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RESEARCH PAPER

Natural variation in *Arabidopsis thaliana* reveals shoot ionome, biomass, and gene expression changes as biomarkers for zinc deficiency tolerance

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Abstract

Zinc (Zn) is an essential nutrient for plants, with a crucial role as a cofactor for many enzymes. Approximately one-third of the global arable land area is Zn deficient, leading to reduced crop yield and quality. To improve crop tolerance to Zn deficiency, it is important to understand the mechanisms plants have adopted to tolerate suboptimal Zn supply. In this study, physiological and molecular aspects of traits related to Zn deficiency tolerance were examined in a panel of 19 *Arabidopsis thaliana* accessions. Accessions showed a larger variation for shoot biomass than for Zn concentration, indicating that they have different requirements for their minimal Zn concentration required for growth. Accessions with a higher tolerance to Zn deficiency showed an increased expression of the Zn deficiency-responsive genes ZIP4 and IRT3 in comparison with Zn deficiency-sensitive accessions. Changes in the shoot ionome, as a result of the Zn treatment of the plants, were used to build a multinomial logistic regression model able to distinguish plants regarding their Zn nutritional status. This set of biomarkers, reflecting the *A. thaliana* response to Zn deficiency and Zn deficiency tolerance, can be useful for future studies aiming to improve the performance and Zn status of crop plants grown under suboptimal Zn concentrations.

Key words: Biofortification, biomarker, mineral concentration, plant ionome, shoot growth, zinc usage index.

Introduction

Zinc (Zn) is an essential micronutrient required for plant growth and development. Many agricultural soils in the Middle East, India, and parts of Australia, America, and Central Asia are Zn deficient, often due to poor Zn availability caused by high pH in calcareous soils. Zn-deficient soils affect both crop yield and quality, and can also result in human malnutrition through the intake of food containing low concentrations of Zn and other micronutrients (Cakmak, 2007; Alloway, 2009). The World Health Organization (WHO) and the Food and Agriculture Organization (FAO) of the United Nations estimate that about one-third of the world’s population suffers from some form of Zn deficiency.
Plants exposed to Zn deficiency show reduced growth. Severe deficiency results in extensive leaf chlorosis, wilting, stunting, leaf curling, and reduced root elongation, while mild stress results in chlorosis in young leaves and early senescence of older leaves (Marschner, 1995). In *Arabidopsis thaliana*, all of these symptoms, as well as delayed flowering, are observed when plants are grown under Zn deficiency (Talukdar and Aarts, 2007). Zn deficiency also affects the function of enzymes such as copper/zinc superoxide dismutase (Cu/Zn SOD) and carbonic anhydrase (CA), leading to an accumulation of reactive oxygen species (ROS), which cause oxidative damage and a reduction in photosynthesis (Clemens, 2010; Ibarra-Laclette et al., 2013).

The threshold Zn concentration below which plants are considered to be Zn deficient is ~15–20 μg g⁻¹ dry biomass. This can vary from species to species and between plants of the same species (Marschner, 1995; White and Broadley, 2011). The ability of a plant to grow and yield under Zn-limiting conditions compared with ideal growth conditions is defined as Zn efficiency. It is based on the difference in relative growth or yield between plants grown under control and Zn-deficient conditions. Another parameter used is the Zn usage index (ZnUI), which quantifies the amount of dry matter produced per mg of Zn in the plant. The ZnUI is useful for the comparison of plant genotypes which do not show significant differences in Zn concentration, but differ in biomass production under Zn deficiency (Siddiqi and Glass, 1981; Cakmak et al., 1998; Good et al., 2004; Genc et al., 2006).

To avoid problems associated with inappropriate Zn supply, plants have developed an efficient homeostasis mechanism. Different genes act in the uptake of Zn from soil, distribution over different organs, tissues, cells, and organelles, and (re)mobilization through the plant, to control Zn homeostasis (Sinclair and Krämer, 2012). While the actual Zn deficiency sensor is not yet known, the Zn deficiency response in *A. thaliana* seems to start with the activation of the transcription factors bZIP19 and bZIP23, the function of which is essential for plants to survive Zn deficiency (van de Mortel et al., 2006; Assunção et al., 2013).

Zn is among the essential elements which make up the plant ionome (Salt et al., 2008). Previous studies have shown that the plant ionome profile reflects the physiological state of plants under various genetic, developmental, and environmental backgrounds, and can be used as a biomarker for a particular physiological condition (Huang and Salt, 2016). Ionome-based biomarker models have been used to determine differences in the plant nutritional status among large sets of different genotypes and experimental batches (Baxter et al., 2008a). Natural variation for the concentration of elements comprising the plant ionome has been studied in *A. thaliana*, revealing important mineral homeostasis mechanisms in plants (Rus et al., 2006; Loudet et al., 2007; Baxter et al., 2008a, 2010; Kobayashi et al., 2008; Morrissey et al., 2009; Chao et al., 2012; Pineau et al., 2012; Koprivova et al., 2013).

To improve efficiently the performance of crops grown under suboptimal Zn conditions and increase the Zn content in their edible parts, it is of paramount importance to understand the physiological and molecular mechanisms underlying plants’ tolerance to Zn deficiency. Aspects of natural variation for Zn deficiency tolerance have been described for several plant species, including *A. thaliana* (Graham et al., 1992; Rengel and Graham, 1996; Cakmak et al., 1998; Hacisalihoglu et al., 2004; Genc et al., 2006; Ghandilyan et al., 2012; Karim et al., 2012). However, to date, a detailed study on natural variation of plants’ tolerance to Zn deficiency involving both physiological and molecular mechanisms has not been performed. In this study, we evaluated natural variation among 19 diverse *A. thaliana* accessions to identify physiological and molecular traits involved in the tolerance to Zn deficiency. It shows that high-throughput screening of genetic variation for Zn deficiency tolerance can be simplified by focusing on the combination of changes in the ionome profile, the minimum Zn concentration required for growth, and the expression level of Zn deficiency-responsive genes.

## Materials and methods

### Plant material and hydroponic growth

A set of 19 *A. thaliana* accessions was selected based on their diverse site of origin (Supplementary Table S1 at JXB online). Seeds were surface-sterilized with chlorine vapour and sown in Petri dishes on wet filter paper followed by a 4 d stratification treatment at 4 °C in the dark, to promote uniform germination. Seeds were transplanted to 0.5% (w/v) agar-filled tubes, the bottoms of which were cut off, and placed in a modified half-strength Hoagland nutrient solution for hydroponic growth (Assunção et al., 2003): 3 mM KNO₃, 2 mM Ca(NO₃)₂, 1 mM NH₄H₂PO₄, 0.5 mM MgSO₄, 1 μM KCl, 25 μM H₂BO₃, 2 μM MnSO₄, 0.1 μM CuSO₄, 0.1 μM (NH₄)₆Mo₇O₂₄·4H₂O, and 20 mM Fe(II)-EDTA. The solution pH was set at 5.5 using KOH and buffered with 2 mM MES. Plants were grown hydroponically in two experiments performed separately. In experiment one, referred to as the mild Zn deficiency experiment, we compared plants grown for 41 d under control (2 μM ZnSO₄) and mild Zn deficiency (0.05 μM ZnSO₄). In experiment two, referred to as the severe Zn deficiency experiment, we compared plants grown for 31 d under control (2 μM ZnSO₄) and severe Zn deficiency (no Zn added). Plants were grown in a climate-controlled chamber set at 70% relative humidity, with a 12 h day (120 μmol photons m⁻² s⁻¹) and 20 °C/15 °C day/night temperatures. The hydroponic system consisted of plastic trays (46 × 31 × 8 cm) holding 9 litres of nutrient solution, covered with a non-translucent 5 mm thick plastic lid with evenly spaced holes in a 7 × 10 format holding the agar-filled tubes with plantlets. The nutrient solution was refreshed once a week. Shoot fresh weight (SFW) was measured in all samples during harvest.

Shoot fresh weight (SFW) was measured in all samples during harvest. Some samples were immediately frozen in liquid nitrogen and stored at −80 °C for gene expression and element concentration analysis. The shoot dry weight (SDW) of these samples was calculated based on a fresh weight/dry weight correction factor obtained from additional plants which were dried for 72 h at 60 °C. For each trait, the treatment versus control relative values were determined as Rel_trait=(trait Zn deficiency/trait control)×100. The ZnUI was calculated based on the following formula:

\[
\text{ZnUI} = \frac{\text{shoot biomass (mg)}}{\text{shoot Zn concentration (ppm)}}
\]
For each treatment, the shoot ionome profile was determined for five biological replicates of each *Arabidopsis thaliana* accession. Samples were first dried for 72 h at 60 °C, transferred to 96-well plates with tubes containing one 5 mm glass bead, and homogenized for 5 min at 30 Hz with a Qiagen 96-well plate mixer mill. A 2–4 mg aliquot of leaf tissue was transferred to Pyrex test tubes (16 × 100 mm) and digested with 0.9 ml of concentrated nitric acid (Baker Instrafreeze Analyzed; Avantor Performance Materials; http://www.avantormaterials.com) for 5 h at 115 °C. Samples were diluted to 10 ml with 18.2 MQ cm Milli-Q water. Elemental analyses were performed with inductively coupled plasma mass spectrometry, ICP-MS (Elan DRC II; PerkinElmer, http://www.perkinelmer.com) for Li, B, Na, Mg, P, S, K, Ca, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Pb, Sr, Mo, and Cd. A reference, composed of pooled samples of digested leaf material, was prepared and included every ninth sample in all sets of 70 samples to correct for variation between and within ICP-MS analysis runs. Seven samples from each sample set were weighed and used during the iterative weight normalization process to estimate the weight of the remaining 63 samples from the set (Danku et al., 2013). The following elements were not added to the nutrient solution: Li, Co, Ni, As, Se, Pb, Sr, and Cd and, except for Cd, their concentrations are not shown.

Gene expression

Gene expression analysis was performed for eight accessions with different ZnUI values selected from the 19 accessions grown under mild Zn deficiency conditions. Frozen leaf material from plants grown under mild and severe Zn deficiency and their respective control treatments was used, in three biological replicates, each consisting of material from three plants. Total RNA was extracted using the method of Onate-Sanchez and Vicente-Carbajosa (2008). cDNA was synthesized from 1 μg of total RNA using the iScript cDNA synthesis kit from BioRad as per the manufacturer’s instructions. Following synthesis, cDNA was diluted 10-fold. Quantitative real-time PCRs (qRT-PCRs) were performed in triplicate with iQ SYBR Green Supermix (BioRad) using an iQ Real Time PCR machine (BioRad). Relative transcript levels of selected genes were determined by qRT-PCR. The expression of *IRT3* (At1g60960), *ZIP3* (At2g32270), *ZIP4* (At1g10970), *bZIP19* (At4g35040), *CSD2* (At2g28190), and *CA2* (At5g14740) was measured. The oligonucleotides used for each gene are shown in Supplementary Table S2. Amplicon lengths were between 80 bp and 120 bp, and all primer combinations had at least 95% efficiency. Reaction volumes were 10 μl (5 μl SYBR green qPCR mix, 300 nmol of each primer, and 4 μl of cDNA template). Cycling parameters were 4 min at 95 °C, then 40 cycles of 15 s at 95 °C and 30 s at 55 °C. Gene expression values were normalized to the reference genes *PXE4* (At5g25760), *SAND* (At2g28390), and 18S. Gene expression levels relative to the average of the reference genes for each accession under mild and severe Zn deficiency and their respective control treatments were calculated based on ΔCT values. Gene expression levels of accessions exposed to mild and severe Zn deficiency, relative to their respective control treatment, were calculated based on ΔΔCT values (Livak and Schmittgen, 2001).

Statistical analysis

For all shoot traits and gene expression levels relative to reference gene expression, a two-way ANOVA was performed to test for significant differences between treatments, accessions, and the interaction between treatments and accessions. A one-way ANOVA was performed to test for significant differences between accessions for relative gene expression values, relative change in SDW, Zn concentration, and Zn content. A one-way ANOVA was also performed to test for significant differences in element concentrations between the four treatments used (mild and severe Zn deficiency and their respective controls). Element concentration values were log10-transformed and a Benjamini–Hochberg correction of the *P*-values was performed. When significant differences were found, a Tukey's HSD post-hoc test with a significance level of 0.05 was performed. Broad-sense heritability (*H*²) was calculated as the ratio between estimated genetic variance and total phenotypic variance (Kruijer et al., 2015).

Results

**Natural variation in Zn deficiency response for physiological and morphological traits**

*Arabidopsis thaliana* accessions were grown hydroponically under control conditions (2 μM ZnSO₄) and either mild (0.05 μM ZnSO₄) or severe Zn deficiency (no Zn added). After 31 d of exposure to severe Zn deficiency, plants showed clear deficiency symptoms compared with plants in the control treatment. This was primarily visible as reduced growth, leaf curling, and the presence of chlorotic and necrotic spots (Fig. 1A, B). After 31 d in the mild Zn deficiency treatment, accessions did not show any sign of Zn deficiency, hence they were grown for an additional 10 d. Even then, only a few accessions had visible symptoms of Zn deficiency, mainly slight chlorosis in leaves and reduced growth (Fig. 1C, D), confirming that the treatment was indeed mild.

Accessions showed significant phenotypic variation for most traits analysed which varied according to the trait and Zn treatment (Supplementary Tables S3, S5). Plants in the severe Zn deficiency treatment had shoot Zn concentrations...
close to the reported minimum required for growth, which is \( \sim 15-20 \mu g g^{-1} \) dry biomass (Marschner, 1995). Shoot Zn concentrations under mild Zn deficiency were approximately twice as high than under severe Zn deficiency (Fig. 2). In addition, plants in the mild Zn deficiency experiment had a higher SDW than plants in the severe Zn deficiency experiment, as they were grown for 10 d longer. From all shoot traits, only Zn concentration was significantly correlated between the controls of the two Zn deficiency experiments, indicating that during the additional 10 d of growth between experiments, other factors such as the growth rate of accessions affected their shoot biomass and Zn content in a different manner (Supplementary Table S8). Accession Cvi-0 had to be excluded from further analysis as it had established poorly and too many plants were lost, especially from the mild Zn deficiency experiment.

In both Zn deficiency treatments, most accessions showed reduced SDW relative to their respective control treatments, while few had a higher SDW and apparently were not affected by the reduced Zn supply (Fig. 3A, B). Nearly all accessions had a reduction in shoot Zn concentration of \( \sim 80\% \) in both Zn deficiency treatments relative to their respective controls (Fig. 3C, D). Also, accessions with high shoot Zn concentrations were not always among the ones with a high shoot total Zn content, due to differences in SDW. Tsu-0, Col-0, and Mt-0 were the best performing accessions under mild Zn deficiency in terms of having similar Zn concentrations to the other accessions and higher SDW across the Zn deficiency and control treatments. Thus, these accessions seem to be able to maintain growth under Zn deficiency, albeit with some reduction in shoot Zn concentration. Conversely, Pa-2, C24, and Li-5:2 performed poorly under mild Zn deficiency, with a strong reduction in growth in comparison with the other accessions though with a small reduction in shoot Zn concentrations in both Zn deficiency treatments. These accessions appear to have a poor ability to take up Zn under both control and Zn-deficient conditions, which results in a limited capacity to grow and to maintain cellular Zn levels. Only accession Bor-4 showed an increase in SDW under severe Zn deficiency relative to its control treatment, even though this was not statistically different from most of the other accessions (Fig. 3A; Supplementary Table S8). Bor-4 also showed an increase in SDW under mild Zn deficiency, as did Shah. However, it is important to note that these two accessions were among the ones with the lowest SDW in their respective control treatments, which could explain their lower sensitivity to Zn deficiency.

Fig. 1. Comparison of A. thaliana accessions grown under control and severe or mild Zn-deficient conditions. Representative examples of A. thaliana accessions grown in hydroponic medium under Zn-sufficient control conditions (2 µM ZnSO\(_4\)) (A and C) or severe (no Zn added) (B) and mild Zn deficiency (0.05 µM ZnSO\(_4\)) (D). Plants in (A) and (B) are grown for 31 d; plants in (C) and (D) are grown for 41 d. Accessions from left to right in rows from top to bottom: C24, Per-1, Tsu-0, Mc-0, Hau-0, Mt-0, Shah, Kas-2, Bor-4, Wag-3, Ocs-1, Pa-2, Li-5:2, Ge-0, Can-0, Var 2-1, Ler-1, Cvi-0, Bur-0, and Col-0. Scale bars indicate 2 cm.
Accessions with contrasting tolerance to Zn deficiency show differences in the expression of Zn deficiency-responsive genes

The ZnUI was used to determine the amount of biomass produced per unit of tissue Zn concentration (Fig. 4). In accordance with the results previously shown for SDW and Zn concentration, the accessions Mt-0 and Tsu-0 had the highest ZnUI values for both Zn deficiency treatments, and C24 and Pa-2 had the lowest values. Even though only in the mild Zn deficiency treatment, these accessions had significantly higher or lower ZnUI values when compared with the other accessions (Supplementary Table S6). Eight accessions with different ZnUI values in the mild Zn deficiency treatment were then selected to examine whether natural variation for Zn deficiency tolerance is reflected at the gene expression level. Mild Zn deficiency was favoured over the severe treatment as the variation between accessions for SDW was larger in the mild treatment. In addition, mild Zn deficiencies are more likely to be found in nature. The accessions Tsu-0 and Col-0 had high ZnIU values, accessions Ge-0, Bur-0, and Can-0 were intermediate, and Pa-2, C24, and Per-1 had low ZnIU values. Accessions with higher ZnUI values were considered to be more tolerant to Zn deficiency (Fig. 4).

The expression of six genes involved in the plant Zn deficiency and oxidative stress response was determined in Zn deficiency-tolerant and -sensitive accessions (Fig. 5; Supplementary Fig. S1). bZIP19 encodes one of the two redundant bZIP transcription factors which control the Zn deficiency response in A. thaliana. We also looked at the expression of the IRT3, ZIP4, and ZIP3 transcriptional targets genes of bZIP19, all encoding ZIP-like Zn transport proteins, strongly induced following Zn deficiency (Assunção et al., 2010). The expression of the CSD2 gene, encoding a Cu/Zn SOD which needs Zn as a structural component to function (Sharma et al., 2004), and the CA2 gene, encoding a carbonic anhydrase (CA) requiring Zn as cofactor, was also determined. CSD2 is needed for detoxification of superoxide radicals, while CA2 facilitates the diffusion of CO2 through the liquid phase of the cell to the chloroplast, important for photosynthesis (Randall and Bouma, 1973; Li et al., 2013). Both CSD2 and CA2 are expected to decrease in expression under Zn deficiency exposure due to the reduced concentration of Zn in the cells (Ibarra-Laclette et al., 2013).

There was a significant effect of both the mild and severe Zn deficiency treatments on the expression level of most studied genes. The exceptions were bZIP19 and CA2 in the severe Zn deficiency treatment (Supplementary Table S3). The Zn deficiency-responsive genes IRT3, ZIP4, and ZIP3 were up-regulated in all accessions under both Zn deficiency treatments, confirming that the plants sensed Zn deficiency (Fig. 5; Supplementary Fig. S1). ZIP4 and IRT3 in particular were in general more highly expressed in the more Zn deficiency-tolerant accessions than in the more Zn deficiency-sensitive accessions, with Tsu-0 especially showing strong induction of these genes under severe Zn deficiency. The expression of ZIP3, which is predominantly expressed in roots (van de Mortel et al., 2006), is the least prominent of the three Zn transporter genes in shoots. The expression levels of CSD2 and CA2 were generally low and variable in both Zn deficiency treatments, but these genes are down-regulated especially under mild Zn deficiency. The Zn deficiency-tolerant accessions Ge-0 and Bur-0 had the highest induction of the CA2 and CSD2 genes under severe Zn deficiency (Fig. 5). Significant accession by treatment interaction was found for all genes tested, except for bZIP19, in at least one of the Zn deficiency experiments (Supplementary Table S3), indicating that gene expression differences between the response of accessions to Zn deficiency are pronounced.

To understand further the relationship between the expression levels of Zn deficiency-responsive genes and...
Zn deficiency tolerance traits, a correlation analysis was performed. Under severe Zn deficiency, we found a significant positive correlation between the expression levels of \( IRT3 \) and \( CSD2 \) with ZnUI and of \( ZIP4 \) with SFW (Supplementary Table S9).

Zn deficiency affects the shoot ionomic profile of \( A. \) thaliana accessions

The shoot ionome of the 19 \( A. \) thaliana accessions was then determined. Box plots of the combined results per element showed a substantial variation between treatments for almost all the elements measured (Fig. 6; Supplementary Table S7). Significant differences between treatments were observed for Zn, Mg, Mo, Cu, and Cd concentrations in both the mild and severe Zn deficiency experiments. B, Na, and Ca concentrations were significantly different between treatments only in the mild Zn deficiency experiment, and Mn and Fe concentrations only in the severe Zn deficiency experiment. When comparing Zn concentrations across the four treatments, there was a significant difference between severe and mild deficiency but not between their respective control treatments. \( H^2 \) values were calculated to estimate the genetic contribution to the observed phenotypic variation (Table 1). \( H^2 \) values were generally higher in the mild compared with the severe Zn deficiency experiment and in plants exposed to Zn deficiency in comparison with their control treatments. The heritability for ZnUI was highest in the mild Zn deficiency treatment, suggesting that under those conditions a large part of the observed variation is due to genetic differences between accessions. Fe concentration had the lowest heritability in both control treatments, whereas Mo concentration had the highest heritability across the treatments. Even though the Zn concentrations of plants grown under severe Zn deficiency were very low, there was substantial heritability for both Zn concentration and Zn content, with values of 0.49 and 0.41, respectively, indicating that the minimal Zn concentration/content levels are subject to genetic variation.
Classification of the plant Zn deficiency state using multinomial logistic regression

The univariate model (i.e. with a single element as the only predictor) performed poorly as a predictor of plant nutritional status, for most elements, and often mistakenly identified plants under Zn deficiency as being control (Table 2). As expected, only the Zn concentration was able to separate the three classes very well, with prediction accuracies ranging from 0.92 for the plants under severe Zn deficiency to 0.99 for the control plants. Cu also had a good prediction performance for severe Zn deficiency, while Ca was the only element (apart from Zn) that identified a substantial number of the plants under mild Zn deficiency (Table 2). Mg, Mn, Fe, and Mo performed only marginally well, having some ability to identify plants under severe and mild Zn deficiency. For the other elements (B, P, S, and K), the univariate model performed no better than a naïve classifier that would always predict control conditions.

The penalized multivariate model, fitted on all elements except Zn, performed similarly to the univariate model fitted with Zn alone, the latter having a higher accuracy for the controls and mild Zn deficiency treatments and less for the severe Zn-deficient plants (Table 2).

Discussion

The natural variation in the response of A. thaliana to two levels of Zn deficiency was examined, with a focus on physiological and molecular traits. Analysis of genetic variation indicated that: (i) accessions vary for the minimum Zn requirement for growth; (ii) tolerance to Zn deficiency seems to be related to an increased expression of genes encoding Zn transmembrane transporter proteins (ZIP4 and IRT3); and (iii) Zn deficiency results in changes in the plant ionome which can be used as a biomarker to predict the plant’s physiological condition.

Natural variation of growth and Zn concentration in response to severe and mild Zn deficiencies

The tested A. thaliana accessions showed substantial diversity for all traits studied in both Zn deficiency experiments (Figs 1–3). Extreme accessions were identified for all traits, confirming the existing large natural variation in the A.
Fig. 5. Normalized gene expression levels of bZIP19, IRT3, ZIP3, ZIP4, CSD2, and CA2 in rosette leaves of eight A. thaliana accessions under Zn deficiency (Zn–) and control treatments (Zn+ control) in the severe (left) and mild Zn deficiency experiments (right). Accessions are ranked from left to right according to decreasing Zn usage index values under mild Zn deficiency (see Fig. 4). Plants were grown in hydroponic medium under Zn-sufficient control conditions (2 μM ZnSO₄) and severe (no Zn added) or mild Zn deficiency (0.05 μM ZnSO₄), for 31 d or 41 d, respectively. Error bars represent the SE; one-way ANOVA and pairwise comparisons between accessions are provided in Supplementary Tables S3 and S4.
The mild Zn deficiency treatment is more suitable to reveal genetic variation underlying a plant’s response to Zn nutrition, with higher heritability for most traits in comparison with the other treatments (Table 1). The disadvantage of using this mild treatment was that plants were 10 d older than in the severe Zn deficiency treatment, resulting in the initiation of flowering in some accessions. Such a change in development could include remobilization of minerals...
from older to younger organs (e.g. from rosette leaves to developing fruits); however, Waters and Grusak (2008) previously showed that the contribution of remobilization is probably <10% of the seed mineral content, so we considered this not much of a disturbing factor. In addition, this treatment seems better in representing Zn-deficient conditions likely to be encountered by *A. thaliana* in nature, with an average Zn concentration in leaves of 26 ppm in comparison with 18 ppm in the severe Zn deficiency treatment. To support this, Ghandilyan et al. (2012) observed leaf average Zn concentration of 40 ppm when using a Zn-deficient and nutrient-poor soil originating from Eskisehir, Central Anatolia in Turkey to grow the *A. thaliana* Ler×Cvi RIL population. Furthermore, the harshness of the severe Zn deficiency treatment seems to be beyond the genetic capacity of most accessions to tolerate, based on the extensive chlorosis displayed by nearly all accessions in this treatment and their very low average leaf Zn concentration, which was within or below the minimum Zn concentration range of 15–20 ppm required for growth as suggested by Marschner (1995).

Heritabilities of most traits were higher in the severe and mild Zn deficiency treatments than in their respective controls, further supporting the observed large genetic variation for all traits in response to the Zn deficiency treatments. Contrary to these observations, Ghandilyan et al. (2012) reported lower heritability values for shoot biomass and most element concentrations in *A. thaliana* plants grown in Zn-deficient soil compared with control conditions. Yet, other studies show that heritabilities for the same trait can change according to the growth conditions (Ghandilyan et al., 2009; Richard et al., 2011; Baxter et al., 2012), hence the importance of taking heritability into account when selecting growth conditions most amenable to detect genetic variation for a specific trait.

The control treatments of the two Zn deficiency experiments were significantly correlated with respect to the Zn concentration, but not for SDW and Zn content (Supplementary Table S8). This is probably due to differences in growth rate between the *A. thaliana* accessions during the additional 10 d of growth in the mild Zn deficiency experiment. Previous studies have shown that growth rate is highly variable among plants, being affected by both internal and external factors such as developmental processes and environmental conditions (El-Lithy et al., 2004; Zhang et al., 2012). Differences in growth rate between accessions in the mild and severe Zn deficiency experiments are probably caused by differences in the initiation of flowering. Most accessions in the control treatment of the mild Zn deficiency experiment were flowering or bolting at the harvesting day; while only three accessions, of the control treatment, were flowering in the severe Zn deficiency experiment at the harvesting day, which was 10 d earlier than in the mild Zn deficiency experiment (Fig. 1A, C).
Table 1. Broad-sense heritability ($H^2$) values for the traits measured in A. thaliana accessions grown under severe and mild Zn deficiency and their respective Zn sufficiency (control) conditions

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<td>0.45</td>
<td>0.59</td>
<td>0.53</td>
<td>0.58</td>
</tr>
<tr>
<td>[K]</td>
<td>0.51</td>
<td>0.65</td>
<td>0.46</td>
<td>0.48</td>
</tr>
<tr>
<td>[Ca]</td>
<td>0.72</td>
<td>0.69</td>
<td>0.42</td>
<td>0.52</td>
</tr>
</tbody>
</table>

SFW, shoot fresh weight (g); SDW, shoot dry weight (mg); ZnUI, Zn usage index; S2ZnC, shoot total Zn content (µg); and [X], mineral element concentrations (µg g$^{-1}$ DW).

Table 2. Estimated prediction performance values for elements used in the logistic regression model to predict plant nutritional Zn status

<table>
<thead>
<tr>
<th>Univariate models</th>
<th>Control</th>
<th>Zn deficiency</th>
<th>Average</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Severe</td>
<td>Mild</td>
</tr>
<tr>
<td>B</td>
<td>0.914</td>
<td>0.002</td>
<td>0.0837</td>
</tr>
<tr>
<td>Mg</td>
<td>0.901</td>
<td>0.364</td>
<td>0.5442</td>
</tr>
<tr>
<td>P</td>
<td>1</td>
<td>0</td>
<td>0.6905</td>
</tr>
<tr>
<td>S</td>
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<td>0</td>
<td>0.0061</td>
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<tr>
<td>K</td>
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<td>0</td>
<td>0.0102</td>
</tr>
<tr>
<td>Ca</td>
<td>0.949</td>
<td>0</td>
<td>0.2673</td>
</tr>
<tr>
<td>Mn</td>
<td>0.882</td>
<td>0.412</td>
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<tr>
<td>Fe</td>
<td>0.977</td>
<td>0.278</td>
<td>0.5608</td>
</tr>
<tr>
<td>Cu</td>
<td>0.877</td>
<td>0.716</td>
<td>0.6276</td>
</tr>
<tr>
<td>Zn</td>
<td>0.996</td>
<td>0.92</td>
<td>0.9857</td>
</tr>
<tr>
<td>Mo</td>
<td>0.911</td>
<td>0.204</td>
<td>0.509</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Multivariate models</th>
<th>All elements except Zn</th>
<th>All elements</th>
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<tr>
<td></td>
<td>0.8738</td>
<td>0.7750</td>
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<td>0.6596</td>
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<td></td>
<td>0.9681</td>
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</table>

Physiological and molecular mechanisms of Zn deficiency tolerance in A. thaliana

Arabidopsis thaliana accessions showed a larger variation for relative change in SDW than in Zn concentration under both Zn deficiency treatments (Fig. 3). This indicates the presence of genetic variation for their minimum Zn requirement and for the ability to tolerate low Zn concentrations. This is not unique for A. thaliana though. Also for barley, bread and durum wheat, common bean, and rice, different genotypes are reported to have similar shoot Zn concentrations with different levels of Zn deficiency tolerance (Cakmak et al., 1998; Rengel, 2001; Gene et al., 2002; Hacisalihoglu et al., 2003; Wissuwa et al., 2006; Sadeghzadeh et al., 2009). Further indications that A. thaliana accessions vary for the minimum Zn requirement is shown by a few accessions with slightly higher SDW in the Zn-deficient treatment relative to its control.

The ability to enhance the root Zn uptake and the root to shoot Zn transport are among the proposed mechanisms underlying tolerance to Zn deficiency (Rengel, 2001; Broadley et al., 2007), but the Zn deficiency signal may come from shoots. Indeed, accessions considered tolerant to Zn deficiency had a higher expression of the Zn deficiency-responsive genes ZIP4 and IRT3 in shoots (Fig. 5). These genes, encoding Zn transmembrane transporters (Grotz et al., 1998), are transcriptionally responsive to Zn deficiency and mainly expressed in roots, but are also expressed in shoot tissue in response to low Zn, suggesting a role in both Zn uptake and distribution (Lin et al., 2009; Jain et al., 2013).

Our findings indicate that higher tolerance to Zn deficiency may be the result of an increased, or more efficient, shoot Zn re-allocation capacity, and that natural variation for it may reflect variation in the expression of these and other Zn transport genes in A. thaliana.

Previous studies have shown that tolerance to Zn deficiency can also be affected by the plant’s capacity to deal with the high levels of ROS produced under low Zn conditions (Rengel, 2001; Sinclair and Krämer, 2012). In this study, a relationship was found between the expression of CA2 and ZnUI (Supplementary Table S9). Further studies examining the ability of plants to tolerate ROS under Zn deficiency and other mechanisms not included in this study, but thought to contribute to tolerance to Zn stress, will be useful for a more complete understanding of the mechanisms involved in plant tolerance to Zn deficiency (Cakmak et al., 1996; Rengel, 2001; Gao et al., 2005; Gene et al., 2006; Hoffland et al., 2006; Wissuwa et al., 2006; Chen et al., 2009; Impa et al., 2013a, b).

This should include examining the ability of plants to increase the bioavailability of Zn$^{2+}$ ions in the soil, to improve the root system architecture to scavenge larger soil volumes, and a more efficient utilization, compartmentalization, and remobilization of Zn.

Model to predict Zn deficiency status based on the concentration of other elements

Exposing A. thaliana plants to different levels of Zn deficiency also affects the homeostasis of other elements, which made it possible to develop an MLR model able to predict the Zn deficiency status of a plant based on changes in other elements (Table 2). This approach is analogous to the model used by Baxter et al. (2008b) to predict the physiological status of A. thaliana plants exposed to Fe or P deficiency. Contrary to the MLR model developed for Zn deficiency, Baxter et al. (2008b) found that changes in Fe concentration alone had no power to detect Fe deficiency, and detection was
Conclusion

This study demonstrates that several physiological and molecular mechanisms underlie differences in Zn deficiency tolerance in *A. thaliana*. These include the minimum Zn concentration required for growth and the ability to take up and translocate Zn by inducing the expression of Zn deficiency-responsive genes. ZnUI, the reduction in SDW, and the expression level of Zn deficiency-responsive genes, such as ZIP4 and IRT3, are useful proxies to evaluate plant tolerance to Zn deficiency in future studies. A mild Zn deficiency condition is more amenable for genetic studies than a severe stress, with higher heritability values for most studied traits and providing a more natural condition, at least for *A. thaliana*. Finally, the shoot ionome profile is a useful predictor of the plant Zn deficiency status. Changes in Zn concentration alone or in combination with other elements have an excellent capacity to detect physiological plant Zn deficiency in the absence of other visible symptoms. While we have shown this now for *A. thaliana*, a model plant species, the application of our findings will be in crops. For crop species, the application of our findings will be in crops.

Acknowledgements

We gratefully acknowledge Maarten Koornneef for his critical comments on earlier versions of this manuscript. During the review and publication process of this manuscript Dr John Danku died. He will be greatly missed by his family, friends and colleagues and his untimely death is a significant loss to the field of ionomics. This work was supported by the Centre for BioSystems Genomics and grant 93512008 of the ZonMWZentrum program, both initiatives under the auspices of the Netherlands Genomics Initiative, and by the EU-COST Action FA0905.

References


**Supplementary data**

Supplementary data are available at *JXB* online.

Table S1. Detailed information of the *A. thaliana* accessions used in this study.

Table S2. Oligonucleotide PCR primer sequences.

Table S3. Two-way ANOVA of shoot dry weight, Zn concentration, total Zn content, and gene expression of *A. thaliana* accessions exposed to control and Zn deficiency.

Table S4. Tukey pairwise multiple comparison between accessions for shoot dry weight, shoot Zn concentration, shoot Zn content, and gene expression levels (relative to reference genes) in the severe and mild Zn deficiency experiments.

Table S5. One-way ANOVA of differences between *A. thaliana* accessions exposed to control and Zn deficiency for relative changes in shoot dry weight, Zn concentration, total Zn content, Zn usage index, and gene expression.

Table S6. Tukey pairwise multiple comparison between accessions for relative change in shoot dry weight, shoot Zn concentration, shoot Zn content, and gene expression values in the severe and mild Zn deficiency treatments relative to their respective control treatments.

Table S7. One-way ANOVA of the log10-transformed shoot element concentrations to determine significant differences between treatments.

Table S8. Pearson correlation coefficients for the comparison of shoot traits measured in the 19 *A. thaliana* accessions grown under severe or mild Zn deficiency and their respective control treatments.

Table S9. Pearson correlation coefficients for the comparison of gene expression with shoot traits measured in eight *A. thaliana* accessions grown under severe or mild Zn deficiency and their respective control treatments.

Fig. S1. Relative gene expression in leaves of eight *A. thaliana* accessions grown under severe and mild Zn deficiency, compared with their respective control treatments.


Oñate-Sánchez L, Vicente-Carbajosa J. 2008. DNA-free RNA isolation protocols for Arabidopsis thaliana, including seeds and siliques. BMC Research Notes 1, 93.


