New Insights into Xanthan Synergistic Interactions with Konjac glucomannan: A Novel Interaction Mechanism proposal

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Abstract

The interactions of xanthans containing precise acetate and pyruvate concentration with Konjac glucomannan (KGM) were studied at different sodium chloride and polymer concentrations. A new unified model of the interaction is proposed, taking into account previous models in the literature. This study suggests that the interactions occur by two distinct mechanisms dependent on xanthan conformation. These interactions are not mutually exclusive and may co-exist and hence produce complicated traces. Consequently two types of gel which melt at different temperature ranges can be formed. Depending on the xanthan helix coil transition temperature, one or both of the synergistic states may exist in the hydrocolloid blend. The proposed model has been tested rheologically and using Differential Scanning Calorimetry by varying salt concentration and using samples containing different functional group concentrations.
Introduction

Mixed polysaccharide gels are a current active area of research. Methods continue to be improved to control and describe the mechanism of interaction and the properties of mixed gels.

One polysaccharide which is well-known for gelation dependent on synergistic interaction with mannan based gums, is xanthan. Xanthan gum is an extracellular polysaccharide used as a food additive and rheological modifier, however it does not form true gels on its own (Ross-Murphy et al., 1983). Xanthan forms self-supporting gels in mixtures with galactomannans of low galactose content, such as locust bean gum, or with glucomannans such as konjac glucomannan (KGM). Similar to xanthan, these polysaccharides have a β-(1→4) linkage linear backbone.

Konjac glucomannan, which is used in the work here, is found in the tuber of the Amorphophallus konjac plant (40% by dry weight). This glucomannan is made-up of a linear backbone of blocks of β-(1→4)-linked mannopyranose units, which are interposed with linear β-(1→4)-linked blocks of glucopyranose units. The usual ratio of glucose and mannose in KGM lies between molar ratios of 1:1.5 and 1:1.6 with a high degree of substitution (5-10%) of the OH groups on the sugars by acetyl groups. Despite being shown to be a determinant of solubility and gelling properties, in particular the strength of the gel (Shatwell et al. 1990), the assumption was made that this property was constant throughout the experiments presented here and had negligible effect on the final conclusions.

There are many hypotheses concerning the mechanism of gelation between xanthan and galacto- and glucomannans. One of the earliest proposed mechanisms (Morris et al., 1977; Dea et al., 1977) was that the unsubstituted regions of the mannan backbone attach to the surface of the xanthan helix, i.e. interaction occurs when xanthan is in the ordered conformation. This hypothesis has been supported by further work (Mannion et al., 1992; Annable et al., 1994; Copetti et al., 1997). Annable et al.1994 in particular studied the xanthan and KGM interaction, emphasizing that the interaction occurred only after ordering of xanthan chains either in the presence or absence of electrolyte. They also suggested that in the presence of electrolyte xanthan self-association was promoted at the expense of xanthan-KGM associations.

In a different approach, Tako and Nakamura (1986), Kitamura et al. (1991) and Paradossi et al. (2002) suggested the possibility of interaction between the side chains of xanthan and unsubstituted segments of the backbone of the mannan polysaccharide.

However, it has also been suggested that the gel formation is due to intermolecular binding between the galacto/glucomannan and the disordered backbone of the xanthan molecule, rather than to the 5-fold helix surface (Cairns et al., 1986; Cairns et al., 1987; Brownsey et al., 1988; Cheetham and Mashimba, 1988; Morris, 1992).

In addition, as interactions are known to occur at room temperature i.e. well below the transition temperature (T_m) and when xanthan is in the helical conformation, it has been suggested that low amounts of the disordered form may exist among the ordered chains making the interaction with the disordered form possible at room temperature (Fitzsimons et al., 2008). In this hypothesis the xanthan order-disorder transition is an equilibrium process occurring over a significant range of temperature, and the level of disordered xanthan will depend on the temperature of the xanthan sample relative to T_m (Norton et al., 1984). Conversely, another possibility is that small amounts of the ordered form will exist among
Although the interaction, there is still controversy over whether it is disordered, the same heterotypic structure will be formed directly, as this is enthalpically more favourable than interaction with the normal 5-fold helix. This interaction may therefore be with both ordered and disordered xanthan conformations. If xanthan is in the disordered conformation the direct backbone-to-backbone binding occurs with KGM, however, if the xanthan molecule is ordered then its conformation will be altered to that required for efficient binding (Morris and Foster, 1994). Direct backbone to backbone binding is rather ill-defined but will presumably involve energetically favourable interactions between the carbohydrate subunits and perhaps involves the formation of a 2 fold structure (Cairns et al., 1986; Millane and Wang, 1990) which can sterically directly interact with mannans. It is proposed that junction zone in xanthan-KGM mixture can then convert to a 6 fold helical structure on cooling with a helical pitch of 5.6 nm compared to that of 4.7 nm (5 fold) observed for xanthan alone (Cairns et al., 1987; Brownsey et al., 1988; Ridout et al., 1998).

A final hypothesis suggests that either ordered or disordered segments but not both, may participate in interactions with galacto and glucomannans (Williams et al. 1991; Mannion et al., 1992; Lundin and Hermansson, 1995; Rinaudo et. al., 1999; Goycoolea et. al., 2001). Williams et. al. (1991) suggested that in the absence of electrolyte KGM interacts with disordered xanthan chains whilst in the presence of 40 mM NaCl, it interacts with ordered xanthan chains. They also suggested that in the absence of salt the gels are slightly stronger.

In the literature to date more work has been carried out on the interactions of xanthan with galactomannans, such as Locust Bean Gum, than glucomannans such as KGM, however similar mechanisms have been suggested. Mannion et al. (1992) proposed that xanthan and galactomannans may interact by two distinct mechanisms. The first requires heating of the mixture to within 70°C of the Tm of xanthan, and gives higher storage moduli dependent on the galactose content of the galactomannan. The second mechanism takes place at room temperature and gives weaker, more flexible gels, whose rheological properties do not depend on galactose content, and in which the xanthan helix is retained in the post-interaction complex. For glucomannans, Fitzsimons et. al (2008) studied the effect of thermal history on xanthan and KGM interactions at different salt levels. They concluded that the same molecular interactions occur on mixing solutions of xanthan and KGM at ambient temperature as on cooling from high temperature, however the unheated mixture was weaker and less cohesive.

Clearly then, although the interactions between xanthan and galacto and glucomannans have been investigated over the past four decades, there is still controversy concerning the mechanism of gelation. The aim of this work is to attain a better understanding of the interaction mechanism between xanthan and KGM, by studying xanthans having different but well defined levels of acetate and pyruvate, and changing polymer and sodium chloride concentrations in order to manipulate the xanthan transition temperature and test its relevance to the mechanism of synergistic interaction. In future work we intend to describe the interaction between galactomannans such as locust bean gum and xanthan, however in the present work we have chosen to study konjac glucomannan in some detail, as this polymer exhibits less variability in structure due to the absence of sidechains.
MATERIALS AND METHODS

A range of xanthan samples having variable pyruvate and acetate contents (see Table 1) was kindly provided by DuPont. Glucomannan (KGM) was supplied by the Shimizu Chemical Corporation. The mannose to glucose ratio for the KGM used here was 1.7:1 with 3.5% of galactose (dry weight basis) also being present. The weight average molecular weight was 2.06 X 10^6 g/mol. The value for the G/M ratio, of 1:1.7 was measured independently in a total sugar analysis and not supplied by the manufacturer. The sugar analysis was carried out by methanalysis of the polysaccharide followed by trimethylsilylated-derivatization of the methyl glycosides. Analysis was by Gas Chromatography with a Flame Ionisation Detector (GC-FID). The galactose was detected in the total sugar analysis. It is known that there can be up to 5% of galactose branching in konjac mannans (Buckeridge et al. 2000), however this relatively small content is presumed to have little effect on the interactions with other biopolymers. Moreover this fraction should be constant throughout the experiments further reducing any effect on the interaction between xanthan and KGM.

The molecular weight of the KGM was determined using a standard treatment and filtration method followed by injection into a Size Exclusion Chromatography Multi Angle Laser Light Scattering (SECMALS) system. 2.06 X10^6 is at the upper end but within the range for commercial samples (Parry 2010).

Sodium azide was supplied by Acros organic, Geel Belgium and used as a bactericide at a concentration of 0.05 % in all samples. Sodium chloride was obtained from Fisher Scientific UK Ltd. Loughborough, UK.

Manufacture of xanthans

The method of producing xanthans of different contents of pyruvate was as follows. The SA/SP and SA/HP samples were a result of natural variations in the concentration of the functional groups produced during the fermentation process by Xanthomonas Campestris. Medium and high pyruvate batches were then selected from the available range of concentrations. The low pyruvate content of the SALP sample was produced by subjecting the fermentation broth to heat treatment in acidic conditions. The method of producing xanthans of different contents of acetate was based upon heat treatment under alkaline conditions of xanthan with naturally occurring high acetate levels (Abbaszadeh et al., 2015). All concentrations are expressed in %wt/total wt unless otherwise stated.

Preparation of solutions

Solutions were prepared by cold dispersion of the polymers in distilled water. Samples of one polymer were then mixed using a magnetic flea whilst heating to 90 ºC and held at that temperature for 20 min. Upon cooling to room temperature individual polymer samples were mixed together in a ratio of 1:1 and left over night.

High Sensitivity Differential Scanning Calorimetry (DSC)

Experiments were performed in a Setaram Micro DSC III (Setaram, Caluire, France) using cells made from Hastelloy, capable of holding approximately 0.8 ml and sealed using Hastelloy.
screw tops with viton “O” rings. Solutions were placed with the minimum of disturbance of
the viscous structure into the cell and initially cooled to a starting temperature of 10 °C.
Samples were heated and cooled at rates of 1ºC min⁻¹ from 10 to 100°C. Normally 2 complete
heating and cooling cycles were run. The reference cell was filled with water and matched for
overall heat capacity with the sample. This ensured that the calorimeter was balanced and
that the heat flow signal was centred at approximately the zero level which gave the most
sensitive result.

**Small deformation mechanical spectroscopy**
Rheological testing of the solutions and gels was performed in oscillation using cone and
plate geometry (4 cm diameter; 1.581° cone angle) with an Anton Paar Physica MCR-301
rheometer. Samples were loaded with minimal disturbance at room temperature, coated
around the periphery of the sample with light silicone oil to minimise the loss of water, and
left unperturbed for 30 min before measurements were made. A temperature sweep was
obtained by heating at 1 °C min⁻¹ and measurements made at a frequency of 1Hz and a strain
of 0.5 %. This ensured that data was obtained in the linear viscoelastic region.

**RESULTS AND DISCUSSION**
The influence of salt, acetate and pyruvate content on the xanthan transition temperature
and the interaction with KGM
Konjac glucomannan (KGM) at low ionic strengths is known to form a gel after mixing with
xanthan (Shatwell et al. 1990; Annable et al., 1994) and shows a limited interaction with stable
xanthan helices (Foster and Morris, 1994). Figure 1 and 2 show DSC second heats of different
types of xanthan, individually and in mixtures with KGM at a total concentration of 0.5 %wt.
Goycoolea et al. (1995) studied the stoichiometry of mixtures of xanthan-KGM and discovered
a 1:1 mix was optimal, which is the ratio used in this study. The mix of SA/SP-KGM shows the
previously seen endotherm by Annable et al. (1994) using DSC. The wide and biphasic helix-
coil transition (Figure 1a) for SA/SP (48°C to 80°C; enthalpy = 6.2 J/g) has changed to a
narrower single peak transition (52°C to 67°C enthalpy = 5.2 J/g; 10.4J/g if we assume all the
heat is due to the xanthan) after mixing with KGM. The SA/HP and LA/HP samples, in which
the T_m for xanthan alone was approximately 52°C and 43°C respectively, form a new
endothermic peak with almost the same temperature range and enthalpy as the SA/SP sample
in combination with KGM and the xanthan transition peak disappears (Figure 1b and c).

The SA/LP xanthan shows a different trend (Figure 2a). The xanthan transition, at high
temperature, remains detectable when combined with KGM, however a smaller endotherm
in the region of 40°C represents the synergistic interaction. This kind of transition has been
reported previously by Annable et al. (1994) at an ionic strength of 40mM NaCl rather than
10mM NaCl. By increasing the ionic strength to 40mM NaCl (Figure 2b), the SA/HP sample
now shows a similar trend to the SA/LP sample at the lower ionic strength of 10mM NaCl, with
a synergistic peak in the region of 40-58°C, slightly higher than the SA/LP synergistic peak, and
a separate, discernible xanthan helix-coil transition.

The effect of xanthan helical stability on the interaction
The synergistic peak at around 40°C is also detectable for the most stable of the samples, SA/VLP (Figure 2c), which indicates that the existence of the terminal mannose alone is enough to produce the low temperature synergistic junction zone. However, Foster and Morris (1994) did not find a synergistic peak for the polytetramer and attributed this to the enthalpic stability of the polytetramer helix (ΔH ≈ 8.5 J/g). The similarity of the enthalpy of transition for SA/VLP (ΔH ≈ 8.4 J/g) which does show the synergistic peak and the polytetramer may indicate a different reason, probably structural, for the lack of synergistic interaction between polytetramer and KGM. The present results (Figure 2) indicate that even though the xanthan helix-coil transition temperatures are almost matched by changing the sodium chloride concentration, the destabilising effect of pyruvate on the helical compactness produces slightly higher temperature melting i.e. more stable synergistic gels. As is shown in Figure 2 by increasing the pyruvate content respectively in SA/VLP, SA/LP and SA/HP samples, the temperature of synergistic gelation (T_{gel}) is increased from 41 °C for SA/VLP to 43°C for SA/LP and 49 °C for SA/HP.

Recently Fitzpatrick et al. (2012) emphasized that the interaction between xanthan and KGM in mixed gels occurs only if the xanthan molecules are in the ordered helical form. However as can be seen in Figure 3 upon cooling the exothermic peak of the disorder-order transition of VLA/HP xanthan alone occurs at a low temperature, below ~40°C compared with the synergistic exotherm (~60°C) i.e. xanthan is obviously in a disordered conformation when interaction occurs at temperatures above the onset of xanthan helix formation.

The combination of DSC and rheology data

Figure 4 shows combined DSC and rheology traces for xanthan/KGM mixtures at a total polymer concentration of 1 wt%. The high and low pyruvylated samples show a similar trend to that seen for the lower concentration samples (0.5 wt%) using both DSC and rheology, i.e. for a high pyruvylated sample after mixing with KGM the xanthan transition peak is replaced with a new peak at around 60°C and for a low pyruvylated sample the xanthan transition, at high temperature, remains detectable by DSC when combined with KGM, however a smaller endotherm in the region of 40°C represents the synergistic interaction.

The rheometry data on Figure 4a and b show the elastic modulus (G') on the second heating as a function of temperature with the Y-axis shifted to aid comparison. The onset temperatures for the synergistic endothermic DSC peaks are mirrored by the temperatures at which the onset of gel melting is seen. Interestingly, at the higher concentration of the SA/SP sample, the broad transition endothermic peak from 59°C to 84°C at 1%wt and 10mM NaCl is replaced with two smaller peaks at around 60°C and 40°C after mixing with KGM (Figure 4a). Figure 4b shows DSC and rheology data for samples mixed with KGM but at 40mM NaCl, again at a concentration of 1%wt. As expected, by increasing the ionic strength the xanthan helix-coil transition temperature is increased for all of the samples. It is essential to realise that this increase in temperature with increasing ionic strength applies to the xanthan helix-coil transition remaining after the interaction and not to the interaction peak itself. For the high pyruvylated sample SA/HP at 40mM NaCl, the 60°C synergistic peak seen at 10mM NaCl is replaced with a new peak in the region of 40-50°C, slightly higher than the SA/LP synergistic peak, and a still visible xanthan helix-coil transition. For the SA/SP-KGM mixture at 40mM
NaCl, the remnant xanthan T\textsubscript{m} increased to \textasciitilde{}90°C and the 60°C endotherm disappeared, however the 40-50°C endotherm was detectable. For the SA/LP-KGM mixture the xanthan T\textsubscript{m} increased to \textasciitilde{}94°C but the temperature of gelation (T\textsubscript{gel}) remained detectable at 40°C.

The synergistic endothermic peak onset temperatures observed by DSC are commensurate with the temperatures at which the onset of gel melting is measured by rheology. These effects have previously been reported as the effect of salt in decreasing the T\textsubscript{gel}; the more stable the xanthan helix, produced by higher salt contents or lower pyruvate levels, the more limited the interactions which occur with co-synergists.

A more detailed consideration of the nature of the interaction

Two types of mechanism are proposed, supported by evidence showing the binding of xanthan to KGM from rheology and DSC data. By taking into account previously proposed models and utilising a range of proprietary xanthans with known molecular structure, a unified model for the interaction of xanthan and co-synergist has been proposed. According to this model xanthan interacts with KGM at room temperature by two mechanisms (Type A and Type B) (cf. Mannion et al., 1992 and Fitzsimons et al., 2008). This can occur simultaneously and form gels which melt in two temperature ranges. The mechanisms are discussed below.

The storage modulus and the temperature of gelation

Figure 5 shows the elastic modulus (G') as a function of temperature whilst cooling at a rate of 1°C/min\textsuperscript{-1} for xanthan-KGM mixtures at a total polymer concentration of 0.5% w/w. For the highly pyruvylated xanthan samples SA/HP and LA/HP, G' starts to increase sharply below about 60°C independent of acetate level. By increasing the ionic strength to 40mM NaCl, the SA/HP sample shows a consequent reduction in T\textsubscript{gel} to \textasciitilde{}50°C, commensurate with the endotherm shown in Figure 4b. The T\textsubscript{gel} for SA/LP at 10 mM NaCl also matched the temperature of the endotherm seen in Figures 2a and 4a. The HP synergistic gels show higher final elastic moduli at 20°C, indicating that either there are more junction zones in the gel formed at 60°C, when the xanthan is still disordered, or the homotypic xanthan-xanthan interactions are enhanced in the HP samples (data not shown) and play an important role in stabilising the synergistic gel. Given that there is no indication of the individual xanthan transition \textless{}60°C in these mixtures and the fact that the mixed gel stoichiometry corresponds with such mixing ratios, this argues against the latter assumption, and suggests all xanthan molecules are occupied in the synergistic interactions.

The relevance of the xanthan transition temperature

As has been indicated already, the SA/SP xanthan sample, which exhibits a broad DSC peak for xanthan alone at 10mM NaCl, shows two waves of G' increase when mixed with KGM (Figure 4a); one starting around 60°C similar to that seen for samples containing high pyruvate levels and low ionic strength and another one about 45°C, in the same region as that for SA/HP at 40 mM NaCl, which suggests a different mechanism for its interaction with KGM. As has been shown on Figure 1a, the broad exotherm of the coil-helix transition for SA/SP occurs over a wide range centred around 60°C, where gelation is seen for the HP xanthans. Therefore, at the point of the first wave of G' increase (60°C) some of the xanthan molecules are ordered,
with approximately 50% of the transition still to take place. The two waves of G' increase therefore indicate two different modes of xanthan-KGM interaction within one mixture. These mechanisms are described in the following discussion.

**Type A and Type B interactions**

In the Type A interaction, xanthan retains its helical conformation when interacting with KGM and the gel melts at a lower temperature, ~30-45°C. In the Type B interaction, the 2-fold conformation of disordered xanthan is responsible for the synergistic binding with KGM. The possibility of this transition has been shown by computer simulation (Millane and Wang 1990; Chandrasekaran and Radha, 1997). The interaction occurs via backbone-to-backbone formation of junction zones and the formed gel melts at the higher temperature of ~60°C. The Type B interaction can start with both ordered and disordered xanthan conformations. If xanthan is in the disordered conformation just before the synergistic interaction, for example the VLA/HP sample at low ionic strength, direct binding (backbone-to-backbone) occurs with KGM, however, if the xanthan molecule is ordered such as in the SA/LP sample, a type A interaction is expected. The SA/SP sample at 10mM NaCl has a xanthan coil-helix transition with a mid-point around 60°C therefore for this xanthan both Types of interaction, A and B occur.

**Charge effects**

Agoub et al. (2007) reported that a progressive reduction in pH raises the temperature of the xanthan disorder–order transition as measured by DSC, and decreases the gelation temperature of xanthan/KGM mixtures. They demonstrated that an increase in G' for mixtures of commercial xanthan with KGM at pH values of 4.5 and 4.25 occurs in two discrete steps on cooling. The first occurs at the temperature observed here for the same mixtures at neutral pH and the second occurs over the lower temperature range observed here for mixtures of KGM with pyruvate free xanthan.

They conclude that these two “waves” of gel formation are attributed to the interaction of KGM with, respectively, xanthan sequences that had retained a high content of pyruvate substituents, and sequences depleted in pyruvate by acid hydrolysis. They also reported that the increases in G' were not dependent on the xanthan transition temperature.

Figures 4c and d show the temperature sweeps for xanthan samples on cooling. High pyruvate xanthan samples have one strong ascending wave at ~60°C (Type B); the low pyruvate xanthans show one wave at ~45°C (Type A) and SA/SP has both types A and B. However, pH was not varied here in contrast with the work of Agoub et al. (2007). Similar to the effect of increased salt levels, increasing the acidity which is equivalent to an increase in proton ion concentration, reduces the charge on the pyruvate and consequently increases the temperature of the conformational transition of xanthan. However the temperature of the interaction endotherm decreases.

What has not been reported (Agoub et al., 2007) is the result of the first heating of the samples, i.e. mixing the polymers cold and loading them cold into the rheometer. As can be seen on Figure 6, the modulus for all xanthan samples during the first heat, shows both types
of waves of increase but with different intensities. Linearizing the logarithmic scale for the modulus facilitates the comparison of the magnitude of the waves.

**The Effect of heat treatment and xanthan transition temperature on the interaction mechanism**

Figure 6a and 7b, for a high pyruvate sample in which the xanthan disorder-order transition temperature is below 60°C show a significant increase in the Type B interaction after the first heating (Figure 7b). Heating intensifies type B interactions and provides new opportunities for the formation of xanthan-co-synergist networks which may lead to a more unified network arrangement on cooling. Interestingly, after the first heat, the samples in which the xanthan helix-coil transition temperatures were higher than ~60°C (Type B interaction occurs at ~60°C), e.g. SA/LP, no longer exhibit a type B wave or drop in the modulus at around 50°C with heating. Only the SA/SP-KGM mixture shows both types of interactions on cooling. This behaviour has already been explained by the temperature at which the helix to coil transition is found. As can be seen on Figure 1, the SA/SP xanthan disorder-order transition has started just before the Type B interaction temperature (~60°C) and has ended after it. At the point of the first wave of G’ increase (60°C) some of the xanthan molecules are ordered. Therefore upon cooling, the xanthan in this sample can participate in both types of interactions.

**Verifying the proposed model**

In order to verify the proposed model of A and B type interactions, the transition temperature for SA/HP was adjusted by changing the salt content so that the mid-point occurred at the same temperature as the SA/SP sample i.e. the transition started below and finished above 60°C (Figure 7a and b).

The SA/HP-KGM mixture in 15 mM NaCl shows both types of interaction on cooling comparable to the SA/SP-KGM mixture.

On increasing the salt level to 40 mM NaCl, the preferred interaction is Type A mechanism, especially after the first heat (Figure 4). Williams et al. (1991) suggested that disordered xanthan interacts with KGM at very low ionic strength and by increasing the ionic strength to 40 mM the interaction occurs only with ordered xanthan. Figure 7 compares the DSC curve for SA/HP-KGM mixture at 0.5% and 1% total polymer concentration, on cooling. As can be seen at 1%, the low temperature peak ~45°C is detectable and represents Type A interactions. However, at this polymer concentration and ionic environment there is also evidence of a small amount of Type B interaction, with the additional higher temperature transition being that of the xanthan coil-helix transition.

As a further test, and using a different approach, 40 mM NaCl has been added after preheating a mixture of xanthan SA/HP+KGM at 45°C before letting the mixture cool. After adding salt at 45°C, the cooled mixture was loaded on the rheometer and a temperature sweep performed (Figure 7). At the temperature of salt addition, a gel due to the type A interaction would melt but that due to a type B interaction would be stable. Consequently the salt did not have an effect on the type B transition. After cooling and upon first heating the
mixture shows the evidence of both Type A and B interaction, however, after the first heating
and melting of the Type B gel, the salt is now distributed in the solution and on cooling the
Type B transition has disappeared and only the type A gel remains.

A summary of the different types of interaction is shown on figure 8.

It is interesting to examine the proposal as to whether unique models can be constructed
from information of this type. DSC data in particular deals with heat flow and whilst providing
useful information does not give a unique or indeed any picture as to what is occurring in the
sample. Essentially many models or pictures can be constructed from the information.
Additional techniques, such as rheology, are used here to provide a picture of the molecular
events. Future work must include techniques such as NMR, X-ray diffraction measurements
or titration calorimetry, if these can be applied to the systems studied here, to look at the
molecular interactions in more detail.

In conclusion

The main conclusions for this investigation are as follows:

A new model for interaction of xanthan and KGM is proposed in which 2 possible interaction
mechanisms may exist singly or together. In the Type A interaction, xanthan keeps its helical
conformation and the formed gel melts in a lower temperature range of approximately 30-45°
C. The type B interaction occurs with xanthan in a disordered conformation and the formed
gel melts at the higher temperature of approximately 60°C. The xanthan transition
temperature is the key parameter in determining the Type of interaction (Figure 8) and can be
manipulated by changing environmental conditions such as ionic concentration, pH and
functional group content.

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Figures

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Figure 1 DSC 2nd Heats at a scan rate of 1°C min⁻¹ for a) SA/SP b) SA/HP and c) LA/HP xanthan both alone and in 1:1 mixtures with KGM at an ionic strength of 10 mM NaCl and a total concentration of 0.5%.
Figure 2 DSC 2nd Heats at a scan rate of 1ºC min⁻¹ for a) SA/LP xanthan at an ionic strength of 10 mM NaCl b) SA/HP xanthan at an ionic strength of 40 mM NaCl c) SA/VLP xanthan at an ionic strength of 40 mM NaCl; both alone and in 1:1 mixtures with KGM at a total concentration of 0.5%. The peak and limiting temperatures of the synergistic interaction peak are shown by arrows and dotted lines respectively.
Figure 3 DSC 1st and 2nd heating and cooling scans at a scan rate of 1°C min⁻¹ for VLA/HP xanthan at an ionic strength of 10 mM NaCl both alone and in 1:1 mixtures with KGM at a total polymer concentration of 0.5 % wt. Artefacts such as shown in the lowest trace (VLA/HP + KGM first heat) are commonly seen on first heating runs as the material is not completely uniformly dispersed. Consequently where possible information is taken from 2nd traces. Arrows show the onset temperatures of xanthan helix/coil transition and synergistic interaction on cooling.
Figure 4 $G'$ for the 2nd heat of VLA/HP xanthan ($\bigcirc$), SA/HP xanthan ($\square$), SA/SP xanthan ($\Diamond$) and SA/LP xanthan ($\triangle$) and corresponding DSC curves in 1:1 mixtures with KGM and at a total polymer concentration of 1% wt. at a) 10 mM NaCl and b) 40 mM NaCl. and $G'$ on cooling and corresponding DSC curves for the same named curves as in a) and b) at c) 10 mM NaCl and d) at 40 mM NaCl; all at a total polymer concentration of 1%.

Figure 5 $G'$ on cooling for SA/HP xanthan ($\square$), LA/HP xanthan ($\bigcirc$), SA/SP xanthan ($\Diamond$), SA/LP xanthan ($\triangle$) all at 10 mM NaCl and SA/HP xanthan (X) at 40 mM NaCl; in 1:1 mixtures with KGM and at a total polymer concentration of 0.5%.
Figure 6 a) (G' for 1st heats and b) 2nd heats of VLA/HP xanthan (○), SA/HP xanthan (□), SA/SP xanthan (◊) and SA/LP xanthan (△) in 1:1 mixtures with KGM at a total polymer concentration of 1% and an ionic strength of 10 mM NaCl. Temperature regions for type A and B interactions are shown.
Figure 7  a) $G'$ for the 1st heat and  b) 2nd heat of SA/HP xanthan in 15 mM NaCl (□) and SA/SP xanthan in 10 mM NaCl (◊) both in 1:1 mixtures with KGM and at a total concentration of 1%. Also shown are the corresponding DSC heat flow curves. DSC 1st heating traces can often show artefacts such as the jump marked here with an arrow and similar to that shown on figure 3. These can be due to non-uniform distribution, settling and convection in the DSC cell. c) Heating and cooling scans for SA/HP xanthan in 40 mM NaCl in 1:1 mixtures with KGM at total concentrations of 0.5 and 1%. d) 1st heating and cooling.
scans for SA/HP xanthan preheated at 45°C in 40 mM NaCl in 1:1 mixtures with KGM at a total concentration of 1%.

**Figure 8** Proposed mechanisms for the interaction of xanthan with Konjac glucomannan.