCTTCATACATGAATACATCAGCGGATACTACAGAGTGTATCTT			672
Query 2984	ATTTCCCTGGAAAACGTTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATAT			3043
Sbjct 673	ATTTCCCTGGAAAACGTTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATAT			732
Query 3044	TTACCTGTATAGTGTACTTTCATGTTAGGATTGAAGCAAAGGCAGATGCCTTCTTCGTTA			3103
Sbjct 733	TTACCTGTATAGTGTACTTTCATGTTAGGATTGAAGCAAAGGCAGATGCCTTCTTCGTTA			792
Query 3104	TGATGTTTACCCTTATGATGGTGGCTTATTAGCCAGTTCATGGCACTGGCCATAGCAG			3163
Sbjct 793	TGATGTTTACCCTTATGATGGTGGCTTATTAGCCAGTTCATGGCACTGGCCATAGCAG			852
Query 3164	CAGGTGAGAGTGTGGTTCTGTAGCAACACTTCTCATGACCATCTGTTTGTGTTTATGA			3223
Sbjct 853	CAGGTGAGAGTGTGGTTCTGTAGCAACACTTCTCATGACCATCTGTTTGTGTTTATGA			912
Query 3224	TGATTTTTTTCAGGCTGTGGTCAATCTCACAACCATTGCATCTTGGCTGTATGGCTTC			3283
Sbjct 913	TGATTTTTTTCAGGCTGTGGTCAATCTCACAACCATTGCATCTTGGCTGTATGGCTTC			972
Query 3284	AGTACTTCAGCATTCCACGATATGGATTTACGGCTTTCAGCATAAATGAATTTTGGGAC			3343
Sbjct 973	AGTACTTCAGCATTCCACGATATGGATTTACGGCTTTCAGCATAAATGAATTTTGGGAC			1032
Query 3344	AAAACCTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGAACATGCAACATGTA			3403
Sbjct 1033	AAAACCTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGAACATGCAACATGTA			1092
Query 3404	CTGGCGAAGAAATTTGGTAAAGCAGGGCATCGATCTCTACCCTGGGGCTTGTGGAAGA			3463
Sbjct 1093	CTGGCGAAGAAATTTGGTAAAGCAGGGCATCGATCTCTACCCTGGGGCTTGTGGAANA			1152
Query 3464	ATCACGTGGCCTTGGCTTGTATGATTGTTATTTTCCTCACAAATGGCTACCTGAAATGT			3523
Sbjct 1153	ATCACGTGGCCTTGGCTTGTATGATTGTTATTTTCCTCACAAATGGCTACCTGAAATGT			1212
Query 3524	TATTTCTTAAAAAATATTCTTAAATGGATTCTAGAGGGCCCGTTAAACCCGCTGATCA			3583
Sbjct 1212	TATTTCTTAAAAAANNITCTTAAATGGATTCTAGAGGGCCCGTTAAACCCGCTGATCA			1272
Query 3584	GCCTCGACTGTGCTTCTAGTTGCCAGCCATCTGTTGTTGCCCTCCCCCGTGCCTTCC			3643
Sbjct 1272	GCCTCAANNNGGCTTNA-TTCCNGCCANCTGTTGTTNGCCNNNCCCC-GTG-CTTCC			1328
Query 3644	TTGACCTTGAAGGTGCCACTCCACTGTCCTTTCTAATAAA		3686	
Sbjct 1329	TTGACCNNGGAGGGGCC-CNNCCNGGTCCTTCC-AAATAAA		1369	

## Seq482

Score	Expect	Identities	Gaps	Strand
2375 bits(1286)	0.0	1309/1323(99%)	0/1323(0%)	Plus/Plus
Query 2960	GCGGATACTACAGAGTGCATCTTATTTCCTTGGAAAACTGTTATCTGATTATTACCCA			3019
Sbjct 13	GCGGANACTACAGAGTGCATCTTATTTCCTTGGAAAACTGTTATCTGATTATTACCCA			72
Query 3020	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			3079
Sbjct 73	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			132
Query 3080	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTCAGCCA			3139
Sbjct 133	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTCAGCCA			192
Query 3140	GTTCCATGGCACTGGCCATAGCAGCAGGTGAGAGTGTGGTTTCTGTAGCAACACTTCTCA			3199
Sbjct 193	GTTCCATGGCACTGGCCATAGCAGCAGGTGAGAGTGTGGTTTCTGTAGCAACACTTCTCA			252
Query 3200	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGCTGTTGGTCAATCTCACAACCA			3259
Sbjct 253	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGCTGTTGGTCAATCTCACAACCA			312
Query 3260	TTGCATCTTGGCTGTGCATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCTT			3319
Sbjct 313	TTGCATCTTGGCTGTGCATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCTT			372
Query 3320	TGCAGCATAATGAATTTTTGGGACAAAACCTTCTGCCAGGACTCAATGCAACAGGAAACA			3379
Sbjct 373	TGCAGCATAATGAATTTTTGGGACAAAACCTTCTGCCAGGACTCAATGCAACAGGAAACA			432
Query 3380	ATCCTTGTAACTATGCAACATGACTGGCGAAGAATATTTGGTAAAGCAGGGCATCGATC			3439
Sbjct 433	ATCCTTGTAACTATGCAACATGACTGGCGAAGAATATTTGGTAAAGCAGGGCATCGATC			492
Query 3440	TCTCACCCCTGGGCTTGTGGAAGAATCACGTGGCCTTGGCTTGTATGATTGTTATTTTCC			3499
Sbjct 493	TCTCACCCCTGGGCTTGTGGAAGAATCACGTGGCCTTGGCTTGTATGATTGTTATTTTCC			552
Query 3500	TCACAAATGCCATACC TGAATTTGTTATTTCTTAAAAAATATTCTTAAATGGATTCTAGA			3559
Sbjct 553	TCACAAATGCCATACC TGAATTTGTTATTTCTTAAAAAATATTCTTAAATGGATTCTAGA			612
Query 3560	GGGCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTT			3619
Sbjct 613	GGGCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTT			672
Query 3620	GTTTGCCCCCTCCCCGTGCCTTCTTGCCTTGGAAAGGTGCCACTCCCCTGTCTTTTCC			3679
Sbjct 673	GTTTGCCCCCTCCCCGTGCCTTCTTGCCTTGGAAAGGTGCCACTCCCCTGTCTTTTCC			732
Query 3680	TAATAAAATGAGGAAATTCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTgggggt			3739
Sbjct 733	TAATAAAATGAGGAAATTCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTgggggt			792
Query 3740	ggggtgggCAGGACAGCAAGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGAT			3799
Sbjct 793	ggggtgggCAGGACAGCAAGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGAT			852
Query 3800	GCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTAGGGGGTATCCC			3859
Sbjct 853	GCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTAGGGGGTATCCC			912
Query 3860	CACGCGCCCTGTAGCGGCATTAAGCGCGGGCGGTGTGGTGGTTACGCGCAGCGTGACC			3919
Sbjct 913	CACGCGCCCTGTAGCGGCATTAAGCGCGGGCGGTGTGGTGGTTACGCGCAGCGTGACC			972
Query 3920	GCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCTTCTCGCC			3979
Sbjct 973	GCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCTTCTCGCC			1032
Query 3980	ACGTTTCGCGGCTTTCCCGTCAAGCTCTAAATCGGGGCATCCCTTAGGGTCCGATTT			4039
Sbjct 1033	ACGTTTCGCGGCTTTCCCGTCAAGCTCTAAATCGGGGCATCCCTTAGGGTCCGATTT			1092
Query 4040	AGTGCTTTACGGCACCTCGACCCAAAAAATTTGATTAGGGTGTGGTTACGTTAGTGGG			4099
Sbjct 1093	AGTGCTTTACGGCACCTCGACCCAAAAAATTTGATTAGGGTGTGGTTACGTTAGTGGG			1152
Query 4100	CCATCGCCCTGATAGACGGTTTTTCGCCCTTGGAGTCCACGTTCTTTAATAGT			4159
Sbjct 1153	CCATCGCCCTGATAGACGGTTTTTCGCCCTTGGAGTCCACGTTCTTTAATAGT			1212
Query 4160	GGACTCTTGTCCAAACTGGAACAACACTCAACCTATCTCGGTCTATTCTTTGATTTA			4219
Sbjct 1213	GGACTCTTGTCCAAACTGGAACAACACTCAACCTATCTCGGTCTATTCTTTGATTTN			1272
Query 4220	TAAGGGATTTGGGGATTTTCGGCCTATTGGTTAAAAAATGAGCTGATTTAACAAAAATTT			4279
Sbjct 1273	TAAGGGATTTGGGGATTTTCGGCCTATTGGTTAAAAAATGAGCTGATTTAACAAAAATTT			1332
Query 4280	AAC 4282			
Sbjct 1333	AAC 1335			

## 6.2.6 A397S/V401A/L539A – Mut1

## T7 promoter (F)

Score	Expect	Identities	Gaps	Strand
2519 bits(1364)	0.0	1377/1387(99%)	1/1387(0%)	Plus/Plus
Query 912	AGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTT	CGAGAAGGGAGGTGGAAGCGG	971	
Sbjct 17	AGCTGGTACCGCCACCATGGCATGGAGCCACCCACAGTT	CGAGAAGGGAGGTGGAAGCGG	76	
Query 972	TGGAGGCTCAGGAGGCGAGCGCATGGTCCCACCCCAGTTT	GAAAAGCTTGCCACCATGGA	1031	
Sbjct 77	TGGAGGCTCAGGAGGCGAGCGCATGGTCCCACCCCAGTTT	GAAAAGCTTGCCACCATGGA	136	
Query 1032	CAAAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCT	TGGGCAAGCTGGAACGTGTC	1091	
Sbjct 137	CAAAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCT	TGGGCAAGCTGGAACGTGTC	196	
Query 1092	TGGGTGCGAACAGGGCTTGCACGAGATCAAGCTGCTGGG	CAAAGGAACATCGCCGCCGA	1151	
Sbjct 197	TGGGTGCGAACAGGGCTTGCACGAGATCAAGCTGCTGGG	CAAAGGAACATCGCCGCCGA	256	
Query 1152	CGCCGTGGAAGTGCCTGCCCCAGCCCGCTGCTGGGCGG	ACCAGAGCCACTGATGCAAGC	1211	
Sbjct 257	CGCCGTGGAAGTGCCTGCCCCAGCCCGCTGCTGGGCGG	ACCAGAGCCACTGATGCAAGC	316	
Query 1212	CACCGCTGGCTCAACGCCACTTTACCAGCCTGAGGCC	CATCGAGGAGTTCCCTGTGCC	1271	
Sbjct 317	CACCGCTGGCTCAACGCCACTTTACCAGCCTGAGGCC	CATCGAGGAGTTCCCTGTGCC	376	
Query 1272	AGCCCTGCACCACCCAGTGTCCAGCAGGAGAGCTTT	ACCAGCCAGGTGCTGTGAAACT	1331	
Sbjct 377	AGCCCTGCACCACCCAGTGTCCAGCAGGAGAGCTTT	ACCAGCCAGGTGCTGTGAAACT	436	
Query 1332	GCTGAAAGTGGTGAAGTTCGGAGAGGTCAACAGCTACC	AGCAGCTGGCCGCCCTGGCCGG	1391	
Sbjct 437	GCTGAAAGTGGTGAAGTTCGGAGAGGTCAACAGCTACC	AGCAGCTGGCCGCCCTGGCCGG	496	
Query 1392	CAATCCCGCCGCCACCGCCCGTGAACCCGCCCTGAG	CGGAAATCCCCTGCCCATTTCT	1451	
Sbjct 497	CAATCCCGCCGCCACCGCCCGTGAACCCGCCCTGAG	CGGAAATCCCCTGCCCATTTCT	556	
Query 1452	GATCCCCGTCACCGGGTGGTGTCTAGCTTGGCGCCGT	GGGGGGCTACGAGGGCGGGCT	1511	
Sbjct 557	GATCCCCGTCACCGGGTGGTGTCTAGCTTGGCGCCGT	GGGGGGCTACGAGGGCGGGCT	616	
Query 1512	CGCCGTGAAAGAGTGGCTGCTGGCCCACGAGGGCC	CACAGACTGGGCAAGCTGGGCTGGG	1571	
Sbjct 617	CGCCGTGAAAGAGTGGCTGCTGGCCCACGAGGGCC	CACAGACTGGGCAAGCTGGGCTGGG	676	
Query 1572	CGAATTCATGCTTCCAGTAATGTCGAAGTTTTATCCC	AGTGTCAACAAGGAAACACCAA	1631	
Sbjct 677	CGAATTCATGCTTCCAGTAATGTCGAAGTTTTATCCC	AGTGTCAACAAGGAAACACCAA	736	
Query 1632	TGGCTTCCCGCGACAGCTTCCAATGACCTGAAGGC	ATTACTGAAGGAGCTGTGTTAAG	1691	
Sbjct 737	TGGCTTCCCGCGACAGCTTCCAATGACCTGAAGGC	ATTACTGAAGGAGCTGTGTTAAG	796	
Query 1692	TTTTCAATACATCTGCTATCGAGTAAAACGAAGAGT	GGCTTTCTACCTTGTGAAAACC	1751	
Sbjct 797	TTTTCAATACATCTGCTATCGAGTAAAACGAAGAGT	GGCTTTCTACCTTGTGAAAACC	856	
Query 1752	AGTTGAGAAAGAAATATTATCGAATATCAATGGG	ATCATGAAACCTGGTCTCAACGCCAT	1811	
Sbjct 857	AGTTGAGAAAGAAATATTATCGAATATCAATGGG	ATCATGAAACCTGGTCTCAACGCCAT	916	
Query 1812	CCTGGGACCCACAGGTGGAGGCAAACTTTCGTTATT	AGATGCTTAGCTGCAAGGAAAGA	1871	
Sbjct 917	CCTGGGACCCACAGGTGGAGGCAAACTTTCGTTATT	AGATGCTTAGCTGCAAGGAAAGA	976	
Query 1872	TCCAAGTGGATTATCTGGAGATGTTCTGATAAATGG	AGCACCACGACCTGCCAATCTCAA	1931	
Sbjct 977	TCCAAGTGGATTATCTGGAGATGTTCTGATAAATGG	AGCACCACGACCTGCCAATCTCAA	1036	
Query 1932	ATGTAATTCAGGTTACGTGGTACAAGATGATGTTGT	GATGGGCACCTGACGGTGGAGAGA	1991	
Sbjct 1037	ATGTAATTCAGGTTACGTGGTACAAGATGATGTTGT	GATGGGCACCTGACGGTGGAGAGA	1096	
Query 1992	AAACTTACAGTCTCAGCAGCTCTTCGGCTTGCAACA	ACTATGACGAATCATGAAAAAAA	2051	
Sbjct 1097	AAACTTACAGTCTCAGCAGCTCTTCGGCTTGCAACA	ACTATGACGAATCATGAAAAAAA	1156	
Query 2052	CGAACGGATTAAACAGGGTCATTCAAGAGTTAGGT	CTGGATAAAGTGGCAGACTCCAAGGT	2111	
Sbjct 1157	CGAACGGATTAAACAGGGTCATTCAAGAGTTAGGT	CTGGATAAAGTGGCAGACTCCAAGGT	1216	
Query 2112	TGGAACCTCAGTTTATCCGTGGTGTCTGGAGGAGA	AAAAAGGACTAGTATAGGAAT	2171	
Sbjct 1217	TGGAACCTCAGTTTATCCGTGGTGTCTGGAGGAGA	AAAAAGGACTAGTATAGGAAT	1276	
Query 2172	GGAGCTTACTGATCCTTCCATCTTGTCTTGGATG	AGCCTACAACCTGGCTTAGACTC	2231	
Sbjct 1277	GGAGCTTACTGATCCTTCCATCTTGTCTTGGATG	AGCCTACAACCTGGCTTAGACTC	1336	
Query 2232	AAGCACAGCAAAATGCTGTCTTTTGTCTCTGAAA	AGGATGCTAAG-CAGGGACGAACAA	2290	
Sbjct 1337	AAGNNCAGCAAAATGCTGTCTTTTGTCTCTGAAA	AGGATGCTAANCCAGGGACNAANAA	1396	
Query 2291	TCATCTT 2297			
Sbjct 1397	TCANCTT 1403			

## SeqF1

Score	Expect	Identities	Gaps	Strand
2333 bits(1263)	0.0	1289/1302(99%)	5/1302(0%)	Plus/Plus
Query 1876	AGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTCAAATGT			1935
Sbjct 24	AGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTCAAATGT			83
Query 1936	AATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGACTCTGACGGTGAGAGAAAAC			1995
Sbjct 84	AATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGACTCTGACGGTGAGAGAAAAC			143
Query 1996	TTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACTATGACGAATCATGAAAAAAAACGAA			2055
Sbjct 144	TTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACTATGACGAATCATGAAAAAAAACGAA			203
Query 2056	CGGATTAACAGGGTCAATCAAGAGTTAGGCTGGATAAAGTGGCAGACTCCAAGGTTGGA			2115
Sbjct 204	CGGATTAACAGGGTCAATCAAGAGTTAGGCTGGATAAAGTGGCAGACTCCAAGGTTGGA			263
Query 2116	ACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAAGGACTAGTATAGGAATGGAG			2175
Sbjct 264	ACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAGAAAAGGACTAGTATAGGAATGGAG			323
Query 2176	CTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACGGCTTAGACTCAAGC			2235
Sbjct 324	CTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACGGCTTAGACTCAAGC			383
Query 2236	ACAGCAAATGCTGCTCTTTTGTCTGAAAAGGATGCTAAGCAGGGACGAAACAATCATC			2295
Sbjct 384	ACAGCAAATGCTGCTCTTTTGTCTGAAAAGGATGCTAAGCAGGGACGAAACAATCATC			443
Query 2296	TTTCCATTTCATCAGCCTCGATATCCATCTTCAAGTTGTTTGTAGCCTCACCTTATTG			2355
Sbjct 444	TTTCCATTTCATCAGCCTCGATATCCATCTTCAAGTTGTTTGTAGCCTCACCTTATTG			503
Query 2356	GCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGATACTTTGAATCA			2415
Sbjct 504	GCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGATACTTTGAATCA			563
Query 2416	GCTGGTTATCACTGTGAGGCCATAAATAACCCCTGCAGACTTCTTCTGGACATCATTAAAT			2475
Sbjct 564	GCTGGTTATCACTGTGAGGCCATAAATAACCCCTGCAGACTTCTTCTGGACATCATTAAAT			623
Query 2476	GGAGATTCCTACTGCTGTGGCATTAAACAGAGAAGAAGACTTTAAAGCCACAGAGATCATA			2535
Sbjct 624	GGAGATTCCTACTGCTGTGGCATTAAACAGAGAAGAAGACTTTAAAGCCACAGAGATCATA			683
Query 2536	GAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTATGTCAACTCC			2595
Sbjct 684	GAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATTAGCGGAGATTATGTCAACTCC			743
Query 2596	TCCTTCTACAAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAAGAAGAAG			2655
Sbjct 744	TCCTTCTACAAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAAGAAGAAG			803
Query 2656	AAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTATCAACTCAGATGG			2715
Sbjct 804	AAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTATCAACTCAGATGG			863
Query 2716	GTTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCAAGGCTCTATAGCTCAGATC			2775
Sbjct 864	GTTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCAAGGCTCTATAGCTCAGATC			923
Query 2776	ATTGTCACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAAATGAT			2835
Sbjct 924	ATTGTCACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAAATGAT			983
Query 2836	TCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTGACGACCAACAGTGTTC			2895
Sbjct 984	TCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTGACGACCAACAGTGTTC			1043
Query 2896	AGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACATGAATAC			2955
Sbjct 1044	AGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACATGAATAC			1103
Query 2956	ATCAGCGGATACTACAGAGTGCATCTATTTCCCTGGAAAACTGTATCTGATTATTA			3015
Sbjct 1104	ATCAGCGGATACTACAGAGTGCATCTATTTCCCTGGAAAACTGTATCTGATTATTA			1163
Query 3016	CCCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTG			3075
Sbjct 1164	CCCATGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTG			1223
Query 3076	AAGCCAAAGGCAGATGCCCTTCTCGTTATGATGTTTACCC- TTATGAT- GGTTGGCTTATT			3133
Sbjct 1224	AAGCCAAAGGCAGATGCCCTTCTCGTTATGAGGTTTACCCCTTATGAAGGGTGGCTTNTT			1283
Query 3134	CAGCCAGTTCCATGG-CACT-GGCCATAGCAGC-AGGTCAGA 3172			
Sbjct 1284	CAGCCAGTTCCATGGNCNCTNGGCCATAGCAACCAGGTCNGA 1325			

## SeqF2

Score	Expect	Identities	Gaps	Strand
2410 bits(1305)	0.0	1362/1394(98%)	10/1394(0%)	Plus/Plus
Query 2324	TCTTCAAGTTGTTTGTATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGC			2383
Sbjct 14	TCTTC-AGTTGTTTGTATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGC			72
Query 2384	CTGCTCAGGAGGCCCTGGGATACCTTTGAATCAGCTGGTTATCACTGTGAGGCCATAAATA			2443
Sbjct 73	CTGCTCAGGAGGCCCTGGGATACCTTTGAATCAGCTGGTTATCACTGTGAGGCCATAAATA			132
Query 2444	ACCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATTCCACTGCTGTGGCATTAAACA			2503
Sbjct 133	ACCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATTCCACTGCTGTGGCATTAAACA			192
Query 2504	GAGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCA			2563
Sbjct 193	GAGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCA			252
Query 2564	TAGAAAAATAGCGGAGATTTATGTCAACTCCTCCTTCTACAAGAGACAAAAGCTGAAT			2623
Sbjct 253	TAGAAAAATAGCGGAGATTTATGTCAACTCCTCCTTCTACAAGAGACAAAAGCTGAAT			312
Query 2624	TACATCAACTTTCCGGGGTGAGAAGAAGAAGATCACAGCTTCAAGGAGATCAGCT			2683
Sbjct 313	TACATCAACTTTCCGGGGTGAGAAGAAGAAGATCACAGCTTCAAGGAGATCAGCT			372
Query 2684	ACACCACCTCCTTCTGTCACTCAACTCAGATGGGTTCTAAGCGTTCATTCAAAAACCTGC			2743
Sbjct 373	ACACCACCTCCTTCTGTCACTCAACTCAGATGGGTTCTAAGCGTTCATTCAAAAACCTGC			432
Query 2744	TGGGTAATCCCAGGCCCTATAGCTCAGATCATTGTCCAGTCGTAAGGACTGGTTA			2803
Sbjct 433	TGGGTAATCCCAGGCCCTATATCTCAGATCATTGCCACAGTCGTAAGGACTGGTTA			492
Query 2804	TAGGTGCCATTACTTTGGGCTAAAAAATGATTCTACTGGAATCCAGAACAGAGCTGGGG			2863
Sbjct 493	TAGGTGCCATTACTTTGGGCTAAAAAATGATTCTACTGGAATCCAGAACAGAGCTGGGG			552
Query 2864	TTCTCTTCTCCTGACGACCAACCAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTG			2923
Sbjct 553	TTCTCTTCTCCTGACGACCAACCAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTG			612
Query 2924	TGGTAGAGAAGAAGCTCTTCATACATGAATACATCAGCGGATACTACAGAGTGTATCTT			2983
Sbjct 613	TGGTAGAGAAGAAGCTCTTCATACATGAATACATCAGCGGATACTACAGAGTGTATCTT			672
Query 2984	ATTTCCCTGGAAAACGTTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATAT			3043
Sbjct 673	ATTTCCCTGGAAAACGTTATCTGATTTATTACCCATGAGGATGTTACCAAGTATTATAT			732
Query 3044	TTACCTGTATAGTGTACTTCACTGTTAGGATTGAAGCCAAAGGCAGATGCCTTCTTCGTTA			3103
Sbjct 733	TTACCTGTATAGTGTACTTCACTGTTAGGATTGAAGCCAAAGGCAGATGCCTTCTTCGTTA			792
Query 3104	TGATGTTTACCCTTATGATGGTGGCTTATTACGCCAGTTCATGGCACTGGCCATAGCAG			3163
Sbjct 793	TGATGTTTACCCTTATGATGGTGGCTTATTACGCCAGTTCATGGCACTGGCCATAGCAG			852
Query 3164	CAGGTGAGAGTGTGGTTCTGTAGCAACACTTCTCATGACCATCTGTTTGTGTTTATGA			3223
Sbjct 853	CAGGTGAGAGTGTGGTTCTGTAGCAACACTTCTCATGACCATCTGTTTGTGTTTATGA			912
Query 3224	TGATTTTTTTCAGGCTGTGGTCAATCTCACAAACATTGCATCTTGGCTGTATGGCTTC			3283
Sbjct 913	TGATTTTTTTCAGGCTGTGGTCAATCTCACAAACATTGCATCTTGGCTGTATGGCTTC			972
Query 3284	AGTACTTCAGCATTCCACGATATGGATTTACGGCTTTCAGCATAATGAATTTTTGGGAC			3343
Sbjct 973	AGTACTTCAGCATTCCACGATATGGATTTACGGCTTTCAGCATAATGAATTTTTGGGAC			1032
Query 3344	AAAACCTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGAACATGCAACATGTA			3403
Sbjct 1033	AAAACCTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGAACATGCAACATGTA			1092
Query 3404	CTGGCGAAGAAATTTGGTAAAGCAGGGCATCGATCTCTACCCTGGGGCTTGTGGAAGA			3463
Sbjct 1093	CTGGCGAAGAAATTTGGTAAAGCAGGGCATCGATCTCTACCCTGGGGCTTGTGGAANA			1152
Query 3464	ATCACGTGGCCTTGGCTTGTATGATTGTTATTTCCCTACAATGGCTACCTGAAATTTGT			3523
Sbjct 1153	ATCACGTGGCCTTGGCTTGTATGATTGTTATTTCCCTACAATGGCTACCTGAAATTTGT			1212
Query 3524	TATTTCTTAAAAAATATTCTTAAATGGATTCTAGAGGGCCCGTTTAAACCCGCTGATCA			3583
Sbjct 1213	TATTTCTTAAAAAATATTCTTAAATGGATTCTAGAGGGCCCGTTTAAACCCGCTGATCA			1272
Query 3584	GCCTCGACTGT-GCCCTTCAAGTGGCAGCCATCTGTT-GTTTGGCCCTCCCCC-GTGCCT			3640
Sbjct 1273	GCCTCGACTGTNGCCTTCTAGTTGCCNGCCATCTGNTGGTTTGGCCNTCCCNNGGCT			1332
Query 3641	TCCTTG-ACCCTGGAA-GGTGCCACTCCCCTGT-CCTTTCTAATAAAA-TGAGG-AAA			3695
Sbjct 1333	TCCTTGGACCCTGGAAAGGTGCCNCCNNTGGNCTTTCNAAAAAANTGAGGNAAA			1392
Query 3696	TTGCAT-CGCATTG 3708			
Sbjct 1393	TTGCATNCCCATG 1406			

## Seq482

Score	Expect	Identities	Gaps	Strand
2427 bits(1314)	0.0	1354/1380(98%)	2/1380(0%)	Plus/Plus
Query 2960	GCGGATACTACAGAGTGCATCTTATTTCCTTGGAAAACTGTTATCTGATTATTACCCA			3019
Sbjct 13	GCGGNNACTACAGAGTGCATCTTATTTCCTTGGAAAACTGTTATCTGATTATTACCCA			72
Query 3020	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			3079
Sbjct 73	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			132
Query 3080	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTACAGCA			3139
Sbjct 133	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTACAGCA			192
Query 3140	GTTCCATGGCACTGGCCATAGCAGCAGGTGAGAGTGTGGTTTCGTAGCAACACTTCTCA			3199
Sbjct 193	GTTCCATGGCACTGGCCATAGCAGCAGGTGAGAGTGTGGTTTCGTAGCAACACTTCTCA			252
Query 3200	TGACCATCTGTTTTGTGTTTATGATGATTTTTTTCAGGTCGTGGTCAATCTCACAAACA			3259
Sbjct 253	TGACCATCTGTTTTGTGTTTATGATGATTTTTTTCAGGTCGTGGTCAATCTCACAAACA			312
Query 3260	TTGCATCTTGGCTGTTCATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCTT			3319
Sbjct 313	TTGCATCTTGGCTGTTCATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCTT			372
Query 3320	TGCAGCATAATGAATTTTTGGGACAAAACCTTCTGCCAGGACTCAATGCAACAGGAAACA			3379
Sbjct 373	TGCAGCATAATGAATTTTTGGGACAAAACCTTCTGCCAGGACTCAATGCAACAGGAAACA			432
Query 3380	ATCCTTGTAACTATGCAACATGTACTGGCGAAGAAATTTGGTAAAGCAGGGCATCGATC			3439
Sbjct 433	ATCCTTGTAACTATGCAACATGTACTGGCGAAGAAATTTGGTAAAGCAGGGCATCGATC			492
Query 3440	TCTCACCTGGGGCTTGTGGAAGAATCACGTGGCCTTGGCTTGTATGATTGTTATTTTCC			3499
Sbjct 493	TCTCACCTGGGGCTTGTGGAAGAATCACGTGGCCTTGGCTTGTATGATTGTTATTTTCC			552
Query 3500	TCACAATGCGCTACCTGAAATGTTATTTCTTAAAAAATATTCTTAAATGGATTCTAGA			3559
Sbjct 553	TCACAATGCGCTACCTGAAATGTTATTTCTTAAAAAATATTCTTAAATGGATTCTAGA			612
Query 3560	GGGCCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCCTTAGTGGCAGCCATCTGTT			3619
Sbjct 613	GGGCCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCCTTAGTGGCAGCCATCTGTT			672
Query 3620	GTTTGGCCCCCCCCGTGCCCTTCCCTGACCCGGAAGGTGCCACTCCCACTGTCCCTTCC			3679
Sbjct 673	GTTTGGCCCCCCCCGTGCCCTTCCCTGACCCGGAAGGTGCCACTCCCACTGTCCCTTCC			732
Query 3680	TAATAAAATGAGGAAATGTCATCGCATGTCTGAGTAGGTGTCATTCTATTCTgggggt			3739
Sbjct 733	TAATAAAATGAGGAAATGTCATCGCATGTCTGAGTAGGTGTCATTCTATTCTgggggt			792
Query 3740	gggggtgggCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGAT			3799
Sbjct 793	gggggtgggCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCATGCTGGGGAT			852
Query 3800	GCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTCTAGGGGGATCCC			3859
Sbjct 853	GCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTCTAGGGGGATCCC			912
Query 3860	CACGCGCCCTGTAGCGGCGCATTAAAGCGCGCGGGTGGTGGTTACGCGCAGCGTGACC			3919
Sbjct 913	CACGCGCCCTGTAGCGGCGCATTAAAGCGCGCGGGTGGTGGTTACGCGCAGCGTGACC			972
Query 3920	GCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCCCTTCTCGCC			3979
Sbjct 973	GCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTCTTCCCTTCCCTTCTCGCC			1032
Query 3980	ACGTTCCGCCGGCTTCCCGTCAAGCTCTAAATCGGGGATCCCTTTAGGGTCCGATTT			4039
Sbjct 1033	ACGTTCCGCCGGCTTCCCGTCAAGCTCTAAATCGGGGATCCCTTTAGGGTCCGATTT			1092
Query 4040	AGTGCTTACGGCACCTCGACCCCAAAAAACTTGATTAGGGTATGGTTACAGTAGTGGG			4099
Sbjct 1093	AGTGCTTACGGCACCTCGACCCCAAAAAACTTGATTAGGGTATGGTTACAGTAGTGGG			1152
Query 4100	CCATCGCCCTGATAGACGGTTTTTCGCCCTTGGACGTTGGAGTCCACGTTCTTAAATAGT			4159
Sbjct 1153	CCATCGCCCTGATAGACGGTTTTTCGCCCTTGGACGTTGGAGTCCACGTTCTTAAATAGT			1212
Query 4160	GGACTCTTGTCCAAACTGGAACAACACTCAACCCATCTCGGTCTATTCTTTTGAATTA			4219
Sbjct 1213	GGACTCTTGTCCAAACTGGAACAACACTCAACCCATCTCGGTCTATTCTTTTGAATTA			1272
Query 4220	TAAGGGATTTTGGGGATTTGGCCCTATGGTTAAAAAATGAGCTGATTAACAAAAATTT			4279
Sbjct 1273	TAAGGGATTTTGGGGATTTGGCCCTATGGTTAAAAAATGAGCTGATTAACAAAAATTT			1332
Query 4280	AACGCGAATTAATTCGTGGAATGTGT-GTCAGTTAGGGTGTGGAAAGTCCCAGGCTCC			4338
Sbjct 1333	ACNGCGAATTAATTCNTGGAATG-GTGGTCNNTTAGGGTNTGGAAAGTNCACAGGNTCC			1391

## 6.2.7 L405A/I543A/V546A – Mut2

## T7 promoter (F)

Score	Expect	Identities	Gaps	Strand
2518 bits(1363)	0.0	1379/1391(99%)	0/1391(0%)	Plus/Plus
Query 914	CTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGAAGCGGTG			973
Sbjct 16	CTGGTACCGCCACCATGGCATGGAGCCACCCACAGTTCGAGAAGGGAGGTGGAAGCGGTG			75
Query 974	GAGGCTCAGGAGGCAGCGCATGGTCCCACCCAGTTTGAAAAGCTTGCCACCATGGACA			1833
Sbjct 76	GAGGCTCAGGAGGCAGCGCATGGTCCCACCCAGTTTGAAAAGCTTGCCACCATGGACA			135
Query 1034	AAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCGGGCAAGCTGGAAGTGTCTG			1093
Sbjct 136	AAGACTGCGAAATGAAGCGCACCCCTGGATAGCCCTCGGGCAAGCTGGAAGTGTCTG			195
Query 1094	GGTGCGAACAGGGCTGCACGAGATCAAGCTGCTGGGCAAGGAACATCTGCCGCCGACG			1153
Sbjct 196	GGTGCGAACAGGGCTGCACGAGATCAAGCTGCTGGGCAAGGAACATCTGCCGCCGACG			255
Query 1154	CCGTGGAAGTGCCTGCCCGAGCCGCTGCTGGGCGGACAGAGCCACTGATGCAGGCCA			1213
Sbjct 256	CCGTGGAAGTGCCTGCCCGAGCCGCTGCTGGGCGGACAGAGCCACTGATGCAGGCCA			315
Query 1214	CCGCTGGCTCAACGCCTACTTTCACAGCCTGAGGCCATCGAGGAGTTCCTGTGCCAG			1273
Sbjct 316	CCGCTGGCTCAACGCCTACTTTCACAGCCTGAGGCCATCGAGGAGTTCCTGTGCCAG			375
Query 1274	CCCTGCACCCACAGTGTCCAGCAGGAGAGCTTTACCCGCCAGGTGCTGTGGAACATGC			1333
Sbjct 376	CCCTGCACCCACAGTGTCCAGCAGGAGAGCTTTACCCGCCAGGTGCTGTGGAACATGC			435
Query 1334	TGAAAGTGGTGAAGTTCGGAGAGGTCAATCAGCTACCAGCAGCTGGCCGCCCTGGCCGGCA			1393
Sbjct 436	TGAAAGTGGTGAAGTTCGGAGAGGTCAATCAGCTACCAGCAGCTGGCCGCCCTGGCCGGCA			495
Query 1394	ATCCCGCCGCCACCGCCGCGTGA AAAACCGCCCTGAGCGGAAATCCCGTGCCCAATTCGA			1453
Sbjct 496	ATCCCGCCGCCACCGCCGCGTGA AAAACCGCCCTGAGCGGAAATCCCGTGCCCAATTCGA			555
Query 1454	TCCCTGCCACCGGGTGGTGTCTAGCTCTGGCGCCGTTGGGGGCTACGAGGGCGGGCTCG			1513
Sbjct 556	TCCCTGCCACCGGGTGGTGTCTAGCTCTGGCGCCGTTGGGGGCTACGAGGGCGGGCTCG			615
Query 1514	CCGTGAAAGAGTGGCTGTGGCCACGAGGGCCACAGACTGGGCAAGCTGGGCTGGCG			1573
Sbjct 616	CCGTGAAAGAGTGGCTGTGGCCACGAGGGCCACAGACTGGGCAAGCTGGGCTGGCG			675
Query 1574	AATTCATGTCTCCAGTAATGTGGAAGTTTTATCCCAAGTGCACAAGGAAACACCAATG			1633
Sbjct 676	AATTCATGTCTCCAGTAATGTGGAAGTTTTATCCCAAGTGCACAAGGAAACACCAATG			735
Query 1634	GCTTCCC CGCAGAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTGTTAAGTT			1693
Sbjct 736	GCTTCCC CGCAGAGCTTCCAATGACCTGAAGGCATTTACTGAAGGAGCTGTGTTAAGTT			795
Query 1694	TTCATAACATCTGCTATCGAGTAAAACGAAGAGTGGCTTTCACCTTGTGAAAACAG			1753
Sbjct 796	TTCATAACATCTGCTATCGAGTAAAACGAAGAGTGGCTTTCACCTTGTGAAAACAG			855
Query 1754	TTGAGAAAGAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGCCATCC			1813
Sbjct 856	TTGAGAAAGAAATATTATCGAATATCAATGGGATCATGAAACCTGGTCTCAACGCCATCC			915
Query 1814	TGGGACCCACAGGTGGAGGCAAATCTTCGTTATAGATGCTTAGCTGCAAGGAAAGATC			1873
Sbjct 916	TGGGACCCACAGGTGGAGGCAAATCTTCGTTATAGATGCTTAGCTGCAAGGAAAGATC			975
Query 1874	CAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACACGACCTGCCAACTTCAAAT			1933
Sbjct 976	CAAGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACACGACCTGCCAACTTCAAAT			1035
Query 1934	GTAATTCAGGTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGAAA			1993
Sbjct 1036	GTAATTCAGGTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGAAA			1095
Query 1994	ACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAAAAAAAACG			2053
Sbjct 1096	ACTTACAGTTCTCAGCAGCTCTTCGGCTTGCAACAACATGACGAATCATGAAAAAAAAACG			1155
Query 2054	AACGGATTAACAGGGTCATTCAAGAGTTAGGCTTGGATAAAGTGGCAGACTCCAAGGTTG			2113
Sbjct 1156	AACGGATTAACAGGGTCATTCAAGAGTTAGGCTTGGATAAAGTGGCAGACTCCAAGGTTG			1215
Query 2114	GAACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAGAAAAAGGACTAGTATAGGAATGG			2173
Sbjct 1216	GAACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAGAAAAAGGACTAGTATAGGAATGG			1275
Query 2174	AGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCTACAACCTGGCTTAGACTCAA			2233
Sbjct 1276	AGCTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCTACAACCTGGCTTAGACTCAA			1335
Query 2234	GCACAGCAAATGCTGTCTTTTGTCTTGGAAAGGATGCTAAGCAGGGACGAACAATCA			2293
Sbjct 1336	GNCACAGCAAATGCTGTCTTTTGTCTTGGAAAGGATGCTAANCGGGACAANCAATCA			1395
Query 2294	TCTTCTCCATT 2304			
Sbjct 1396	NCTTNNCCATT 1406			

## SeqF1

Score	Expect	Identities	Gaps	Strand
2409 bits(1304)	0.0	1342/1364(98%)	6/1364(0%)	Plus/Plus
Query 1876	AGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTCAAATGT			1935
Sbjct 21	AGTGGATTATCTGGAGATGTTCTGATAAATGGAGCACCACGACCTGCCAACTTCAAATGT			80
Query 1936	AATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGAAAAC			1995
Sbjct 81	AATTCAGGTTACGTGGTACAAGATGATGTTGTGATGGGCACTCTGACGGTGAGAGAAAAC			140
Query 1996	TTACAGTTCTCAGCAGCTCTTCGGCTTGAACAACATGACGAATCATGAAAAAAAACGAA			2055
Sbjct 141	TTACAGTTCTCAGCAGCTCTTCGGCTTGAACAACATGACGAATCATGAAAAAAAACGAA			200
Query 2056	CGGATTAACAGGGTCATTCAAGAGTTAGGCTTGGATAAAGTGGCAGACTCCAAGGTTGGA			2115
Sbjct 201	CGGATTAACAGGGTCATTCAAGAGTTAGGCTTGGATAAAGTGGCAGACTCCAAGGTTGGA			260
Query 2116	ACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAAAGGACTAGTATAGGAAATGGAG			2175
Sbjct 261	ACTCAGTTTATCCGTGGTGTCTGGAGGAGAAAAAAGGACTAGTATAGGAAATGGAG			320
Query 2176	CTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTTAGACTCAAGC			2235
Sbjct 321	CTTATCACTGATCCTTCCATCTTGTCTTGGATGAGCCTACAACCTGGCTTAGACTCAAGC			380
Query 2236	ACAGCAAATGCTGTCTTTTGTCTCTGAAAAGGATGCTAAGCAGGGACGAACAATCATC			2295
Sbjct 381	ACAGCAAATGCTGTCTTTTGTCTCTGAAAAGGATGCTAAGCAGGGACGAACAATCATC			440
Query 2296	TTCTCCATTTCATCAGCCTCGATATTCATCTTCAAGTTGTTTGATAGCCTCACCTTATTG			2355
Sbjct 441	TTCTCCATTTCATCAGCCTCGATATTCATCTTCAAGTTGTTTGATAGCCTCACCTTATTG			500
Query 2356	GCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGATACTTTGAATCA			2415
Sbjct 501	GCCTCAGGAAGACTTATGTTCCACGGGCTGCTCAGGAGGCTTGGGATACTTTGAATCA			560
Query 2416	GCTGGTTATCACTGTGAGGCCATAATAACCTGCAGACTTCTTCTGGACATCATTAAAT			2475
Sbjct 561	GCTGGTTATCACTGTGAGGCCATAATAACCTGCAGACTTCTTCTGGACATCATTAAAT			620
Query 2476	GGAGATTCCTACTGCTGGCATTAAACAGAGAAGAAGACTTTAAAGCCACAGAGATCATA			2535
Sbjct 621	GGAGATTCCTACTGCTGGCATTAAACAGAGAAGAAGACTTTAAAGCCACAGAGATCATA			680
Query 2536	GAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATAGCGGAGATTATGTCAACTCC			2595
Sbjct 681	GAGCCTTCCAAGCAGGATAAGCCACTCATAGAAAAATAGCGGAGATTATGTCAACTCC			740
Query 2596	TCCCTTACAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAAGAAGAAG			2655
Sbjct 741	TCCCTTACAAGAGACAAAAGCTGAATTACATCAACTTCCGGGGGTGAGAAGAAGAAG			800
Query 2656	AAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTATCAACTCAGATGG			2715
Sbjct 801	AAGATCACAGTCTTCAAGGAGATCAGCTACACCACCTCCTTCTGTATCAACTCAGATGG			860
Query 2716	GTTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCGAGCCTCTATAGCTCAGATC			2775
Sbjct 861	GTTTCTAAGCGTTCATTCAAAAACCTGCTGGGTAATCCCGAGCCTCTATAGCTCAGATC			920
Query 2776	ATTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAATGAT			2835
Sbjct 921	ATTGTACAGTCTGACTGGGACTGGTTATAGGTGCCATTTACTTTGGGCTAAAAATGAT			980
Query 2836	TCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTGACGACCAACCAGTGTTC			2895
Sbjct 981	TCTACTGGAATCCAGAACAGAGCTGGGGTCTCTTCTTCTGACGACCAACCAGTGTTC			1040
Query 2896	AGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACATGAATAC			2955
Sbjct 1041	AGCAGTGTTCAGCCGTGGAACCTTTGTGGTAGAGAAGAAGCTTTCATACATGAATAC			1100
Query 2956	ATCAGCGGATACTACAGAGTGTATCTTATTTCCCTGGAAAACTGTTATCTGATTTATTA			3015
Sbjct 1101	ATCAGCGGATACTACAGAGTGTATCTTATTTCCCTGGAAAACTGTTATCTGATTTATTA			1160
Query 3016	CCCATGAGGATGTTACCAAGTATTATATTACCTGTATAGTGTACTTCATGTTAGGATTG			3075
Sbjct 1161	CCCATGAGGATGTTACCAAGTATTATATTACCTGTATAGTGTACTTCATGTTAGGATTG			1220
Query 3076	AAGCCAAAGGCAGATGCCCTTCTCGTTATGATGTTTACCCTTATGAT-GGTGGCTTATTC			3134
Sbjct 1221	AAGCCAAAGGCAGATGCCCTTCTCGTTATGATGTTTACCCTTATGATGGGTGGCTTATTC			1280
Query 3135	AGCCAGTTCCATGG-CACTGG-CCATAGCAGCAGGTGAGAGTGT-GGTTCTGTAGCAAC			3191
Sbjct 1281	AGCCAGTTCCATGGGCACTGGNCCATAGCAGCAGGTGAGANNNGGGTTTCNGNAGCAAN			1340
Query 3192	AC-TTCTCATGACCATCTGTTTGTGTTTATG-ATGATTTTTTC			3233
Sbjct 1341	NCNTTTCATGACCGCCTGTTTGGCTTTATGNANAATTTTTTC			1384

## SeqF2

Score	Expect	Identities	Gaps	Strand
2386 bits(1292)	0.0	1338/1365(98%)	2/1365(0%)	Plus/Plus
Query 2324	TCTTCAAGTTGTTTGATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGC			2383
Sbjct 14	TCTTC-AGTTGTTTGATAGCCTCACCTTATTGGCCTCAGGAAGACTTATGTTCCACGGGC			72
Query 2384	CTGCTCAGGAGGCTTGGGACTTTGAATCAGCTGGTTATCACTGTGAGGCCATAAATA			2443
Sbjct 73	CTGCTCAGGAGGCTTGGGACTTTGAATCAGCTGGTTATCACTGTGAGGCCATAAATA			132
Query 2444	ACCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATTCCACTGCTGTGGCATTAAACA			2503
Sbjct 133	ACCCTGCAGACTTCTTCTTGGACATCATTAATGGAGATTCCACTGCTGTGGCATTAAACA			192
Query 2504	GAGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCA			2563
Sbjct 193	GAGAAGAAGACTTTAAAGCCACAGAGATCATAGAGCCTTCCAAGCAGGATAAGCCACTCA			252
Query 2564	TAGAAAAATAGCGGAGATTTATGTCAACTCCTCCTTCTACAAAGAGACAAAAGCTGAAT			2623
Sbjct 253	TAGAAAAATAGCGGAGATTTATGTCAACTCCTCCTTCTACAAAGAGACAAAAGCTGAAT			312
Query 2624	TACATCAACTTTCGGGGGTGAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCT			2683
Sbjct 313	TACATCAACTTTCGGGGGTGAGAAGAAGAAGATCACAGTCTTCAAGGAGATCAGCT			372
Query 2684	ACACCACCTCCTTCTGTCACTCAACTCAGATGGGTTTCTAAGCGTTCATTCAAAAACCTGC			2743
Sbjct 373	ACACCACCTCCTTCTGTCACTCAACTCAGATGGGTTTCTAAGCGTTCATTCAAAAACCTGC			432
Query 2744	TGGGTAATCCCCAGGCTCTATAGCTCAGATCATTGTCACAGTCGTAAGGACTGGTTA			2803
Sbjct 433	TGGGTAATCCCCAGGCTCTATAGCTCAGATCATTGTCACAGTCGTAAGGACTGGTTA			492
Query 2804	TAGGTGCCATTTACTTGGGCTAAAAAATGATTCTACTGGAATCCAGAACAGAGCTGGGG			2863
Sbjct 493	TAGGTGCCATTTACTTGGGCTAAAAAATGATTCTACTGGAATCCAGAACAGAGCTGGGG			552
Query 2864	TTCCTTCTCCTGACGACCAACAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTG			2923
Sbjct 553	TTCCTTCTCCTGACGACCAACAGTGTTCAGCAGTGTTCAGCCGTGGAACCTTTG			612
Query 2924	TGGTAGAGAAGAAGCTCTTACATACATGAATACATCAGCGGATACTACAGAGTGCATCTT			2983
Sbjct 613	TGGTAGAGAAGAAGCTCTTACATACATGAATACATCAGCGGATACTACAGAGTGCATCTT			672
Query 2984	ATTTCCCTGGAAAACGTATCTGATTATTACCCATGAGGATGTTACCAAGTATTATAT			3043
Sbjct 673	ATTTCCCTGGAAAACGTATCTGATTATTACCCATGAGGATGTTACCAAGTATTATAT			732
Query 3044	TTACCTGTATAGTGTACTTCACTGTAGGATTGAAGCCAAAGGCAGATGCCTTCTTCGTTA			3103
Sbjct 733	TTACCTGTATAGTGTACTTCACTGTAGGATTGAAGCCAAAGGCAGATGCCTTCTTCGTTA			792
Query 3104	TGATGTTACCCCTATGATGGTGGCTTATTCAGCCAGTCCATGGCACTGGCCATAGCAG			3163
Sbjct 793	TGATGTTACCCCTATGATGGTGGCTTATTCAGCCAGTCCATGGCACTGGCCATAGCAG			852
Query 3164	CAGGTCAGAGTGTGGTTCTGTAGCAACACTTCTCATGACCATCTGTTTGTGTTTATGA			3223
Sbjct 853	CAGGTCAGAGTGTGGTTCTGTAGCAACACTTCTCATGACCATCTGTTTGTGTTTATGA			912
Query 3224	TGATTTTTTTCAGGTCGTGGTCAATCTCACAAACATTGCATCTTGGCTGTTCATGGCTTC			3283
Sbjct 913	TGATTTTTTTCAGGTCGTGGTCAATCTCACAAACATTGCATCTTGGCTGTTCATGGCTTC			972
Query 3284	AGTACTTCAGCATTCCACGATATGGATTTACGGCTTTCAGCATAAATGAATTTTGGGAC			3343
Sbjct 973	AGTACTTCAGCATTCCACGATATGGATTTACGGCTTTCAGCATAAATGAATTTTGGGAC			1032
Query 3344	AAAACCTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGTAACTATGCAACATGTA			3403
Sbjct 1033	AAAACCTTCTGCCAGGACTCAATGCAACAGGAAACAATCCTTGTAACTATGCAACATGTA			1092
Query 3404	CTGGCGAAGAATATTTGGTAAAGCAGGGCATCGATCTCTACCCTGGGGCTTGTGGAAGA			3463
Sbjct 1093	CTGGCGAANAATATTTGGTAAAGCAGGGCATCGATCTCTACCCTGGGGCTTGTGGAAGA			1152
Query 3464	ATCACGTGGCCTTGGCTTGTATGATTGTTATTTTCTCACAAATTGCCTACCTGAAATGT			3523
Sbjct 1153	ATCACGTGGCCTTGGCTTGTATGATTGTTATTTTCTCACAAATTGCCTACCTGAAATGT			1212
Query 3524	TATTTCTTAAAAAATATTCTTAAATGG-ATTCAGAGGGCCCGTTTAAACCCGCTGATC			3582
Sbjct 1213	TATTTCTTAAAAAATATTCTTAAATGGAAATTCAGAGGGCCCGTTTAAACCCGCTGATC			1272
Query 3583	AGCCTCGACTGTGCCCTTAGTGGCAGCCATCTGTTGTTTGGCCCTCCCCCGTGCCTTC			3642
Sbjct 1273	AGCCTCGACGGGGCTTNNNGTTGCCNGCCATCGGTTGTTGGCCCTCCCCCGTGCCTTC			1332
Query 3643	CTTGACCCTGGAAGGTGCCACTCCCCTGCTTTCCTAATAAAA		3687	
Sbjct 1333	CTTGACCCTGGAAGGGGCCNCCCTTNNCCTTCCAAAAAAA		1377	

## Seq482

Score	Expect	Identities	Gaps	Strand
2398 bits(1298)	0.0	1352/1383(98%)	8/1383(0%)	Plus/Plus
Query 2960	GCGGATACTACAGAGTGCATCTTATTTCCTTGGAAAACGTGTTATCTGATTATTACCCA			3019
Sbjct 12	GCGGNNACTACAGAGTGCATCTTATTTCCTTGGAAAACGTGTTATCTGATTATTACCCA			71
Query 3020	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			3079
Sbjct 72	TGAGGATGTTACCAAGTATTATATTTACCTGTATAGTGTACTTCATGTTAGGATTGAAGC			131
Query 3080	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTACGCCA			3139
Sbjct 132	CAAAGGCAGATGCCTTCTTCGTTATGATGTTTACCCTTATGATGGTGGCTTATTACGCCA			191
Query 3140	GTTCCATGGCACTGGCCATAGCAGCAGGTGAGAGTGTGGTTTCTGTAGCAACTTCTCA			3199
Sbjct 192	GTTCCATGGCACTGGCCATAGCAGCAGGTGAGAGTGTGGTTTCTGTAGCAACTTCTCA			251
Query 3200	TGACCATCTGTTTGTGTTTATGATGATTTTTTTCAGGTCGTGGTCAATCTCACAAACA			3259
Sbjct 252	TGACCCTCTGTTTGTGTTTATGATGATTTTTTTCAGGTCGTGGTCAATCTCACAAACA			311
Query 3260	TTGCATCTTGGCTGTGATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCT			3319
Sbjct 312	TTGCATCTTGGCTGTGATGGCTTCAGTACTTCAGCATTCCACGATATGGATTTACGGCT			371
Query 3320	TGCAGCATAATGAATTTTTGGGACAAAACCTCTGCCAGGACTCAATGCAACAGGAAACA			3379
Sbjct 372	TGCAGCATAATGAATTTTTGGGACAAAACCTCTGCCAGGACTCAATGCAACAGGAAACA			431
Query 3380	ATCCTTGTAACTATGCAACATGTACTGGCGAAGAATATTGGTAAAGCAGGGCATCGATC			3439
Sbjct 432	ATCCTTGTAACTATGCAACATGTACTGGCGAAGAATATTGGTAAAGCAGGGCATCGATC			491
Query 3440	TCTCACCTGGGGCTTGTGGAAGAATCACGTGGCTTGGCTTGTATGATTGTTATTTTCC			3499
Sbjct 492	TCTCACCTGGGGCTTGTGGAAGAATCACGTGGCTTGGCTTGTATGATTGTTATTTTCC			551
Query 3500	TCACAATGGCTACCTGAAATGTTATTTCTTAAAAAATATTCTTAAATTGGATTCTAGA			3559
Sbjct 552	TCACAATGGCTACCTGAAATGTTATTTCTTAAAAAATATTCTTAAATTGGATTCTAGA			611
Query 3560	GGGCCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTT			3619
Sbjct 612	GGGCCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTT			671
Query 3620	GTTTGGCCCCCCCCGTGCCTTCTTGGACCCCTGGAAGGTGCCACTCCCCTGTCTTTCC			3679
Sbjct 672	GTTTGGCCCCCCCCGTGCCTTCTTGGACCCCTGGAAGGTGCCACTCCCCTGTCTTTCC			731
Query 3680	TAATAAAATGAGGAAATTCATCGCATGCTGTCTGAGTAGGTGTCATTCTATTCTgggggt			3739
Sbjct 732	TAATAAAATGAGGAAATTCATCGCATGCTGTCTGAGTAGGTGTCATTCTATTCTgggggt			791
Query 3740	ggggggggCAGGACAGCAAGGGGAGGATTGGGAAGCAATAGCAGGCATGCTGGGGAT			3799
Sbjct 792	ggggggggCAGGACAGCAAGGGGAGGATTGGGAAGCAATAGCAGGCATGCTGGGGAT			851
Query 3800	GCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTTAGGGGGATCCC			3859
Sbjct 852	GCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGGCTTAGGGGGATCCC			911
Query 3860	CACGCGCCCTGTAGCGGCGCATTAAAGCGCGCGGGTGTGGTGGTTACGCGCAGCGTGACC			3919
Sbjct 912	CACGCGCCCTGTAGCGGCGCATTAAAGCGCGCGGGTGTGGTGGTTACGCGCAGCGTGACC			971
Query 3920	GCTACACTTGCAGCGCCCTAGCGCCCGCTCTTTCGCTTCTTCCCTTCTTCTCGCC			3979
Sbjct 972	GCTACACTTGCAGCGCCCTAGCGCCCGCTCTTTCGCTTCTTCCCTTCTTCTCGCC			1031
Query 3980	ACGTTCCCGGCTTTCCCGTCAAGCTCTAAATCGGGGCATCCCTTAGGGTCCGATTT			4039
Sbjct 1032	ACGTTCCCGGCTTTCCCGTCAAGCTCTAAATCGGGGCATCCCTTAGGGTCCGATTT			1091
Query 4040	AGTGCTTTACGGCACCTCGACCCCAAAAACCTGATTAGGGTGTGGTTACGTTAGTGGG			4099
Sbjct 1092	AGTGCTTTACGGCACCTCGACCCCAAAAACCTGATTAGGGTGTGGTTACGTTAGTGGG			1151
Query 4100	CCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTAAATAGT			4159
Sbjct 1152	CCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTAAATAGN			1211
Query 4160	GGACTCTTTGTTCCAAACTGGAACAACACTCAACCTATCTCGGCTATTCTTTGATTTA			4219
Sbjct 1212	GAACTCTTTGTTCCAAACTGGAACAACACTCAACCTATCTCGGCTATTCTTTGATTTA			1271
Query 4220	TAAAGGATTTGGGGA-TTTC-GGCTATTGGTT-AAAAAATGAGCTGATTTAACAAAA			4276
Sbjct 1272	TAAAGGATTTGGGGA-TTTC-GGCTATTGGTT-AAAAAATGAGCTGATTTAACAAAA			1331
Query 4277	TTTAAACG-CGAATTAATCTGTGGAAATGTGT-GTCAG-TTAGGGTGTGGAAA-GTCCCC-			4331
Sbjct 1332	TTTAAACGCGAATTAATNNNTGGAANGGNTNGTCAGNTTAGGNGTGGAAAAGTCCCC			1391
Query 4332	AGG 4334			
Sbjct 1392	AGG 1394			