

25 **Abstract:**

26 Caveolae regulate many cardiovascular functions and thus could be of interest in relation to
27 pre-eclampsia, a pregnancy specific disorder characterised by hypertension and proteinuria.
28 We examined placental mRNA and protein expression/localisation of the caveolae
29 components Caveolin 1-3, Cavin 1-4 as well as eNOS/ iNOS in normotensive control (n=24)
30 and pre-eclamptic pregnancies (n=19). Placental mRNA expression of *caveolin-1*, *cavin 1-3*,
31 was lower and *eNOS* expression was increased in pre-eclampsia ($P<0.05$ for all). Additionally
32 Caveolin-1 protein expression was also reduced in pre-eclampsia ($P=0.007$); this could be an
33 adaptive response in pre-eclampsia, possibly to attenuate the oxidative
34 stress/inflammation.

35 **Keywords:** Hypertension; cavin; caveolin; pre-eclampsia; placenta.

36

37 Introduction

38 Pre-eclampsia is a hypertensive disorder of pregnancy. As the placenta receives no
39 autonomic input, it relies upon vasoactive mediators to regulate its vascular reactivity. Nitric
40 oxide (NO) plays an integral role in controlling vascular resistance within the placenta; a
41 disruption of this pathway has been identified in pre-eclampsia [1]. NO production is
42 catalysed by the conversion of L-arginine to NO by NO synthases (NOS), two isoforms being
43 present in the placenta: endothelial and inducible NOS (eNOS /iNOS)[2].

44 Caveolae are invaginations of the plasma membrane present in most mammalian cell types
45 [3]. Caveolins (Cav-1, Cav-2, Cav-3) and cavins (1 to 4), participate in the formation of the
46 caveolae and the coordination of the signal transduction [4]. Cavins (adapter proteins) are
47 responsible for caveolae assembly and Cav protein expression and stabilisation [5]. Four
48 isoforms of cavins have been identified (cavin-1 to 4).

49 Caveolae and Cavs, in particular *Cav-1* expressed in endothelial cells (EC), have been shown
50 to have regulatory roles in pathological angiogenesis and in vascular disease such as
51 atherosclerosis and pulmonary hypertension [6]. Studies have confirmed protein expression
52 of Cav-1 and 2 in the endothelium placental capillaries and syncytiotrophoblast in term
53 human placental tissue [7-9]. Cav-1 is an organiser of redox-sensitive signalling pathways,
54 specifically involved in reactive oxygen species (ROS)-dependent signalling events [10]. Co-
55 localisation of NADPH oxidase with eNOS in Cav-1 rich-caveolae in ECs both sustains ROS-
56 mediated activation of eNOS by Angiotensin II and simultaneously promotes eNOS
57 uncoupling.

58 We examined placental mRNA and protein expression/localisation of the caveolae
59 components and eNOS/iNOS in normotensive control and pre-eclamptic women.

60

61 **Materials and Methods**

62 Two groups of white European women (24 normotensive, 19 pre-eclamptic) were analysed;
63 detailed demographics and outcome data have previously been published [11]. The study
64 was approved by the Nottingham University Hospitals Ethics Committee (LREC-Q2090312)
65 and written, informed consent was obtained. Pre-eclampsia was stringently defined as per
66 the International Society for the Study of Hypertension in Pregnancy guidelines [12]; full
67 depth placental tissue samples were collected [11].

68 Evaluation of the mRNA expression was conducted as per previously published methods
69 [11]. Immunostaining of paraffin-embedded placental sections were performed as
70 previously described [13]. All slides were assessed by the same observers (KS-J & MRH) and
71 quantified using the Positive Pixel Algorithm of Aperio Image Scope software [13].

72

73 **Results**

74 The placental mRNA expression of *Cav-1-3*, *cavin-1-4*, *eNOS* and *iNOS* is presented in Figure
75 1a-c. *Cav-1* and *cavin-1-3* had significantly lower expression in pre-eclamptic women ($P < 0.05$
76 for all; Figs. 1a & 1b) whereas *Cav-2* and *3* and *cavin-4* were not statistically different
77 between groups ($P > 0.05$ for all). *eNOS* was increased in pre-eclampsia ($P = 0.045$; Fig. 1c)
78 but *iNOS* did not differ between groups ($P > 0.05$).

79 The protein expression and localisation in placental tissue are shown in Figure. 2.
80 Immunohistochemical staining for Cav and cavin isoforms were localised around fetal
81 vessels and fibroblasts, with staining also in syncytiotrophoblasts. Both eNOS and iNOS
82 expression was localised to the syncytiotrophoblast, with some staining in the endothelium.
83 Cav-1 placental protein expression was significantly reduced in pre-eclampsia (median [IQR]:
84 0.78 [0.73, 0.82] vs. 0.87 [0.82, 0.90] respectively; $P = 0.001$). No significant differences were
85 observed for any other proteins ($P > 0.05$).

86

87

88 **Discussion**

89 This is the first report of a detailed expression profile of Cavs and cavins, together with both
90 eNOS and iNOS in placentae from women who had pre-eclampsia. This study demonstrates
91 that the mRNA expression of *cavin-1-3* and *Cav-1* are down regulated in this tissue. *eNOS*
92 mRNA is upregulated in pre-eclamptic placentae in agreement with the literature [14]. As
93 with previous studies [15, 16], no other differences in eNOS protein expression were
94 observed between groups. The placental localisation of Cav-1 and eNOS in this study also
95 coincides with previous observations [7-9].

96 The reduction in placental Cav-1 protein expression in pre-eclamptic pregnancies may have
97 effects on eNOS uncoupling via Angiotensin type 1 receptor (AT1R). We have previously
98 reported increased placental AT1R protein expression in pre-eclampsia [17]; a partial down
99 regulation in Cav-1 could reduce eNOS uncoupling through attenuation of NADPH oxidase
100 assembly [18] and activation of eNOS in response to Angiotensin II, whilst still maintaining
101 functional eNOS at the membrane, as has been reported in ECs [19]. This could explain why
102 eNOS protein, but not mRNA expression, was unchanged between groups. Caveolae have
103 been implicated as mediators of vascular inflammation, as well as determinants of
104 intracellular redox status; the latter accomplished by facilitating the formation of ROS and
105 decreasing NO bioavailability in response to EC injury or inflammatory stimuli [20].

106 We have previously reported increased maternal Thiobarbituric acid reactive substances
107 (TBARS) [11] and placental oxidative stress markers (xanthine oxidase and NADPH oxidase)
108 [21] and reduced placental antioxidant glutathione peroxidase activities in the women with
109 pre-eclampsia [11]. The reduction of Cav-1 in pre-eclampsia could be an adaptive response,

110 independent of eNOS, to attenuate the increased oxidative stress and inflammation, as seen
111 in models of atherosclerosis [22].

112 Disruption and progressive loss of Cav-1 have been associated with pulmonary hypertension
113 outside of pregnancy [18], but detailed analysis in relation to normotensive pregnancy is
114 sparse [7-9] and lacking altogether in pre-eclampsia. In order to determine if similar
115 differences antedate the clinical onset of the disease, future longitudinal studies are needed
116 to determine whether the results are cause or effect. Examination of first and second
117 trimester placentae would enable us to trace the ontogeny of mRNA and protein expression
118 of caveolae through pregnancy.

119

120 **Acknowledgments:** We thank all the women who participated in the study and the
121 midwives and doctors whose support made this study possible. We also thank Dr.
122 Geneviève Escher and Mr Yosef Mansour for proof reading the manuscript and help with
123 images. Some of this work was funded by Tommy's Charity (Charity number: 1060508),
124 CAPES/CNPq, Brazil (MRH); CEPF is a CNq researcher and KS-J was funded by a Society for
125 Endocrinology Summer studentship.

126

127 **Conflict of Interest:** No conflict of interest for all authors.

128

129 **References**

- 130 [1] Ghabour MS, Eis AL, Brockman DE, Pollock JS and Myatt L. Immunohistochemical
131 characterization of placental nitric oxide synthase expression in preeclampsia. *Am J Obstet*
132 *Gynecol.* 1995;173(3 Pt 1):687-94.
- 133 [2] Alderton WK, Cooper CE and Knowles RG. Nitric oxide synthases: structure, function and
134 inhibition. *Biochem J.* 2001;357(Pt 3):593-615.
- 135 [3] Parton RG. Caveolae and caveolins. *Curr Opin Cell Biol.* 1996;8(4):542-8.
- 136 [4] Briand N, Dugail I and Le Lay S. Cavin proteins: New players in the caveolae field.
137 *Biochimie.* 2011;93(1):71-7.
- 138 [5] Liu L and Pilch PF. A critical role of cavin (polymerase I and transcript release factor) in
139 caveolae formation and organization. *J Biol Chem.* 2008;283(7):4314-22.
- 140 [6] Mathew R. Cell-specific dual role of caveolin-1 in pulmonary hypertension. *Pulm Med.*
141 2011;2011:573432.
- 142 [7] Byrne S, Cheent A, Dimond J, Fisher G and Ockleford CD. Immunocytochemical
143 localization of a caveolin-1 isoform in human term extra-embryonic membranes using
144 confocal laser scanning microscopy: implications for the complexity of the materno-fetal
145 junction. *Placenta.* 2001;22(6):499-510.
- 146 [8] Lyden TW, Anderson CL and Robinson JM. The endothelium but not the
147 syncytiotrophoblast of human placenta expresses caveolae. *Placenta.* 2002;23(8-9):640-52.
- 148 [9] Linton EA, Rodriguez-Linares B, Rashid-Doubell F, Ferguson DJ and Redman CW. Caveolae
149 and caveolin-1 in human term villous trophoblast. *Placenta.* 2003;24(7):745-57.
- 150 [10] Ushio-Fukai M and Alexander RW. Caveolin-dependent angiotensin II type 1 receptor
151 signaling in vascular smooth muscle. *Hypertension.* 2006;48(5):797-803.

- 152 [11] Mistry HD, Wilson V, Ramsay MM, Symonds ME and Broughton Pipkin F. Reduced
153 selenium concentrations and glutathione peroxidase activity in pre-eclamptic pregnancies.
154 Hypertension. 2008;52:881-8.
- 155 [12] Brown MA, Lindheimer MD, de Swiet M, Van Assche A and Moutquin JM. The
156 classification and diagnosis of the hypertensive disorders of pregnancy: statement from the
157 International Society for the Study of Hypertension in Pregnancy (ISSHP). Hypertens
158 Pregnancy. 2001;20(1):IX-XIV.
- 159 [13] Mistry HD, McCallum LA, Kurlak LO, Greenwood IA, Broughton Pipkin F and Tribe RM.
160 Novel expression and regulation of voltage-dependent potassium channels in placentas
161 from women with preeclampsia. Hypertension. 2011;58(3):497-504.
- 162 [14] Dotsch J, Hogen N, Nyul Z, Hanze J, Knerr I, Kirschbaum M and Rascher W. Increase of
163 endothelial nitric oxide synthase and endothelin-1 mRNA expression in human placenta
164 during gestation. Eur J Obstet Gynecol Reprod Biol. 2001;97(2):163-7.
- 165 [15] Corthorn J, Germain AA, Chacon C, Rey S, Soto GX, Figueroa CD, Muller-Esterl W, Duarte
166 I and Valdes G. Expression of kallikrein, bradykinin b2 receptor, and endothelial nitric oxide
167 synthase in placenta in normal gestation, preeclampsia, and placenta accreta. Endocrine.
168 2006;29(3):491-9.
- 169 [16] Matsubara S, Takizawa T, Takayama T, Izumi A, Watanabe T and Sato I. Immuno-
170 electron microscopic localization of endothelial nitric oxide synthase in human placental
171 terminal villous trophoblasts-normal and pre-eclamptic pregnancy. Placenta. 2001;22(8-
172 9):782-6.
- 173 [17] Mistry HD, Kurlak LO and Broughton Pipkin F. The placental renin-angiotensin system
174 and oxidative stress in pre-eclampsia. Placenta. 2013;34(2):182-6.

175 [18] Chen F, Barman S, Yu Y, Haigh S, Wang Y, Dou H, Bagi Z, Han W, Su Y and Fulton DJ.
176 Caveolin-1 is a negative regulator of NADPH oxidase-derived reactive oxygen species. *Free*
177 *Radic Biol Med.* 2014;73:201-13.

178 [19] Lobysheva I, Rath G, Sekkali B, Bouzin C, Feron O, Gallez B, Dessy C and Balligand JL.
179 Moderate caveolin-1 downregulation prevents NADPH oxidase-dependent endothelial nitric
180 oxide synthase uncoupling by angiotensin II in endothelial cells. *Arterioscler Thromb Vasc*
181 *Biol.* 2011;31(9):2098-105.

182 [20] Layne J, Majkova Z, Smart EJ, Toborek M and Hennig B. Caveolae: a regulatory platform
183 for nutritional modulation of inflammatory diseases. *J Nutr Biochem.* 2011;22(9):807-11.

184 [21] Williams PJ, Mistry HD, Innes BA, Bulmer JN and Broughton Pipkin F. Expression of
185 AT1R, AT2R and AT4R and their roles in extravillous trophoblast invasion in the human.
186 *Placenta.* 2010;31(5):448-55.

187 [22] Frank PG, Lee H, Park DS, Tandon NN, Scherer PE and Lisanti MP. Genetic ablation of
188 caveolin-1 confers protection against atherosclerosis. *Arterioscler Thromb Vasc Biol.*
189 2004;24(1):98-105.

190
191
192
193
194
195
196

197 **Figure Legends**

198 **Figure 1:** Normalised mRNA expression (copy number) of a) Cav-1-3; b) cavin-1-4 and c)
199 eNOS and iNOS in placentae from normotensive and pre-eclamptic pregnancies. Boxplots
200 represent median [interquartile range].

201 **Figure 2:** Placental protein expression and localisation of Cav-1, 2 and 3; cavin-1, 2, 3 and 4;
202 eNOS and iNOS , in normotensive control and pre-eclamptic women. Expression was
203 significantly downregulated in pre-eclamptic placentae ($P<0.05$). Positive staining was
204 localised mainly around fetal vessels (black arrows) with some weak staining in
205 syncytiotrophoblasts (red arrow).

206