1	
2 3	Characterization of Southeast Asia mangoes (<i>Mangifera indica</i> L) according to their physicochemical attributes
4	Tamunonengiyeofori Lawson ^{a, b} , Grantley W. Lycett ^b , Asgar Ali ^a , Chiew Foan Chin ^{a*}
5	
6	^a School of Biosciences, Faculty of Science, The University of Nottingham Malaysia Campus,
7	Jalan Broga, 43500 Semenyih, Selangor, Darul Ehsan, Malaysia
8	^b Division of Plant and Crop Sciences, School of Biosciences, University of Nottingham, Sutton
9	Bonington Campus, Loughborough, LE12 5RD, UK
10	
11	*Corresponding author: Chiew Foan Chin,
12	Email: chiew-foan.chin@nottingham.edu.my
13	Phone: +6 (03) 8924 8216
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	

26 Abstract

25

Mango (Mangifera indica.L.) is an economically important fruit crop grown in the tropics. One 27 of the important traits of mango for successful commercial production is the storage quality of 28 the fruit. This study was conducted to evaluate the postharvest qualities of three mango 29 (Mangifera indica) varieties namely 'Chokanan', 'Golden phoenix' and 'Water lily' grown in 30 Southeast Asia regions. The study found that variety and ripening stage had an impact on the 31 32 postharvest qualities. In general, an increase in weight loss, L* value and soluble solids concentration (SSC) along with a reduction in titratable acidity (TA), firmness and hue value as 33 ripening progressed were observed irrespective of the variety. Analysis of variance and 34 multivariate analysis were used to characterize the ripening process. This study provides useful 35 information for devising strategies in postharvest handling and implementation of breeding 36 37 programs for mango crop improvement.

Abbreviations: N, Newtons; SSC, soluble solid content; TA, titratable acidity; PCA, principal
component analysis

40 Keywords: Ethylene production; fruit ripening; *Mangifera indica* L; respiration rate;
41 physicochemical characteristics; varieties.

- 42
- 43
- 44
- 45
- 46

48 1 Introduction

Mango (Mangifera indica L.) is one of the most important tropical fruit crops with significant 49 commercial value. Mango fruit is widely consumed globally due to its juiciness, delicious taste, 50 exotic flavor and nutritional value. In addition, mango fruit is a rich source of health promoting 51 compounds such as carotenoids, ascorbic acids, quercetin and mangiferin (Lauricella et al., 52 2017). Currently, Asia is the largest mango-producing region, with a production of 34.6 million 53 tons, which accounts for 74.30 % of global mango production. This is followed by America 54 (13.00 %; 4 million tons), Africa (11.00 %; 3 million tons) and a very little portion from Oceania 55 (0.10 %; 0.04 million tons) (FAOSTAT, 2016). There are thousands of mango varieties which 56 57 are distributed worldwide. Of which, Asia has over 500 fully characterized varieties (Singh et al., 2016). However, only a few of these available mango varieties are traded internationally while 58 most are grown for local consumption (Kuhn et al., 2017). Commercial mango varieties that 59 60 dominate the global export market include 'Tommy Atkins', 'Haden', 'Ataulfo', 'Kent', 'Keitt' and 'Alphonso' (Bally, 2011; Galán Saúco, 2015; Nassur et al., 2015). Mango varieties in 61 Malaysia include 'Chokanan', 'Harumanis', 'Sala', 'Masmuda' and 'Maha 65' amongst others 62 (MOA, 2016). However, these varieties have not attained equal international popularity as 63 compared to Indian or Floridian varieties due to lack of research attention (Abu Bakar and Fry, 64 65 2013).

Fruit ripening involves a spectrum of significant physiological, biochemical and molecular changes that give rise to an edible fruit of desired quality (Barry and Giovannoni, 2007). An increased rate of respiration and ethylene production during ripening has been documented extensively in climacteric fruit such as papaya (Ong *et al.*, 2013) as well as in mango ripening process (Khaliq *et al.*, 2015; Palafox-Carlos *et al.*, 2015; Zerbini *et al.*, 2015). The period of fruit

ripening is also characterized with an increase in sugar content and color changes (Palafox-71 72 Carlos et al., 2015; Ibarra-Garza et al., 2015). Mango peel color changes facilitate the identification of the appropriate maturity stage for harvesting and consumption albeit not all 73 varieties change from green to vellow/orange upon ripening (Yahia, 2011). Mango ripens within 74 4-9 days (variety dependent) (Carrillo-Lopez et al., 2000; Srivastava et al., 2016) although there 75 has been reports on 'Alphonso' and 'Banganapalli' mangoes with a ripening duration of 12-18 76 days from harvest (Deshpande et al., 2017; Nambi et al., 2015). At cold storage (13 °C), mango 77 78 can be stored for up to 2-3 weeks (Carrillo-Lopez et al., 2000).

As postharvest qualities may differ according to varieties, it is necessary to carry out specific 79 80 studies on each local mango variety in order to uncover their potential to become a commercial marketable fruit. Such information will provide an insight into the development of postharvest 81 strategies towards mango fruit quality improvement and open new marketing opportunities to the 82 farmers and to the local industry. To date, only a few published results on the physicochemical 83 and physiological profile of locally produced mangoes in the literature are available (Bejo and 84 Kamaruddin, 2014; Mansor et al., 2011; Khaliq et al., 2015; Zakaria et al., 2012). Therefore, the 85 objectives of this study were to evaluate the effect of ripening on the physicochemical 86 characteristics and physiological behavior of 'Chokanan', 'Golden phoenix' and 'Water lily' 87 88 mango varieties, which are grown in the Southeast Asia regions.

89 2 Materials and Methods

90 2.1 Mango samples

Mature green mangoes (*Mangifera indica* vars. 'Chokanan', 'Golden phoenix' and 'Water lily')
of maturity index 2 (FAMA, 2017) were purchased from a mango farmer in Malacca, Malaysia.

Mango fruit were selected for uniformity in size, shape and absence of external injury. After sorting, fruit were washed, dried and allowed to ripen at ambient temperature $(25 \pm 1 \text{ °C}, 80 \pm 5)$ % relative humidity). Assessment of postharvest quality parameters were observed on arrival (0th day) and at 2 day intervals of the ripening period. At each evaluation time, four replicates consisting of three individual fruit per replicate were randomly sampled for each mango variety. The analyses were conducted at the Postharvest Laboratory, School of Biosciences, University of Nottingham Malaysia Campus.

100 2.2 Determination of physicochemical parameters

Evaluation of physicochemical parameters was carried out as reported by Ali *et al.*, (2016). Weight loss determination was obtained by weighing mango on the 0th day of storage and at 2 day intervals over the storage period. The percentage weight loss was calculated relative to the initial weight.

Peel color was assessed on the basis of the Hunter Lab System using a MiniScan XE Plus 105 colorimeter and presented in the values of L* a* b* and h°. The L* coordinate indicates 106 brightness of color with values ranging from 0 = black to 100 = white. Coordinates, a* and b*, 107 indicate color directions: $+a^*$ is the red direction, $-a^*$ is the green direction, $+b^*$ is the yellow 108 direction, and $-b^*$ is the blue direction. From these values, hue angle (h°) was calculated as h° = 109 tangent⁻¹ b*/a*) where 0° = red purple, 90° = yellow, 180° = blue-green and 270° = blue. Fruit 110 111 firmness was assessed using an Instron Universal Testing Machine (Instron 2519-104, Norwood, MA). Measurements were taken from three points of the equatorial region for each sampled fruit. 112 An average of three readings was obtained and expressed in Newtons (N). The same fruit pulp 113 samples (10 g) used in the firmness evaluation were homogenized using a kitchen blender 114 (Philip, Malaysia) with 40 ml of distilled water, and filtered through a double layer of muslin 115

cloth to extract juice for further analyses. Soluble solid content (SSC) was determined with a
droplet of the filtrate using a Palette Digital Refractometer (Model: PR-32α, Atago Co Ltd.,
Japan) and expressed as a percentage (%). Titratable acidity (TA) was determined by titration of
5ml of filtrate with 0.1 N NaOH to an endpoint of pH 8.1 by two drops of 0.1 % phenolphthalein
indicator. The results are expressed as a percentage of citric acid equivalents.

121 2.3 Respiration and ethylene production

The respiration and ethylene production of mango fruit were carried out as described by Ong et 122 al., (2013). Fruit were placed in a plastic container tightly sealed with a lid. After 1 hour of 123 incubation, 1 ml of gas sample was withdrawn from the headspace and analyzed in the gas 124 chromatograph (GC) (Clarus-500 Perkin-Elmer, USA) equipped with a column (Agilent J&W, 125 DB-5MS column: 30 m in length, 0.25 mm in diameter and 0.25µm in film thickness) with two 126 detectors connected in series; a thermal conductivity detector (TCD) and flame ionization 127 detector (FID) for the quantification of carbon dioxide (CO₂) and ethylene respectively. Helium 128 was used as the carrier gas for thermal conductivity (TCD) and temperatures were 60 °C, 150 °C 129 and 200 °C for the oven, injector and detector respectively. The injector, oven and detector 130 temperatures were 200 °C, 120 °C and 250 °C respectively with nitrogen as the carrier gas for 131 the flame ionization detector (FID). Concentration of the standards used was 1.0 % CO₂ and 1 132 ppm ethylene (C₂H₄). Respiration and ethylene production rate are expressed as nmol $kg^{-1}s^{-1}$ 133 134 according to Banks et al., (1995).

135 2.4 Statistical analysis

The experiments were conducted according to a completely randomized design (CRD) in four replications. For each replicate, three fruit were randomly selected for analysis at each evaluation time. Data were subjected to analysis of variance (ANOVA) using the GENSTAT (18th edition) software. Means were separated using Duncan's Multiple Range Test (DMRT; p < 0.05).
Multivariate analysis was carried out using the XLSTAT (Addinsoft, New York, USA). PCA
was performed to predict the total variability between days of ripening and mango varieties. The
Pearson's correlation coefficient was employed to explore the relationship between the
postharvest parameters.

144 **3** Results

145 **3.1** Changes in physical quality parameters

Based on the external appearance and postharvest changes score, 'Golden phoenix', 'Water lily'and 'Chokanan' mango varieties were found to achieve ripeness at 7, 7 and 9 days respectively.

148 **3.1.1** Weight loss

149 A progressive weight loss was observed during ripening for all the varieties under study (Fig. 1). It increased significantly (p < 0.05) over the ambient storage period. 'Chokanan' variety 150 exhibited a 2.3 % weight loss after two days of storage (Fig 1a). The highest rate of weight loss 151 (6.98 %) was noticed on the 8th day of ripening for 'Chokanan' (a mean loss of 0.76 % per day). 152 As can be seen in Fig. 1b, weight loss in 'Golden phoenix' variety significantly increased (p < p153 0.05) from the 2nd (2.76 %) to 4th day (5.78 %). The percentage weight loss observed on the 4th 154 day was not significantly different (p < 0.05) from that obtained on the 6th day of ripening. At 155 the end of storage, 'Golden phoenix' had lost 7.76 % of initial weight with an average of 1.20 % 156 per day. 'Water lily' lost 2.48 % of its initial weight after two days of storage and this was 157 maintained with significant differences (p < 0.05) until the 6th day (Fig. 1c). At the end of 158 storage, it attained an 8.44 % weight loss which averaged 1.40 % per day. 159

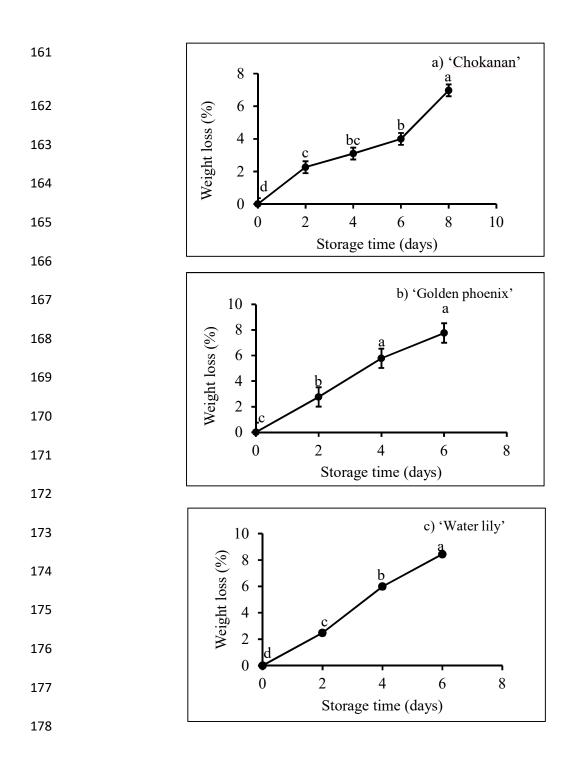


Figure 1: Weight loss of a) 'Chokanan', b) 'Golden phoenix' and c) 'Water lily' mango varieties during storage. Note: Vertical bars indicate standard error of mean of four replicates per variety. Different letters indicate significant difference between storage times at p < 0.05 for each mango variety

183 **3.1.2** Peel color

The external appearance of each variety at the beginning and end of storage is presented in Fig. 184 2. Color parameters as influenced by the ripening period are provided in Table 1. As ripening 185 progressed, the peel color changed from green to slightly or full yellow color depending on 186 variety. The visual skin color of 'Chokanan' changed noticeably to yellow during fruit ripening 187 (Fig. 2). The L* value (lightness) of 'Chokanan' was 53.63 on the 0th day of storage and 188 gradually increased as the fruit ripening advanced (Table 1). When 'Chokanan' was fully ripened 189 190 after eight days, there was a significant (p < 0.05) increase in lightness to 63.78. 'Chokanan' peel color exhibited a decline in hue angle, which started at 118.20 and was maintained with 191 192 significant differences from the 2nd to 8th day of storage (Table 1). An increasing trend was also observed on the peel a* and b* values during ripening. 'Golden phoenix' showed no conspicuous 193 changes of peel color from green to yellow upon ripening (Fig. 2). Lightness (L*) value of the 194 195 'Golden phoenix' peel increased, beginning on the 2nd day and presented no significant changes until the end of storage. Similarly, there was a gradual increase in peel a* value beginning on the 196 2nd day, and higher b* values on day four (Table1). Meanwhile, hue angle dropped 197 progressively from 119.03 to 108.61 during the ripening period. In 'Water lily' variety, hue angle 198 decreased from 120.4 to 103.3 with significant differences (p < 0.05) between the storage times 199 (Table 1). A progressive increase in peel a* value beginning on day two, and higher L* value on 200 day four (Fig. 2) were observed. Similarly, an increasing trend was observed for b* values with 201 significant differences (p < 0.05) between storage time. Overall, the peel colors of the three 202 203 mango varieties under study became lighter (higher L* values), less green (increased a* values) and tended to be more yellow (increased b* values) as ripening time progressed. 204

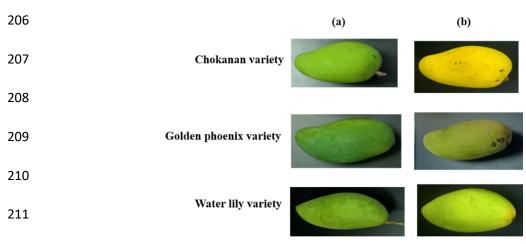


Figure 2. External peel color appearance of mango varieties.

(a) Fruit samples on arrival (day 0) and (b) samples at the end of storage (8th day for 'Chokanan'

and 6th day for 'Golden phoenix' and 'Water lily' respectively).

215

212

Table 1: Changes in peel color in mango (*Mangifera indica* L.) varieties ('Chokanan', 'Golden
phoenix' and 'Water lily') during storage.

218					
219	Variety/	Hue	L* value	a* value	b* value
220	storage time	1140	E value	u vuitue	o vuide
221	'Chokanan'				
222	0	118.20a	53.63d	-16.33d	30.75d
	2	116.61a	58.46c	-15.72d	34.53c
223	4	107.38b	60.22bc	-12.76c	41.38b
224	6	101.45c	62.53ab	-8.96b	43.51b
225	8	89.63d	63.78a	-1.31a	53.27a
226	'Golden phoenix'				
227	0	119.03a	49.38b	-15.71c	28.70b
228	2	116.10b	54.80a	-14.74bc	30.40b
	4	110.49c	54.83a	-13.68ab	36.94a
229	6	108.61c	57.59a	-12.64a	37.96a
230	'Water lily'				
231	0	120.40a	49.00b	-17.53c	29.90d
232	2	117.00b	52.65b	-17.13bc	33.89c
233	4	110.50c	57.85a	-15.38b	41.10b
234	6	103.30d	57.92a	-11.31a	48.20a

Note: L*, a* and b* indicate lightness, indexes of red/green and yellow/blue color of fruit respectively. Hue describes the visual color of the fruit. Values are means of four replicates. Different letters mean significant difference between storage times at p < 0.05 for each mango variety.

3.1.3 Pulp firmness

Over the period of storage time, a loss of pulp firmness was observed in all mango varieties under study. Firmness of 'Chokanan' decreased significantly (p < 0.05) during storage from 138.18N to 12.67N after eight days (Fig. 3a). There were no significant firmness changes during the first two days. A rapid loss of firmness (82.86 %) took place in 'Chokanan' between 2nd and 6th day of storage, with slow changes thereafter. In 'Golden phoenix', decline in firmness which started at 109.22N was maintained with significant differences (p < 0.05) between sampling points (Fig.3b). A significant decrease in firmness had begun on the second day by up to 36 % for 'Golden phoenix'. Firmness values at the end of storage (9.53 N) resulted in total loss of 91.27 % of the firmness recorded compared to the beginning of the study. For 'Water lily' variety, the firmness value decreased significantly during storage from 104.47 to 7.50 N after six days (Fig. 3c). A sharp decline was observed until the 4th day of ripening (16.61 N, 84 % loss), whereas from the 4th to the 6th day of ripening, the loss in fruit firmness remained negligible. At the end of the ripening period, 'Water lily' had lost 92.82 % of its initial fruit firmness.

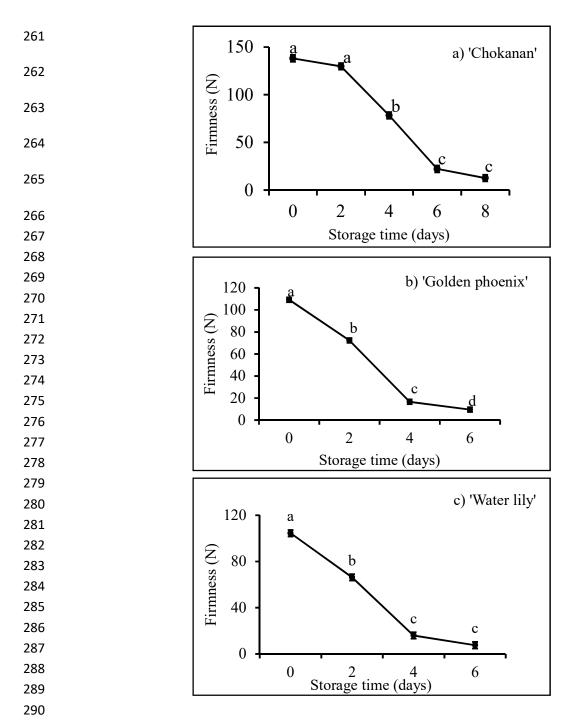


Figure 3. Firmness of a) 'Chokanan' b) 'Golden phoenix' and c) 'Water lily' mango varieties during storage. Note: Vertical bars indicate standard error of mean of four replicates per variety. Different letters indicate significant difference between storage times at p < 0.05 for each mango variety

- 295
- 296
- 297

3.1.4 Titratable acidity (TA) and soluble solids content (SSC)

In general, SSC value increased while TA declined during storage regardless of the variety. Changes in SSC and TA observed are shown in Fig. 4. The initial SSC content for 'Chokanan' was 6.83 % and it peaked (p < 0.05) at 16.80 % on the 8th day of storage (Fig. 4a) when the fruit was ripe (as depicted by the peel coloration; Fig. 2). SSC did not present much variation between storage days. TA decreased from 1.05 % on day zero to 0.26 % on the 8th day of ripening. SSC value in 'Golden phoenix', which started at 7.18 % was maintained with significant differences between the days of ripening (Fig. 4b). However, on the 6th day of storage the highest SSC value (20.30 %) was observed. A decrease in TA was recorded for 'Golden phoenix' from 0.69 % to 0.19 %, which was not statistically different (p > 0.05) between the 2nd and 4th day of storage. In 'Water lily', a significant increase in SSC value beginning on day four was recorded. The value was maintained until the end of the storage (Fig. 4c). However, changes in SSC were negligible between day four and six. While SSC increased, TA decreased from 0.34 % to 0.12 % after six days of ripening.

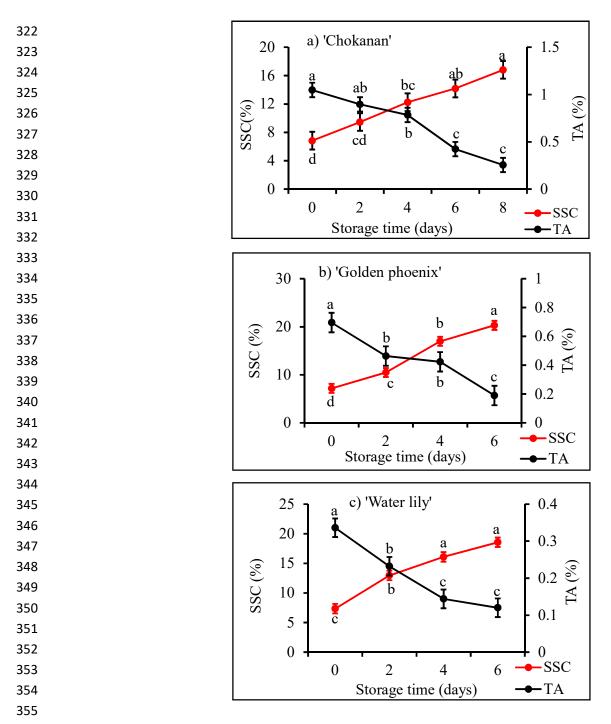


Figure 4. Titratable acidity and soluble solid concentration of (a) 'Chokanan' (b) 'Golden phoenix' and (c) 'Water lily') mango varieties during storage. Note: Vertical bars indicate standard error of mean of four replicates per variety. Different letters indicate significant difference between storage times at p < 0.05 for each mango variety. SSC, soluble solid concentration; TA, titratable acidity

362 3.1.5 Respiration and ethylene production

A typical climacteric pattern of respiration and ethylene production was observed in all mango varieties during ripening (Fig. 5). In 'Chokanan', a respiratory climacteric was apparent on the 4th day of storage and peaked at 579.40 nmol $kg^{-1}s^{-1}$ on the 6th day (Fig. 5a) when fruit exhibited a more yellow peel color. Ethylene production also peaked on the 6th day with a maximum value of 0.010 nmol $kg^{-1} s^{-1}$ and decreased afterwards (Fig. 5a). Respiration rate of 'Golden phoenix' was 279.10 nmol $kg^{-1}s^{-1}$ on day zero reaching a climacteric maximum of 939.3 nmol kg⁻¹ s⁻¹ on the 4th day. This was followed by a decrease to 797.70 nmol kg⁻¹ s⁻¹ on the sixth day (Fig. 5b). Maximum production of ethylene was observed in fruit from the 4th day (0.011 nmol kg⁻¹ s⁻¹) (Fig. 5b). In 'Water lily' a respiratory climacteric was apparent after two days in storage and peaked at 1161.40 nmol $kg^{-1}s^{-1}$ on the 4th day (Fig. 5c). Ethylene production also peaked on the 4th day of storage with a maximum value of 0.013 nmol $kg^{-1} s^{-1}$ (Fig. 5c). At that moment the production peaks, it declined until the end of the storage.

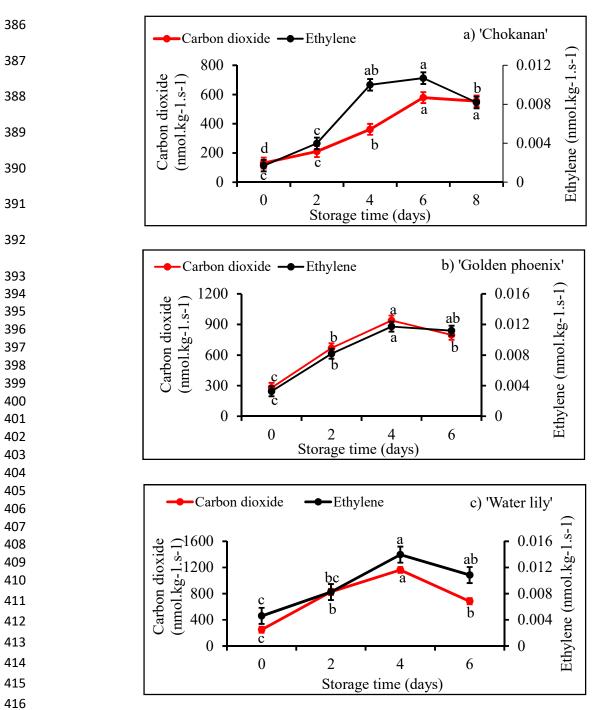


Figure 5: Respiratory pattern and ethylene production of (a) 'Chokanan' (b) 'Golden phoenix' and (c) 'Water lily') mango varieties during storage. Note: Vertical bars indicate standard error of mean of four replicates per variety. Different letters indicate significant difference between storage times at P < 0.05 for each mango variety

422 **3.2** Multivariate analysis of postharvest quality parameters

Pearson's correlation coefficient (r) was employed to explore the relationship between the 423 postharvest quality parameters during fruit ripening. Results are presented in Table 2. 424 Respiration showed a significant (p < 0.05) positive correlation between ethylene (r = 0.84) and 425 SSC (r = 0.67). Ethylene also showed a significant positive correlation with SSC (r = 0.67, p =426 0.012) and a negative correlation with firmness (r= -0.81, p < 0.01) and TA (-0.60, p = 0.029) 427 respectively. Firmness was positively correlated with hue (r = 0.59, p = 0.035) and TA (r = 0.86, 428 p < 0.01) while a negative correlation was shown for b* value (r = -0.76, p < 0.01) and SSC (r = 429 -0.86, p <0.01). 430

431 Furthermore, to obtain a broader view on the postharvest quality changes that occurred during fruit ripening, the whole data set was subjected to principal component analysis (PCA) using the 432 correlation matrix. The first principal component (F1) explained up to 62.18% of total variance 433 and PC2 explained 21.59 %, totaling 83.77 %. The rest of the components varied to a less extent 434 with 16.23 % of total variance. The samples of all varieties were well separated on the PCA 435 biplot (Fig. 6). Samples were separated along the first principal component (F1) based on 436 firmness, SSC, TA, b* value, ethylene and respiration rate. The second PC classified the 437 samples related to their external coloration (hue, L* and a* values). The positive contribution on 438 439 F1 dimension is due to high TA and firmness, whereas the negative contribution is due mainly to high SSC, respiration and ethylene rate. Separation of samples according to their ripening state 440 was achieved on F1 dimension, with unripe fruit located at the right hand side and ripe fruit on 441 the left hand side. In other words, unripe fruit have a higher firmness and TA while ripe fruit 442 have higher SSC. The contribution of b* value tells us that there is a great variability between 443 unripe and ripe fruit of the studied mango varieties based on their yellowness although this is 444

more conspicuous in 'Chokanan' variety (Fig. 2). On the other hand, F2 dimension showed separation related to the variety effect, with 'Chokanan' samples at the top (increased L* and a*) and the other varieties on the lower region (high hue values). However, no clear demarcation was achieved for 'Waterlily' and' Golden phoenix' varieties. This could be due to a lesser variability of the color coordinates (hue, L* and a*) on the F2 dimension between these varieties. The green coloration retained by these varieties ('Waterlily' and 'Golden phoenix') upon ripening supports this possibility (Fig. 2). More positive scores along F2 dimension for 'Chokanan' on the 8th day of storage could be as a result of further accumulation of pigmentation yielding more yellow coloration as ripening progresses.

455 Table 2: Correlation matrix among postharvest quality variables

Variables	Respiration	Ethylene	Firmness	Hue	a*	L*	b*	TA	SSC
Respiration	1	0.84*	-0.81*	-0.26	0.17	0.19	0.52	-0.60*	0.67*
Ethylene		1	-0.79*	-0.47	0.30	0.38	0.74*	-0.59*	0.67*
Firmness			1	0.59*	-0.49	-0.37	-0.76*	0.86*	-0.86
Hue				1	-0.96*	-0.90*	-0.51	0.34	-0.69
a*					1	0.83*	0.31	-0.26	0.58
L*						1	0.39	-0.10	0.57
b*							1	-0.62*	0.71°
ТА								1	-0.64
SSC									1

* indicates significance of correlation at the level of 0.05. SSC, soluble solid concentration; TA,
titratable acidity.

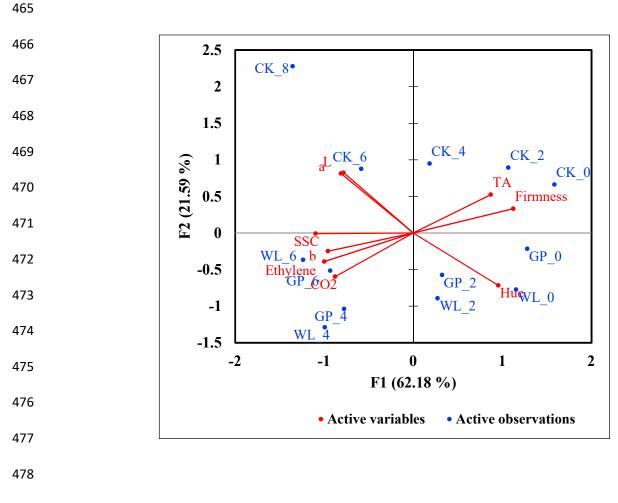


Figure 6. PCA biplot of the postharvest quality attributes in the three mango varieties 'Chokanan'
(CK), 'Golden phoenix' (GP) and Waterlily (WL) on 0th day (0), 2nd day (2), 4th day (4), 6th
day (6) and 8th (8). (L, L* value; a, a* value; b, b* value; CO₂, carbon dioxide; SSC, soluble
solid content; TA, titratable acidity).

485 4 Discussion

486 4.1 Changes in postharvest qualities

487 4.1.1 Weight loss

Weight loss is an aspect that determines the storage life and quality of fruit. Harvested fruit continue to respire and lose water to the environment. In mango, water loss through the stomata and lenticels seems to be the possible reason for physiological weight loss during storage (Brecht and Yahia, 2009). The results from this study are in line with the findings obtained from 'Dashehari' mango (Gupta and Jain, 2012) and other climacteric fruit such as papaya (Ong *et al.*, 2013) at ambient storage. However, the variability among species, varieties, ripening stage and storage conditions could be possible factors explaining the differences (Kader, 2002).

495 4.1.2 Peel color changes

A change in peel color is mainly caused by the degradation of chlorophyll and accumulation of pigments such as carotenoid, xanthophyll and lycopene (Ornelas-Paz *et al.*, 2008). The peel color showed a reduction in hue values and increase in L* value, characterizing the loss of the green color during fruit ripening. However, our results show that peel color is not a stand-alone indicator but rather more useful when combined with other quality parameters throughout ripening period. The results from this study show similar trends to those observed in other mango varieties (Ibarra-Garza *et al.*, 2015; Palafox-Carlos *et al.*, 2015).

503 4.1.3 Loss of fruit firmness

504 'Chokanan' variety has been reported to be a firm variety in comparison with other varieties 505 under study (Jarimopas and Kitthawee, 2007; Vásquez-Caicedo *et al.*, 2002). This is in 506 agreement with our observations for 'Chokanan' variety having a longer storage period. Decreased fruit firmness has been reported in other mango varieties (Jha *et al.*, 2013; Ibarra-Garza *et al.*, 2015; Palafox-Carlos *et al.*, 2015). Flesh firmness is of great concern in mango as it plays an integral role in shelf life, pathogen resistance, transportation and storage of the fresh produce. Loss of flesh firmness has been reported to be associated with the cell wall modification and starch hydrolysis (Muda *et al.*, 1995). The best organoleptic quality of mango fruit is when they are soft with a pulp firmness between 4.5 N and 26. 7 N (Nassur *et al.*, 2015).

513 4.1.4 Titratable acidity and soluble solids concentration

TA and SSC play an important role in both fresh and processing markets of table fruit. The 514 515 patterns of TA and SSC observed in this study have been similarly reported for other mango 516 varieties such as 'Ataulfo' (Palafox-Carlos et al., 2015), 'Haden' (Nassur et al., 2015) and 'Keitt' (Padda et al., 2011). However, the different acidity values obtained in the respective studies 517 reflects variations exist among various mango varieties (Yahia, 2011). Decline in acidity is 518 attributed to their utilization as substrates for respiration and conversion to sugars as ripening 519 520 progresses (Espitia et al., 2012). SSC values for 'Chokanan' variety were similar to those 521 reported by Bejo and Kamarudin, (2011) from the same variety and geographical region. Overall, studies on other mango varieties (Nassur et al., 2015; Ibarra-Garza et al., 2015; Padda et al., 522 2011; Palafox-Carlos et al., 2015) corroborate with the findings generated in this studies. The 523 SSC of all the ripe mango varieties in this study fitted well with the 10-20 % SSC requirement 524 for ripe mangoes (Mitcham, 2012; Yahia, 2011). 525

526 4.1.5 Respiration and ethylene production

527 Climacteric fruits such as mango are characterized by an increase in respiration rate and ethylene 528 production. Based on the results, it can be inferred that the climacteric rise in mango fruit

occurred when it was considerably ripe. Similar patterns have been reported for other mango 529 varieties such as 'Ataulfo' (Palafox-Carlos et al., 2015) and 'Cogshall' (Nordey et al., 2016). In 530 contrast, 'Amrapali' and 'Dasheri' mangoes did not follow a climacteric pattern (Reddy and 531 Srivastava, 1999). Similar ethylene production rates recorded in this study were reported for 532 other mango varieties such as 'Carabao' (Cua and Lizada, 1990), 'Kesington pride' (Lalel et al., 533 2003) and 'Ataulfo' (Palafox-Carlos et al., 2015). As observed by these authors, the outburst of 534 ethylene may precede, coincide or lag behind the respiratory peak during mango ripening. The 535 comparison of the respiration profiles and the ethylene production for the three mango varieties 536 revealed that the two physiological processes occurred in a similar way. 537

Furthermore, the mangoes investigated in this study were comparable to the globally traded 538 mango varieties (Appendix Table 1). The tropical mango varieties under study did not differ 539 540 greatly from the commercial mangoes reported so far in terms of their postharvest quality 541 parameters including pulp firmness, soluble solids and titratable acidity. On the other hand, the peel color of 'Golden phoenix' and 'Water lily' varieties tended to be higher, characterizing by 542 543 their green fruit coloration (Fig. 2) upon ripening compared to the other varieties. Since peel color is one of the most important visual attributes in mango that drives marketability and 544 consumption (Jha et al., 2013; Nassur et al., 2015), this quality of not attaining a full yellow 545 coloration may influence the consumer acceptance of 'Golden phoenix' and 'Water lily' 546 mangoes in the international market (Jha et al., 2013; Nassur et al., 2015). Regarding firmness 547 and SSC, the Southeastern mangoes fitted well at 4.5 – 26 N pulp firmness (Nassur *et al.*, 2015) 548 and 10 - 20 % SSC (Mitcham, 2012; Yahia, 2011) requirement for ripe mangoes. Even though 549 'Golden phoenix' and 'Water lily' mangoes show green coloration upon ripening, utilization of 550 these varieties in the pulping industry for mango purée and juices may be a good option because 551

of the soluble solid concentration and acidity level (Nambi et al., 2015; Vásquez-Caicedo et al.,

553 2002). Taken together, the results of this study offer new insights to uncover the potential of the

554 investigated mango varieties to become commercially marketable fruits.

555 4.2 Multivariate studies

Pearson's correlation coefficient was employed to explore the relationship between the 556 postharvest quality parameters during fruit ripening. The positive relationship between SSC and 557 b* value can be explained by the observation that as ethylene, respiration and SSC increases 558 during ripening, the fruit becomes less acidic and firm. The negative correlation between hue and 559 the other color coordinates (b* and L* values) is expected because as a mango fruit ripens, these 560 values increases with pigment accumulation leading to a reduced hue value (fruit becoming 561 brighter and more yellow). Correlation of some postharvest parameters observed in this study are 562 in line with studies in mango (Nambi et al., 2015) and tomato (Aoun et al., 2013). Hue was not 563 significantly correlated with respiration and ethylene, which agree with the observation by Ketsa 564 et al., (1999) who found that 'Tongdum' mangoes, which remained green upon ripening had 565 high ethylene production compared with 'Nam Dok Mai' mangoes, which turn completely 566 567 yellow. Similar discrimination based on fruit ripening stages as observed in this study has been reported in other mango varieties (Nambi et al., 2015; Padda et al., 2011) and banana (Valérie 568 Passo Tsamo et al., 2014). As the fruit ripened, there was a shift from right to left along F1 (Fig. 569 6) with increase in SSC, yellowness (b* value), ethylene and respiration rate. In this study, 570 decrease in acidity and firmness in unripe fruit, was also characterized by a shift from right to 571 572 left, reflecting the ripening process in the mango varieties. The two principal components played 573 an important role in explaining the total variation of the external appearance in this study since color coordinates (hue, L*, b*and a* values) were distributed over the PCA biplot. The lack of 574

separation between 'Golden phoenix' and 'Water lily' mangoes on the F2 dimension could be 575 due to a lesser variability of the changes in color coordinates (hue, L* and a* values) between 576 'Golden phoenix' and 'Water lily' varieties as ripening progressed over the storage time. There 577 is a huge variability of postharvest attributes among mango varieties. For this reason, a common 578 classification of postharvest qualities is not suitable for all mango varieties (Nambi et al., 2016). 579 Multivariate comparisons clearly indicated the correlation between the physicochemical 580 581 parameters and their relationship in different mango fruit varieties during the ripening period. 582 The present postharvest studies to assess the phenotypic variabilities in the mango fruit varieties would be useful indicators for postharvest quality determination. 583

584 5 Conclusion

This study showed that variety and ripening period had an impact on the postharvest qualities on 585 586 mango fruit. Considering the high genetic variability of the mango varieties, additional investigations at the biochemical and molecular levels are recommended to provide a more 587 588 complete picture of what occurs at ripening. Besides understanding ripening behavior, it would be beneficial to integrate the results of this study with additional investigations that also take into 589 consideration different harvesting times, location and postharvest storage conditions. Such 590 information will provide an insight into the development of postharvest management strategies 591 towards mango fruit quality improvement and open new marketing opportunities to the farmers 592 and the local industry. Multivariate analysis has shown to be a valuable tool in making decisions 593 and view variable/variety interrelations, thus facilitating mango selection and utilization strategy. 594 Consumer perception for the fruit is an important factor that influences the marketability of fruits 595 such as mango. As such, further investigation on these mango varieties aiming at the evaluation 596 of their sensory properties will provide valuable information which could be used by growers, 597

598 plant breeders, exporters and marketing agents to facilitate increased utilization and export of

599 varieties that would be acceptable by consumers globally. Nevertheless, the information

600 provided in this study would likely to open up promising possibilities in the world market trade

601 for Southeast Asian mangoes which are locally common but globally rare.

602 Acknowledgements

This work was supported by the CFF-UNMC Doctoral Training Partnership under the FoodPlus
program [Food P1-016]. The authors wish to thank Dr. Ajit Singh for his invaluable help in the
statistical analysis.

606 Declarations of interest: none

607 **References**

- Abu Bakar, M.F.A., Fry, J., 2013. A review on underutilized indigenous 'Bambangan'
 (*Mangifera pajang*) fruit as a potential novel source for functional food and medicine.
 Journal of Medicinal Plant Research 7, 3292-3297.
- Ali, A., Hei, G.K., Keat, Y.W., 2016. Efficacy of ginger oil and extract combined with gum
 arabic on anthracnose and quality of papaya fruit during cold storage. Journal of Food
 Science and Technology 53, 1435-1444. doi:10.1007/s13197-015-2124-5
- Aoun, A.B., Lechiheb, B., Benyahya, L., Ferchichi, A., 2013. Evaluation of fruit quality traits of
- 615 traditional varieties of tomato (*Solanum lycopersicum*) grown in Tunisia. African Journal of
- 616 Food Science 7, 350-354. doi:10.5897/ajfs2013.1067
- Bally, I.S.E., 2011. Advances in research and development of mango industry. Revista Brasileira
- 618 de Fruticultura 33, 57-63. doi:10.1590/s0100-29452011000500008

- Banks, N., Cleland, D., Cameron, A., Beaudry, R., Kader, A.A, 1995. Proposal for rationalized
 system of units for post-harvest research in gas exchange. Hortscience 30, 1129-1131.
- Barry, C.S, Giovannoni, J.J., 2007. Ethylene and fruit ripening. Journal of Plant Growth
 Regulation 26, 143-159.
- Bejo, S.K., Kamaruddin, S., 2014. 'Chokanan' mango sweetness determination using nondestructive image processing technique. Australian Journal of Crop Science 8, 475-480.
- Brecht, J.K., Yahia, E.M., 2009. Postharvest Physiology, in: Richard, L. (Ed) The mango botany,
 production and uses. CABI, Wallingford, U.K, pp. 484–528.
- Carrillo-Lopez, A., Ramirez-Bustamante, F., Valdez-Torres, J.B., Rojas-Villegas, R., Yahia,
 E.M., 2000. Ripening and quality changes in mango fruit as affected by coating with an
 edible film. Journal of Food Quality 23, 479-486. doi:10.1111/j.1745-4557.2000.tb00573.x
- Cua, A.U., Lizada, M.C.C., 1990. Ethylene production in the Carabao mango (*Mangifera Indica*L.) fruit during maturation and ripening. Acta Horticulturae 169-180.
 doi:10.17660/actahortic.1990.269.22
- Deshpande, A., Anamika, K., Jha, V., Chidley, H., Oak, P., Kadoo, N., Pujari, K., Giri, A.,
 Gupta, V., 2017. Transcriptional transitions in Alphonso mango (*Mangifera indica* L.)
 during fruit development and ripening explain its distinct aroma and shelf life
 characteristics. Scientific Reports 7, 8711. doi:10.1038/s41598-017-08499-5.
- Espitia, P.J.P., Soares, N.F.F., Botti, L.C.M., Melo, N.R., Pereira, O.L., Silva, W.A., 2012.
 Assessment of the efficiency of essential oils in the preservation of postharvest papaya in an

- antimicrobial packaging system. Brazilian Journal of Food Technology 15, 333-342.
 doi:10.1590/s1981-67232012005000027
- FAMA, Federal Agricultural and Marketing Authority, 2017. Grading standards and
 specification of fruits and vegetables http://www.fama.gov.my/documents/10157/af9f2d40c116-4a3e-8c8ccc1c897bb039 (accessed 6th May, 2017).
- FAOSTAT, Food and Agriculture Organization of the United Nations: Statistics Division, World
 mango production in 2016. http://www.fao.org/faostat/en/#data/QC (accessed 20th
 November, 2017).
- Galán Saúco, V., 2015. Current situation and future prospects of worldwide mango production
 and market. Acta Horticulturae 69-84. doi:10.17660/actahortic.2015.1066.7
- Gupta, N., Jain, S.K., 2014. Storage behavior of mango as affected by post-harvest application of
 plant extracts and storage conditions. Journal of Food Science and Technology 51, 24992507. doi:10.1007/s13197-012-0774-0
- 652 Ibarra-Garza, I.P., Ramos-Parra, P.A., Hernández-Brenes, C., Jacobo-Velázquez, D.A., 2015.
- Effects of postharvest ripening on the nutraceutical and physicochemical properties of mango (*Mangifera indica* L. cv 'Keitt'). Postharvest Biology and Technology 103, 45-54.
- 655 doi:10.1016/j.postharvbio.2015.02.014
- Jarimopas, B., Kitthawee, U., 2007. Firmness properties of mangoes. International Journal of
 Food Properties 10, 899-909. doi:10.1080/10942910701221731

- Jha, S.N., Jaiswal, P., Narsaiah, K., Kaur, P.P., Singh, A.K., Kumar, R., 2013. Textural
 properties of mango cultivars during ripening. Journal of Food Science and Technology 50,
 1047-1057. doi:10.1007/s13197-011-0431-z
- Kader, A.A., 2002. Postharvest biology and technology: an overview, in: Kader, A.A. (Ed.)
 Postharvest Technology of Horticultural Crops. University of California, Agriculture and
 Natural Resources, Oakland, CA, pp. 39-47
- Ketsa, S., Phakawatmongkol, W., Subhadrabhandhu, S., 1999. Peel enzymatic activity and
 colour changes in ripening mango fruit. Journal of Plant Physiology 154, 363-366.
 doi:10.1016/s0176-1617(99)80181-3
- Khaliq, G., Mohamed, M.T.M., Ali, A., Ding, P., Ghazali, H.M., 2015. Effect of gum arabic
 coating combined with calcium chloride on physico-chemical and qualitative properties of
 mango (*Mangifera indica* L.) fruit during low temperature storage. Scientia Horticulturae
 190, 187-194. doi:10.1016/j.scienta.2015.04.020
- Kuhn, D.N., Bally, I.S.E., Dillon, N.L., Innes, D., Groh, A.M., Rahaman, J., Ophir, R., Cohen,
 Y., Sherman, A., 2017. Genetic map of mango: A tool for mango breeding. Frontiers in
 Plant Science 8, 577. doi:10.3389/fpls.2017.00577
- Lalel, H.J.D., Singh, Z., Tan, S.C., Agustí, M., 2003. Maturity stage at harvest affects fruit
 ripening, quality and biosynthesis of aroma volatile compounds in 'Kensington Pride'
 mango. The Journal of Horticultural Science and Biotechnology 78, 225-233.
 doi:10.1080/14620316.2003.11511610

678	Lauricella, M., Emanuele, S., Calvaruso, G., Giuliano, M., D'Anneo, A., 2017. Multifaceted
679	health benefits of Mangifera indica L. (mango): The inestimable value of orchards recently
680	planted in Sicilian rural areas. Nutrients 9, 525. doi:10.3390/nu9050525

- Mansor, A., Mahmod, O., Nazari, A., 2014. Fuzzy ripening mango index using RGB colour
 sensor model. Research Journal of Art Science & Commerce 5, 1-9
- 683 Mitcham, B., 2012. Mango ripening [PowerPoint slides]. Retrieved from
 684 http://ucce.ucdavis.edu/files/datastore/234-2168.pdf
- Muda, P., Seymour, G.A., Errington, N., Tucker, G.A., 1995. Compositional changes in cell wall
 polymers during mango fruit ripening. Carbohydrate Polymers 26, 255-260.
 doi:10.1016/0144-8617(95)00028-6
- MOA, Ministry of Agriculture and Agro based Industry, Malaysia, 2016. Mango.
 http://www.moa.gov.my/buah-buahan (accessed 15th December, 2017).
- Nambi, V., Thangavel, K., Shahir, S., Chandrasekar, V., 2016. Color Kinetics During Ripening
 of Indian Mangoes. International Journal of Food Properties 19, 2147-2155.
 doi:10.1080/10942912.2015.1089281
- Nambi, V.E., Thangavel, K., Jesudas, D.M., 2015. Scientific classification of ripening period and
 development of color grade chart for Indian mangoes (*Mangifera indica* L.) using
 multivariate cluster analysis. Scientia Horticulturae 193, 90-98.
 doi:10.1016/j.scienta.2015.05.031.
- Nassur, R.C., González-Moscoso, S., Crisosto, G.M., Lima, L.C., Vilas Boas, E.V., Crisosto,
 C.H., 2015. Describing quality and sensory attributes of 3 mango (*Mangifera indica* L.)

- cultivars at 3 ripeness stages based on firmness. Journal of Food Science 80, S2055-S2063.
 doi:10.1111/1750-3841.12989.
- Nordey, T., Léchaudel, M., Génard, M., Joas, J., 2016. Factors affecting ethylene and carbon
 dioxide concentrations during ripening: Incidence on final dry matter, total soluble solids
 content and acidity of mango fruit. Journal of Plant Physiology 196-197, 70-78.
 doi:10.1016/j.jplph.2016.03.008
- Ong, M.K., Forney, C.F., Alderson, P.G., Ali, A., 2013. Postharvest profile of a Solo variety
 'Frangi' during ripening at ambient temperature. Scientia Horticulturae 160, 12-19.
 doi:10.1016/j.scienta.2013.05.026.
- Ornelas-Paz, J., Yahia, E.M., Gardea, A.A., 2008. Changes in external and internal color during
 postharvest ripening of 'Manila' and 'Ataulfo' mango fruit and relationship with carotenoid
 content determined by liquid chromatography–APcI+-time-of-flight mass spectrometry.
 Postharvest Biology and Technology 50, 145-152. doi:10.1016/j.postharvbio.2008.05.001
- Padda, M.S., do Amarante, C.V., Garcia, R.M., Slaughter, D.C., Mitcham, E.J., 2011. Methods
 to analyse physico-chemical changes during mango ripening: A multivariate approach.
 Postharvest Biology and Technology 62, 267-274. doi:10.1016/j.postharvbio.2011.06.002.
- Palafox-Carlos, H., Yahia, E.M., Islas-Osuna, M., Gutierrez-Martinez, P., Robles-Sánchez, M.,
 González-Aguilar, G., 2012. Effect of ripeness stage of mango fruit (*Mangifera indica* L.,
- cv. 'Ataulfo') on physiological parameters and antioxidant activity. Scientia Horticulturae
- 718 135, 7-13. doi:10.1016/j.scienta.2011.11.027.
- Reddy, Y.T.N., Srivastava, G.C., 1999. Ethylene biosynthesis and respiration in mango fruit
 during ripening. Indian Journal of Plant Physiology 4, 32–35.

721	Singh, N., Mahato, A., Jayaswal, P., Singh, A., Singh, S., Singh, N., Rai, V., Mithra, A.,
722	Gaikwad, K., Sharma, N., Lal, S., Srivastava, M., Prakash, J., Kalidindi, U., Singh, S.,
723	Singh, A., Khan, K., Mishra, R., Rajan, S., Bajpai, A., Sandhya, B., Nischita, P.,
724	Ravishankar, K., Dinesh, M., Kumar, N., Jaiswal, S., Iquebal, M., Kumar, D., Rai, A.,
725	Sharma, T., 2016. Origin, diversity and genome sequence of mango (Mangifera indica L.).
726	Indian Journal of History of Science 51. doi:10.16943/ijhs/2016/v51i2.2/48449.

- Srivastava, S., Singh, R.K., Pathak, G., Goel, R., Asif, M.H., Sane, A.P., Sane, V.A., 2016.
 Comparative transcriptome analysis of unripe and mid-ripe fruit of *Mangifera indica* (var.
 'Dashehari') unravels ripening associated genes. Scientific Reports 6, 32557.
 doi:10.1038/srep32557.
- Valérie Passo Tsamo, C., Andre, C., Ritter, C., Tomekpe, K., Ngoh Newilah, G., Rogez, H.,
 Larondelle, Y., 2014. Characterization of *Musa* sp. fruit and plantain banana ripening stages
 according to their physicochemical attributes. Journal of Agricultural and Food Chemistry
 62, 8705-8715. doi:10.1021/jf5021939
- Vásquez-Caicedo, A., Neidhart, S., Pathomrungsiyounggul, P., Wiriyacharee, P., Chattrakul, A.,
 Sruamsiri, P., Manochai, P., Bangerth, F., Carle, R., 2002. Physical, chemical and sensory
 properties of nine Thai mango cultivars and evaluation of their technological and nutritional
 potential, in: International symposium sustaining food security and managing natural
 resources in Southeast Asia Challenges for the 21st century.
- Yahia, E.M., 2011. Mango (*Mangifera indica* L.), in: Yahia, E.M, Postharvest biology and
 technology of tropical and subtropical fruit: cocona to mango. Woodhead Publishing
 Limited, pp. 492B-586

743	Zakaria, A	., Shakaff, A.,	Masnan, 1	M., Saad,	F., Adom,	A., Ahmad,	M., Jaafar,	M., Abdullah	, A.
-----	------------	-----------------	-----------	-----------	-----------	------------	-------------	--------------	------

- Kamarudin, L., 2012. Improved maturity and ripeness classifications of Magnifera Indica
- cv. 'Harumanis' mangoes through sensor fusion of an electronic nose and acoustic sensor.
- 746 Sensors 12, 6023-6048. doi:10.3390/s120506023.
- 747 Zerbini, P.E., Vanoli, M., Rizzolo, A., Grassi, M., Pimentel, R.M., Spinelli, L., Torricelli, A.,
- 748 2015. Optical properties, ethylene production and softening in mango fruit. Postharvest
- 749 Biology and Technology 101, 58-65. doi:10.1016/j.postharvbio.2014.11.008.

750

751

Appendix Table 1: Comparison of postharvest quality attributes between Southeast Asianmangoes and globally traded mangoes.

A	(C1.1	'Golden	'Water	6 4 1 1	(A 4 1 C - 2	·II. 1	(17 . 44)	'Tommy
Attribute	'Chokanan'	Phoenix'	lily'	'Alphonso'	'Ataulfo'	'Haden'	'Keitt'	Atkins'
Firmness (N)	12.67 ^a	9.53 ^a	7.50 ^a	0.93 ^b	7.84 ^c /11.70 ^d	8.82 ^c	5.30 ^e	5.88 °
SSC (%)	16.80 ^a	20.30 ^a	18.55 ^a	19.41 ^b	18.84 ° /21.60 ^d	13.87 °	17.30 ^e	19.54 °
TA (%)	0.26 ^a	0.19 ^a	0.12 ^a	0.01 ^b	0.56 °	0.10 ^c	0.20 ^e	0.18 °
Hue*	89.63 ^a	108.61 ^a	103.30 ^a		85.00 ^d /89.80 ^c	67.42 ^c		89.88 ^c
L*	63.78 ^a	57.59 ^a	57.97 ^a	64.30 ^b	59.31 ^c /75.00 ^d	57.80°		59.53°

754

Note: Subscript letters: (a) Data obtained from the present study; (b) Nambi et al., 2015; (c)

756 Nassur et al., 2015; (d) Palafox-Carlos et al., 2012; (e) Padda et al., 2011