- 1 Exploring drivers of litter decomposition in a greening Arctic: Results from a
- 2 transplant experiment across a tree-line
- 3 Running head: Decomposition in a greening Arctic
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22 Key words: vegetation change, litter, decomposition, Arctic, tundra, forest, snow

23 Abstract

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Decomposition of plant litter is a key control over carbon (C) storage in the soil. The biochemistry of the litter being produced, the environment in which the decomposition is taking place, and the community composition and metabolism of the decomposer organisms exert a combined influence over decomposition rates. As deciduous shrubs and trees are expanding into tundra ecosystems as a result of regional climate warming, this change in vegetation represents a change in litter input to tundra soils and a change in the environment in which litter decomposes. To test the importance of litter biochemistry and environment in determining litter mass loss, we reciprocally transplanted litter between heath (Empetrum nigrum), shrub (Betula nana) and forest (Betula pubescens) at a sub-arctic tree-line in Sweden. As expansion of shrubs and trees promotes deeper snow, we also used a snow fence experiment in a tundra heath environment to understand the importance of snow depth, relative to other factors, in the decomposition of litter. Our results show that B. pubescens and B. nana leaf litter decomposed at faster rates than E. nigrum litter across all environments, while all litter species decomposed at faster rates in the forest and shrub environments than in the tundra heath. The effect of increased snow on decomposition was minimal, leading us to conclude that microbial activity over summer in the productive forest and shrub vegetation is driving increased mass loss compared to the heath. Using B. pubescens and E. nigrum litter, we demonstrate that degradation of carbohydrate-C is a significant driver of mass loss in the forest. This pathway was less prominent in the heath, which is consistent with observations that tundra soils typically have high concentrations of 'labile' C. This experiment suggests that further expansion of shrubs and trees may stimulate the loss of undecomposed carbohydrate-C in the tundra.

- 48 *Introduction*
- 49 Climate warming in the Arctic of 1 4 °C since 1960 (Serreze and Francis 2006,
- 50 Serreze and Barry 2011) has resulted in large areas of tundra becoming more
- 51 productive, with some landscapes showing increases in aboveground biomass of 10 g
- 52 m⁻² yr⁻¹ (Epstein et al. 2012). In many of these areas, shrubs and trees have been
- observed to increase in cover and height (Myers-Smith et al. 2011, Elmendorf et al.
- 54 2012) and are generally thought to contribute to the increase in "greenness" that is
- observed from space (Tape et al. 2006). Earth system models have predicted that
- 56 increased productivity in arctic ecosystems will increase carbon (C) sequestration at the
- 57 biome level (Cramer et al. 2001, Qian et al. 2010, Todd-Brown et al. 2013) through
- 58 increased litter-fall. However, these predictions are at odds with observations in the
- 59 Arctic of lower soil organic matter (SOM) storage under shrub and tree species than
- adjacent tundra systems (Wilmking et al. 2006, Hartley et al. 2012, Parker et al. 2015).
- This suggests that we do not yet fully understand the interactions between plant
- 62 functional types (PFTs), litter input and decomposition rates and ecosystem carbon
- 63 cycling in the Arctic.
- Plant litter is the primary input of C into soil (Aber and Melillo 2001); its
- decomposition contributes towards humic substances which can lead to the formation of
- stable soil organic matter (SOM) (Melillo et al. 1989, Sollins et al. 1996). Along with
- 67 physico-chemical environmental controls (i.e. temperature, humidity, pH, mineralogy),
- the species identity and functional type are key to determining the rate of
- decomposition of their litter and eventual contribution to SOM (Dorrepaal et al. 2005,
- 70 Cornelissen et al. 2007, Cornwell et al. 2008, Brovkin et al. 2012). More specifically,
- 71 the chemical composition of litter is important in determining its decomposition in any
- 72 given environment (Coûteaux et al. 1995) with low carbon: nitrogen and high cellulose:

- 73 lignin content favoring faster decomposition (Melillo et al. 1989). The decomposition
- of litter can be highly dependent on the interaction between litter species identity and
- 75 the decomposer environment (Freschet et al. 2012, Keiser et al. 2014). Understanding
- 76 the decomposition of different litter types in relevant contrasting environments will give
- insight into how litter decomposition may be altered under future global change.
- 78 Empetrum nigrum is widespread across arctic and alpine tundras of Fennoscandia and
- boreal forests across Eurasia (Bell and Tallis 1973, Tybirk et al. 2000, Büntgen et al.
- 80 2014). Decomposition of E. nigrum leaf litter is very slow due to its production of
- allelopathic compounds (Wardle et al. 1998, Gallet et al. 1999) and high concentrations
- of the lipid polymer cutin, which is particularly slow to break down (Tegelaar et al.
- 1989, Rasse et al. 2005) as a result of a well-developed waxy cuticle (Bliss 1962,
- Hetherington et al. 1984). In addition, its physical structure (small, needle-like leaves
- with low specific leaf area (Tybirk et al. 2000, Kleyer et al. 2008, Kattge et al. 2011)),
- so is also likely contribute to slow decomposition in the field. By contrast, leaf litter of
- 87 deciduous shrubs and trees decomposes faster than that of evergreen species such as *E*.
- 88 nigrum (Aerts et al. 2006, Cornwell et al. 2008, McLaren et al. 2017). Litter inputs are
- also known to stimulate the decomposition of SOM (Subke et al. 2004), in particular,
- 90 high quality litter inputs from deciduous boreal systems are linked to faster
- 91 biogeochemical cycling and lower soil carbon stocks than evergreen systems (Melvin et
- al. 2015). A replacement of ericaceous evergreen species with deciduous shrubs and
- 93 forests could thus stimulate litter decomposition and eventually higher turnover of
- 94 SOM.
- 95 Previous work at the arctic tree-line has found that local site characteristics –
- 96 specifically, the presence or absence of forest cover exerted the strongest control on
- 97 the decomposition of *B. pubescens* leaf litter, with higher rates of decomposition in

birch forests than nearby tundra heaths (Sjögersten and Wookey 2004). This vegetation contrast was apparently more important than differences in regional climate (in contrast to the findings of other studies; Dorrepaal et al., 2005; Cornelissen et al., 2007) and experimental warming. The authors hypothesized that litter moisture in the birch forest was important in enhancing decomposition rates, but other abiotic factors such as deeper snow cover and therefore warmer winter soils and more active microbial communities (Grogan and Jonasson 2006, Blok et al. 2016) could also contribute to this. Contrasting decomposition rates between forest and tundra sites may therefore reflect the combined influence of several factors, both biotic and abiotic, the disentangling of which remains challenging. Saprotrophic fungi that grow in litter horizons of forest floors have the capacity to degrade a large range of simple and complex plant-derived structural molecules and are therefore key to the decomposition of litter (Hatakka 1994, Rytioja et al. 2014, Talbot et al. 2015). Decomposition in tundra soils, by contrast, may be under different controls, where strong environmental pressure, such as low temperature (Robinson 2001) and a 'closed' C and N cycle dominated by ericoid mycorrhizal fungi (Read and Perez-Moreno 2003), may restrict the growth and activity of other fungi. A comparison of the components of soil C in forest and tundra heath supports this view, showing that tundra has a more 'labile' signature, with more poorly-decomposed, cellulose-related fractions than the soil of mountain birch forest (Sjögersten et al. 2003). This would suggest that there is less fungal activity in the tundra, especially that of 'brown-rot' fungi which target cellulose as their primary energy source (Talbot et al. 2015). An expansion of forests could result in increased metabolism of previously poorly-decomposed litter should the appropriate decomposer community become present.

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122	Using a decomposition experiment whereby litter from the dominant species of three
123	important vegetation types (forest, shrub and tundra heath) was reciprocally
124	transplanted across a sub-arctic tree-line, we aimed to understand the key drivers of
125	decomposition rates in this ecosystem. We tested the following specific hypotheses:

- Litter from the more productive vegetation types (forest and shrub) decomposes at
 the fastest rates, regardless of the local soil environment;
- The forest and shrub environments are more favorable than tundra heath for the
 decomposition of all litter types, irrespective of origin;
- 3. Deep winter snow and associated soil microclimates, which are characteristic of
 forest and shrub environments, increase litter decomposition compared to heath
 environments.

1344. *Materials and methods*

Sites description

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The study area spans a 2 km², permafrost-free landscape around the sub-arctic/alpine 136 tree-line at Nissunsnuohkki (Abisko area, Sweden; ca. 68°18'N 18°49' E, ~600 m asl). 137 The tree-line is formed by mountain birch (Betula pubescens Ehrh. ssp czerepanovii 138 139 (Orlova) Hämet Ahti), with an ericaceous understorey, and the ecotone typically comprises of a thick layer of shrub vegetation before transitioning to tundra heath 140 141 dominated by Empetrum nigrum L. ssp hermaphroditum (Hagerup) Böcher and 142 Vaccinium vitis-idaea L. The intermediate shrub zone is dominated by Betula nana L. and grey willow (Salix) species (typically Salix glauca, often accompanied by Salix 143 144 lanata; other Salix spp., including S. hastata and S. lapponum, occur less frequently). Soil pH in the organic horizon is 4.5 ± 0.1 at forest and 4.3 ± 0.1 at heath locations in 145 146 the Abisko area (Table 1). Twelve independent, short (<100 m) transects were 147 established across the multiple forest patches in the tree-line study area. Transect 148 lengths ranged from 52 to 97 m depending on the sharpness of the forest - heath ecotone 149 transition. The soils at all sites are well-drained (Sjögersten and Wookey 2002) with 150 standing-water only observable for a short number of days every year at snow melt (Parker, Personal Observation). Care was taken to select vegetation transitions that were 151 not influenced by local topography, for example where water and snow accumulation 152 due to dips and hollows dominate site conditions, and avoiding steep slopes (mean 153 elevation change from heath to forest plots of 2.7 m). For more details on study sites, 154 155 see Parker et al. (2015). Three plots (approximately 2 m²) were established along each transect in order to 156 represent the transition in vegetation from heath to forest. These were designated: 157

tundra heath (H), shrub (S) and forest (F) (see Table 1 for further plot details). H plots were chosen for an open heath environment with low *B. nana* cover and a low canopy height, and with vegetation dominated by *E. nigrum*. S plots were identified as areas dominated by *B. nana* with shrub height characteristically between 40 and 60 cm. F plots were chosen to be in areas dominated by *B. pubescens*, approximately 10 to 15 m inside the forest edge.

Snow fences and snow depth measurements

Five replicate 3.5 m wide, 1.5 m high snow fences were erected on tundra heath sites between 0.1 and 1 km north of the transect sites (Fig. S1). They were erected before snowfall in 2012 and in 2013 (and lowered during the summer to avoid shading the vegetation and influencing evapotranspiration), and designed to create snow drifts of comparable depth to the typical seasonal snow-cover at F and S plots on the transects. To replicate the snow at F plots, plots were set up 2 m to the leeward side of the fence, 7 m for the S plots and 20 m for the H plots (no extra snow). Snow depths were measured at both snow fence and transect plots, once each between 14/3 and 29/3 in 2013 and between 29/3 and 30/3 in 2014. At each of the transects, snow depth was recorded at five points taken within 1.5 m of the logged position of the litter bags (the horizontal accuracy of the GPS unit was 3 m). At the snow fences this was not necessary due to the exact known location of the litter bags under the snow, and one measurement was taken per plot. The snow fence treatment that replicated shrub snow depths increased snow depths by 17 cm (compared to 19 cm in the shrub sites). The snow fence plots that replicated snow found in the forests increased snow depth by 55 cm (compared to 46 cm in the forest sites (Table 1)).

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Litter bags

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Litter was collected from four different transects at the Abisko study site from 183 2/9/2012- 12/9/2012. Freshly fallen B. pubescens and B. nana litter was collected from 184 the top of the litter layer, taking care to exclude older litter (which was easily 185 identified). E. nigrum litter was collected by carefully removing senesced leaves from 186 the stem of extracted *Empetrum* shoots. Only recently senesced leaves were taken (light 187 brown colour, 2-4 years old according to growth scars). Litter was collected from the 188 189 'home' plots in which each species is dominant; i.e. B. pubescens from F plots, B. nana 190 from S plots, and E. nigrum from H plots. All litter was sorted to remove any adhering particles or litter from other species, and air dried at 40°C for 72 hours. For each 191 species, 0.5 ± 0.01 g of litter was weighed into 7 x 7 cm polyester mesh bags with a 0.3 192 193 mm mesh size and heat sealed. Note that the relatively small mesh size required to 194 contain the E. nigrum litter will exclude many soil and litter fauna. All litter bags were placed in the field on 17/9/2012. Six bags of each species were placed at every plot on 195 196 all 12 transects and at snow fences. Care was taken to ensure that every bag had good 197 contact with the L horizon at each plot. Two corners of each bag were fastened to the 198 ground using stainless steel pins and all bags were tied with nylon thread to nearby vegetation. Bags were also deployed in the same manner on the leeward side of the 199 200 snow fences. Ten additional 0.5-g samples of each species were oven dried at 60°C for 72 hours, and the mass of undecomposed litter at the initiation of field emplacement 201 202 was corrected according to the residual moisture of air-dried litter. On 13/6/2013 (269 days of incubation), 24/7/2013 (310 days), 16/9/2013 (365 days), 203 204 20/6/2014 (641 days) and 18/10/2015 (1126 days) one litter bag of each species (one to 205 two on the final harvest, see later text)) at each plot at both transect and snow fence sites was retrieved from the field and oven dried at 60°C for 72 hours. Once ingrown 206

vegetation was removed, the remaining litter was extracted, weighed, and percentage mass remaining calculated. Due to the duration of field emplacement (>3 years) some litter bags were lost or disturbed (9.8 %); at the final harvest, if two bags were remaining at a plot and both bags were not damaged, a mean percentage remaining of the two was calculated.

Solid state CPMAS ¹³C NMR

Five samples of *B. pubescens* and *E. nigrum* in either the H or F sites at the 641-day harvest were taken forward for solid state ¹³C nuclear magnetic resonance CPMAS ¹³C NMR (cross-polarization/magic angle spinning ¹³C nuclear magnetic resonance spectroscopy) and elemental (C and N) analysis. Samples were randomly selected within each of the four groups Samples were randomly selected from a pool of 12 samples within each of the four groups (species (*B. pubescens, E. nigrum*) and site (Forest, Heath) combinations). For both species, three randomly selected undecomposed litter samples (from a pool of ten undecomposed samples at the beginning of the experiment) were taken forward for CPMAS ¹³C NMR. This totalled 26 samples taken for CPMAS ¹³C NMR. *Betula pubescens* and *E. nigrum* was selected for the for CPMAS ¹³C NMR analysis as they had the most contrasting decomposition rates.

CPMAS ¹³C NMR spectra were obtained using a Bruker Avance 300 spectrometer (Bruker Analytik GmbH, Rheinstetten, Germany). 2500 scans were obtained from

(Bruker Analytik GmbH, Rheinstetten, Germany). 2500 scans were obtained from approximately 0.25 g of ball-milled leaf material of each sample, packed into a cylindrical zirconia rotor with approximately 0.02 g Tetrakis (trimethylsilyl) silane (TKS) packed on top and sealed with a Bruker Kel-F drive cap (Bruker Analytik GmbH, Rheinstetten, Germany). The scanning parameters were as follows: 200 MHz

frequency, 1000 ms contact time, 1.5 s relaxation time, 5500 Hz spinning speed, and line broadening of 50 Hz. Chemical shift values were obtained compared to TKS. Total signal intensities from NMR spectra were integrated into eight major chemical shift regions (Table 3).

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FTIR-NMR spectra transformation

Diffuse reflectance Fourier transform infrared (FTIR) spectroscopy in combination with multivariate statistical techniques represents a robust and low-cost way of predicting major properties of various materials including NMR-observed chemistry (Forouzangohar et al. 2015). We applied FTIR spectroscopy to build a predictive model from the 26 samples with NMR spectra. This model was later used to predict change in litter organic chemistry for the final harvest. For these 26 samples, FTIR spectra were acquired on a Bruker Vertex 70 (Bruker Optics, Billerica, MA, USA) equipped with a wide-range Si beam splitter and mid infrared detector with Csl windows and a Pike Autodiff (Pike Technologies, Madison, WI USA) diffuse reflectance accessory for finely ground samples from undecomposed and 641-day harvests which already had associated NMR spectra (n = 26), as well as on 20 samples from the 1126-day harvest that did not have associated NMR spectra. Consistent with the sample selection for NMR, 5 replicates of each treatment were randomly selected from the 1126-day harvest (n = 20). Spectra were acquired on finely ground material over 6000-180 cm⁻¹ with a resolution of 4 cm⁻¹. For each sample, 60 scans were collected and averaged using the OPUS software package (Bruker Optics) and then corrected for background signal (average of 60 scans) and transformed into absorbance spectra.

The acquired FTIR spectra were truncated to 4000-630 cm⁻¹ and normalized using the standard normal variation (SNV) transformation. A partial least squares regression (PLSR) analysis was used to predict the eight major NMR chemical shift regions on the 26 samples that had associated NMR data. Given the small sample size (n = 26), a full cross-validation procedure was used. The PLSR analysis was able to produce good 5factor models for the dominant chemical shift regions, with less reliability for the regions with only minor contributions (Table S1). These models were then used to predict the signal intensity in each chemical shift region, along with prediction errors (De Vries & Ter Braak, 1995), for the unknown samples that decomposed for 1126 days in the field. All data processing and analysis was performed using the Unscrambler X software (CAMO Software AS, Oslo Norway). To aid in the interpretation of the ¹³C NMR data, the distribution of signal intensity from each of the chemical shift regions (Table 3) at each time point (undecomposed, 641-day, 1126-day) was used in a molecular mixing model (Baldock et al. 2004) which calculates the best linear fit of the distribution of NMR signal intensity of five major biochemical components (carbohydrates, protein, lignin, lipids and carboxyl C). After analysis by CPMAS ¹³C NMR (undecomposed and 641-day), samples were separated from TKS, ensuring no contamination of the sample, and were analysed for carbon and nitrogen content after combustion in a Vario EL Cube elemental analyser (Elementar, Hanau, Germany). After FTIR analysis, the 1126-day samples were analysed for carbon and nitrogen content using a Flash 2000 CN analyser (Thermo Scientific, Waltham, MA, USA). The carbon content data were then applied to the actual mass of the litter remaining and estimated fractions of C components to calculate the mass of carbon remaining in each component.

Statistical analysis

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Decay constants (*k*) were calculated for the loss of litter mass of every replicate species and site combination on both the snow fence and natural transect experiments according to the negative exponential litter decay model

 $ln (M_t/M_0) = -kt$ equation (1)

where M_0 is the initial dry mass of the sample and M_t is the mass at time t (years). The first two harvests (269 days and 310 days) were omitted for this calculation because they do not fit the long-term exponential decay model as a result of low mass loss in the first winter. Differences in k between site (heath, shrub and forest (or snow level in the case of the snow fence experiment)) and species (E. nigrum, B. nana and B. pubescens) were compared using a linear mixed effects model in the 'nlme' package (Pinheiro et al. 2017) of the R statistical software (R Development Core Team 2016). In the linear mixed effects model, 'Transect' was expressed as a random intercept factor due to unquantified baseline differences in decomposition between transects. The interaction between 'site' and 'species' was found not to be statistically significant in the original model (P = 0.64) and was therefore removed from the analysis (Crawley 2007). Pairwise comparisons of decomposition rates between different levels of species and site types were carried out by comparing Least-Square means derived from the statistical models with a Tukey HSD test.

The mass remaining and the percentage of undecomposed samples remaining of carbohydrates, lipids and lignin estimated from NMR spectra were analysed using a three-way ANOVA with time, site (heath and forest) and species (*B. pubescens* and *E. nigrum*) as treatment effects. The percentage data were arcsin-square root transformed

prior to analysis. All analyses were carried out using R v3.3.1. (R Development Core

303 Team 2016).

304 Results

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Litter decomposition rate

- Decomposition rates differed significantly between species on both the natural transects 306 (P < 0.001, Table 2) and at the snow fence experiment (P < 0.001, Table 2). Betula 307 pubescens, with an average decomposition constant of 0.25 year⁻¹ across all sites, 308 decomposed significantly faster than both B. nana (0.18 year⁻¹ (P < 0.001)) and E. 309 nigrum (0.15 year⁻¹ (P < 0.001)) (Fig. 1a), B. nana decomposed faster than E. nigrum (P310 311 = 0.0018). The host site (in which litter was decomposing) was also highly significantly related to decomposition rates in the litter transplant experiment (P < 0.001, Fig. 1a, 312 313 Table 2). On average, across litter types, litter decomposed marginally faster in the forest (decomposition constant = 0.21 year^{-1}) than in the shrub sites (0.20 year^{-1}) (P =314 0.06) and heath sites (0.18 year⁻¹ (P < 0.001)). Overall, decomposition was faster in the 315 shrub sites than the heath sites (P = 0.011). There was no effect of different snow 316 treatments on litter decomposition rates in the snow fence experiment (P = 0.9 Fig. 1b, 317 Table 2). At the end of the experiment, B. pubescens in the forest and shrub plots had 318 319 the least mass remaining (51 % each (Fig. 2)) and E. nigrum in the heath had the most (71 % (Fig. 2)). 320
- 321 ¹³C NMR and carbon components of litter
- Prior to decomposition, *E. nigrum* and *B. pubescens* differed substantially in the relative contributions of different regions of their NMR spectra, with *E. nigrum* dominated by alkyl-containing compounds and *B. pubescens* dominated by O- alkyl-containing

compounds (Table 3). These initial proportional differences in NMR spectra were still apparent after litter had decomposed after 641 and 1126 days in the field (Table 3). The proportion of O-alkyl compounds in both litter types reduced through time whilst alkyls remained stable as a proportion of the litter remaining in both litter types, resulting in an increase in Alkyl:O-alkyl ratio (Table 3). The C:N ratio of fresh B. pubescens litter was (60.8) under half of that measured in E. nigrum (138.3). Over time the C:N ratio decreased rapidly for both litter types, especially in the forest plots where C:N ratio at the end of the experiment reduced to 23.6 and 50.8 in for B. pubescens and E. nigrum respectively (compared to 31.9 and 64.3 at the heath plots (Table 3)). Prior to decomposition, litter from *B. pubescens* contained 1.7 times more carbohydrate-C than E. nigrum, whereas E. nigrum had 4.9 times more lipid-C in its biomass compared to B. pubescens. Amounts of lignin were similar between the litter types (Fig. 3). After incubation in the field, there was a highly significant effect of site (F = 28, P < 0.001 (Table S2)) and species of litter (F = 26, P < 0.001 (Table S2)) on the mass of carbohydrates remaining in litter, whereby this mass was lower in litter decomposing in forest plots and B. pubescens contained higher amounts of carbohydrates than E. nigrum, respectively (Fig. 3a). In the forest, litter carbohydrates initially decomposed rapidly between 0 and 614 days, and then stabilized at approximately 40 % (B. pubescens, Fig. S3a) and 50 % (E. nigrum, Fig. S3a), after which there was only marginal mass loss (Fig. 3). In contrast, the decomposition of litter carbohydrates in the heath followed a more linear pattern, with slower decomposition to 614 days, which then continued to 1126 days. The final percentage mass remaining of carbohydrates of both B. pubescens (49 %) and E. nigrum (54 %) at the end of the experiment in the heath was within 10 % and 6 %, respectively, of the litter in the forest, despite slower initial decomposition rates (Fig. S3a).

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Due to very high alkyl-C contents in E. nigrum litter, the mass of lipids modelled to be present in this litter was also very high (Fig. 3b), resulting in a highly significant relationship between species type and mass of lipids in extracted litter samples (F = 690, P < 0.001). There was also a strong effect of site on mass of lipids, with lower amounts remaining in both E. nigrum and B. pubescens at the forest plots (F = 15, P <0.001 (Table S2)). When expressed as a proportion of the original lipid mass, the results show a strong effect of 'species' (F = 18, P < 0.001 (Table S2)) and 'site' (F = 12, P =0.002 (Table S2)); B. pubescens had 60 % of lipid mass remaining in the forest and 70 % in the heath, whereas E. nigrum had 82 % remaining in the forest and 96 % in the heath (Fig. S3b). Lignin was present in low amounts in litter (Fig. 3c) and there were no significant differences in mass of lignin remaining over the study duration between site (F = 0.4, P= 0.5 (Table S2)) or species (F = 0.0003, P = 0.98 (Table S2)), but there was a significant decline in mass with time (F = 11, P = 0.002 (Table S2)). Although initial amounts of lignin were low (Fig. 3c), it decomposed in all species-site treatments to about 50 % of its original amount (Fig. S3c)

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Discussion

The greater decomposition rates of *B. pubescens* and *B. nana* than *E. nigrum* regardless of decomposition environment clearly support the first hypothesis that litter from an arctic tree and shrub species decomposes at a faster rate than the typical heath species, *E. nigrum*. This difference is consistent with the differences in C stocks in the environments that these species dominate respectively i.e. low C stocks in forest and high C stocks in tundra heath (Hartley et al. 2012, Parker et al. 2015).

Litter of E. nigrum, a key species of tundra heaths, decomposed very slowly. This is likely due to high levels of aliphatic compounds (alkyls) which make up the lipids of its waxy cuticle (Bliss 1962, Hetherington et al. 1984). Lipid levels in E. nigrum litter were over four times higher than in B. pubescens, and showed very low rates of mass loss, especially in the tundra heath environment. Whilst our methods cannot distinguish between plant- vs. microbe-derived alkyls (Baldock et al. 1997), it is clear that these compounds are contributing substantially to the persistence of E. nigrum litter in this experiment. The strong contribution of lipids to long-term SOC storage in tundra heaths is also corroborated by the components of C found in the SOM of ericaceous tundra around Abisko (Sjögersten et al. 2003), which also contained high levels of alkyls. This link between aliphatic compounds in E. nigrum litter and a resulting alkyl signature in the soil has also been found in Norwegian tundra heath systems (Väisänen et al. 2015), emphasizing that this could be a significant driver of high SOM storage in tundra. Although we could not explicitly address the potential role of the physical structure of the litter studied here, it is important in determining decomposition rates (Cornelissen et al. 1999). E. nigrum has a far lower surface area: mass ratio (Specific leaf area) than the Betula species used in this study (Kleyer et al. 2008) which may render the substrate more immediately available to decomposer communities. In contrast to E. nigrum, B. pubescens lost substantial mass in the initial stages of decomposition. The measurements of remaining carbon suggest that this initial rapid decomposition was due to the metabolism and breakdown of the initially high levels of carbohydrates (predominately O-alkyls). This loss of carbohydrates is a likely contributing factor to rapid turnover of C and ultimately low storage of C in the soil in deciduous arctic and boreal ecosystems (Parker et al. 2015, Melvin et al. 2015). Carbohydrates in B. pubescens litter decomposed to a similarly low residual level in the

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tundra as in the forest, even though their initial decomposition was not as rapid. This supports the hypothesis that litter identity is central to its eventual decomposability (Coûteaux et al. 1995, Cornelissen et al. 2007), irrespective of *in situ* processing rates. We also examined the decomposition rates of leaf litter from B. nana, a shrub species which has been observed to be expanding its range over arctic tundra in response to climate change (Tape et al. 2006, Myers-Smith et al. 2011). This litter also lost significantly more mass than E. nigrum, and observations of high soil C flux from these shrub systems (Parker et al. 2015) may in part be explained by this more rapid leaf litter turnover. However, B. nana decomposed at slower rates than B. pubescens, which could be due to a number of factors including differences in specific leaf area (a facet of physical structure; note earlier paragraph), N content and structural C compounds. Indeed, with regards to the litter chemistry Väisänen et al. (2015) reported carbohydrate concentration of 39 % and alkyl to O-alkyl ratio around 0.51 indicating that the intermediate decomposition rates of *B. nana* may be attributed to its intermediate levels of carbohydrates (Väisänen et al. 2015). Based on our observed species-specific decomposition rates, any expansion of B. pubescens forests is likely to increase leaf litter decomposition in tundra to a greater extent than an expansion of B. nana, but both are likely to increase C cycling rates if only PFT (deciduous) of the litter input is considered. The second overarching hypothesis of this study, that litter would decompose fastest in the forest and shrub environments compared with the heath, was supported by the majority of the data, with the exception of the shrub B. nana. Our snow fence experiment gives some insight into separating the influence of abiotic (snow depth, temperature and, potentially, moisture) effects on decomposition from the confounding biological factors (i.e. vegetation/microbial). There were no increases in litter loss with

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increased winter snow depth over the 2 years of study, concurring with findings of another study in arctic tundra (DeMarco et al. 2014) but not those of Blok et al. (2016). As the experimentally manipulated snow depth did not influence decomposition rates, we must conclude that the naturally deep snow cover was not the driver behind the rapid decomposition which we observed in the forest. We however, do not rule out a longer-term effect of many years of snow cover on microbial communities and resulting decomposition rates. Litter moisture is an abiotic factor that we could not take directly into account in the present study. It is known to be important in controlling microbial activity and litter turnover in boreal forests (Schimel et al. 1999), and low surface moisture in heath ecosystems has been implicated in slowing decomposition (Sjögersten and Wookey 2004). We acknowledge that there are abiotic controls other than snow depth that we have not accounted for, but conclude that the major differences in decomposition that we observe along the tree-line are due to microbial and biochemical differences. We propose that the rapid decomposition of carbohydrate rich litter in the forest was driven by two interlinked processes: Firstly, there is a rich and active fungal community (especially brown-rot fungi) in the litter horizons of the forest (Lindahl et al. 2007) capable of producing an array of enzymes that can target initially available celluloserelated structures (Talbot et al. 2015) until this source of C is depleted. Secondly, there is a biochemically favorable environment that 'primes' the decomposition of cellulose in the forest plots due, in part, to the high-cellulose content of previous litter-falls. Temperature (Pietikainen et al. 2005) and pH (Rousk and Bååth 2011) are important in determining fungal and bacterial growth rates, but soil pH and thaw-season soil temperature is remarkably similar across the study ecotones (Table 1). This leaves the biochemical environment as a key remaining factor explaining why fungi may grow

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449 well in the birch forests. Experimental additions of cellulose have been found to 450 increase fungal growth (Subke et al. 2004, Meidute et al. 2008) and enzyme production (Talbot and Treseder 2012). Thus, it is feasible that in the mountain birch forests in the 451 452 present study there are tight linkages between the carbohydrate rich litter, increased fungal activity and rapid turnover of C (Parker et al. 2015). 453 The production of allelopathic compounds by E. nigrum is a process that can have 454 ecosystem-wide influence (Wardle et al. 1998). Production of poly-phenolic secondary 455 compounds by E. nigrum has been linked to inhibited activity of soil fungi and animals 456 and as a result lowered decomposition rates and increased build-up of SOM (Wardle et 457 458 al. 1998, Tybirk et al. 2000). Slow decomposition rates of E. nigrum in the present study may partially be due to remaining residues of allelopathic compounds on the litter 459 and in the surrounding litter in the heath. However, it should be noted that the forest 460 461 sites also have high cover of E. nigrum across the understory (Parker et al. 2015) yet carbon turnover is very high compared with the heath. Although assessing the 462 importance of allelopathy across the sub-arctic tree-line is not in the scope of this work, 463 464 it may have important controls over decomposition. 465 Betula pubescens litter in the forest plots decomposed to half of its original mass within 18 months, with limited further mass loss for the remainder of the time in the field. This 466 467 is consistent with observations that the most labile components of litter are decomposed 468 initially, whilst remaining litter residue starts to form soil organic matter (Melillo et al. 469 1989, Sjögersten and Wookey 2004). This prompts the question; how is carbon 470 processed after this initial mass loss, bearing in mind that standing stocks of soil organic matter are very low in these forests (Hartley et al. 2012, Parker et al. 2015)? In 471 boreal forests, ectomycorrhizal fungi (EMF) grow in the organic and mineral horizons 472 473 below the litter (Lindahl et al. 2007) and have been shown to be able to stimulate

474 decomposition of macromolecular complexes through the production of extracellular 475 enzymes, specifically, peroxidases (Bödeker et al. 2014, Lindahl and Tunlid 2015). Although other pathways are plausible, we propose that the decomposition of litter in 476 477 this forest ecosystem is characterized by an initial rapid mass loss due to metabolism by saprotrophic fungi and bacteria of relatively simple organic molecules e.g. 478 carbohydrates, and a subsequent steadier decomposition by EMF of the remaining more 479 480 complex compounds. Taken together, this could result in a thin organic soil horizon and low net C storage in the ecosystem (Hartley et al. 2012). 481 482 This study has shown that litter of a common tundra heath species, E. nigrum, 483 decomposes faster in forest or shrub environments than in tundra heath, and that this decomposition will be driven in the first instance by carbohydrate loss. As forests are 484 expanding in range and cover in some areas of the sub-Arctic (Tømmervik et al. 2009, 485 486 Rundqvist et al. 2011, Hofgaard et al. 2013) and shrubs have been observed be increasing in community dominance in many locations across the arctic tundra (Tape et 487 al. 2006, Myers-Smith et al. 2011), the findings of the current study have important 488 implications for the future of arctic C stocks. If tundra heath soils, rich in less-489 decomposed forms of C (Sjögersten et al. 2003), are colonized by deciduous forest, 490 491 with its associated fungal community (including EMF which are also potentially efficient decomposers (Lindahl and Tunlid 2015)), then this C will be rapidly 492 493 metabolized and a significant part of the C currently stored in tundra heath will be released to the atmosphere. This would represent a positive feedback to climate 494 495 warming. In conclusion, the dominant litter types across the forest-heath ecotone decomposed 496 faster of litter in the most productive ecosystems. We hypothesize that this is due to a 497 498 carbohydrate-rich input of litter from the birch canopy and the presence of a

decomposer community that can metabolize this relatively labile source of C. Using a snow fence experiment on tundra soils, we show that the effect of increased snow in the forest compared to the heath alone is modest and that the effect of environment on decomposition rates in the forest is likely exerted via microbial metabolism over the summer. We raise the hypothesis that microbially-accessible litter C from tundra heath species is vulnerable to decomposition should more productive deciduous species further expand onto heaths, resulting potentially in a net emission of CO_2 to the atmosphere.

Acknowledgements

This work was supported by NERC (UK Natural Environment Research Council) research Studentship training grant NE/J500434/1. The research leading to these results has also received funding from the European Union Seventh Framework Programme [FP7/2007-2013] under grant agreement n° 262693 [INTERACT]. We would like to thank staff at Abisko Scientific Research Station warmly for hosting us and for supporting this work. Thanks to Duncan Cameron and Gareth Phoenix for support at the University of Sheffield. We thank Karina Clemmensen and Björn Lindahl for constructive comments on a previous version of this paper. Thanks to two anonymous reviewers for constructive comments.

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Table 1: Site characteristics along transects at Abisko (means ± 1 SEM, n = 12).

'Canopy height' refers to the actual vegetation canopy for Heath, and Shrub communities, and the understorey of the Forest (where mountain birch trees - *Betula pubescens* - comprise the canopy). Snow depths measured over transects are paired in either 2013 or 2014 with snow depth data from the snow fence experiment, at plots which were selected to mimic snow depth along the transect. Vegetation and soil data (except temperature data) are adapted from Parker et al. (2015). Soil temperature data are average seasonal temperatures at 5 cm depth across six of the twelve transects. The start of each season is defined by soil temperatures deviating and remaining above (Summer) or below (Winter) 0 °C.

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	Year	Property	Heath	Shrub	Forest		
		Distance from Heath (m)		28.3 ± 2.9	67.6 ± 5.9		
		Canopy height (cm)	14.7 ± 0.7	32.0 ± 2.4	19.0 ± 1.7		
Vegetation		B. pubescens density (trees ha ⁻¹)			785.0 ± 109.0		
		B. nana cover (%)	21.2 ± 2.7	60.3 ± 4.8	8.0 ± 2.2		
		E. nigrum cover (%)	65.4 ± 3.3	66.9 ± 4.7	45.4 ± 4.2		
		pH (organic horizon)	4.3 ± 0.1	4.4 ± 0.1	4.5 ± 0.1		
		Organic horizon carbon (kg m ⁻²)	7.0 ± 0.8	3.0 ± 0.5	2.0 ± 0.3		
		Mineral horizon carbon (kg m ⁻²)	2.0 ± 0.3	3.3 ± 1.3	2.5 ± 0.4		
Soil	2012-13	Summer temperature (°C)	5.4 ± 0.3	5.1 ± 0.3	5.5 ± 0.2		
	2012-13	Winter temperature (°C)	-3.9 ± 0.2	-1.3 ± 0.2	-1.1 ± 0.2		
	2013-14	Summer temperature (°C)	6.6 ± 0.3	6.6 ± 0.6	7.1 ± 0.2		
	2013-14	Winter temperature (°C)	-2.5 ± 0.5	-1.0 ± 0.1	-0.2 ± 0.1		
2012-13		Snow depth at transects	13.1 ± 1.8	35.4 ± 4.0	46.8 ± 3.4		
Charr	2012-13	Snow depth at snow fences (cm)	13.9 ± 2.2	22.6 ± 2.9	58.5 ± 13.3		
Snow	2012 14	Snow depth at transects	14.4 ± 3.5	29.7 ± 5.3	72.2 ± 9.1		
	2013-14	Snow depth at snow fences (cm)	13.0 ± 1.5	39.0 ± 8.7	78.2 ± 10.4		

Table 2: The effect of species of litter and incubation site on decomposition rate (k) on the natural transects ('Site' represents differences both in abiotic factors (e.g. snow cover, thermal and moisture regimes) and biotic factors e.g. microbial community and others)) and at the snow fences (where 'Environment' initially represents differences in abiotic factors associated with altered snow only).

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Natural Transects										
Factor	d.f.	F	P							
Species	2,89	94.4	< 0.001							
Site	2,89	13.3	< 0.001							

Snow fence experiment										
Factor	d.f.	F	P							
Species	2,36	86.9	< 0.001							
Snow	2,36	0.2	0.9							

Table 3: Percentage contributions of chemical shift regions to ¹³C NMR spectra, Alkyl:

O-Alkyl ratios and C:N ratios of litter samples of *Betula pubescens* and *Empetrum*nigrum that were decomposing in forest or heath environments at 0 days

(undecomposed), 614 days and 1126 days. Error values signify ± 1 SEM (n = 5 for decomposed field samples, n = 3 for undecomposed samples).

		0 days 641 days							1126 days						
				F	orest	į	J	Heat	h]	Fore	st	I	I eatl	1
	Me	an ±	SE	Me	an± S	SE	Me	ean±	SE	Me	ean±	SE	Me	an±	SE
Betula pubescens															
Alkyl (0-45 ppm)	15.5	±	0.3	20.9	±	1.2	18.5	±	0.2	20.8	±	1.4	25.8	±	7.3
N-Alkyl/Methoxyl (45-60 ppm)	5.1	±	0.1	6.6	±	0.6	6.2	±	0.1	6.6	±	0.1	6.6	±	0.2
O-Alkyl (60-95 ppm)	47.6	±	0.9	38.3	±	1.6	45.7	±	0.7	38.3	±	1.0	40.2	±	4.8
Di-O-Alkyl (95-110 ppm)	11.3	±	0.2	8.7	±	0.5	10.4	±	0.1	8.7	±	0.3	8.9	±	1.2
Aryl (110-145 ppm)	11.1	±	0.8	11.4	±	0.8	9.6	±	0.3	11.3	±	0.4	9.4	±	0.6
O-Aryl (145-165 ppm)	4.2	±	0.2	4.5	±	0.8	3.6	±	0.3	4.7	±	0.4	3.1	±	0.5
Amide/Carboxyl (165-190 ppm)	5.1	±	0.3	9.5	±	1.9	6.0	±	0.4	9.7	±	0.7	5.9	±	1.1
Alkyl/O-Alkyl	0.3	±	0.0	0.5	±	0.0	0.4	±	0.0	0.5	±	0.0	0.8	±	0.4
C:N	60.8	±	4.3	31.5	±	1.9	49.7	±	0.9	23.6	±	1.3	31.9	±	3.2
Empetrum nigrum															
Alkyl (0-45 ppm)	43.9	±	1.0	50.3	±	1.7	51.6	±	1.3	52.3	±	2.3	54.4	±	0.9
N-Alkyl/Methoxyl (45-60 ppm)	4.7	±	0.2	5.0	±	0.3	5.5	±	0.2	6.0	±	0.1	6.0	±	0.1
O-Alkyl (60-95 ppm)	26.9	±	1.0	21.4	±	1.1	24.8	±	0.6	21.3	±	1.7	21.7	±	0.5
Di-O-Alkyl (95-110 ppm)	6.2	±	0.1	4.8	±	0.4	5.0	±	0.1	4.4	±	0.5	4.4	±	0.2
Aryl (110-145 ppm)	9.9	±	0.1	9.6	±	0.3	7.9	±	0.4	8.7	±	0.2	7.9	±	0.1
O-Aryl (145-165 ppm)	3.9	±	0.3	4.0	±	0.4	2.4	±	0.3	2.9	±	0.1	2.4	±	0.1
Amide/Carboxyl (165-190 ppm)	4.4	±	0.3	4.9	±	0.5	2.8	±	0.6	4.3	±	0.1	3.2	±	0.2
Alkyl/O-Alkyl	1.6	±	0.1	2.4	±	0.2	2.1	±	0.1	2.6	±	0.4	2.5	±	0.1
C:N	138.3	±	3.0	74.6	±	4.5	111.6	±	5.0	50.8	±	3.9	64.3	±	3.1

Figure Legends

Figure 1: Decomposition constants (k) of E. nigrum, B. nana and B. pubescens litter across (a) transects across natural tree-lines from heath to forest and (b) under three different snow depths simulating snow accumulation found at different vegetation types: Heath (control), + Snow (Shrub) and ++ Snow (Forest). Error bars represent \pm 1 SEM (transects n = 12, snowfences n = 5).

Figure 2: Percentage mass remaining of litter over time of three different species: (a,d) *Empetrum nigrum*, (b,e) *Betula nana*, (c,f) *Betula pubescens* in either distinct vegetation communities (heath, shrub or forest), distributed across natural transects (a,b,c), or under three different snow depths simulating snow accumulation found at different vegetation types: Heath (control), + Snow (Shrub) and ++ Snow (Forest) (d,e,f). Error bars represent \pm 1 SEM (transects n = 12, snowfences n = 5). The extent of the shaded areas on the x axis indicates the length of the snow covered season in the first two years of study.

Figure 3: Mass of (a) Carbohydrates, (b) Lipids and (c) Lignin of in *Betula pubescens* (grey diamonds) and *Empetrum nigrum* litter (black squares) in forest (open shapes) and heath (closed shapes) environments at initial levels (0 days), and after 614 and 1126 days of decomposition (t5). Error bars represent \pm 1 SEM (initial litter: n = 3, decomposed samples: n = 5). Bold lettering in the inset text indicates significant (*P* <0.05) factors and interactions in three way analysis of variance; number of asterisks indicate level of significance: *** P < 0.001, ** P < 0.01, * P < 0.05, see table S2 for further statistics relating to these data.





