Structural Associations of Symptomatic Knee Osteoarthritis

Laura A. Stoppiello,1 Paul I. Mapp,1 Deborah Wilson,2 Roger Hill,2 Brigitte E. Scammell,1 and David A. Walsh3

Objective. Structural changes of osteoarthritis (OA) may occur in the absence of pain. In this study, we aimed to identify histopathologic features that are associated with symptomatic knee OA.

Methods. Medial tibial plateaus and synovium samples were obtained at the time of total knee replacement (TKR) surgery for OA (advanced OA group) or were obtained postmortem from subjects who had not sought medical attention for knee pain during the last year of life (non-OA control group). To identify features of OA, we compared the patients with advanced OA with the age-matched non-OA controls (n = 26 per group).

To identify OA features associated with symptoms, we compared two additional groups of subjects who were matched for severity of chondropathy (n = 29 per group): patients undergoing TKR for symptomatic OA (symptomatic chondropathy group) and postmortem subjects with similar severity of chondropathy who were asymptomatic during the last year of life (asymptomatic chondropathy group). The histologic features of the samples were graded, and immunoreactivities for macrophages (CD68) and nerve growth factor (NGF) in the synovium were quantified. The cellular localization of synovial NGF was determined by double immunofluorescence analysis.

Results. Advanced OA cases displayed more severe changes in the synovium (synovitis, increased synovial NGF, and CD68-immunoreactive macrophages) and cartilage (loss of cartilage surface integrity, loss of proteoglycan, tidemark breaching, and alterations in chondrocyte morphology) than did the non-OA controls. Synovial NGF was localized predominantly to fibroblasts and to some macrophages. The symptomatic chondropathy group displayed greater levels of synovitis, synovial NGF, and loss of cartilage integrity, in addition to alterations in chondrocyte morphology, than did the asymptomatic chondropathy group (P < 0.05 for each comparison).

Conclusion. Synovitis, increased synovial NGF, alterations in chondrocyte morphology, and loss of cartilage integrity are features of knee OA that may be associated with symptoms.

Osteoarthritis (OA) is the most common joint disease in the elderly population. Pathologic features include articular cartilage degradation, synovial inflammation, osteophytes, and bone marrow lesions. Pain is the main reason sufferers seek clinical assistance. Patients with end-stage OA may eventually benefit from total knee replacement (TKR) surgery.

Although joint pathology makes an important contribution to OA pain, the precise relationship between pain and structural pathology is not entirely clear (1,2). Radiographic features of OA are often weakly associated with pain, suggesting that structural, cellular, or biochemical changes that directly mediate pain may not be easily detected on radiographs. Imaging studies have indicated possible associations between pain and synovitis (3–6) or subchondral changes (6–9), but associations with histopathology have not been explored in detail. Such studies have previously been limited by age limitations.
differences between OA patients and normal controls and by limited access to joint tissue from people with OA whose pain is less than that which would lead them to undergo joint replacement surgery.

Synovitis may contribute to OA knee pain. The cause of synovitis in OA is incompletely understood, but has been observed in both early (10) and late disease (11). Synovitis may mediate pain through the production of factors that sensitize or activate sensory nerves.

Recent evidence has suggested an important role of nerve growth factor (NGF) in mediating inflammatory pain through increasing nociceptor sensitivity or facilitating sensory nerve growth (12). NGF blockade was shown to improve the degree of pain from knee OA (13–15), indicating a causal link between NGF and OA pain.

NGF is elevated in a variety of rheumatic diseases, particularly in the synovial fluid of patients with inflammatory arthritis (16,17). NGF is expressed by synovial macrophages from patients with rheumatoid arthritis, spondyloarthritis, and to a lesser extent, in OA (17), as well as by synovial fibroblasts from patients with OA (18). However, the expression and cellular profile of NGF in synovial biopsy samples from people with knee OA has not been explored in detail. Synovial fibroblasts can be visualized using immunohistochemistry with antibodies directed against Hsp47, a collagen-specific molecular chaperone (19). Macrophages can be identified by the transmembrane glycoprotein CD68 (20).

Our aim in the present study was to first validate the measurement of histopathologic characteristics of knee OA by comparing samples obtained at the time of TKR surgery for OA (advanced OA) with samples collected postmortem from age-matched non-OA controls. Second, we aimed to identify histopathologic features associated with symptomatic knee OA by comparing cases with similar macroscopic chondropathy, half of whom had sought medical care for knee pain and had undergone TKR (symptomatic chondropathy) and the other half had not sought medical care for knee pain but had died of an unrelated illness (asymptomatic chondropathy). We hypothesized that symptomatic OA is mediated by specific structural and biochemical changes within the knee joint, whereas other changes may be coincidental or even protective.

PATIENTS AND METHODS

Patient samples. The joint tissue repository of the Arthritis Research UK Pain Centre, which contains samples from >1,700 subjects, was screened first to select tissues obtained at the time of TKR for OA (advanced OA group) and tissues obtained postmortem from age-matched subjects who had not sought medical attention for knee pain during the last year of life (non-OA control group) (n = 26 per group) and second to select tissues from patients who had undergone TKR for symptomatic OA (symptomatic chondropathy group) and tissues with similar macroscopic chondropathy scores obtained postmortem from subjects who had not sought medical attention for knee pain during the last year of life (asymptomatic chondropathy group) (n = 29 per group). One TKR case was included in both the advanced OA group and the symptomatic chondropathy group and was therefore counted twice.

Patients undergoing TKR fulfilled the American College of Rheumatology classification criteria for OA (21) at the time of surgery. Subjects from whom samples were obtained postmortem were recently deceased, had no history of rheumatoid arthritis or pseudogout, and had not previously sought help for knee pain during the last year of life, as determined by interviews with the relatives and review of case notes. Exclusion criteria for non-OA controls consisted of a history of OA, Heberden’s nodes identified on clinical examination, macroscopic chondropathy lesions of grade 3 or 4 in the medial tibiofemoral compartment, or osteophytes on direct visualization of the dissected knee.

Body mass index (BMI; kg/m²) in the TKR cases was calculated from measurements of height and weight that were documented in the cases notes. BMI in the postmortem cases could not be calculated because height and weight measurements were not available.

Informed consent was obtained from the TKR patients and from the next of kin of the postmortem subjects. The study was approved by the UK National Research Ethics Service (Nottingham Research Ethics Committee 1 [05/Q2403/24] and Derby Research Ethics Committee 1 [11/H0405/2]).

Sample processing. Midcoronal sections of the middle one-third of the medial tibial plateau were fixed in neutral-buffered formalin and then decalcified in 10% EDTA in 10 mM Tris buffer (pH 6.95; at 4°C) prior to embedding in wax. Surgeons and technicians were instructed to collect synovium from the medial joint line. Synovial tissues were fixed in formalin and embedded in wax without decalcification or were frozen without fixation in melting isopentane and embedded in OCT mounting medium (Raymond A. Lamb Ltd.).

Macrosopic chondropathy score. Following tissue harvesting, the articular surfaces of the medial and lateral tibial plateaus and femoral condyles were evaluated for the extent and severity of loss of surface integrity by a single assessor (RH), as described by Walsh et al (22) based on the method of Ayral et al (23). Briefly, articular surface defects were graded from 0 (normal, smooth, unbroken surface) to 4 (subchondral bone exposure), and the proportion of each articular surface area corresponding to each grade was used to calculate a chondropathy score (range 0–100). Scores for each of the 4 compartments were summed to give a total tibiofemoral chondropathy score (range 0–400). Surgical damage precluded the calculation of macroscopic chondropathy scores for some TKR cases.

Radiographic OA severity score. Radiographic severity scores for OA were derived using preoperative posteroanterior knee radiographs as previously described (22). An atlas of line drawings of the knee joint was used to grade medial and lateral joint space narrowing and osteophytes (24). The scores for joint space narrowing (range 0–6) and osteophytes (range
Table 1. Clinical and pathologic characteristics of the study groups*

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Non-OA control (n = 26)</th>
<th>Advanced OA (n = 26)</th>
<th>Asymptomatic chondropathy (n = 29)</th>
<th>Symptomatic chondropathy (n = 29)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>61 (48–67)</td>
<td>61 (50–67)</td>
<td>71 (66–79)</td>
<td>68 (61–74)†</td>
</tr>
<tr>
<td>% male</td>
<td>62</td>
<td>31†</td>
<td>55</td>
<td>31 (28–37)</td>
</tr>
<tr>
<td>BMI, kg/m²</td>
<td>NA</td>
<td>32 (29–40)</td>
<td>NA</td>
<td>31 (177–215)†</td>
</tr>
<tr>
<td>Macropscopic chondropathy score (range 0–400)</td>
<td>51 (40–69)</td>
<td>NA</td>
<td>199 (177–215)†</td>
<td>223 (185–234)‡</td>
</tr>
<tr>
<td>Total radiographic score (range 0–18)‡</td>
<td>NA</td>
<td>12 (10–13)</td>
<td>NA</td>
<td>12 (10–13)</td>
</tr>
<tr>
<td>JSN score (range 0–6)</td>
<td>NA</td>
<td>3 (4–5)</td>
<td>NA</td>
<td>5 (4–5)</td>
</tr>
<tr>
<td>Osteophyte score (range 0–12)</td>
<td>NA</td>
<td>7.5 (5–8)</td>
<td>NA</td>
<td>8 (5.7–8)</td>
</tr>
<tr>
<td>Histologic features</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total Mankin score (range 0–14)¶</td>
<td>5 (3–7)</td>
<td>8 (7–9)‡</td>
<td>7 (7–8)</td>
<td>8 (7.5–9)</td>
</tr>
<tr>
<td>Cartilage surface integrity (range 0–6)</td>
<td>2 (2–3)</td>
<td>4 (4–4)‡</td>
<td>4 (3–4)</td>
<td>4 (4–4)‡</td>
</tr>
<tr>
<td>% with breached tidemark</td>
<td>35</td>
<td>68†</td>
<td>66</td>
<td>67</td>
</tr>
<tr>
<td>Proteoglycan loss (range 0–4)</td>
<td>1 (1–1)</td>
<td>1 (1–2)‡</td>
<td>1 (1–2)</td>
<td>1 (1–2)</td>
</tr>
<tr>
<td>Chondrocyte morphology (range 0–3)</td>
<td>1 (0–3)</td>
<td>2.5 (2–3)†</td>
<td>3 (1.5–3)</td>
<td>3 (2–3)†</td>
</tr>
<tr>
<td>% with subchondral bone marrow replacement</td>
<td>0</td>
<td>19</td>
<td>18</td>
<td>32</td>
</tr>
<tr>
<td>Synovitis (range 0–3)</td>
<td>0 (0–0)</td>
<td>0 (0–3)†</td>
<td>0 (0–0.25)</td>
<td>2 (0–3)‡</td>
</tr>
<tr>
<td>Fractional areas**</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Macrophage</td>
<td>3.4 (1.8–4.6)</td>
<td>4.8 (3.1–8.7)†</td>
<td>7.4 (4.2–13.2)</td>
<td>13.1 (6.6–15.3)</td>
</tr>
<tr>
<td>Nerve growth factor</td>
<td>1.4 (0.4–1.8)</td>
<td>4.0 (1.5–8.3)†</td>
<td>8.7 (5.9–12.4)</td>
<td>12.9 (9.4–18.6)†</td>
</tr>
</tbody>
</table>

* Tissues were obtained at the time of total knee replacement for osteoarthritis (OA) or were obtained postmortem from cadaver donors. Data for the body mass index (BMI) was available for 23 patients with advanced OA and 20 patients with symptomatic chondropathy. (One patient with TKR was included in both groups and was therefore counted twice.) Statistical comparisons for the immunohistochemical and histomorphometric data were not made between the features of OA and the features of symptomatic OA subgroups because staining was carried out in separate experiments. Except where indicated otherwise, values are the median (interquartile range). NA = not available.
† P < 0.05 versus the corresponding subgroup (i.e., non-OA control group or asymptomatic chondropathy group).
‡ P < 0.001 versus the non-OA control group.
§ The total radiographic score is the sum of the scores for tibiofemoral joint space narrowing (JSN) and osteophytes.
¶ The total Mankin score is the sum of the scores for cartilage surface integrity, tidemark breaching, proteoglycan loss, and chondrocyte morphology. Tidemark breach is expressed as the percentage of subjects in whom cartilage integrity permitted assessment (25 in the advanced OA group and 24 in the symptomatic chondropathy group).
# P < 0.01 versus the corresponding subgroup.
** Fractional areas are the percentage of the synovial area occupied by CD68-immunoreactive macrophages or nerve growth factor–immunoreactive cells. Fractional areas are reported only for samples that displayed synovial lining (n = 15–26 per group).

0–12) were summed to provide a total radiographic OA severity score (range 0–18). Radiographs were not available for postmortem cases.

**Histologic assessment and grading.** Formalin-fixed tissue sections (5 μm) were stained with hematoxylin and eosin or, to visualize proteoglycan content in the medial tibial plateau, with Safranin O–fast green. Articular cartilage changes of OA were graded as 4 components according to the Mankin scoring system (25): cartilage surface integrity was graded on a scale of 0–6 (where 0 = normal and 6 = complete disorganization), tidemark integrity was graded 0 or 1 (where 0 = intact and 1 = crossed by vessels), chondrocyte morphology was graded on a scale of 0–3 (where 0 = normal and 3 = hypocellular), and proteoglycan loss was graded on a scale of 0–4 (where 0 = normal, no loss of Safranin O staining and 4 = complete loss of staining). The presence of subchondral bone marrow replacement by fibrovascular tissue was assessed in each case as either present (pathologic) or absent (normal). Synovial inflammation was graded on a scale of 0–3 (where 0 = normal and 3 = severe inflammation) by assessing the degree of synovial lining hyperplasia, inflammatory cell infiltrate, and cellularity (11).

**Immunohistochemistry.** Sections of formalin-fixed synovium underwent antigen retrieval (1 mg/ml of pepsin in 0.5M acetic acid at 37°C for 2 hours), incubated with monoclonal mouse anti-human CD68 (clone PG-M1; Dako), followed by biotinylated horse anti-mouse IgG secondary antibody (Vector). NGF immunoreactivity was visualized after citrate buffer antigen retrieval (90°C for 20 minutes). Sections were blocked with 5% bovine serum albumin (BSA) containing goat serum, followed by incubation with rabbit monoclonal antibody to NGF (clone EP1320Y; Abcam) and biotinylated goat anti-rabbit IgG secondary antibody (Vector). Visualized by both immunoreactivities used avidin–biotin complex peroxidase (Vector) with nickel-enhanced diaminobenzidine development (26).

Synovium samples from 3 TKR patients, 3 postmortem cases with high levels of chondropathy, and 2 non-OA controls were simultaneously double-stained for NGF and either macrophage or fibroblast markers. Frozen synovial tissue sections were fixed in 4% paraformaldehyde, followed by acetone and then were blocked with 5% BSA containing goat serum and horse serum. Sections were incubated overnight with a mixture of antibodies directed toward NGF (as above) and either CD68 (as above) or Hsp47 (M16.10A; Enzo Life Sciences), followed by incubation with a mixture of secondary antibodies: Texas Red–labeled horse anti-mouse IgG and fluorescein-labeled goat anti-rabbit IgG (Vector). Sections were counter-
stained with DAPI, and autofluorescence was blocked with 1%
Sudan black (Sigma-Aldrich) in 70% ethanol. Sections were
mounted in phosphate buffered saline/glycerol.

**Image analysis.** All histologic and immunohistologic
scoring was carried out by a single observer (LAS) who was
blinded with regard to the diagnostic group and clinical details.
A Zeiss Axioscop-50 microscope was used. Macrophage and
NGF fractional areas were quantified using a 3-CCD video
camera and KS300 image analysis software. Areas of the
synovium with the highest density of positive staining for CD68
or NGF were identified. Briefly, an image was captured, and
area of interest, which included the synovial lining and sublin-
ing, was delineated. Positive staining was differentiated from
background by thresholding the image according to hue in
order to create a mask. The area of positive staining and the
total area were measured. The fractional area was determined
as the percentage of synovial area positive for CD68 or NGF
immunoreactivity within the total area. Data were collected
using a 20× objective lens in 4 different fields of view. Only
samples with synovial lining were included in the analyses.

**Statistical analysis.** Data were analyzed using SPSS
v.21 software. The Mann-Whitney U test was used to compare
TKR and postmortem groups for each variable, except for age
(compared by t-tests) and the presence of subchondral bone
marrow replacement (compared by chi-square test) between
groups. Associations between BMI and outcome measures
were evaluated using linear regression, adjusting for age and
experimental group. Statistical significance was accepted when
the P value was less than 0.05. Logistic regression was used to
adjust for sex differences in the non-OA control and age-
matched advanced OA group comparisons and for age differ-
ences in the asymptomatic and symptomatic chondropathy
group comparisons.

The intraobserver reliability for the histology and
fractional area scores was assessed by resoring 20–25 ran-
domly selected cases on 2 separate occasions 6 months apart.
For reproducibility analyses, the kappa statistic (StatsDirect
software) (27) was used for cartilage, subchondral bone, and
synovial scores. The intraclass correlation coefficient (SPSS
software) was used for the fractional area values.

---

**Figure 1.** Histopathologic features of non-osteoarthritic (non-OA) (A–D) and OA (E–J) knees. Non-OA knees typically displayed mild cartilage
surface irregularities of the superficial zone, with normal proteoglycan staining (A), whereas in some patients with advanced OA of the knee, there
were deep clefts in the articular cartilage surface and severe proteoglycan loss (E) (Safranin O–fast green stained). Channels can be seen in the
calcified cartilage beneath an intact tidemark in a non-OA knee (B), whereas an OA knee shows a channel breaching the tidemark and entering the
noncalcified cartilage (F) (hematoxylin and eosin [H&E] stained). Normal subchondral bone marrow spaces are filled with fatty tissue in a non-OA
knee (C), whereas subchondral bone marrow is replaced by fibrovascular tissue in an OA knee (G) (H&E stained). Mild synovial lining hyperplasia
is shown in a sample from a non-OA knee (D), whereas severe synovitis, characterized by hyperplasia and hypertrophy of the synovial lining and
sublining hypercellularity, can be seen in an OA knee (H) (H&E stained). Macrophage immunoreactivity for CD68 (I) and immunoreactivity for
nerve growth factor (J) in OA knees was localized to the synovial lining and sublining regions (staining [blue-black] developed with nickel-enhanced
diaminobenzidine). Arrows indicate the synovial surface. Bars = 500 µm.
Dotted lines indicate the synovial surface. Bars

Red (Hsp47 and CD68 [red]) and fluorescein (NGF [green]) staining. C shown in a cell that is not immunolabeled for CD68 (D, NGF.

Indicated in OA. Immunoreactivity of macrophages in synovium from a patient with

Figure 2. Cellular localization of nerve growth factor (NGF) immunoreactivity in human synovium. A, Hsp47 immunoreactivity localized to cells with fibroblast morphology in the lining and sublining regions of synovium from a patient with osteoarthritis (OA). Arrow indicates a fibroblast colocalized to NGF (same cell as indicated in B); arrowhead indicates a fibroblast not colocalized to NGF. B, NGF immunoreactivity colocalized with Hsp47 (arrow) and in a cell that is not immunolabeled for Hsp47 (arrowhead) in the section shown in A. C, CD68 immunoreactivity of macrophages in synovium from a patient with OA. Arrow indicates a macrophage colocalized to NGF (same cell as indicated in D); arrowhead indicates a macrophage not colocalized to NGF. D, NGF immunoreactivity colocalized with CD68 (arrow) and in a cell that is not immunolabeled for CD68 (arrowhead) in the section shown in C. Double immunohistochemistry was visualized by Texas Red (Hsp47 and CD68 [red]) and fluorescein (NGF [green]) staining. Dotted lines indicate the synovial surface. Bars = 50 μm.

RESULTS

Histopathologic features of knee OA. Patient demographics are reported in Table 1, and histopathologic features are illustrated in Figure 1. Samples displayed a range of cartilage changes, including mild clefts to the articular surface with normal proteoglycan content (Figure 1A), as compared with deep clefts and loss of proteoglycan in some advanced OA cases (Figure 1E). Channels from the subchondral bone were found in close proximity to the tidemark (Figure 1B) and also breached the tidemark to invade the noncalcified cartilage (Figure 1F). Subchondral bone spaces mostly contained normal fatty tissue (Figure 1C); however, in some cases, this was replaced by fibrovascular tissue (Figure 1G). Synovial inflammation was sometimes marked, with lining hyperplasia and lymphoid aggregates (Figure 1H). CD68+ (Figure 1I) and NGF+ (Figure 1J) cells were evident in the synovial lining and sublining regions and surrounded the blood vessels.

NGF immunoreactivity in the synovium was predominantly localized to Hsp47+ spindle-shaped and mononuclear cells, consistent with fibroblast morphology (Figures 2A and B). However, some NGF+Hsp47− and NGF−Hsp47+ cells were also observed. Some CD68+ macrophages in the lining and sublining regions also displayed NGF immunoreactivity (Figures 2C and D). Similar distributions were found in advanced OA patients and non-OA controls.

Intraobserver reliability for histologic scoring was moderate or substantial, as determined using the kappa statistic (range 0.53–0.76). For fractional area measurements, the intraclass correlation coefficient was 0.85, indicating good reproducibility.

Patients with advanced OA displayed more severe cartilage surface changes, proteoglycan loss, tidemark breaching, changes in chondrocyte phenotype, and synovitis than did non-OA controls (P < 0.05 for each comparison) (Figure 3). Tidemark breaching was observed in 17 of 25 (68%) of the advanced OA patients and 9 of 26 (35%) of the non-OA controls (P = 0.012). Subchondral fibrovascular replacement was present in 5 of 26 (19%) of the advanced OA patients and was absent from the non-OA controls; however, significance was lost after adjusting for sex differences. The synovial fractional areas positive for CD68 and NGF immunoreactivities were greater in patients with advanced OA than in the non-OA controls (P = 0.031 and 0.035, respectively) (Figures 3E and F).

Histopathologic features of symptomatic knee OA. Both the asymptomatic and symptomatic chondropathy groups displayed evidence of structural OA, with higher macroscopic chondropathy scores than the non-OA controls (Table 1). Patients in the symptomatic chondropathy group displayed macroscopic chondropathy features similar to the subjects in the asymptomatic chondropathy group, indicating appropriate matching (Table 1). Severe macroscopic chondropathy lesions graded as 3 or 4 (deep cartilage fibrillation or subchondral bone exposure, respectively) were present in the medial tibiofemoral compartment of 28 of 29 (97%) subjects with asymptomatic chondropathy and 29 of 29 (100%) patients with symptomatic chondropathy.

Loss of cartilage surface integrity and alterations in chondrocyte morphology were greater in the symptomatic chondropathy group than in the asymptomatic chondropathy group (Table 1 and Figures 4A and B), whereas proteoglycan loss, tidemark integrity, and subchondral bone marrow replacement by fibrovascular tissue did not differ significantly between groups.
Figure 3. Histopathologic features of osteoarthritis (OA). Scatterplots illustrate the differences between the non-OA control group and the advanced OA group for the scores on cartilage surface integrity (A), proteoglycan loss (B), chondrocyte morphology (C), and synovitis (D), and for the synovial fractional area positive for CD68-immunoreactive macrophages (E) and nerve growth factor (NGF) (F). Each symbol represents an individual subject; horizontal lines and error bars show the median and interquartile range. \(* = P < 0.05; ** = P < 0.01; *** = P < 0.001\) versus the non-OA control group.

Figure 4. Histologic associations of symptomatic osteoarthritis (OA). Scatterplots illustrate the differences between subjects with similar macroscopic chondropathy scores who did not (asymptomatic chondropathy) or did (symptomatic chondropathy) undergo total knee replacement surgery. The cartilage surface integrity score (A), chondrocyte morphology score (B), synovitis score (C), and synovial fractional area positive for nerve growth factor (NGF) (D) were greater in the symptomatic chondropathy group, but differences in the synovial fractional area positive for CD68-immunoreactive macrophages (E) did not reach statistical significance. Each symbol represents an individual subject; horizontal lines and error bars show the median and interquartile range. \(* = P < 0.05; ** = P < 0.01\) versus the asymptomatic chondropathy group.
Synovitis scores were greater in patients in the symptomatic chondropathy group than in subjects in the asymptomatic chondropathy group (Table 1 and Figure 4C). Severe inflammation with perivascular lymphoid aggregates was observed in 8 of 29 symptomatic chondropathy patients (28%) compared with 1 of 29 asymptomatic chondropathy subjects (3%) \((P = 0.021)\). The synovial area positive for NGF was greater in symptomatic chondropathy patients than in asymptomatic chondropathy patients (Table 1 and Figure 4D). Differences in macrophage infiltration between groups did not reach statistical significance (Table 1 and Figure 4E). We used logistic regression analysis to examine whether the association between symptomatic OA and synovitis was explained by increased synovial NGF immunoreactivity and/or macrophage fractional area. Associations between synovitis and symptomatic chondropathy remained significant in each regression model \((P \leq 0.008)\).

BMI (kg/m²) was calculated for both TKR groups (advanced OA and symptomatic chondropathy). Data were available for 42 of 54 patients (78%). Of these, 29 patients (69%) were classified as obese (BMI \(\geq 30\)), 11 patients (26%) were classified as overweight (BMI 25–29.9), and 2 patients (5%) were a healthy weight (BMI 18.5–24.9). Linear regression was used to determine associations between BMI and outcome measures, combining data from both TKR groups. Age and experimental group were used as independent covariates. BMI was not significantly associated with any histopathologic or immunohistologic parameter.

**DISCUSSION**

We found that a range of histopathologic features in the synovium and tibial plateaus were associated with OA. Of these, synovitis, increased synovial NGF immunoreactivity, chondrocyte changes, and loss of cartilage surface integrity were associated with symptomatic OA, as compared with postmortem subjects with similar macroscopic joint surface appearances who had not sought TKR during life. Increased synovitis and synovial NGF expression by fibroblasts and some macrophages may be key features associated with pain in OA.

The prevalence and severity of OA are strongly associated with increasing age, raising doubts in earlier studies as to whether the histopathologic features are specifically associated with OA, rather than being characteristics of healthy aging. We addressed this by carefully age-matching the patients with advanced OA and the non-OA controls, a process that required a large source repository from which to select cases.

Synovitis, the densities of CD68-immunoreactive macrophages or NGF-immunoreactive cells within the synovium, proteoglycan loss, chondrocyte changes, loss of tidemark integrity, and tidemark breaching were all greater in patients with advanced OA than in age-matched non-OA controls. This finding indicates that in this study, these pathologic features of OA were not explained by aging alone. However, each of these features was also observed to a lesser extent in our age-matched control group, indicating the presence of either “preclinical” early OA in the control group or the presence of normal structural features in the advanced OA group that are exaggerated by the disease. A key question, therefore, is which pathologic features are associated with clinical OA?

The relevance of structural changes to OA symptoms has been difficult to determine. Evidence of synovitis on magnetic resonance imaging (MRI) has been associated with pain in people who have OA or those who are at high risk of developing OA (5). Furthermore, a recent study demonstrated that fluctuation in synovitis corresponds to a fluctuation in knee pain, providing further evidence of the contribution of synovitis to symptomatic OA (28). Patients with greater amounts of synovitis display more severe chondropathy, and synovitis may predict progressive structural damage in knee OA (22,29). It can therefore be difficult to be certain as to whether synovitis itself is a cause of pain or whether it acts as a surrogate for other aspects of OA pathology. Our work extends these previous findings by indicating that synovitis is associated with symptomatic OA independently of the severity of chondropathy.

In the present study, we explored the characteristics of synovitis that have been associated with symptomatic OA. We confirmed the previous findings of increased macrophage infiltration in OA (10), but the presence of macrophages may not be sufficient to explain OA pain. Levels of NGF are elevated in the synovial fluid of patients with rheumatoid arthritis (16,30), and NGF is present to a lesser extent in patients with OA (17), but a previous study did not localize NGF to the synovium in the single OA case reported (31). Nevertheless, monoclonal antibodies directed against NGF have been shown to reduce pain and improve function in people with OA (13–15).

We showed that in OA, NGF immunoreactivity is present in the synovium, predominantly in Hsp47+ fibroblasts, and in some macrophages. NGF immunoreactivity was greater in the synovial tissue of the symptomatic chondropathy group than in that of the asymptomatic chondropathy group. These findings suggest an important contribution of fibroblast-derived...
NGF in the generation of OA pain. The proinflammatory cytokines interleukin-1β and tumor necrosis factor α (TNFα) can stimulate the release of NGF from synovial fibroblasts, and NGF can modulate the release of TNFα (18). NGF may act on sensory nerves to increase pain in OA. After we adjusted for NGF and/or CD68 immunoreactivity, the significant association of synovitis with symptomatic OA remained, suggesting that increased NGF and macrophage infiltration, either alone or together, do not completely explain the pain. The production of pain mediators, for example, by fibroblasts, may yet be driven by macrophages or other aspects of inflammation.

Previous histopathologic studies of OA have often relied on samples obtained at joint replacement surgery, which display a range of OA structural severity (22). The severity of chondropathy overlaps to a small extent between postmortem cases and surgical cases (22), and therefore, our study of pathologic associations in symptomatic OA required selection of cases with matched degrees of macroscopic chondropathy from a large tissue repository. Radiographic scores in our symptomatic chondropathy group were similar to those in our advanced OA group, and macroscopic chondropathy scores in the symptomatic and asymptomatic chondropathy groups were significantly higher than those in the non-OA control group, indicating that both of the chondropathy groups had pathologic features of knee OA.

Although the two chondropathy groups were matched for similarity of macroscopic chondropathy scores, microscopic loss of cartilage surface integrity was greater in the symptomatic chondropathy group. Chondropathy cases were selected on the basis of similar total macroscopic chondropathy scores of the tibiofemoral joint, where the appearance of the 4 articular surfaces was evaluated: the medial and lateral femoral condyle and tibial plateau. Histologic grading was examined in one midcoronal section of the middle one-third of the medial tibial plateau because this is an important weight-bearing area that is characteristically affected by OA. Macroscopic and microscopic analyses therefore reflect distinct but related aspects of OA pathology (22).

Not all histopathologic features associated with OA were associated with symptomatic OA. Tidemark breaching was found in twice as many asymptomatic chondropathy cases than non-OA controls, suggesting that it may be an early feature of OA, but it was no more common in the symptomatic chondropathy group than the asymptomatic chondropathy group. In contrast, osteochondral vascularity was previously associated with pain behavior in rats with OA following transection of the medial meniscus (32). Our current data suggest that tidemark breaching alone is insufficient to cause OA symptoms and that additional factors, such as sensory innervation or local expression of NGF, may be necessary mediators of pain from osteochondral channels (12). Our current study provides novel insights into the incidence of tidemark breaching in a subgroup of asymptomatic postmortem cases with macroscopic chondropathy similar to patients who underwent TKR. Further work would be required to determine the time course and mechanisms by which channels breaching the tidemark may predispose to pain.

Fibrovascular replacement of the subchondral bone marrow was twice as common in the symptomatic chondropathy group as it was in the asymptomatic chondropathy group, but this difference did not reach statistical significance after adjusting for age differences between groups. Bone marrow replacement has been associated with bone marrow lesions on MRI (33), which in turn, has been associated with pain in knee OA (7,34). Our study may have had insufficient power to detect differences between groups, and further work would be required to define the structural and biochemical features of bone marrow lesions that may explain their association with pain.

Obesity is a major risk factor for OA (35–37) and has been associated with pain (38) and cartilage changes (39,40). The majority of patients with advanced OA in our cohort were classified as overweight or obese, but histopathologic factors associated with symptoms were not significantly associated with BMI in our study.

Our study is necessarily subject to a number of limitations. We assumed that the subjects in the asymptomatic chondropathy group had experienced less pain than the patients in the symptomatic chondropathy group, since to the best of our knowledge, these subjects had not sought medical attention for knee pain during the last year of life. Clinical data acquisition in postmortem cases relies on case notes, where knee pain may not have been reported, and interviews with bereaved relatives, who may not have been aware of the deceased’s symptoms. Histologic analysis was performed on only 1 section of osteochondral tissue per case, and the sample was obtained from the midcoronal section of the middle one-third of the medial tibial plateau, a key weight-bearing site. It is possible that there may be different structural associations with symptoms at other sites within the joint. Furthermore, our patients with symptomatic OA all had late-stage disease (undergoing TKR), and different structural features may be associated with pain in early OA.

Our study design avoided confounding by age in our characterization of features associated with OA, but
we were unable to simultaneously match for sex. Similarly, matching for the severity of chondropathy precluded age matching between our symptomatic and asymptomatic chondropathy groups, even when using a source repository containing samples from >1,700 cases. However, we adjusted for age or sex differences in our analyses.

We classified postmortem cases as having OA based on the presence of cartilage changes comparable to those seen in patients undergoing TKR for OA, and we selected small, homogeneous patient groups in order to undertake detailed histopathologic experiments. Our work extends imaging and arthroscopic studies indicating the relative importance of synovitis (3,5) and macroscopic cartilage changes (41,42) to symptoms in the broader OA population.

We have identified histopathologic features of knee joint tissues that are associated with OA; however, not all of these features may cause pain. Symptomatic OA was associated with synovitis, synovial NGF immunoreactivity, changes in chondrocyte morphology, and loss of cartilage surface integrity as compared with the asymptomatic chondropathy group showing a similar macroscopic joint surface appearance who had not sought TKR during life. Different pathologic features of OA typically coincide, and better understanding of the structural features that are associated with symptomatic OA would help in refining strategies for developing structure-modifying treatments (disease-modifying OA drugs) by focusing on those which are most likely to improve symptoms. Synovitis and synovial NGF expressed by fibroblasts and some macrophages may be key mediators associated with OA knee pain.

ACKNOWLEDGMENTS

We are grateful to the patients, orthopedic surgeons, and Bereavement Centre colleagues at Sherwood Forest Hospitals NHS Foundation Trust for providing the postmortem and surgical tissue for our repository. We thank the staff of the Histopathology Department at Sherwood Forest Hospitals NHS Foundation Trust for processing the tissues.

AUTHOR CONTRIBUTIONS

All authors were involved in drafting the article or revising it critically for important intellectual content, and all authors approved the final version to be published. Ms Stoppiello had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Study conception and design. Stoppiello, Mapp, Wilson, Hill, Scammell, Walsh.

Acquisition of data. Stoppiello.

Analysis and interpretation of data. Stoppiello, Mapp, Scammell, Walsh.

REFERENCES


19. Izquierdo E, Canete JD, Celis R, Del Rey MJ, Usatequi A, Marsal...


