

***Achatina (Lissachatina) fulica* BOWDICH: ITS
MOLECULAR PHYLOGENY, GENETIC VARIATION IN
GLOBAL POPULATIONS, AND ITS POSSIBLE ROLE IN
THE SPREAD OF THE RAT LUNGWORM
Angiostrongylus cantonensis (CHEN)**

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ABSTRACT

The Giant African Snail, *Achatina (Lissachatina) fulica* Bowdich, 1822, is a tropical crop pest species with a widespread distribution across East Africa, the Indian subcontinent, Southeast Asia, the Pacific and the Caribbean. It is also a known intermediate host of the rat lungworm, *Angiostrongylus cantonensis*, which can infect humans and cause eosinophilic meningitis. The phylogenetic position of *A. fulica* within the Achatinoidea and the Achatinidae was investigated using segments of the nuclear ribosomal (r) RNA cluster, actin and histone 3 genes and the mitochondrial CO1 and 16S rRNA genes. Results from molecular data support the monophyly of the Achatinidae based on the taxa surveyed as well as the morphological distinction of the Eastern *Achatina (Lissachatina)* from the Western and Central *Achatina (Achatina)*; *Lissachatina* should therefore be elevated to genus status. The results also show non-monophyly of the Coeliacidae, Ferussaciidae and Subulinidae; the taxonomy of these families must therefore be reassessed. The extent of genetic diversity in global *A. fulica* populations was also determined using an SSCP molecular marker developed from the 16S rRNA gene. Results reveal only one haplotype (C) emerged from East Africa and spread globally. The rat lungworm (*Angiostrongylus cantonensis*) has a parallel distribution with *A. fulica*, and the possible role of the snail in the spread of the parasite is investigated using a molecular marker derived from the small subunit (SSU) rRNA gene. A survey of the parasite within the route of dispersal of *A. fulica* detected *A. cantonensis* only in the Philippines and the French Polynesian territory of Tahiti, the latter of which being the first reported case of *A. cantonensis* infection for *Achatina fulica* in that territory. Due to the limited sampling of the snail and the patchy distribution of the parasite, there are insufficient data at this time to assess the role of *Achatina fulica* in the spread of *Angiostrongylus cantonensis*.

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CHAPTER 1 – Introduction

The tropical Giant African Land Snail *Achatina (Lissachatina) fulica* Bowdich, 1822 is one of the most extensively studied snails because of its economic, ecological and medical importance (Mead, 1979). It belongs to a family of African snails, the Achatinidae (Mollusca, Gastropoda, Stylommatophora, Achatinoidea), that includes more than 200 species in 13 genera (Schileyko, 1999). *Achatina fulica* is a major crop pest species that originated in East Africa but has been spreading across the globe since before the 1800's primarily through human activities (Mead, 1961, 1979; Raut & Barker, 2002). The World Conservation Union (IUCN) has listed *Achatina fulica* as one of the world's 100 most invasive species (Lowe *et al.*, 2000). The snail also serves as an intermediate host of the rat lungworm *Angiostrongylus cantonensis* (Chen) (see Alicata, 1966). Humans get accidentally infected by ingesting the 3rd juvenile stage of this parasite, derived from the snail intermediate host, which leads to eosinophilic meningoencephalitis (EME) or angiostrongyliasis, a disease of the central nervous system (Marquardt *et al.*, 2000). Although much is already known about *Achatina fulica*, many issues regarding the snail remain unaddressed. These include its phylogenetic relationship with other members of the Achatinoidea and the Achatinidae, the validity of the subgenus *Lissachatina* based on molecular data, the genetic variation present in introduced populations of *A. fulica* across the globe, and its potential role in the spread of the parasite *Angiostrongylus cantonensis*.

1.1. Taxonomy of *Achatina fulica*: Mollusca, Gastropoda, Stylommatophora, Achatinoidea, Achatinidae

Achatina fulica belongs to the phylum Mollusca, the second largest animal phylum in the world, with an estimated total species diversity ranging from less than 50,000 to as much as 200,000 and inhabiting freshwater, marine and terrestrial habitats (van Bruggen, 1995). Although members of this phylum exhibit diversity in form as exemplified by snails, clams, octopods, squids, chitons and the tusk shells, this group possesses three unique hallmarks that distinguish its members from other animal phyla. These hallmarks are: (1) a muscular foot for locomotion; (2) a fleshy skin fold called a mantle that secretes a calcareous shell; and (3) a feeding organ called a radula. Some molluscan groups may have lost one or more of these hallmarks during the course of evolution, but their ancestors clearly had all three as shown by fossil records (Ruppert *et al.*, 2004). Other characteristics of molluscs are the lack of true segmentation, the reduction of the body cavity, and the presence of spiral cleavage during early development (van Bruggen, 1995).

Terrestrial slugs and snails like *Achatina fulica* belong to the Gastropoda, the largest taxonomic class in the Mollusca, in which members have a characteristic head-foot region and a visceral mass (Ruppert *et al.*, 2004). Gastropods undergo torsion during larval development that leads to the 180° rotation of the visceral mass and brings the ctenidia or gills and the excretory organs to the anterior region above and behind the head (Barker, 2001). There are an estimated 30,000-35,000 species of terrestrial snails and slugs (Solem, 1984), and many of these have become very important to Man either as a food source, parasite vectors, ornaments or even tools (Barker, 2001). The majority of terrestrial snails and slugs, including *A. fulica*, do not have gills but instead have a vascularised chamber or a 'lung' within the mantle cavity, a characteristic of the

subclass Pulmonata. The groupings within the pulmonates are still in a state of flux as disagreements still persist regarding interpretations of evolutionary pathways based on conchological and anatomical characters, but most workers agree on the monophyly of the (sub)order Stylommatophora. Stylommatophorans like *A. fulica* characteristically have two pairs of caudal tentacles, with the upper pair bearing eyes at the tip (Ruppert *et al.*, 2004); a contractile pneumostome that minimises contact between the environment and the pallial cavity that houses the lung, kidney and ureter; and the lack of an operculum that covers the aperture (Barker, 2001). Around 71-92 families constitute the Stylommatophora (Emberton *et al.*, 1990). Recent molecular evidence from (1) sequences from the rRNA cluster (Wade & Mordan, 2000); (2) primary sequence data of mitochondrial genes, particularly the absence of entire stem/loop structures in some domains of the mitochondrial 16S rRNA gene (Lydeard *et al.*, 2000); (3) sequence data provided by the cytochrome c oxidase subunit I (COI) gene (Remigio & Hebert, 2003); and (4) rare genomic changes in the trnP and COI genes (Grande *et al.*, 2004) all support the monophyly of the Stylommatophora.

1.2.1. Classification of the Stylommatophora based on morphological data

There is considerable debate as to how the Stylommatophora should be subdivided into groups that reflect true evolutionary relationships. The widely used but criticised Pilsbry-Baker System divides the Stylommatophora into four infraorders based on the excretory system (Pilsbry, 1900; Baker, 1955). The main excretory organ is made up of three parts: (1) a nephridium or nephridial sac (=kidney); (2) a distal ureteric pouch or orthureter; and (3) a separate ureter represented either as an open ciliated groove or a closed tube. Based on variations of these structures, the four infraorders are as follows: (1) Orthurethra (those with nephridium, orthureter with an

anterior nephropore, and a ureter developed as a groove, the proximal part of which lies along the rectal face of the nephridium, Figure 1.1A); (2) Mesurethra (those with neither an orthureter on the nephridium nor a closed ureter, Fig. 1.1B); (3) Sigmurethra (those without an orthureter but with a ureter that runs along the anterior portion of the nephridium and connects to the pallial cavity before terminating at the pneumostome, Fig. 1.1C); and (4) Heterurethra (those without an orthureter, with the nephridium extending transversely, and a ureter that runs along the face of the kidney before following the rectum and terminating at the pneumostomal opening Fig. 1.1D). It has been hypothesised that the orthurethran type of excretory system is the most 'primitive' or ancient while the other types are derived from it (Pilsbry, 1900; Baker, 1955; Barker, 2001). Based on this classification system, *A. fulica* is included in the Sigmurethra.

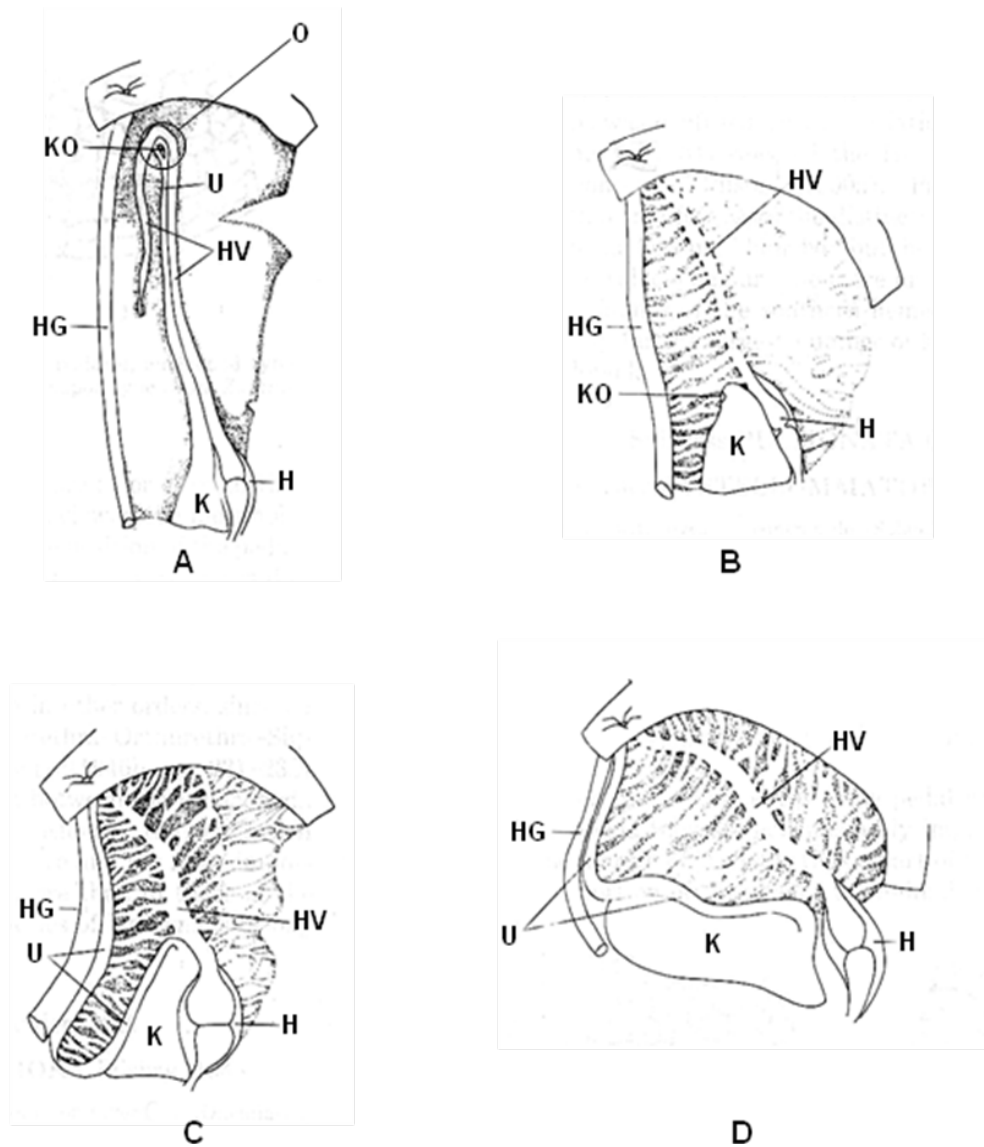


Figure 1.1: The Pilsbry-Baker system of classification of the Stylommatophora based on the structures of the excretory organ. K – nephridium (=kidney); U – ureter; KO – renal orifice or nephropore; O – orthureter; HG – hindgut; HV – principal pulmonary vein; H – heart. The four infraorders of the Stylommatophora are (A) – Orthurethra; (B) – Mesurethra; (C) – Sigmurethra; (D) – Heterurethra. (From Solem, 1959)

Pilsbry's classification system is not without its critics (Simroth & Hoffmann, 1908-1928; Thiele, 1929-1935). Tillier (1989) proposed an alternative classification system based on the differentiation within the renal organ. He retained the Orthurethra as one suborder but divided the other stylommatophorans into two different suborders: the Brachynephra in which the shortening of the kidney took place followed by closure

of the ureter; and the Dolichonephra, in which the ureter closed first before the kidney shortened. He also based his classification scheme on the various groups' current distributions that corresponded to regions that were once part of either of the two hypothetical supercontinents that resulted from the breakup of the Pangaeon landmass 200 million years ago. The Orthurethra have a mixed Laurasian (Europe, North America and Asia) and Gondwanian (South America, Africa, India, Australia) distribution, the Brachynephra includes representatives that are wholly or partly Gondwanian, and the Dolichonephra have members with Laurasian links. In this classification scheme, *A. fulica* is placed in the Dolichonephra. However, Nordsieck (1992) criticised Tillier's system and believed that overemphasis was given to the differentiation within the renal organ. Instead, he divided the Stylommatophora into two suborders: the Orthurethra that bear an orthurethran excretory system and have a primarily Laurasian origin, and the Sigmurethra with a non-orthurethran system and mixed Laurasian and Gondwanian origins. He agreed with Pilsbry's assumption of the basal position of the Orthurethra. Nordsieck's Sigmurethra also includes *Achatina fulica*.

1.2.2. Relationships within the Stylommatophora based on molecular data

Molecular markers are increasingly being used to infer phylogenetic relationships among groups of taxa because of the advantages they provide. Molecular data are derived from discrete heritable characters, are unambiguous, can easily be subjected to quantitative analyses and homology assessment, may provide characters that can be used to compare distantly related taxa, and are abundant (Graur & Li, 2000).

Although stylommatophoran taxa have been used in a variety of molecular studies examining evolutionary relationships within the Mollusca, relatively few studies

have focused on the relationships within the Stylommatophora themselves. Studies that include a handful of representative stylommatophorans, with emphasis on relationships at deeper levels within the Mollusca and the position of the Stylommatophora therein, include those of Winnepenninckx *et al.* (1998) on the small subunit (SSU) ribosomal (r) RNA gene; Thollessen (1999) on the 16S rRNA gene; Lydeard *et al.* (2000) on the secondary structures of the 16S rRNA gene; Wade & Mordan (2000) on the rRNA cluster; Remigio & Hebert (2003) on the COI gene; Colgan *et al.* (2003) on the SSU rRNA, COI and histone 3 genes; Grande *et al.* (2004) on the rare genomic changes in the trnP and COI genes; and Passamanek *et al.* (2004) on the LSU and SSU rRNA genes.

Studies focusing on the evolutionary relationships within the Stylommatophora include those of Armbruster *et al.* (2005) and Wade *et al.* (2001; 2006). Armbruster *et al.* (2005) surveyed 18 species from eight families of the Stylommatophora using the combined coding regions of the histone 3 and histone 4 genes. Their study demonstrated the basal position of the Helicidae relative to Punctidae, Clausiliidae, Pupillidae, Enidae, Vertiginidae, Cochlicopidae, and Valloniidae. *Achatina fulica* was not represented in their study. Wade *et al.* (2001, 2006) conducted a more comprehensive phylogenetic study of the Stylommatophora with initially 104 species from 50 families including *A. fulica* (2001), which they expanded to 160 species encompassing 61 families (2006). They utilised DNA sequence data from the nuclear rRNA gene cluster spanning a 1460 bp region of the 5.8S rRNA gene, the internal transcribed spacer (ITS) 2 region and the large subunit (LSU) ribosomal (r) RNA gene and comprising 823 unambiguously aligned nucleotides used for phylogenetic analysis (Figure 1.2). Although the results they obtained were in general agreement with currently accepted taxonomic families, a clear and unexpected dichotomy of the

Stylommatophora into two clades was also revealed: the strongly supported ‘achatinoids’ (99% NJ bootstraps for both the 2001 and 2006 studies) and the weakly supported ‘non-achatinoids’ (65% NJ bootstraps for the 2001 study and 63% NJ bootstraps for the 2006 study). Both clades included members with Laurasian and Gondwanian distributions, implying that the radiation of the Stylommatophora predated the breakup of Pangaea into Laurasia and Gondwana some 200 million years ago. This also implies that the sigmurethran type of excretory system is ancestral with all the other types being derived from it, contradicting Pilsbry’s assumption of a basal Orthurethra and suggesting that the Orthurethra is a derived group.

Additionally, several studies have focused on specific taxonomic groups within the Stylommatophora, as exemplified by Dutra-Clarke *et al.* (2001) on the Succineidae using the SSU rRNA gene; Holland & Hadfield (2004) on endemic Hawaiian Achatinellinae using the COI gene; Steinke *et al.* (2004) on the western Palearctic Helicidae also using the COI gene; and Tongkerd *et al.* (2004) on the Thai Pupillidae using the LSU and the 16S rRNA genes.

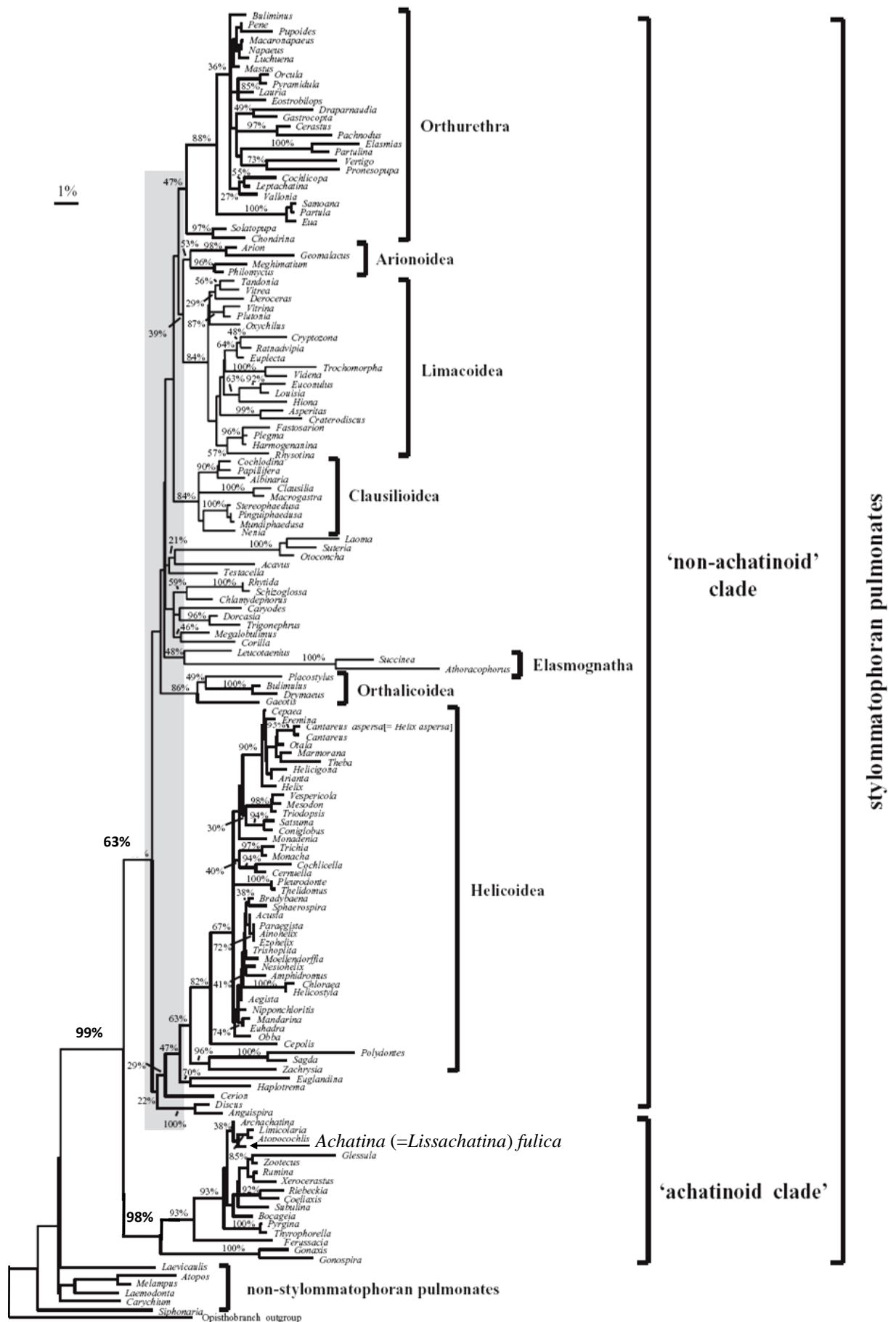


Figure 1.2: Neighbor-joining phylogenetic tree of the Stylommatophora based on the large subunit rRNA gene (823 unambiguously aligned nucleotide sites). The phylogeny shows dichotomy of the group into the 'achatinoid' (bootstrap support=99%) and the 'non-achatinoid' (bootstrap support=63%) clades. (From Wade et al., 2006.)

1.2.3. The ‘achatinoid clade’ and the Achatinoidea

The ‘achatinoid clade’ of Wade *et al.* (2001, 2006) based on the rRNA cluster consisted of two superfamilies: the Streptaxoidea and the Achatinoidea. Traditionally, the Streptaxoidea comprises only a single family, the Streptaxidae. They are the only carnivorous group within the ‘achatinoid clade,’ and have a long buccal mass and short oesophagus reflecting their carnivorous adaptation (Tillier, 1989). The other families in Wade *et al.*’s ‘achatinoid clade’ belong to the superfamily Achatinoidea, which included representatives of the Achatinidae, Coeliacidae, Ferussaciidae, Subulinidae, Glessulidae and Thyrophorellidae. Various authors have different views as to which families constitute the Achatinoidea. Solem’s (1978) Achatinacea (=Achatinoidea) comprised the Achatinidae, Ferussaciidae, Megaspiridae, Spiraxidae and Subulinidae (including the coeliacids and glessulids as the subfamilies Coeliacinae and Glessulinae, respectively) while Abbott (1989) regarded Coeliacidae as a family in its own right while additionally including the monotypic Thyrophorellidae. Tillier’s (1989) Achatinoidea comprised the Achatinidae, Ferussaciidae, Subulinidae (including the subfamilies Coeliacinae and Glessulinae), Thyrophorellidae as well as the Streptaxidae, Succineidae and the Oleacinidae (Spiraxidae and Testacellidae). Tillier treated the Achatinoidea as those with a closed ureter, symmetrical cerebro-pedal connectives, short cerebral commissure, and contiguous left parietal and visceral ganglia; however, he also included the Succineidae because of superficial features it shares with the Ferussaciidae and the presence of a heterurethran type of excretory organ. Vaught (1989) incorporated the Achatinidae, Coeliacidae, Ferussaciidae, Subulinidae, (including the Glessulidae which Vaught regarded as subfamily Glessulinae) and Thyrophorellidae in her Achatinoidea. Schileyko (1999) placed only the Achatinidae in the Achatinoidea, whereas Bouchet & Rocroi (2005) included the Achatinidae,

Ferussaciidae, Subulinidae (which also incorporated the coeliacids and glessulids as the subfamilies Coeliacinae and Glessulinae) and Micractaeonidae. In Wade *et al.*'s (2006) study, the Spiraxidae (represented by *Euglandina*) and Succineidae (represented by *Succinea*) fell within the 'non-achatinoid' clade and not within the Achatinoidea, thus contradicting Solem (1978) and Abbott (1989) for their inclusion of the Spiraxidae in the Achatinoidea and Tillier (1989) for his inclusion of the Oleacinidae (Spiraxidae and Testacellidae) and the Succineidae. The Micractaeonidae, which are presumably distantly related to the Ferussaciidae (Schileyko, 1999), were not represented in the Wade *et al.* study. Table 1.1 summarises the various authors' groupings of the Achatinoidea.

Table 1.1: The Achatinoidea according to various authors in comparison with the Achatinoidea by Wade *et al.* (2006) based on molecular data (rRNA cluster).

Solem (1978)	Abbott (1989)	Tillier (1989)	Vaught (1989)	Schileyko (1999)	Bouchet & Rocroi (2005)	Wade <i>et al.</i> (2006)
Achatinidae Ferussaciidae Megaspiridae Spiraxidae Subulinidae (including Coeliacinae & Glessulinae)	Achatinidae Coeliacidae Ferussaciidae Spiraxidae Subulinidae Thyrophorellidae	Achatinidae Ferussaciidae Oleacinidae* Streptaxidae Subulinidae (including Coeliacinae & Glessulinae) Succineidae Thyrophorellidae	Achatinidae Coeliacidae Ferussaciidae Subulinidae (including Glessulinae) Thyrophorellidae	Achatinidae	Achatinidae Ferussaciidae Micractaeonidae Subulinidae (including Coeliacinae & Glessulinae)	Achatinidae Coeliacidae Ferussaciidae Glessulidae Subulinidae Thyrophorellidae

* Oleacinidae – Spiraxidae + Testacellidae

The Achatinoidea in Wade *et al.*'s (2006) study were represented by 14 taxa, and their relationship was evaluated using only a short fragment (823 unambiguously aligned nucleotides) of the rRNA cluster. Increasing the number of taxa and using more genes could shed more light on the relationships of the various achatinoid groups and the validity of the taxonomic designation of the families under the Achatinoidea.

1.2.4. The Achatinidae

The family Achatinidae is a group of land snails confined to Sub-Saharan Africa, with the exception of *Achatina fulica*, which has been introduced by Man to areas outside Africa (Mead, 1979; Raut & Barker, 2002). Their shells are mostly dextral, higher than wide and are fusiform, ovoid or pillar-shaped (Schileyko, 1999). Classification within the family is based on conchological features (Bequaert, 1950) and the highly variable reproductive tract (Mead, 1991). There are three subfamilies within the Achatinidae: (1) the Callistopeplinae in which members have medium sized shells with a truncated columellar margin and a vas deferens that is not attached to the penis sheath (Mead, 1994); (2) the Limicolarinae in which members also have medium sized shells but with a non-truncated columellar margin and a vas deferens that is attached to the penis sheath (Schileyko, 1999); and (3) the Achatininae with medium to large shells with truncated columellar margin and a vas deferens that is also attached to the penis sheath (Mead, 1994). Variation also exists within the subfamilies. In the Achatininae, for instance, the East African *Achatina* (*Lissachatina*), to which *A. fulica* belongs, has a smooth nepionic whorl (Bequaert, 1950) and a half-dome apex (Mead, 1995) as well as a muscular bulboid enlargement of the basal vagina (Mead, 1991; 1995) while the West and Central African *Achatina* (*Achatina*) has a sculptured nepionic whorl (Bequaert, 1950), a plateaued apex (Mead, 1995) and no muscular bulboid enlargement in the basal vagina (Mead, 1991; 1995).

Despite the huge amount of morphological data available on the Achatinidae, no systematic molecular approach has yet been attempted to correlate molecular data with the morphological data, in particular to assess the designation of *Achatina fulica* under the subgenus *Lissachatina* based on conchological and anatomical features.

1.3. The biology of *Achatina fulica*

Achatina fulica is a large snail with a shell length ranging from 5 to 10 cm, with some specimens even reaching 20 cm. The conical shell is light brown in colour, though the colour pattern may vary (Schotman, 1989). The presence of streaks is associated with a dominant allele such that homozygous recessive individuals have unstreaked shells (Allen, 1983). However, variation in shell morphology in terms of size, shape and colour exists and has been largely attributed to environmental conditions (Mead, 1961).

A typical *A. fulica* has a life span of 5-6 years, becoming sexually mature as early as five months. Although hermaphroditic, *A. fulica* cross-fertilises and lays eggs 8-20 days after mating. The number of eggs laid can vary depending on the age of the snail but can reach up to 1800 in a year in a tropical setting. If conditions become unfavourable, the snail can aestivate by burrowing underground and covering its shell opening with a calcareous membrane, called an epiphragm, until such time as the environment improves (Mead, 1979, Raut & Barker, 2002).

1.4. The biology of introduced species

Achatina fulica is a classic example of an introduced species. Introduced species, also known as exotic species, are those found outside their natural range due to human activity (Primack, 2006). Species may be introduced deliberately to benefit Man, with examples including agricultural plants and animals for human consumption, decorative plants for gardening, and animals for hunting or fishing. Other species may be introduced unintentionally such as parasites or pests found in deliberately introduced species and those that “hitchhike” with transported goods (Freeland, 2005).

Species introduced by Man to new areas can have a profound effect on the ecosystem of that area as well as on the introduced species itself. They can prey upon, infect, outcompete or hybridise with native species or alter the habitat at the expense of the native species (Simberloff *et al.*, 2005), thus providing additional selective pressure on the native species (Suarez & Tsutsui, 2008). A balancing act must therefore come into play between the native species and the introduced species, and sometimes even between the introduced species and the local environment of the invaded habitat; otherwise, either the native or introduced species will face displacement or extinction (Suarez & Tsutsui, 2008). The disturbance caused by introduced species has become problematic in many areas such that they have been regarded as the second greatest threat to biodiversity, next only to habitat loss (Freeland, 2005). In some cases, however, introduced species simply spread into new areas that are human-disturbed from which most native species have already disappeared due to habitat loss (Cowie & Robinson, 2003).

Not all species become easily established once translocated into a new area, but characteristics such as a rapid reproduction rate, high fecundity and generalist food and habitat requirements can increase the success of an introduced species (Cowie, 2000). Organisms that become invasive are also most likely to possess traits that facilitate their transport by humans, the ability to withstand the rigours of transport, the capacity to tolerate varying environmental conditions, and the predilection to thrive in human disturbed areas (Suarez & Tsutsui, 2008). As mentioned previously, species may be introduced deliberately or inadvertently, although those in the former category may have a greater chance of being established, particularly if the introduction is perceived to have some economic benefits. Deliberate introductions involve individuals that are

cared for and are brought to new areas in large numbers, thus improving their chance of survival once they are released in the wild (Cowie & Robinson, 2003).

The success of an introduced species in a new area can also be influenced by the genetic composition of its population. In many cases, introduced species are represented by a few individuals with a reduced amount of genetic variation when compared to their source population, a phenomenon called a founder effect. After many generations, a population bottleneck ensues where genetic variation is considerably reduced and allele frequencies undergo massive shifts (Dlugosch & Parker, 2008). As a consequence, some beneficial adaptive traits that could otherwise improve the survival and fitness of the species in the new habitat may be lost (Kolbe *et al.*, 2007). However, this low genetic variability as a result of founder events and bottlenecking could be counteracted by multiple introductions from different source populations (Dlugosch & Parker, 2008), as was shown in the cheatgrass *Bromus tectorum* (Novack & Mack, 1993) and the *Anolis* lizards (Kolbe *et al.*, 2007).

1.5. *Achatina fulica* as an introduced species and its dispersal from East Africa

Man has always been drawn to the Giant African Land Snail for reasons including its large size, supposed medicinal properties and its potential as a human or animal food source (Mead, 1979; Kliks & Palumbo, 1992; Raut & Barker, 2002). It is for these reasons that *Achatina fulica* has been spreading globally primarily through human factors, and its success as an introduced species can be attributed to several factors.

First, the biology of *Achatina fulica* makes it eminently suitable as an introduced species. The snail has a high reproductive capacity, producing between 10 and 400 eggs per clutch and as many as 1800 eggs per year; they also become sexually mature

between 5 and 8 months (Raut & Barker, 2002). *Achatina fulica*'s high reproductive capacity, in addition to the tendency of people to release the snails into the wild, would help to explain the rapid spread of *Achatina fulica* into new areas, as in Brazil where it was introduced as recently as 1988 but has since spread (Thiengo *et al.*, 2007). The Giant African Land Snail is a voracious herbivore that feeds on a wide range of cultivated plants and even weeds and indigenous plants, thus making it a serious crop pest (see Raut & Barker, 2002 for a comprehensive list of economically important food crops and ornamental and medicinal plants that are susceptible to *Achatina fulica*). It also scavenges its food from detritus and decaying plant material, which may comprise up to 75% of its diet (Raut & Barker, 2002). Its non-specific food requirements further leads to its success as an introduced species.

Second, *Achatina fulica* possesses traits that facilitate its transport by humans. For instance, the snails can easily be transported in consigned cargoes, whether accidentally or on purpose, and survive the journey of several days with little adverse effect on the "hitchhikers." This was demonstrated by a tourist who came from Hawaii and inadvertently brought a live snail to the mainland USA over a period of ten days (Mead, 1979). During these periods of long distance travel, the snails can undergo aestivation to avoid desiccation (Mead, 1961). Furthermore, *A. fulica* has a wide tolerance for different environmental conditions despite being a tropical snail (Mead, 1979; Raut & Barker, 2002). They have been found to survive temperatures as low as 2⁰ C in India and altitudes as high as 1500 meters in Malaysia (Raut & Barker, 2002). In addition, they can thrive in a range of soil pH, moisture and plant cover conditions as well as withstand a diverse group of predators (Mead, 1979). Lastly, *A. fulica* has frequently been associated with human disturbed areas such as agricultural lands and

gardens, though they have also been found in primary and secondary forests in Hawaii, the Bonin Islands, India, Southeast Asia and New Caledonia (Raut & Barker, 2002).

Third, *Achatina fulica* is commonly introduced deliberately and is therefore transported in large numbers and properly cared for, which then increases its chance of survival. In Brazil, *Achatina fulica* was introduced in 1988, probably from Indonesia, when it was heralded as an alternative source of meat. These snails were then distributed for commercial purposes but were subsequently released when people eventually lost interest. As a consequence, Brazil is currently experiencing an explosive stage of the invasion that is characterised by large individuals that are prevalent in urban areas, particularly in gardens (Thiengo *et al.*, 2007). Other deliberate introductions include those in Borneo where duck farmers used the snails as feed (Jarrett, 1931), those in Nepal where the snails were introduced in local gardens and venerated for their religious significance (Budha & Naggs, 2008) and those in the Indo-Pacific where Japanese soldiers and merchants before and during the Second World War used the snails as food and sometimes as pets (Kliks & Palumbo, 1992; Civeyrel & Simberloff, 1996).

Although currently distributed in many areas around the globe, it is possible to elucidate the route of dispersal of *Achatina fulica* since before the 1800's. The snail is thought to be indigenous to East Africa, specifically Kenya and Tanzania. It is believed to have been introduced into Madagascar and Mauritius in the early 19th century (Bequaert, 1950) and subsequently to have spread to the Indian subcontinent, Southeast Asia, most islands of the Pacific (Mead, 1961; Kliks & Palumbo, 1992), the Caribbean (Schotman, 1989) and South America (Paiva, 1999; Thiengo *et al.*, 2007; Borrero *et al.*, 2009). It has also been recently detected in West Africa (Raut & Barker, 2002). There are several pathways or activities that are available to the snail that enables it to spread

quickly to new areas. These are the accidental transfer by humans, usually by hitchhiking onto agricultural machinery and even in cars and trucks (Cowie & Robinson, 2003) and the intentional transfer by humans as pets, for food consumption or for other uses (Mead, 1979; Kliks & Palumbo, 1992; Cowie & Robinson, 2003).

To date, no systematic evaluation of the genetic variation of *Achatina fulica* across global populations has been undertaken. Such data could indicate the number of genetic types that have been introduced in the snail's new range. Furthermore, results from such a global survey would reveal if introduced populations of *A. fulica* are undergoing genetic bottlenecks as a consequence of a lack of genetic variation, or if multiple introductions from various source populations took place that could potentially counteract the effects of bottlenecking.

1.6. Parasites of molluscs

Parasitism is a relationship between two organisms where the parasite thrives on or within the host, which is harmed in some way (Roberts & Janovy, 2005). Many organisms are known to parasitise molluscs. These parasites use molluscs either as intermediate hosts, where the parasite develops but does not reach sexual maturity, as definitive or final hosts where the parasite reaches the adult stage, or as permanent hosts where the entire development of the parasite takes place in the host (Malek & Cheng, 1974; Roberts & Janovy, 2005). Examples of known parasites of molluscs include bacteria such as *Mycobacterium* in the gastropod *Helisoma anceps* (Malek & Cheng, 1974) and *Aeromonas hydrophila* in *Achatina fulica* (Dean *et al.*, 1970), protozoa such as *Hartmanella* sp. in the gastropod *Biomphalaria* (Malek & Cheng, 1974), trematodes such as *Schistosoma japonicum* in the gastropod *Oncomelania quadrasi* (Malek & Cheng, 1974) and *Echinostoma* in the apple snail *Pomacea* (Hollingsworth & Cowie,

2006), and copepod arthropods such as *Mytilocola intestinalis* in the blue mussel *Mytilus edulis* (Malek & Cheng, 1974) and nematodes such as *Oslerus ostratus* in the pulmonate slug *Laevicaulis alte* and *Nemhelix bakeri* in the pulmonate snail *Helix aspersa* (Grewal *et al.*, 2003). Molluscs act as intermediate hosts for several medically important parasites, as exemplified by trematodes such as *Schistosoma* that leads to schistosomiasis (a liver disease characterised by an enlarged liver and spleen, diarrhea, and bloody urine) (Roberts & Janovy, 2005), *Fasciola* that induces fasciolariasis (necrosis of the liver) (Roberts & Janovy, 2005), *Echinostoma* that causes echinostomiasis (an intestinal disease that leads to headache, dizziness, gastric pain, anemia and diarrhea) (Hollingsworth & Cowie, 2006) as well as nematodes such as the rat lungworm *Angiostrongylus cantonensis* that leads to eosinophilic meningioencephalitis (see below).

1.7. Nematode parasites of *Achatina fulica*

Various nematodes are associated with molluscs (Grewal *et al.*, 2003; Morand *et al.*, 2004), including *Achatina fulica*. Members of the phylum Nematoda are typically bilaterally elongated and tapering at both ends. They also have a characteristic body cavity called a pseudocoelom that is derived embryologically from the blastocoel, which normally disappears during gastrulation in eucoleomate animals but not in nematodes. These animals possess a non-cellular body covering called the cuticle that is shed four times during their lifetime through moulting; these worms therefore have four juvenile stages (sometimes incorrectly referred to as larval stages) that resemble the final adult stage in form. Nematodes include sexually dimorphic species (e.g. the rat lungworm *Angiostrongylus cantonensis*) as well as hermaphroditic species (e.g. the soil nematode *Caenorhabditis elegans*). Other characteristics include a complete digestive system,

absence of circular muscles in the body cavity, and in the case of sexually dimorphic species, the presence of females that are generally larger and males that have a more curled tail (Roberts & Janovy, 2005).

Molluscs are infected by nematodes either as intermediate hosts of the juvenile worms or as definitive or final hosts of the adult worms (Grewal *et al.*, 2003; Morand *et al.*, 2004). In total, 108 species have been found to infect gastropods, with 61 using these gastropods as intermediate hosts and 47 using gastropods as final hosts (Grewal *et al.*, 2003). Evolutionarily speaking, it has been postulated that parasitic nematodes started out as facultative parasites in which free-living forms accidentally infected animals (Adamson, 1986). Morand *et al.* (2004) used Blaxter *et al.*'s (1998) phylogeny of the Nematoda to map the occurrence of parasitism in terrestrial molluscs among the different nematode groups. They hypothesised that nematode parasitism in terrestrial molluscs occurred independently at least five times.

Very limited data are available on nematode species associated with *Achatina fulica*. Nematodes shown to be associated with *A. fulica* include *Oslerus ostratus*, which uses *A. fulica* as an intermediate host (Grewal *et al.*, 2003), *Rhabditis* sp., most likely a facultative parasite of the snail (Viyada, 2005), and the medically important *Angiostrongylus cantonensis*, the rat lungworm, which uses the snail as an intermediate host (see Mead, 1979 for a detailed list of *A. fulica* survey for *A. cantonensis*).

1.8. *Angiostrongylus cantonensis*: the nematode that hitched along

Angiostrongylus cantonensis is a parasite of rodents that requires a gastropod intermediate host such as *Achatina fulica* to complete its life cycle. The nematode's definitive or final hosts are murid rodents such as the black rat (*Rattus rattus*), the brown rat (*Rattus norvegicus*), the Pacific rat (*Rattus exulans*), the oriental house rat

(*Rattus tanezuni*), the Philippine forest rat (*Rattus everetti*) and the malabaric bandicoot rat (*Bandicota malabarica*) (Alicata, 1966; Westerlund & Chamberlain, 1969; Marquardt *et al.*, 2000). Humans can also get infected by acquiring the 3rd juvenile stage of the worm from several of its snail or slug intermediate hosts, including *Achatina fulica* (Alicata, 1966; Marquardt *et al.*, 2000), which leads to a disease of the nervous system called eosinophilic meningoencephalitis (EME) or angiostrongyliasis (Marquardt *et al.*, 2000).

1.8.1. The taxonomy of *Angiostrongylus cantonensis*: Nematoda, Rhabditea, Strongylida, Metastrongyloidea, Angiostrongylidae

Angiostrongylus cantonensis belongs to the phylum Nematoda, a group including some of the most abundant animals in the world. There are more than 25,000 described species of nematodes, with 10,000 known to be free-living and more than 15,000 known to be parasitic on animals (Poulin & Morand, 2000; Hugot *et al.*, 2001). Roberts & Janovy (2005) predicted that current figures pertaining to the diversity of the Nematoda are underestimates and that there may be more species of nematodes than there are of insects, which Ruppert *et al.* (2004) estimate to be around 30 million. Nematodes can be divided into two main classes, the Enoplea and the Rhabditea, based on the presence of certain sensilla or small sense organs. Members of the Enoplea possess pouch-like anterior sensilla called amphids whereas members of the Rhabditea, including *Angiostrongylus*, have ventrally coiled amphids; many rhabditeans also possess sensilla called phasmids near the posterior end. Within the Rhabditea is the order Strongylida that includes long, slender worms with males having the characteristic copulatory bursa supported by sensory rays (Roberts & Janovy, 2005). Within the Strongylida is the superfamily Metastrongyloidea whose members utilise mammals as

definitive hosts, with many occupying the host lungs during the adult stage (Anderson, 2000).

The genus *Angiostrongylus* belongs to the family Angiostrongylidae in the Metastrongyloidea. Members of this family have an adult stage that possesses a posterior vulva (Anderson, 2000) but not a buccal cavity or lips at the mouth. They reside in the lungs of the mammalian final hosts and require an invertebrate intermediate host (Roberts & Janovy, 2005). *Angiostrongylus* itself was regarded by Ubelaker (1986) as a heterogeneous group, and he suggested that, on the basis of the morphological differences in the copulatory bursa in adult males as well as differences in their mammalian final host specificity, the genus should be split into five distinct genera including *Parastrongylus* (which Ubelaker was resurrecting and was first described by Baylis in 1928 using *Parastrongylus tateronae* as the type species) as well as *Angiostrongylus* (first described by Baillet in 1866 using *Angiostrongylus vasorum* as type species). In *Parastrongylus*, the lateral rays of the copulatory bursa arise from a single common trunk in adult males whereas those in *Angiostrongylus* arise separately (Fig. 1.3). In terms of their final hosts, *Parastrongylus* utilises murid rodents whereas *Angiostrongylus* infects carnivores like dogs, foxes and cats (Ubelaker, 1986). Based therefore on these characteristics, Ubelaker reclassified *Angiostrongylus cantonensis* as *Parastrongylus cantonensis*. Other members of *Parastrongylus*, according to Ubelaker, include *Parastrongylus costaricensis* (Central and South America, Cuba and southern North America), *P. dujardini* (Europe) and *P. malaysiensis* (Malaysia). Species retained by Ubelaker in the genus *Angiostrongylus* are *A. vasorum* in foxes and dogs (Africa, Europe, North America and South America) (Anderson, 2000) and *A. chabaudi* in wild cats (central Italy) (Ubelaker, 1986).

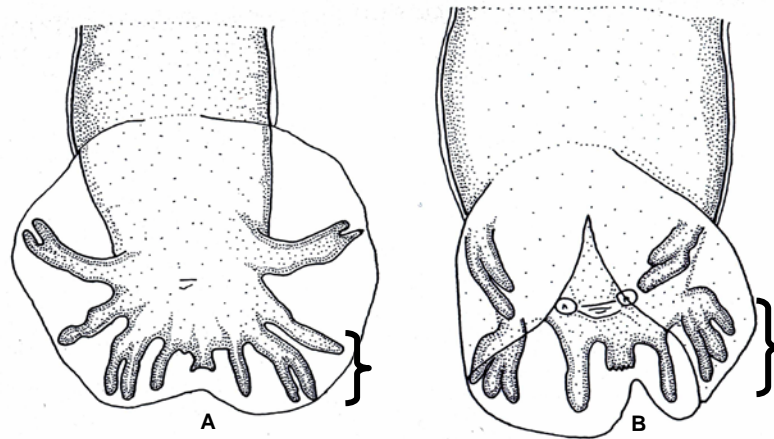


Figure 1.3: Adult male bursa of (A) *Angiostrongylus*, i.e. *A. vasorum*, and (B) *Angiostrongylus* (=Parastrongylus), i.e. *A. cantonensis*. Brackets show lateral rays. Note that the lateral rays arise from a common trunk in (B) but not in (A). From Ubelaker (1986).

The recognition of *Parastrongylus* as a separate genus, however, is not widely accepted and the name *Angiostrongylus* is still used to refer to the genus of *A. cantonensis* and the other species mentioned in the previous paragraph (Kliks & Palumbo, 1992). Molecular data to confirm or refute Ubelaker's designation are also wanting.

1.8.2. Morphology of *Angiostrongylus cantonensis*

Adult female *A. cantonensis* range from 17 to 33 mm long and 0.28 to 0.5 mm wide, whereas the males tend to be smaller, being only 15 to 22 mm long and 0.25-0.35 mm wide. The worms feed on blood, thus giving a red appearance to their intestines. In females, the intestines intertwine with the uterine tubules, which lend a characteristic barber pole appearance (Marquardt *et al.*, 2000; Roberts & Janovy, 2005). The infective 3rd juvenile stage of the parasite is considerably smaller, around 425-524 µm long and 23-34 µm wide. Although the juvenile is similar morphologically to other

species of lungworms like *Aulurostrongylus abstrusus* and *Anafilaroides rostratus*, it can be subtly distinguished by the fine point termination of the tail (Ash, 1970).

1.8.3. The life cycle of *Angiostrongylus cantonensis* (Figure 1.4)

The mature adult worms reside for approximately two weeks in the subarachnoid space separating the meninges of the rat brain. After that period, they move into the circulatory system through the venous system until they reach the heart where the females begin laying their eggs. The arterial blood then brings these eggs to the lungs where they get lodged in the alveoli. As the 1st juvenile stage emerges from the eggs, they are passed from the lungs into the trachea until they reach the gut and stay there for 42-45 days before being eliminated through the faeces (Marquardt *et al.*, 2000; Kliks & Palumbo, 1992). A snail or slug intermediate host then gets infected by these juveniles either by ingesting contaminated rodent faeces or by burrowing of the nematodes through the gastropod body wall or respiratory pores (Hollingsworth & Cowie, 2006). These nematodes then progress into the 2nd and 3rd juvenile stages after about 18 days (Marquardt *et al.*, 2000) where they reside mostly in the head-foot region of the gastropod intermediate host, although they can also be found in the lungs, liver and kidneys (Hollingsworth & Cowie, 2006). The 3rd stage juveniles are passed on to definitive hosts such as rats when they ingest infected gastropod intermediate hosts. In the absence of a definitive host ingesting the intermediate host, the 3rd stage juveniles can become quiescent and remain in the intermediate host tissue for months (Hollingsworth & Cowie, 2006). Occasionally, crabs, prawns and planarians act as paratenic hosts in which they passively carry 1st to 3rd juvenile stage worms; consumption of such paratenic hosts can pass on the 3rd stage juvenile worms (Kliks & Palumbo, 1992; Hollingsworth & Cowie, 2006). Once inside the definitive host, the

nematodes normally take only one to two days before they reach the central nervous system through the blood where they mature into adult worms after about 13 days (Kliks & Palumbo, 1992; Marquardt *et al.*, 2000).

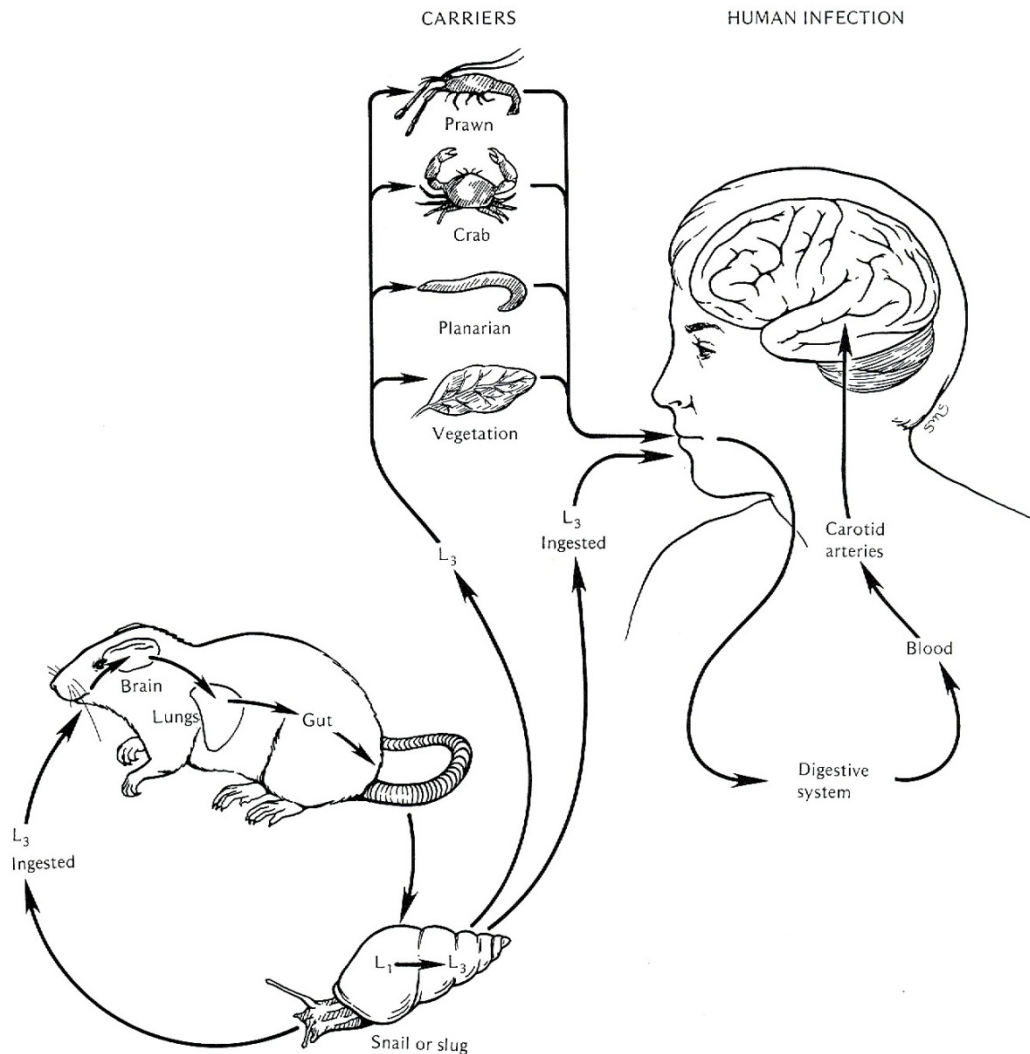


Figure 1.4: Life cycle of *Angiostrongylus cantonensis*. (From Marquardt *et al.*, 2000).

1.8.4. Natural intermediate hosts of *Angiostrongylus cantonensis*

Apart from *Achatina fulica*, there are several other gastropods that serve as natural intermediate hosts of *A. cantonensis* (Anderson, 2000). Gastropods so far identified as hosts include the following: *Hemiplecta sagittifera*, *Helicostyla*

macrostoma, *Cyclophorus* sp., *Chlorea fibula* (Westerlund & Chamberlain, 1969), *Imerinia plebia* (Salazar & Cabrera, 1969), *Bellamya ingallsiana*, *Bradybaena similis*, *Cipangopaludina chinensis*, *Deroceras laeve*, *Euglandina rosea*, *Girasia peguensis*, *Indoplanorbis exustus*, *Laevicaulis alte* (also referred to by the name of *Veronicella altae* in the literature), *Macrochlamys resplendens*, *Microparmarion malayanus*, *Opeas japonicum*, *Pupina complanata*, *Pila ampullacea*, *P. scutata*, *Quantula striata*, *Sarasinula plebeia* (= *Vaginalus plebeius* in the literature) *Subulina octona* (Anderson, 2000) and *Pomacea canaliculata* (= *Ampullarium canaliculatus* in the literature) (Tsai *et al.*, 2001b). Other gastropods that were found experimentally to be susceptible to *A. cantonensis* infection could potentially become intermediate hosts. They are: *Biomphalaria* spp., *Bithynia* sp., *Bradybaena oceania*, *Bulinis* spp., *Deroceras reticulatum*, *Drepanotrema simmonsii*, *Euglandina rosea*, *Euhadra hickonsis*, *Ferrissia tenuis*, *Fossaria ollula*, *Fruticola despecta*, *Helicina orbiculata*, *Helisoma* sp., *Indoplanorbis exustus*, *Lanistes carinatus*, *Limax arborum*, *L. flavus*, *L. maximus*, *L. marginalis*, *Lymnaea* spp., *Marisa cornuarietis*, *Mesodon thyroidus*, *Onchidium* sp., *Physa acuta*, *Planorbis planorbis*, *Plesiophysa hubendicki*, *Segmentina hemisphaerula*, *Semisalcospira libertina*, *Stagnicola elodes* and *Succinea lauta* (Anderson, 2000).

1.8.5. Medical importance of *Angiostrongylus cantonensis*

Angiostrongylus cantonensis is the causative agent of eosinophilic meningoencephalitis (EME) or angiostrongyliasis in humans. In recent years, the geographic range of the nematode and the number of types of animal it infects have been growing rapidly, making EME an ‘emerging’ disease (Prociv *et al.*, 2000). People accidentally acquire the 3rd juvenile stage through any of the following routes of infection: (1) eating raw or undercooked snails or slugs (Marquardt *et al.*, 2000); (2)

drinking water or food such as raw vegetables or salads contaminated with the parasite such as raw vegetables for salads (Wallace & Rosen, 1969; Marquardt *et al.*, 2000); (3) eating raw or undercooked paratenic hosts like crabs, prawns and (by accident) planarians such as *Platydemus* (Kliks & Palumbo, 1992; Marquardt *et al.*, 2000); or (4) handling infected snails followed by failure to wash hands (Wan & Weng, 2004). Since humans are not the natural definitive hosts of *A. cantonensis*, these worms do not mature into adults inside the human body; however, their presence elicits a whole range of clinical manifestations typical of eosinophilic meningoencephalitis. Symptoms of this disease include headaches, stiffness of the neck, vomiting, paresthesia or abnormal sensation, fever, paralysis and tremors. In response to the presence of the worm, there is also a marked increase in the cerebrospinal fluid and peripheral blood of eosinophils, white blood cells responsible for combating infection and parasites (Kliks & Palumbo, 1992; Marquardt *et al.* 2000; Lee, 2002). Cases of EME are rarely fatal, with patients recovering after the symptoms subside within ten weeks (Hollingsworth & Cowie, 2006). It is not known how many infective 3rd stage juveniles are necessary to elicit EME in humans (Prociv *et al.*, 2000), although fatal cases often involve infection of hundreds or thousands of nematodes, as in the case of one Korean fisherman who died after consuming infected *Achatina fulica* in Pago Pago, American Samoa (Kliks *et al.*, 1982). To date, there is no known antihelminthic drug against *A. cantonensis*, although it is possible to treat infection at an early stage using thiabendazole (Roberts & Janovy, 2005). Most doctors are wary of recommending this drug, however, since worms may be more dangerous dead than alive as they tend to elicit a strong inflammatory reaction that could lead to the formation of a large granuloma and cause further damage (Marquardt *et al.*, 2000; Roberts & Janovy, 2005). On the other hand, mebendazole combined with corticosteroids have been successfully used to treat the symptoms of *A.*

cantonensis-induced meningoencephalitis and shorten the course of infection (Tsai *et al.*, 2001a; Wan & Weng, 2004). Aside from humans, other mammals such as dogs (Mason, 1987), horses (Costa *et al.*, 2000), tamarins (Carlisle *et al.*, 1998) and even a captive white-handed gibbon (Duffy *et al.*, 2004) have also been reported to be susceptible to *A. cantonensis* infection.

1.8.6. The dispersal of *Angiostrongylus cantonensis*

The expanding range of *Angiostrongylus cantonensis* has been attributed largely to the global distribution of rats, although the role of the intermediate hosts such as the Giant African Land Snail should not be ignored. The nematode is believed to have originated either in East Africa (Alicata, 1966) or South or Southeast Asia (Drozd *et al.*, 1975). Alicata (1966) noted the nearly parallel distribution of *A. cantonensis* and *A. fulica* and postulated that the rapid dispersal of the snail brought about the current geographical distribution of the parasite. Drozd *et al.* (1975), on the other hand, argued that murid rats were responsible, owing to the long association of the worm with its definitive hosts and the dispersal of the murid rats from tropical Asia. It would be valuable to determine which hypothesis is correct as this would identify which host is primarily responsible for the expanding range of the nematode; hence, appropriate measures could be set in place to control the host. Testing the hypothesis could be addressed by conducting a comprehensive survey to identify *A. cantonensis* among global populations of the nematode's definitive and intermediate hosts.

1.8.7. Molecular identification of *A. cantonensis* and other nematodes

It is, however, difficult to identify nematodes such as *A. cantonensis* to named species. Few taxonomic experts are available (Floyd *et al.*, 2002), and many worms

lack suitable morphological characters to facilitate identification, particularly in the infective juvenile stages (Newton *et al.*, 1998). An alternative to morphological identification is the use of DNA ‘barcodes’ that are unique to individual species. DNA barcoding makes use of a small section of a DNA sequence from a standardised region of the genome to identify species (Dasmahapatra & Mallet, 2006). A 650-bp fragment of the 5’ end of the mitochondrial cytochrome *c* oxidase subunit I gene (COI) is most commonly used for barcoding in animals (Hajibabaei *et al.*, 2007), although other genes have also been used. For instance, the small subunit (SSU) rRNA gene was employed to rapidly identify individual free-living marine nematodes in Southwest England (Bhadury *et al.*, 2006); the gene was likewise used on the parasitic nematodes of the sardine *Sardinella pilchardus* (Santos *et al.*, 2006). Through DNA barcoding, unidentified individuals of nematodes such as *Angiostrongylus cantonensis* could be assigned to species, irrespective of life cycle stage. This tool is applied here to rapidly identify 3rd stage juvenile *A. cantonensis* from their gastropod intermediate hosts.

1.9. Major objectives

This thesis will address five major issues concerning *Achatina fulica* over five chapters. A separate chapter, Chapter 2, will summarise all the general protocols to be used for the five chapters.

In Chapter 3, the phylogenetic relationships of the six achatinoid families (Achatinidae, Coelioxidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophorellidae) will be evaluated using an expanded coverage of the rRNA gene cluster as well as the nuclear actin and histone 3 genes and the mitochondrial cytochrome *c* oxidase subunit I and the 16S rRNA genes. The monophyly and validity of these taxonomic families will likewise be assessed.

In Chapter 4, comprehensive molecular-based phylogenetic analyses will be provided for the first time on the Achatinidae using the same molecular markers to be used in Chapter 3 in order to correlate molecular data with the available morphological data and to provide evidence of relationships among the members of this family. The designation of *Lissachatina* as a subgenus of *Achatina* will likewise be evaluated based on molecular data.

In Chapter 5, genetic variation among global populations of *Achatina fulica* will be examined using the 16S rRNA gene. This study will investigate the movement of *A. fulica* out of Africa and its subsequent spread throughout the tropics. It will establish if more than one genetic type has been introduced in the snail's new range and whether genetic bottlenecking is taking place in these populations. The lack of genetic variation, if any, could have a profound impact on the success of *A. fulica* as an invasive species.

In Chapters 6 and 7, focus will be on the nematode parasite of *A. fulica*, *Angiostrongylus cantonensis*. A survey of global populations of *A. fulica* for the parasite will be conducted to determine the possible role of the snail in the spread of the parasite. In Chapter 6, a rapid molecular method of identification for *A. cantonensis* from Philippine samples of *A. fulica* as well as the black slug *Laevicaulis alte* will be developed using a segment of the small subunit (SSU) rRNA gene. Using the same marker, a phylogenetic analysis will also be conducted on representative taxa of the genus *Angiostrongylus* to assess if Ubelaker's designation of *Parastrongylus* has merit based on molecular data. In Chapter 7, global populations of *A. fulica* will be screened for the nematode parasite using the molecular marker developed in Chapter 6. The role of the snail in spreading the parasite will be evaluated in this chapter. In addition, the presence of other types of nematodes in global populations of *A. fulica* will also be reported.

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CHAPTER 2 - General Protocols

Summarized in this chapter are general protocols that are referred to in the succeeding chapters. A brief description, along with some basic concepts, accompanies each method.

2.1. DNA extraction methods

DNA from fresh or preserved samples of snails and nematodes was extracted using several extraction protocols. Each method is described below, along with details of the rationale for their use. As the DNA extracts were used for subsequent PCR amplification, the methods emphasized the removal of polysaccharides and other PCR inhibitors (Demeke & Adams, 1992) as well as nuclease enzymes that could cleave DNA (Rolfs *et al.*, 1992).

2.1.1. DNA extraction of snail tissues using the CTAB protocol (modified from Hillis *et al.* (1996))

This standard and cost-effective protocol was used to extract DNA from snail tissues used for phylogenetic analyses in Chapters 3 and 4. The protocol was applied to fresh, fresh frozen and ethanol-preserved specimens. Traditionally, this technique also includes phenol (Rolfs *et al.*, 1992) but this was excluded here as phenol is a very toxic substance.

1. For ethanol preserved tissues, the tissue slices were soaked in 1 ml TE buffer (10 mM Tris-HCl, 1mM EDTA) for approximately 1 hour in order to remove excess ethanol to soften the tissue prior to DNA extraction.
2. Tissue was cut into small pieces, placed into 500 µl of CTAB solution [100mM Tris-HCl pH 8, 20mM EDTA pH 8, 1.4 mM NaCl, CTAB 2% (w/v)] and ground

using sterile glass beads and a plastic pestle. CTAB (cetyltrimethylammonium bromide) is a non-ionic detergent that precipitates polysaccharides and lyses cells (Richards *et al.*, 1995).

3. 20 μ l of Proteinase K (10mg/ml), an enzyme that digests proteins such as nucleases that cleave naked DNA, was added to each tube. This was followed by 10 μ l of β -mercaptoethanol, which precipitates polyphenolics (Rolfs *et al.*, 1992). The tubes were then vortexed then incubated at 55⁰ C for at least one hour until the tissue slices were completely digested.
4. 500 μ l of ice-cold chloroform-isoamyl alcohol (24:1) was added, after which the tubes were inverted several times for 5 minutes. This separated the DNA from proteins (Rolfs *et al.*, 1992). The tubes were then centrifuged for 10 minutes at 13,000 rpm, after which the aqueous phase was transferred to a new tube (~400 μ l). When necessary (i.e. there were a lot of proteins), step 4 was repeated.
5. 2.5 volumes (~1 ml) of ice-cold 95% ethanol and 1/10 volume (~40 μ l) of 3M sodium acetate (NaOAc) were added to the mix followed by overnight incubation at -80⁰ C to precipitate the DNA (Rolfs *et al.*, 1992). The tubes were centrifuged for 15 minutes at 13,000 rpm, then the supernatant was carefully removed.
6. The remaining pellets were washed with 500 μ l of ice-cold 70% ethanol and centrifuged for five minutes at 13,000 rpm to remove salts and small organic molecules. The ethanol was carefully removed afterwards.
7. The remaining pellets were air-dried on a heat block at 45°C for a maximum of 15 minutes. The pellets were then resuspended in 150 μ l TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.5).

8. Each suspension was then boiled for 15 min at 100°C to denature any remaining enzymes in order to prolong the life of the DNA extract.
9. The DNA extracts were stored at -80⁰ C until use.

2.1.2. DNA extraction of snail tissues using the Nucleon® PhytopureDNA™ Purification Kit

Because of its relatively fast and simple use, the Nucleon® PhytopureDNA™ kit was used as an alternative to the CTAB protocol (Section 2.1.1) in extracting DNA from snail tissues used for phylogenetic analyses (Chapters 3 and 4) as well as for the global genetic survey of *A. fulica* populations (Chapter 5). It produced no discernible difference in DNA quality in comparison to the CTAB method. Like CTAB, this kit allowed for the precipitation of polysaccharides. The protocol below generally followed the manufacturers' instructions except that volumes indicated were half of what was recommended while the times prescribed were lengthened.

1. 300 µl of reagent 1 was added to a tube containing approximately 0.1 g snail tissue from the foot muscle that was previously ground using sterile glass beads and plastic pestle. After mixing thoroughly, 100 µl of reagent 2 was added. The tubes were inverted several times until a homogenous mixture was obtained.
2. The tubes were placed in a 65⁰ C water bath for at least 30 minutes with regular manual agitation until the tissues were fully digested.
3. The tubes were then placed in ice for at least 30 minutes.
4. 250 µl of ice-cold chloroform was added followed by 50 µl Phytopure DNA extraction resin suspension that was thoroughly shaken prior to use. The tubes were inverted for 10 minutes followed by centrifugation at 13,000 rpm for 10

minutes. The DNA-containing phase above the resin layer was then transferred into a fresh tube.

5. An equal volume (~400 µl) of cold isopropanol was added, after which the tubes were gently inverted several times to precipitate the DNA. The tubes were centrifuged at 13,000 rpm for 5 minutes to pellet the DNA. The isopropanol was pipetted out and discarded.
6. The DNA pellet was washed with 1.0 ml 70% ethanol, then centrifuged at 13,000 rpm for 5 minutes. The ethanol was pipetted out and discarded.
7. The remaining DNA pellets in the tubes were air-dried for a maximum of 15 minutes at 45-55⁰ C on a heat block.
8. The DNA pellets were resuspended in 150 µl TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.5) and boiled for 15 minutes at 100°C.
9. The DNA extracts were stored at -80⁰ C until further use.

2.1.3. DNA extraction of snail tissues using the NaOH-lysis method

This method was used on the *Achatina fulica* population samples (Chapter 5) and was chosen over the two previous methods because it was considerably cheaper to use even if the quantity of DNA extracted was slightly less than that extracted in the previous methods. This disadvantage was offset by using more tissue, which could be readily obtained when using *A. fulica* population samples.

1. Tissue slices were ground in microfuge tubes containing 200 µl of 0.1 N NaOH using sterile glass beads and a plastic pestle. The samples were boiled at 95-100⁰ C for 20 minutes to lyse the cells.
2. 100 µl of sterile distilled water and 300 µl of chloroform-isoamyl alcohol (24:1) were added. The tubes were vortexed, then centrifuged at 13,000 rpm for 10

minutes. The upper phase (~300µl) was collected and transferred into new tubes.

3. An equal volume of isopropanol (~300µl) was added to precipitate the DNA. The tubes were inverted several times, then stored at -80⁰ C for at least one hour.
4. The tubes were centrifuged at 13,000 rpm for 15 minutes, after which the isopropanol was carefully removed.
5. The pellets were washed with 500 µl of 70% ice cold ethanol then centrifuged for 5 minutes at 13,000 rpm to remove salts and small organic molecules. The ethanol was carefully removed.
6. The DNA pellets were air dried on a heat block at 45°C for a maximum of 15 minutes, after which they were resuspended in 150 µl TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.5). Each suspension was then boiled for 15 minutes at 100°C.
7. The DNA extracts were stored at -80⁰ C until further use.

2.1.4. DNA extraction of single nematodes using NaOH digestion

This method is a modification of the NaOH lysis method used to extract DNA from individual nematodes (Floyd *et al.*, 2002). The method was used on nematodes extracted from the gastropod intermediate hosts (Chapters 6 and 7) as well as on the adult angiostrongylid nematodes used for phylogenetic analyses and as references for molecular barcoding (Chapter 6).

1. Each nematode was placed in a separate tube containing 20 µl 0.25 M NaOH to digest the cuticle and cells. The tubes were centrifuged briefly to ensure that the nematodes were fully immersed in NaOH. This was followed by overnight

- incubation at 25⁰ C (or 3-16 hours; beyond that, over-digestion could lead to less intense or infrequent bands during PCR amplification).
2. The tubes were heated for three minutes at 95⁰ C, then cooled to room temperature. The tubes were centrifuged briefly to collect any liquid along the sides and the lid.
 3. The following were added in succession: 4 µl 1.0 M HCl to neutralize the NaOH; 10 µl 0.5 M Tris-HCl, and 5 µl 2% Triton X-100, a strong detergent that disrupts the cell membranes and effectively lyses the cells.
 4. The tubes were mixed and centrifuged briefly, then heated for 3 minutes at 95⁰C. The samples were cooled to room temperature (pH of digest should be between 8 and 9).
 5. The DNA extracts were stored at -80⁰ C until further use.

2.2. Polymerase chain reaction (PCR)

Polymerase chain reaction (PCR) entails the rapid amplification *in vitro* of specific DNA segments using the same principles involved in DNA replication. This technique was used to amplify fragments of the nuclear ribosomal (r)RNA cluster, actin and histone 3 genes of snails and the small subunit (SSU) rRNA of nematodes as well as the mitochondrial cytochrome c oxidase subunit I (COI) and 16S rRNA genes of snails. Crucial in any successful PCR is the pair of primers that bind to specific regions of the complementary strands and the DNA polymerase that facilitates the amplification. Other components of PCR are the deoxynucleotide triphosphates (dNTPs) that are incorporated in the growing chain, and MgCl₂ that acts as the enzyme co-factor of the DNA polymerase. There are three stages in PCR that are repeated over a number of cycles; these are: (1) denaturation of the double stranded DNA at 94⁰ C; (2)

annealing of the PCR primers to target sites in the DNA at specific temperatures; and (3) extension of the growing strand of synthesized DNA at 72⁰ C through the activity of the DNA polymerase (Reece, 2004).

2.2.1. PCR primers

Primers are short oligonucleotides (17-30 nucleotides) that bind to specific targets in the DNA (Reece, 2004). Several factors need to be considered for PCR primers to successfully amplify their intended targets. First, the melting temperature (T_m) of each primer in the primer pair, calculated as $2(A+T)+4(C+G)$, where A, C, G and T refer to the nitrogenous bases of the oligonucleotide, should be approximately equal so that they anneal to their target sites at almost the same time (Reece, 2004). Second, primers should not have strings of repeated nucleotides within their sequences in order to prevent annealing to non-specific targets that exhibit strings of complementary repeated nucleotides (Reece, 2004). Third, primers should not contain complementary sequences with each other or within themselves as these would result in primer dimers and secondary structures, respectively, which would lead to non-amplification (Reece, 2004). Fourth, the 3' end of the primers should match perfectly the target region for the polymerase enzyme to properly extend the primer beginning at the 3' end (Reece, 2004). Fifth, provided that the 3' ends of the primers match with their targets, some mismatched pairing could form partial bonds between the primer and the target. For example, G-T bonds also occur with some stability. Such a phenomenon could be taken advantage of when designing primers that need to be applied across many taxa in which ambiguous positions could potentially be present in the primer-binding sites. If a position in the target region could either be a C or a T, then the primer would be assigned a G for that position in order to bind to either C or T of the

target. Likewise, if the target contains a position that could either be an A or a G, then the primer that binds to it should preferentially contain a T for that corresponding position (Palumbi, 1996). Sixth, the length of the product being amplified by the primer pair would determine the duration of the extension step of the PCR. For instance, 30 seconds are generally needed to amplify products less than 500 bp, 60 seconds for products between 500 and 1500 bp, and 90 seconds for products longer than 1500 bp (Palumbi, 1996). Lastly, more than one round of PCR could be undertaken to ensure the success of amplification of the target segment. For instance, the product of the first round of PCR using one set of primers could be used as template for the second round using an internal set of primers in a two-step process called nested PCR (Aurelius *et al.*, 1991). This was applied to the rRNA cluster and the actin gene in which the first pair of primers amplified a larger fragment (approximately 4000 bp for the rRNA cluster and around 900 bp for the actin) while the second round made use of primer pairs that bound to targets within the larger fragments. This process facilitated the annealing of the primers in the second round where the target regions were more readily available.

2.2.1.1. Snail PCR primers

2.2.1.1.1. Nuclear genes

2.2.1.1.1.1. Ribosomal RNA (rRNA) gene cluster

The nuclear ribosomal RNA gene family encodes the small and large subunit rRNA genes that are incorporated into the small subunit and large subunit, respectively, of the ribosome that facilitates gene translation into amino acid sequences (Lewin, 2008). The gene family consists of five parts that are transcribed as a single unit: (1) the small subunit rRNA gene (SSU or 16-18S); (2) the first internal transcribed spacer (ITS 1); (3) the 5.8S gene; (4) the second internal transcribed spacer (ITS-2); and (5) the LSU rRNA gene (26-28S). A 5S gene found downstream of the LSU gene is separated from this array by an intron or a non-transcribed spacer and is independently transcribed in eukaryotes (Graur & Li, 2000; Lafontaine & Tollervey, 2001; Klug *et al.*, 2007). The ribosomal RNA gene cluster is repeated in a tandem array over the genome, with the actual number of copies varying across different taxa; for instance, nematodes have 50 to 100 copies, some mammals up to several hundred copies, while plants have up to thousands (Long & Dawid, 1980). The function of these genes in protein translation means that the genes are required in many copies and that these copies should be homogenous. The identity of these copies within a species is maintained by concerted evolution through purifying selection in which new variants are eliminated, thus leading to sequence homogeneity within a species (Gasser & Newton, 2000; Graur & Li, 2000). Variation between copies of the rRNA array in a species is rare; in land snails, variation has only ever been found in the ITS as detected by ambiguous positions in direct sequences of the ITS (Wade, pers comm.). The choice of this gene cluster for

phylogenetic study is ideal for several reasons: (1) it is easy to PCR amplify because more copies are available for the PCR primers to anneal to; (2) the genes are present in all animal taxa because of their conserved function in RNA translation into proteins; and (3) regions within the genes exhibit variable evolutionary rates, with some regions that are very conserved and are thus ideal for primer binding sites, while other regions are variable enough to be phylogenetically informative (Hillis & Dixon, 1991).

The primers listed in Table 2.1 were used to amplify an approximately 4000 nucleotide fragment of the rRNA gene cluster that was employed as a marker for the snail phylogenetic analyses (Chapters 3 and 4). The amplified region included nearly the entire large subunit (LSU) rRNA gene as well as the internal transcribed spacer 2 (ITS 2) region and about 80 nucleotides of the 5.8S rRNA gene (Figure 2.1). Actual differences in length of the amplified products among taxa were predominantly due to variations in length in the ITS 2 region and to a lesser extent the minor variations in the variable regions in the LSU. The rRNA cluster was amplified using nested PCR. The first round made use of the LSU-1 and LSU-12 or the LSU-2 and LSU-12 primer pairs. This served as the 1⁰ PCR for the rRNA gene fragment. Using the primary PCR products as template, the succeeding rounds of PCR (2⁰ PCR) amplified the internal rRNA fragments (fragment A using primers LSU-1/1iii and LSU-3/3iii; fragment B using LSU-2 and LSU-5; fragment C using primers LSU-4ii and LSU-7/7i; fragment D using LSU-6/6ii and LSU-9/9ii; fragment E using LSU-8/8ii and LSU-11/11ii; and fragment F using LSU-10/10i and LSU 12/12i; see also Table 2.1).

Table 2.1: Summary of the rRNA primers

LSU Fragment	Primers	Reference	Fragment Size (bp)
A	LSU-1 (sense): 5'-CTAGCTGCGAGAATTAATGTGA-3' LSU-3 (anti-sense): 5'-ACTTTCCCTCACGGTACTTG-3'	Wade & Mordan (2000); Wade <i>et al.</i> (2001); Wade <i>et al.</i> (2006)	~900-1200 (difference due mostly to variable size of the ITS2 region; see also Fig. 2.1)
	LSU-1iii (sense): 5'-TGCGAGAATTAATGTGAATTGC-3' LSU-3iii (anti-sense): 5'-ACGGTACTTGTCCGCTATCG-3'	designed by C. Wade	
B	LSU-2 (sense): 5'-GGGTTGTTTGGGAATGCAGC-3' LSU-5 (anti-sense): 5'-GTTAGACTCCTTGGTCCGTG-3'	Wade & Mordan (2000); Wade <i>et al.</i> (2001); Wade <i>et al.</i> (2006)	~580
C	LSU-4ii (sense): 5'-GTCGGCATTCCACCCGACC-3' LSU-7 (anti-sense): 5'-GCAGGTGAGTTGTTACACACTC-3' LSU-7i (anti-sense): 5'-GTTGTTACACACTCCTTAGCGG-3'	designed by C. Wade	~700
D	LSU-6 (sense): 5'-AAGGTGCCAAACGCTGACGC-3' LSU-6ii (sense): 5'-GTGCCAAACGCTGACGCTCA-3' LSU-9 (anti-sense): 5'-CAGTCCTCAGAGCCAATCCTT-3' LSU-9ii (anti-sense): 5'-ACCCAGTCCTCAGAGCCAATC-3'	designed by C. Hudelot	~850
E	LSU-8 (sense): 5'-CCATATCCGCAGCAGGTCTC-3' LSU-8ii (sense): 5'-GTGCACAGCCTCTAGTCGATA-3' LSU-11 (anti-sense): 5'-CTGAGCTCGCCTTAGGACAC-3' LSU-11ii (anti-sense): 5'-TCCTCCTGAGCTCGCCTTAG-3'	designed by C. Hudelot	~850
F	LSU-10 (sense): 5'-ATCCGCTCTGAAGACAGTGTC-3' LSU-10i (sense): 5'-GGCCGCGATCCGTCTGAAGA-3' LSU-12 (anti-sense): 5'-TTCTGACTTAGAGGCGTTCAG-3' LSU-12i (anti-sense): 5'-GGCTTCTGACTTAGAGGCGTT-3'	designed by C. Hudelot	~500

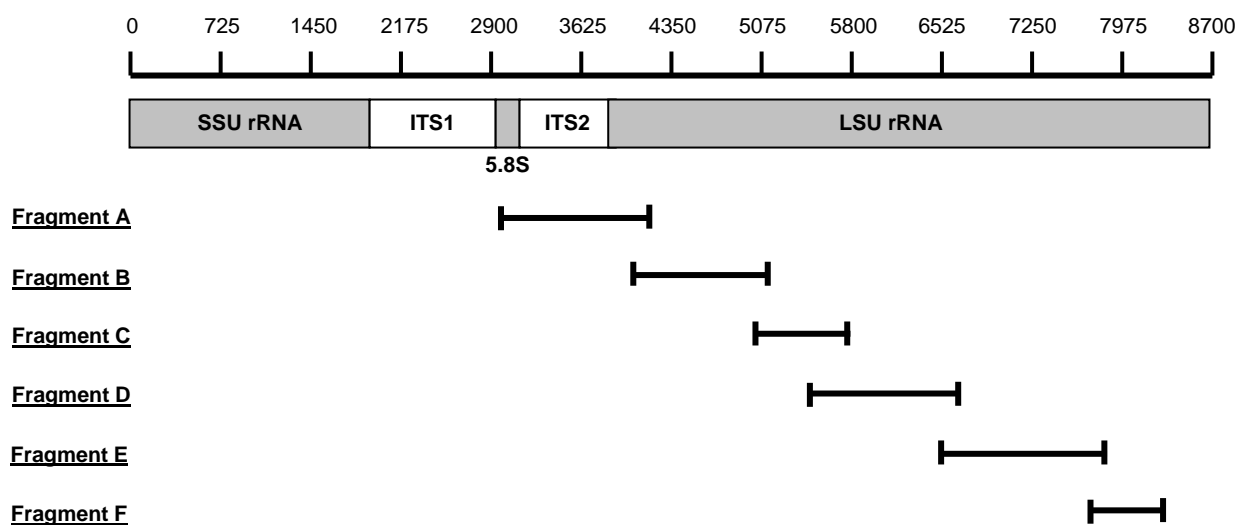


Figure 2.1: Schematic diagram of the rRNA gene cluster based on the complete sequence of *Rattus norvegicus* (GenBank X00133) and showing the position of the primers used in this study.

2.2.1.1.1.2. Actin gene

Actin is a protein involved in various functions such as muscle contraction, cell division and differentiation (Hightower & Meagher, 1986; Hernan, 1993) and is highly conserved across animal groups (Hightower & Meagher, 1986; Palumbi, 1996). Actin is coded by several genes that arose from gene duplication and divergence, resulting in the presence of different forms (isoforms) of the actin protein within an organism (Hightower & Meagher, 1986). These protein isoforms can be classified into two broad categories: the muscle (α) type and the cytoplasmic (β) type (Vandekerckhove & Weber, 1978; Adema, 2002), with the number of isoforms for each category varying between animal groups. In mammals, there are two cytoplasmic actins and four muscle-type actins (Vandekerckhove & Weber, 1978); the same numbers were also found in *Drosophila* (Fyrberg *et al.*, 1981). In molluscs, the number and type of actin isoforms appear to vary among taxa based on the few that have been evaluated to date. DesGroseillers *et al.* (1994) found between three and five genes in the sea hare *Aplysia*

californica that probably coded for the muscle type actin, while Patwary (1996) found 12-15 genes in the bivalve *Placopecten magellanicus* that were also of the muscle type. Carlini *et al.* (2000) surveyed 44 taxa among the coleoid cephalopods and found at least one gene each of the muscle type and cytoplasmic type actins and a third that was most probably a pseudogene. Adema (2002) sequenced one gene of a cytoplasmic actin isoform in each of six planorbid basommatophoran species using cytoplasmic actin-specific primers. However, Southern blotting and hybridization experiments using probes specific for actin yielded up to five genes in *Biomphalaria glabrata* and *Helisoma trivolis*, prompting Adema not to discount the possibility that some of these genes could code for muscle type actin isoforms. Morgan *et al.* (2002) extended the actin survey of the planorbids to 38 taxa using the cytoplasmic actin to correlate the relationship of these snails to the evolution of their *Schistosoma* trematode parasite. Using actin for phylogenetic analyses could pose a problem if the sequences being compared are paralogous, which resulted from gene duplication, rather than orthologous, as in the case of muscle and cytoplasmic actins that arose from gene duplication in the distant past or even within each type due to more recent duplications. Using paralogous sequences could lead to inferring incorrect relationships; awareness of their presence in the sequence data is therefore crucial in any phylogenetic analysis (Bailey *et al.*, 2003). Donald *et al.* (2005) acknowledged the problem of paralogs in actin, but they justified the use of actin to elucidate the phylogeny of marine top shells by employing primers specific for the gene that codes for a single isoform of the cytoplasmic actin and by showing agreement of the actin phylogeny with the 16S rRNA and COI phylogenies. How to address the issues of paralogous sequences in the actin gene is discussed further in Sections 2.9.12 and 2.9.13, pp. 89-91.

The primers listed in Table 2.2 amplified an approximately 900 bp fragment of the actin gene; this fragment was used for the snail phylogenetic analyses in Chapters 3 and 4. Nested PCR was carried out in which a primary PCR product was amplified using the primers ActF2 and ActR1; this fragment was then used as template for the secondary PCR using ActF1 and ActR. If amplification failed, several combinations of primers from the primary and secondary PCR were tried until a product was amplified.

Table 2.2: Summary of the actin primers.

Primers	Reference	Fragment Size (bp)
ActF1 (sense): 5' -TATGTTGGTGATGAGGCTCAG-3'	Morgan <i>et al.</i> (2002)	~900
ActF2 (sense): 5' -GGTATGGGTCAGAAGGACAGCTATG-3' ActR1 (anti-sense): 5' -GAAGCATTCCTGTGGTCAATG-3' ActR (anti-sense): 5' -GATCCACATCTGTTGGAAGGT-3'	Designed by C. Hudelot	

2.2.1.1.1.3. Histone 3 gene

Histone 3 (H3) forms part of the histone core protein octamer that packs the eukaryotic chromatin into bead-like structures (Lewin, 2008). Histone core proteins are some of the most conserved proteins known, suggesting a function that is identical across eukaryotes (Lewin, 2008). Histone genes occur as a tandemly repeating unit comprising the different genes that code for the subunits of the core protein octamer. In *Drosophila*, the repeating unit consists of five genes in the following order, H1, H2A, H2B, H3 and H4; this repeating unit occurs 110 times (Lifton *et al.*, 1977). In bivalve molluscs, a different order was found in the genome of the bivalve mussel *Mytilus galloprovinialis*: H4, H2B, H2A, H3 and H1, with the copy number averaging more

than 200 per haploid genome (Eirin-Lopez *et al.*, 2004). Transcription of the H3 and H4 genes in the stylommatophorans occurs in opposite directions as in other protostomate animals (Armbruster *et al.*, 2005) but not in the bivalve *Mytilus* (Eirin-Lopez *et al.*, 2004). Among the gastropods, Colgan *et al.* (2000) noted a high codon usage bias for the H3 gene. As with the rRNA genes, H3 and H4 genes are subject to concerted evolution, thus leading to sequence homogeneity among copies of the genes within the species (Liao, 1999). The primers listed in Table 2.3 amplified a 331 bp fragment that was also used for the snail phylogenetic analyses in Chapters 3 and 4. Different primer combinations amplified roughly the same fragment.

Table 2.3: Summary of the histone 3 primers

Primers	Reference	Fragment Size (bp)
H3aF (sense): 5'-ATGGCTCGTACCAAGCAGACVGC-3' H3aR (anti-sense): 5'-ATATCCTTRGGCATRATRAGTGAC-3'	Colgan <i>et al.</i> (1998)	328
H3Fm (sense): 5'-ATGGCTCGTACCAAGCAGAC-3' H3Fm1 (sense): 5'-ATGGCTAGAACGAAGCAGAC-3' H3Rm (anti-sense): 5'-TCCTTGGGCATGATGGTGAC-3' H3Rm1 (anti-sense): 5'-CCAACTGAATATCTTTGGGCAT-3'	designed by C. Hudelot	331-340

2.2.1.1.2. Mitochondrial primers

2.2.1.1.2.1. Cytochrome c oxidase subunit I gene

Cytochrome c oxidase is an enzyme that transfers electrons from cytochrome c to O₂ during the electron transport chain in the mitochondrion (Zubay *et al.*, 1995). A typical enzyme has three functional subunits in which subunits I and II contain the

electron carriers (Alberts *et al.*, 2008). Although the amino acid sequence of the subunit I is conserved across the different animal phyla, the nucleotide sequence is subject to silent mutations (Palumbi, 1996), particularly in the 3rd codon positions. In fact, its variability that yields phylogenetic signal and its robust universal primers have made the COI a marker of choice for DNA barcoding in animals (Hebert *et al.*, 2003). Several primer pairs listed in Table 2.4 amplified a fragment that was used in Chapters 3 and 4. Different primer combinations amplified roughly the same fragment.

Table 2.4: Summary of the cytochrome c oxidase subunit I primers

Primers	Reference	Fragment Size (bp)
LCO 1490 (sense): 5' -GGTCAACAAATCATAAAGATATTGG-3' HCO 2198 (anti-sense): 5' -TAAACTTCAGGGTGACCAAAAAATCA-3'	Folmer <i>et al.</i> (1994)	655
STY_LCOi (sense): 5' -TCAACGAATCATAAGGATATTGG-3' STY_LCOii (sense): 5' -ACGAATCATAAGGATATTGGTAC-3' STY_LCOiii (sense): STY_HCO: (anti-sense) 5' -GAATTAAAAATATATACTTCTGGGTG-3'	designed by I. Fontanilla	628-667

2.2.1.1.2.2. 16S ribosomal (r) RNA gene

The 16S rRNA gene transcribes a ribosomal RNA that folds into a secondary structure following base pairing of the nucleotides within it, after which it is incorporated in the mitochondrial ribosome and is used for translation of proteins (Lewin, 2008). Among the Mollusca, 16S exhibits extreme variation in length, with those of the Stylommatophora being the shortest (Lydeard *et al.*, 2000). Two primer pairs are listed in Table 2.5. The first pair (STY_16Sarm and STY_16Sbrm) amplified a 420-450 bp fragment that was used for the snail phylogenies in Chapters 3 and 4. The second pair (16S1i and 16S_SSCP2i) amplified a 293 bp fragment that was used for the

global population survey of *Achatina fulica* in Chapter 5.

Table 2.5: Summary of the 16S rRNA primers

Primers	Reference	Fragment Size (bp)
(for the phylogeny studies-chapters 3 & 4) STY_16Sarm (sense): 5'-CTTCTCGACTGTTTATCAAAAACA-3' STY_16Sbrm (anti-sense): 5'-GCCGGTCTGAACTCAGATCAT-3'	Bonnaud <i>et al.</i> (1994)	420 – 500
(for the SSCP study-chapter 5) 16S1i: 5'-TGACTGTGCAAAGGTAGCATAA-3' 16S_SSCP2i: 5'-CCTAGTCCAACATCGAGGTC-3'	designed by I. Fontanilla	293

2.2.1.2. Nematode PCR primers

The small subunit (SSU) rRNA, which is a part of the ribosomal gene family, has been used to elucidate the phylogeny of the Nematoda (Blaxter *et al.*, 1998; Meldal *et al.*, 2006). The 5' end of the SSU has also been employed as a marker to identify marine nematodes (Bhadury *et al.*, 2006). The primers listed in Table 2.6 amplified four overlapping fragments that included nearly the entire region of the small subunit (SSU) rRNA gene in nematodes (Blaxter *et al.*, 1998); this fragment was used to infer the phylogeny of the Angiostrongylidae relative to other metastrongylids as well as to determine the most suitable marker to identify *Angiostrongylus cantonensis* in Chapter 6. The relative locations of the binding sites for these primers are shown in Figure 2.2.

Table 2.6: The four overlapping primer pairs used to amplify nearly the entire region (approximately 1670 nucleotides) of the SSU rDNA gene.

	Primers	Reference	Fragment size (bp)
Set A	SSU_F_07 (sense): 5'-AAAGATTAAGCCATGCATG-3' SSU_R_09 (anti-sense): 5'-AGCTGGAATTACCGCGGCTG-3'	Blaxter <i>et al.</i> (1998)	480
Set B	SSU_F_11 (sense): 5'-AAGTCTGGTGCCAGCAGCCGC-3' SSU_R_26 (anti-sense): 5'-CATTCCTGGCAAATGCTTTTCG-3'	Blaxter <i>et al.</i> (1998)	360
Set C	SSU_F_24 (sense): 5'-AGRGGTGAAATYCGTGGACC-3' SSU_R_23 (anti-sense): 5'-TCTCGCTCGTTATCGGAAT-3'	Blaxter <i>et al.</i> (1998)	390
Set D	SSU_F_02 (sense): 5'-GGAAGGGCACCACCAGGAGTGG-3' SSU_R_81 (anti-sense): 5'-TGATCCWKCYGCAGGTTTCAC-3'	Blaxter <i>et al.</i> (1998)	600

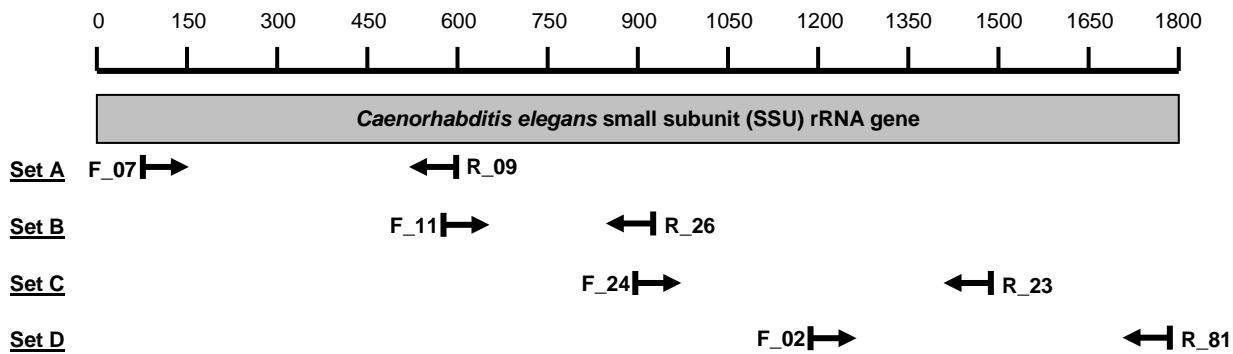


Figure 2.2: Schematic diagram of the SSU rRNA gene based on the complete sequence of *Caenorhabditis elegans* (GenBank X00133) and showing the position of the primers used in this study.

2.2.2. PCR components

Listed in Table 2.7 are the PCR components used and their concentrations. The optimal concentrations varied depending on the size of the gene and the number of copies available per cell. In particular, the concentration of MgCl_2 differed between the nuclear (1.5 mM) and the mitochondrial (2.5 mM) genes.

Table 2.7: PCR components used and their concentrations for the different genes

Components	Initial concentration	Final concentration		Volume (µl)	
Buffer	10X	1X		5.0	
dNTPs	1.25 mM	1 ⁰ PCR for LSU: 300 µM	2 ⁰ PCR for LSU/1 ⁰ & 2 ⁰ PCR for actin/ other genes: 200 µM	1 ⁰ PCR for LSU: 12.0	2 ⁰ PCR for LSU/1 ⁰ & 2 ⁰ PCR for actin/ other genes: 8.0
^a MgCl ₂	50 mM	LSU rRNA/ Actin/ Histone3: 1.5 mM	16S/ Cytochrome c oxidase I: 2.5 mM	LSU rRNA/ Actin/ Histone3: 1.5	16S/ Cytochrome c oxidase I: 2.5
Primer 1	10 mM	200 µM		1.0	
Primer 2	10 mM	200 µM		1.0	
^b Q-solution	5X	1X		10	
Taq	1 unit/µl	0.01 unit/µl		0.5	
Sterile distilled H ₂ O	--	--		To make final volume of 50 µl	

^aSome manufacturers provided PCR buffers that already contained MgCl₂ (e.g. QIAgen™ with 25 mM concentration), while other manufacturers separated the buffer and the MgCl₂ (e.g. Bioline™, which separately provided 50 mM MgCl₂). Care was taken in order to guarantee that the recommended final MgCl₂ concentration was followed.

^bProvided by QIAgen™

2.2.3. PCR running conditions

Different reaction cycles were employed depending on the size of the expected PCR product. For the rRNA gene cluster, a very long extension step (5 minutes) was used in the 1⁰ PCR reaction in order to ensure that the approximately 4000 bp fragment was amplified. For the rRNA 2⁰ PCR reactions, which amplified the shorter internal rRNA fragments, a shorter extension step was used. For all other genes, reaction cycles followed those used for the rRNA 2⁰ PCR reactions.

2.2.3.1. For rRNA 1⁰ PCR

hot start at 96⁰ C for 2 minutes

94 ⁰ C for 30 sec	}	35 cycles
50 ⁰ C for 30 sec		
72 ⁰ C for 5 minutes		

2.2.3.2. For rRNA 2⁰ PCR and all other genes:

hot start at 94⁰ C for 2 minutes

94 ⁰ C for 30 sec	}	38 cycles
^a 45 ⁰ C for 30 sec		
65 ⁰ C for 60 sec		

^a45⁰ C was the optimal annealing temperature for the mitochondrial primers; this temperature also worked well for the nuclear primers

2.3. Gel electrophoresis

Once the PCR products were amplified, they were then separated and visualized through gel electrophoresis. PCR products were loaded in wells in an agarose gel, after which an electric current was applied to move the DNA towards the positive electrode. Different-sized DNA fragments migrate at different rates with smaller fragments moving rapidly through the gel matrix and larger fragments moving more slowly. Different sized PCR products would therefore be found at different positions of the gel at the end of electrophoresis. The DNA molecules were visualized when the ethidium bromide in the agarose gel intercalated between the nitrogen bases of the DNA and fluoresced under UV light (Reece, 2004). Below is the general protocol for agarose gel electrophoresis.

1. For a 1.25% agarose gel, 1.25 g agarose was dissolved in 100 ml TBE buffer [54 g Tris, 27.5 g boric acid and 20 ml 0.5 EDTA dissolved in 1000 ml distilled water to make 5X TBE buffer, then diluted to 1X before use]. The agarose was melted in a microwave oven and the molten gel cast in a container. The combs were added to make the wells.
2. Once the gel had set, the combs were removed and the PCR products loaded on the wells, after which the gel was submerged in a tank containing 1X TBE.
3. A current (100-120 V) was applied through the gel using a portable power supply.
4. Once the PCR products had run their course through the gel, the gel was removed from the tank and placed on a UV transilluminator to visualize the bands.

2.4. Gel extraction

Since most of the PCR amplifications yielded more than one band, it was necessary to separate these bands by electrophoresis prior to purifying the band of the correct size from the agarose gel in preparation for sequencing. Each band in the gel represented a PCR product of a particular size, thus the nucleic acid purified from a particular band is assured of having no other PCR product or primer of a different size. For bands that were very close to each other, care was taken when cutting these bands.

The protocol follows the manufacturers' instructions for the Eppendorf Perfectprep™ Gel Extraction Kit.

1. The PCR product was excised from the agarose gel with a sterile, sharp scalpel and placed in a 1.5ml microfuge tube.

2. The gel slice was weighed. Three volumes of Binding Buffer were added to every volume of the excised gel (100 mg ~ 100 µl).
3. The gel slices were incubated at 50 °C for 10 minutes (or until the gel slice had completely dissolved). To help dissolve the gel, the tubes were inverted every 2-3 minutes during the incubation.
4. One gel volume of isopropanol was added and mixed to precipitate the PCR product.
5. A spin column with a membrane filter was placed in a 2 ml collection tube. To bind the PCR product, the sample was applied to the spin column provided and centrifuged at 13,000 rpm for 1 minute. The flow-through was discarded and the column was placed back into the same collection tube. (Maximum volume capacity of spin column was 800 µl. For sample volumes of more than 800 µl, the remaining samples were also loaded and the collection tube centrifuged again until all the samples were used up.)
6. 750 µl of the wash buffer were added to the column and centrifuged at 13,000 rpm for 1 minute. After discarding the flow-through the column was replaced in the collection tube and centrifuged at 13,000 rpm for another minute to completely remove traces of the Wash Buffer.
7. The spin column was placed into a clean 2 ml collection tube. 30 µl of the Elution Buffer (10 mM Tris-Cl, pH 8.5) was added to the center of the membrane filter and the collection tube centrifuged at 13,000 rpm for 1 minute to elute the PCR product.
8. The spin column was discarded, and the eluted product was stored at -20° C until further use.

2.5. DNA quantification

Eluted PCR products were quantified using a NanoDrop® ND-1000 spectrophotometer prior to DNA sequencing. For cycle sequencing using the ABI PRISM™ Big Dye Version 3.1 Kit (see next section), approximately 1-3 ng concentration was recommended for every 100-200 bp of the PCR product to be sequenced.

2.6. Cycle sequencing

Cycle sequencing was employed to derive the sequence of the purified PCR products. This method uses the principle of dideoxy chain termination (also known as the Sanger method) in determining DNA sequences. In this procedure, a template DNA such as the double stranded PCR product was subjected to amplification of each strand in separate tubes using one primer each. A small amount of fluorescently labeled dideoxynucleotide triphosphates (ddNTPs) were included in the sequencing mix. These ddNTPs differed from the normal dNTPs in that they had an H group rather than an OH group on the 3rd carbon of the ribose sugar. When a ddNTP was incorporated instead of a dNTP in the growing chain, no other dNTP would bind to the 3rd carbon of the ribose sugar in the absence of the OH group, thereby prematurely terminating the sequence. This resulted in a mixture of different sized fragments all terminating in a fluorescently labeled ddNTP. After the sequencing reaction, these fragments were run on a gel in an automated DNA sequencer where the fragments separated according to size. As the samples passed through a certain point along the course of the gel, an argon ion laser fluoresced the dye attached to the samples. Each ddNTP (A, C, G and T) carried a dye that fluoresced at a particular wavelength, which were picked up and registered by a computer as a unique color. A sequence complementary to the template DNA was then

generated as a series of differently colored peaks (Reece, 2004). The protocol below follows the manufacturers' instructions for the ABI PRISM™ Big Dye Version 3.1 Kit except that volumes were reduced by half for most components while less than 20% of the Big Dye was used.

2.6.1. Cycle sequencing components

Listed below are the components used for each sample sequenced. Sequencing of the forward and reverse strands was carried out in separate tubes.

1. 1.5 µl Big Dye Mix (for 500-1000 bp PCR product)
2. 1.6 µl primer (1 µM)
3. purified PCR product (approximately 1-3 ng/ 100-200 bp)
4. sterile distilled water to make a final volume of 10 µl.

2.6.2. Cycle sequencing running conditions

96 ⁰ C for 5 minutes	} 25 cycles
96 ⁰ C for 10 sec	
50 ⁰ C for 5 sec	
60 ⁰ C for 4 minutes	

2.6.3. Precipitation of cycle sequencing samples

Once the cycle sequencing was complete, the samples were precipitated prior to being run in an automated sequencer such as the Applied Biosystems 3730 DNA sequencer. The instructions below follow those recommended by the manufacturers of the ABI PRISM™ Big Dye Version 3.1 Kit but with some modifications.

1. For each 1.5 ml microfuge tube, 2.0 µl 3M NaOAc (pH 4.6) and 50 µl 95% ethanol was added. The entire cycle sequencing mix was then added into the

NaOAc-ethanol mixture. The tubes were vortexed and set aside for at least 30 minutes. This step precipitated the different-sized PCR products in the tube.

2. The tubes were centrifuged for 20 minutes at 13,000 rpm. The supernatant was then carefully removed.
3. The pellet was rinsed with 190 μ l 70% ethanol, then vortexed briefly.
4. The tubes were centrifuged for 5 minutes at 13,000 rpm, after which the supernatant was carefully removed.
5. The tubes were placed with the lid open in a heat block or thermal cycler at 90⁰ C for one minute. The samples were then sent to the Department of Biochemistry of Oxford University or the DNA Sequencing Facility of the Natural History Museum for sequencing.

2.7. Cloning

There are instances in which PCR amplifies more than one copy of a particular gene fragment. This happens if an individual is either a heterozygote for a particular gene or actually possesses several genes. Once the PCR products are sequenced, variable sites among the different copies are manifested as ambiguous sites as PCR does not discriminate between copies as long as they have the same primer-binding sites. Such is the case for actin where several genes coding for actin are sometimes found within the same individual (Hernan, 1993). Where possible, all the different genes should be amplified, cloned and then sequenced in order to identify the presence of paralogous sequences. However, cloning was not possible for the snail phylogenetic analyses in Chapters 3 and 4 due to time and cost constraints, so direct-sequencing from PCR products was employed instead, with ambiguity codes assigned to positions of ambiguity in the direct sequences (see Section 2.9.3, pp. 72-73). These ambiguous sites

were then utilized as ambiguities in the tree-building process. Sequences obtained via cloning from the different copies from a few selected taxa served as a reference in assigning codes for the ambiguous sites. One taxon that did not exhibit any ambiguous sites (i.e. *Leptinaria lamellata*, see Appendix 3.8C, p. 436) was also used for cloning in order to check if any ambiguity was missed in the direct sequence. It should be emphasized that, in the absence of sequencing all the actin clones for each taxon, using direct sequence with ambiguous sites implies using an “average” or consensus sequence for all the possible copies of the actin gene for that species. Caution must therefore be taken in interpreting trees involving taxa with potentially more than one actin gene.

Sequencing different copies of the actin gene separately was achieved by first cloning the PCR products in bacteria using a plasmid vector. A special plasmid used in cloning called the *pUC* plasmid included an ampicillin-resistant gene and a *lacZ* gene that coded for a subunit of the β -galactosidase enzyme that breaks down galactose. Foreign DNA can be inserted into this plasmid within the *lacZ* gene. Once the insert is incorporated in the plasmid, the plasmid was allowed to be taken in through chemical transformation by a *lacZ* mutant *E. coli* that could not produce a functional β -galactosidase enzyme. These bacteria were then allowed to grow in an agar plate containing ampicillin and galactose. Bacteria that took in the plasmid were able to grow on the medium because of the presence of the ampicillin-resistant gene in the plasmid. Bacteria that harbored plasmids with inserts would not exhibit β -galactosidase activity in the presence of galactose because the inserts were placed within the *lacZ* gene, thereby disrupting its transcription. Colonies of these bacteria appeared white when compared to bacteria with functional *lacZ* in the plasmid, which appeared as blue colonies. These white colonies were then picked from the agar and were subcultured in a liquid medium to allow them to grow, thereby producing a vast number of copies of

the insert. The plasmids were then isolated from the bacteria and the inserts direct-sequenced using plasmid specific primers that bound to the flanking regions of the annealing site of the insert. As only one molecule of foreign DNA could be inserted into a plasmid, the inserts derived from a single colony represented a single copy of that insert (Reece, 2004). In this way individual copies of the actin gene were obtained for sequencing.

The protocol below follows the manufacturers' instructions for the Invitrogen™ TOPO Cloning Kit except that the volume of competent cells used was reduced by half.

2.7.1. Agar plate preparation

1. Around 15-20 ml LB agar was poured into a Petri dish and set aside at room temperature to solidify the agar. The agar plate was left slightly open in a drying cabinet for 30 minutes to remove condensation from the lid.
2. 17 µl of 50 mg/ml ampicillin was spread on top of the agar plate before replacing the lid.
3. The agar plate was then warmed at 37⁰ C for 30 minutes.
4. 80 µl of 20 mg/ml X-gal was spread on the agar plate, after which it was incubated at 37⁰ C until use.

2.7.2. Setting up the TOPO™ Cloning Reaction

1. The following were mixed gently and incubated for five minutes at room temperature:

PCR product	1.0 µl	}	provided in the kit
Salt solution	0.5 µl		
water	1.0 µl		
TOPO	0.5 µl		

2. 1 µl of the TOPO cloning reaction from (1) was added to 25 µl of One-Shot™ chemically competent cells and incubated on ice for 5 minutes.
3. The cells were heat-shocked for 30 seconds at 42⁰ C without shaking. This allowed the cells to take in the plasmids. The tubes were immediately transferred in ice.
4. 125 µl of previously defrosted SOC medium from Box 2 was added, and the tube was then capped tightly and shaken horizontally (200 rpm) at 37⁰ C for one hour.
5. 50 µl from each transformation was spread on a pre-warmed (37⁰ C) selective agar plate containing X-gal and incubated overnight at 37⁰ C.
6. White colonies were picked and cultured overnight in 5-ml LB medium containing 50 ug/ml ampicillin.

2.7.3. Plasmid isolation

1. The culture was transferred into a microfuge, after which it was centrifuged for five minutes at 13,000 rpm to discard the supernatant.
2. The process was repeated until most of the culture was used up. [A small amount (~500 µl) was set aside in a separate tube where ~125 µl of 80% sterile

glycerol was added. The tubes were then stored at -80°C so that the culture could be re-grown in the near future should the need arise.]

3. 1.0 ml of cold TE buffer was added, after which the tube was vortexed. The tube was then centrifuged for 5 minutes at 13,000 rpm to discard the supernatant.
4. 100 μl of cold TE buffer was added and the tube was then vortexed. Incubation on ice for a minimum of 15 minutes followed.
5. 200 μl of 0.2 N freshly prepared NaOH / 1% SDS solution was added. This mix lysed the bacterial cells.
6. 150 μl of 3M KOAc, pH 4.8 was added followed by gentle mixing. The tube was mixed gently and left in ice for 5 minutes. This stage precipitated the proteins and neutralized the alkaline pH.
7. The tube was centrifuged for 5 minutes at 13,000 rpm. The supernatant was moved to a separate tube while the pellet was discarded.
8. 10 μl of 2 mg/ml RNase was added, and the tube was then incubated at 37°C for 30 minutes. This degraded RNA.
9. An equal volume ($\sim 450\text{ }\mu\text{l}$) of phenol-chloroform was added, after which the tube was inverted several times. It was then centrifuged for 5 minutes at 13,000 rpm. The aqueous phase was transferred to a new tube.
10. An equal volume ($\sim 400\text{ }\mu\text{l}$) of chloroform was added, after which the tube was inverted several times. It was then centrifuged for five minutes at 13,000 rpm. The aqueous phase was transferred to a new tube.
11. 2.5 volumes ($\sim 1\text{ ml}$) of ice-cold 95% ethanol were added. The tube was placed in -80°C for at least 20 minutes or in -20°C for at least 2 hours and 30 minutes.

It was then centrifuged for 10 minutes at 13,000 rpm. The aqueous phase was discarded.

12. 1.0 ml of 70% ice-cold ethanol was added, and the tube was then centrifuged for 5 minutes at 13,000 rpm before the aqueous phase was discarded.

13. The pellets were dried at 45⁰ C for 15 minutes. These were then eluted in 20 µl sterile distilled water and stored at -20⁰ C until use.

2.7.4. Sequencing of the insert

The protocols for sequencing of the insert were the same as those in Section 2.6 (see pp. 59-61) except that the M13 forward primer (5' – GTAAAACGAC GGCCAGTGAA – 3') and the M13 reverse primer (5' – CAGGAAACAG CTATGACCATGA – 3') were used. These primers bound to sites that flank the annealing site of the insert in the plasmid. This ensured that the sequence, which included some segments of the plasmid DNA, was derived from the insert.

2.8. Single strand conformational polymorphism analysis

Determining variation at the DNA level among populations need not require sequencing every sample. One method of detecting variation is single strand conformational polymorphism analysis (SSCP), which was used to evaluate the variation in global populations of *Achatina fulica* (Chapter 5). Amplified PCR products from a marker gene such as the 16S rRNA was compared for nucleotide variation by denaturing them into single strands and then allowing them to migrate slowly over a long period in a polyacrylamide gel. Single stranded DNA underwent conformational changes that depended on their nucleotide composition. Therefore, similar sized fragments that varied even in just a single nucleotide assumed different conformations

and hence exhibited different migration rates. The DNA was visualized by silver staining in which silver nitrate bound to the DNA. The pattern generated by the two single stranded DNA's on a polyacrylamide gel represented the haplotype of the individual for the amplified genetic marker (Orita *et al.*, 1989). The SSCP technique, however, only works for fragments that are 400 bp or shorter (Sunnucks *et al.*, 2000); in this study, the amplified PCR products derived from the 16S primers were 293 bp long. The subsequent protocol follows that of Orita *et al.* (1989) with some modifications.

2.8.1. Native polyacrylamide gel preparation:

1. The following were mixed in a beaker:

2X MDE™ gel solution (Cambrex Bio Science®)	7.5 ml
1X TBE buffer	18 ml
sterile distilled water	4.0 ml
TEMED	40 µl
ammonium persulfate	400 µl

Note: The ammonium persulfate was added last as this catalyzed the polymerization of the gel.

2. Once the reagents were mixed, the solution was immediately poured into two glass plates stacked on top of each other and separated by 0.4 mm spacers. The gel set-up was then tapped at the bottom to ensure that no bubbles formed. The combs were subsequently added on top of the gel set-up.
3. The gel was allowed to set for at least 40 minutes, after which the combs were gently removed and the wells washed with water.
4. The gel plates were secured to the rig, which was filled with 2.0 L of 0.6X TBE.

2.8.2. Preparation of PCR products for SSCP analysis

1. An equal volume of isopropanol was added to each PCR product. The tubes were then inverted several times, then placed in ice for at least 30 minutes to precipitate the DNA.
2. The tubes were centrifuged for 15 minutes at 13,000 rpm, after which the supernatant was discarded. The pellets were dried at 45-55⁰ C for 15 minutes.
3. The pellets were resuspended in formamide-heavy loading buffer. [Loading buffer: 0.2 g bromophenol blue, 0.2 g CFF, 20 ml 0.5 M EDTA; one part loading buffer diluted in nine parts formamide]
4. The tubes were incubated at 95⁰ C for 10 minutes to denature the DNA into single strands, then the tubes were placed in ice to prevent gradual reannealing.

2.8.3. Loading of samples in the polyacrylamide gel

1. 15 µl of formamide-heavy loading buffer was loaded in each well. The gel was allowed to run at 180 volts for at least one hour to ensure that the gel was both consistent and equilibrated.
2. Once the gel was equilibrated, the samples were then loaded directly from ice into the wells.
3. The samples were allowed to migrate along the gel for 24 hours at 180 V.

2.8.4. Silver staining of SSCP gels

1. The following solutions were prepared:

Fix-stop solution

10% acetic acid	200 ml
Distilled water	1800 ml

Silver nitrate solution

Silver nitrate	2.0 g
Distilled water	1997 ml
Formaldehyde	3.0 ml

Developer

Sodium carbonate	60 ml
Distilled water	1997 ml
Formaldehyde	3.0 ml

2. After the run was complete, the gel was transferred into a staining tray containing 200 ml of fix-stop solution. Under the fume hood, the gel was agitated for 45 minutes to ensure complete fixation of the bands, after which the fix-stop solution was removed.
3. The gel was washed with distilled water three times for 20 minutes or until it attained a smooth appearance, indicating that it was already free of fix-stop solution. The excess water was removed.
4. 200 ml of silver nitrate solution was carefully added on the side of the tray and not directly onto the gel itself to prevent it from blackening. The gel was agitated for 40 minutes to give adequate time for the silver nitrate to bind with the DNA. The silver nitrate solution was removed afterwards into a suitable waste jar containing NaOH pellets to neutralize the solution.

5. The gel was washed with distilled water to remove excess silver nitrate, then the water was discarded into the silver nitrate waste jar.
6. 200 ml of developer was added and the gel agitated until bands were clearly visible.
7. The reaction was stopped by pouring 200 ml of fix-stop solution onto the gel, which was then agitated for an additional 5 minutes to release all the CO₂.
8. The gel was repeatedly washed with 200 ml distilled water until the smell of the acetic acid was no longer detected, after which the excess water was removed.
9. The gel was transferred onto a clean filter paper and covered with cling film. The gel was allowed to dry in a gel dryer for approximately 3 hours at 70 °C.

2.9. DNA sequence analyses

DNA sequences were assembled using the STADEN package version 1.5.3 (Staden, 2000) and aligned manually within the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994). The sequences were then checked for ambiguous nucleotide sites, saturation, and phylogenetic signal before being subjected to phylogenetic estimation. To this end, the optimal model that best fits the dataset was first identified, then a range of tree construction methods were employed.

2.9.1. Aligning sequences

Sequences were brought into the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994) where they were aligned manually. Positions were identified as unambiguously aligned when there was absolutely no question over their positional homology. All other positions for which homology could not be ascertained were excluded from subsequent analyses. For the nuclear rRNA cluster and the

mitochondrial 16S rRNA, the sequences were aligned according to their secondary structure. Initial secondary structure-based alignments for the nuclear rRNA and mitochondrial 16S rRNA for the Stylommatophora were obtained by C. Hudelot with all other sequences brought and aligned into these initial alignments. The initial alignment of the SSU for the Nematoda was provided by M. Blaxter. This alignment was updated to include all nematode SSU sequences available in GenBank (GB Release Numbers 157-158) as well as the new sequences obtained in this study.

2.9.2. Checking for contamination

Every precaution was taken to avoid contamination, which could have occurred either during DNA extraction, PCR amplification of the gene fragment of interest, or through cycle sequencing. These precautions included maintaining a clean work bench, regularly cleaning pipettes with 0.1 N HCl to destroy any contaminant DNA present, and using sterile pipette tips, scalpel blades and reagents. However, sequences obtained were still checked for contamination. A cursory inspection of the alignment was conducted to see if a sequence varied greatly or was more difficult to align than the rest. A BLAST check through NCBI was also carried out to verify if indeed the sequence originated from a contaminant (i.e. the sequence had a closest identity to a non-mollusc such as a human gene or a cross-contaminant with another mollusc). Sequences obtained in this study were also crosschecked against each other to determine whether sequences from different taxa (or even segments of sequences as in the case of the rRNA cluster) exhibited identical sequences; this might indicate cross contamination (though it should be noted that among closely related taxa, relatively conserved regions might reasonably be expected to be identical). In cases of contamination, the entire process from PCR amplification to cycle sequencing was repeated. Where the same

result was obtained, the process was again repeated, this time starting from the DNA extraction.

2.9.3. Assigning ambiguous sites

Sequence positions were scored as ambiguous either because the sequence could not be read reliably (due to poor quality sequence reads), or because there were several copies of the gene (as in the case of actin for which several copies were identified in some achatinoid species; see Chapters 3 and 4). For the actin gene, the identification of ambiguous sites (positions where different copies of the actin gene differed in sequence) was facilitated by using as guides the sequences obtained through cloning from representative taxa. For perfectly clear forward and reverse sequences, an ambiguous site was assigned to a position where two overlapping peaks were seen for that position in both strands. In cases where one strand was clear and the other strand had background noise, then the clear strand was used as the basis for deciding whether the site was ambiguous or not. If both strands exhibited background noise and yet both had the same clear overlapping peaks for a site, then that site was also assigned as ambiguous. Table 2.8 lists the codes for ambiguous nucleotides.

Table 2.8: Codes for ambiguous positions in DNA sequences (Cornish-Bowden, 1985)

Base ambiguity	Code
A or C	M
A or G	R
A or T	W
C or G	S
C or T	Y
G or T	K
A or C or G (not T)	V
A or C or T (not G)	H
A or G or T (not C)	D
C or G or T (not A)	B
A or C or G or T	N

2.9.4. Estimating the average base frequencies and the number of variable and parsimony-informative sites

Sequences for DNA very rarely exhibit identical frequencies for each base due to nucleotide bias. An extreme example of this is exhibited at the 3rd codon position of insects and crustaceans where 95% of the bases are either A or T (Palumbi, 1996). For all sequences under consideration, base frequencies were estimated using the basefreq command in the PAUP* (version 4.0b10) package (Swofford, 2002). The numbers of variable and parsimony-informative sites were calculated using the randtrees command in PAUP*.

2.9.5. Identifying the optimal model for DNA sequence evolution

Multiple ‘hits’, where pre-existing mutations are masked by more recent mutations that occur at the same site, will lead to an underestimate of the actual number of changes that have taken place at a particular site, thus obscuring the phylogenetic relationship of the taxa being compared. It is therefore necessary to apply a model of sequence evolution in order to ‘correct’ for such multiple ‘hits’ (Graur & Li, 2000). Models require certain assumptions as to how variations in DNA sequences evolve. All

possible assumptions for a given situation that are taken into account form a ‘conceptual model’ in which phylogenetic estimation is made. As more assumptions or parameters are incorporated in the model, the more complex it becomes. Several models can be used to account for DNA sequence evolution; these include the JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P (Kimura, 1980), HKY85 (Hasegawa, *et al.* 1985), TN93 (Tamura & Nei, 1993) and general time-reversible (GTR) models (Rodriguez *et al.*, 1990). The JC69 is the simplest model and assumes that all types of change (all substitutions) are equally likely, base frequencies are equal, all sites are equally likely to change and change independently of each other, and base composition is at equilibrium among all the sequences under consideration (Jukes & Cantor, 1969). The K2P is an extension of the JC69 model but allows transitions and transversions to have different substitution rates (Kimura, 1980). Likewise, the F81 model is an extension of the JC69 but allows for unequal base frequencies (Felsenstein, 1981). The HKY85 model allows for different rates of substitution for transitions and transversions as well as allowing for unequal base frequencies (Hasegawa *et al.*, 1985). The TN93 model is an extension of the HKY model but distinguishes between transition rates of purines and pyrimidines (Tamura & Nei, 1993). Finally, the GTR model allows all six pairs of substitution to have different substitution rates as well as allowing for unequal base frequencies (Rodriguez *et al.*, 1990). Rate heterogeneity between sites can also be accounted for by incorporating gamma distributed rates (Γ) into the models (Yang, 1993). Gu *et al.* (1995) proposed to take into account the proportion of invariant sites (I) in the gamma distributed rates, hence the ‘ Γ +I’ model. Yang (2006) describes this model as “pathological” as gamma distribution with an α shape parameter less than 1 already accounts for the invariant sites. Depending on the model under consideration, the base frequencies, rate matrix and shape parameter (α) of the gamma distribution using 16 rate

categories were estimated using likelihood by iteration from an initial neighbor-joining (NJ) tree. The parameters derived from the initial tree were then used to build a new neighbor-joining tree and the parameters re-estimated, repeating the process until no noticeable improvement is seen in the likelihood.

Models are generally selected based on their fit to the sequence data as measured by likelihood values (Kelchner & Thomas, 2007). Normally, addition of parameters in a model increases the likelihood score; this, however, increases complexity and thus the data are spread more thinly, so if there is no significant improvement in likelihood score then there is no justification for using the more complex model. One way to identify which model to use is through a likelihood ratio test (LRT). The LRT is a statistical test that determines the goodness of fit of any two models being compared with a particular dataset. This can be applied to models that are nested since twice the difference in the likelihood scores between two nested models is approximately Chi squared distributed. The formula for this test is given as: $LR = 2 * (\ln L_1 - \ln L_2)$; where $\ln L_1 - \ln L_2$ is the difference in the log likelihood scores between any two nested models being compared. The LRT can then be used to determine if there is a significant difference between the log likelihood scores of the two models by identifying the degrees of freedom and checking for the *P* value in a Chi square table. The number of degrees of freedom is the difference between the number of parameters used by the two models being compared. For example, the GTR and the GTR+ Γ models differ by one parameter (addition of the gamma distribution in the latter); therefore, the number of degrees of freedom for comparing these two models is 1 (Huelsenbeck & Crandall, 1997). Table 2.9 summarizes the number of parameters for a given model of DNA substitution. The model with the best likelihood score was selected but only if it was significantly better than a less complex model; otherwise, the simpler model was used.

Table 2.9: Summary of the number of parameters of the different models of DNA substitution (taken from Morrison, 2006)

Model	Number of Parameters
JC69	0
JC69+ Γ	1
K2P	1
K2P+ Γ	2
F81	3
F81+ Γ	4
HKY85	4
HKY85+ Γ	5
TN93	5
TN93+ Γ	6
GTR	8
GTR+ Γ	9

Twelve different models were evaluated; these were: (1) JC69, (2) JC69+ Γ , (3) K2P, (4) K2P+ Γ , (5) F81, (6) F81+ Γ , (7) HKY85, (8) HKY85+ Γ , (9) TN93, (10) TN93+ Γ , (11) GTR, (12), and GTR+ Γ . Since the F81 and K2P models are not nested, they could not be compared with each other. F81 and K2P could, however, be compared with any other model. Although it was tempting to restrict the model search to the parameter-rich HKY85, TN93 and GTR models as they were determined to be the optimal models by the ModelTest program in 80% of 208 published datasets in 2004 alone (Kelchner & Thomas, 2007), it was more prudent to check the less parameter-rich models to confidently rule them out if the more complex models had significantly higher likelihood scores. The likelihood scores for these models were computed in PAUP*, with the command lines summarized in Appendix 2.1, pp. 360-366.

The application of the LRT described here is similar to that which is applied in the Modeltest program by Posada and Crandall (1998) except that the LRT used in this study allows for a comprehensive comparison of all models under consideration (apart

from non-nested models) whereas Modeltest ‘traverses’ a model space through a series of pairwise comparisons of the different models. For instance, if Modeltest compares the likelihood scores of JC69 and F81 and found the latter to be significantly better, then F81 is selected and compared with HKY85. If HKY85 is better than F81, then HKY85 is selected and compared with GTR. If GTR is better than HKY, then GTR is compared with GTR+ Γ . Otherwise, HKY and TN93 will instead be compared. The problem with this approach is that it does not allow for a comprehensive comparison of all the different models being considered. In the above example, the GTR and TN93 models were not compared, and there is the possibility that TN93 is not significantly better than GTR.

2.9.6. Evaluating the sequences for evidence of saturation

Sequence data could suffer from extreme substitution saturation to such a degree that an optimal model of DNA sequence evolution could no longer correct for multiple hits (Xia *et al.*, 2003). The effect of this phenomenon of substitution saturation in phylogenetic analyses is twofold. First, it underestimates the amount of evolutionary change that took place. Second, homoplasy could occur where descendant sequences appear similar even if the similarity is not brought about by descent from a common ancestor (Graur & Li, 2000). For protein-coding genes, the third codon position has been found to be the most variable, with the 1st and 2nd codon positions more conserved. This is because substitution in either the 1st or 2nd codon position is more likely to lead to amino acid replacement that could affect the function of the protein being coded by the gene; hence, substitution tends to occur at a higher frequency at the 3rd codon position where substitution is mostly synonymous (Graur & Li, 2000). Including the 3rd codon position in phylogenetic analyses is beneficial when close relationships are

examined as it is the most variable position and therefore contains much of the phylogenetic information; removing it, on the other hand, would leave only a few variable sites in the 1st and 2nd codon positions to work with. In the presence of substitution saturation, however, using the 3rd codon position could be more disadvantageous since 3rd codon positions would saturate first (Xia *et al.*, 2003).

Constructing different types of plots based on substitutions have been the standard procedure to check for evidence of saturation in sequence datasets (Morisson, 2006; Xia *et al.*, 2009), and their use abounds in the literature. Three types of plots were constructed. Plotting pairwise uncorrected distances against pairwise corrected distances based on an optimal model (Plot 1), for instance, would normally result in a curve. When distances are small there would be little difference between the uncorrected and corrected distances. As more divergent sequences are compared, however, multiple hits would lead to an underestimate of the actual amount of change in the uncorrected distances relative to the corrected distances where a correction is made to account for these unseen changes. At some point, the sequences being compared would become too divergent to allow correction for multiple hits to be made reliably leading to substitution saturation; when this happens, a plateau is observed, and the actual amount of change taking place would be underestimated (Strimmer & von Haeseler, 2003). Morisson (2006) used this type of plot on the actin and HSP70 genes of *Cryptosporidium*. If both transitions and transversions are plotted against total distance (Plot 2), on the other hand, both would be expected to increase linearly. However, as more divergent sequences are compared, transitions (which generally occur more frequently than transversions, Brown *et al.*, 1982) would be expected to saturate prior to transversions, leading to the transition line curving and eventually reaching a plateau. Transversions (which generally occur less frequently than transitions, Brown *et*

al., 1982) would continue to increase linearly for longer but eventually as sequences become even more divergent even the transversion substitutions would saturate, eventually leading to a curved transversion line and ultimately a plateau. This type of plot was used by Jorgensen *et al.* (2008) on the 16S rRNA and COI genes of apple snails. Plotting transitions against transversions (Plot 3) would result in a linear relationship when sequences are unsaturated. As divergence increases and the transitions saturate, the relationship between transitions and transversions would be non-linear, resulting in a curve in the plot. Tsigenopolous *et al.* (2002) employed this type of plot on the cytochrome b of southern African barbs. It was noted that interpretation of the plots could be somewhat subjective, particularly in determining when a plot has plateaued; it was imperative that all three types of plots were considered alongside one another in evaluating the level of saturation of each dataset.

The command lines in PAUP* for computing the uncorrected and corrected distances as well as the transition and transversion values are listed in Appendix 2.2, p. 367.

In addition to the plots mentioned above, there are other ways to detect the presence of full substitution saturation in a given dataset. One method is the randomization or permutation test in which a set of tree lengths are generated from randomized set of characters derived from the same dataset. If the dataset is saturated, then the tree lengths will not be significantly different from one another (Archie, 1989). Another method is the tree-independent relative apparent synapomorphy analysis (RASA) where synapomorphies or shared characteristics between any two given taxa are statistically evaluated (as expressed by the sum total of all shared character states between two taxa at the exclusion of third taxon) to determine if they result from shared evolutionary history or from convergence (Lyons-Weiler *et al.*, 1996). Both methods,

however, can incorrectly conclude that there is no saturation taking place if there are two very closely related taxa in the dataset even if the other taxa have reached full substitution saturation (Xia, 2009). A third method is the frequency-dependent significance test based on parsimony (see discussion on parsimony in Section 2.9.8, p. 84) in which the distribution of phylogenetically informative sites is statistically tested against the hypothesis that the sequence variation is random, as in the case of full substitution variation (Steel *et al.*, 1993, 1995). This method, however, is affected by long branch attraction, a typical problem of parsimony, where highly divergent taxa tend to group together by chance not because of shared homology but precisely because of homoplasy (Xia, 2009). A fourth method makes use of the index of saturation based on the concept of entropy in information theory where entropy is defined as the measure of uncertainty pertaining to a random variable, in this case the frequency of a nucleotide for a nucleotide site (Xia & Xie, 2001, Xia *et al.*, 2003). The index of substitution saturation (Iss) is the ratio of the observed mean entropy of all the sites in a gene of specific length over the expected entropy of the sequence at full saturation. Once the Iss has reached a critical value (or is not statistically significantly different from this critical value) in which the sequences could no longer be expected to recover the true tree, then the dataset is said to be saturated. However, applying this method becomes problematic if more than 32 taxa are used (Xia, 2009). It is also limited to determining if saturation has taken place for a perfectly symmetrical and completely asymmetrical tree but not anything in between. For all the reasons mentioned, these four methods were therefore not used for testing for evidence of saturation.

2.9.7. Testing for phylogenetic signal

If the tree lengths of all tree topologies generated through maximum parsimony (discussed in Section 2.9.8, p. 84-85) follow a normal distribution, then the differences in tree lengths are due to chance rather than homology; otherwise, the distribution is skewed enough that very few alternative solutions exist near the shortest tree, thus indicating the presence of a phylogenetic signal. A g_1 test can measure how skewed a dataset is when applied to a set of tree lengths based on parsimony. Critical values for the g_1 measure of skewness for such datasets (Table 2.10) were derived by Hillis and Huelsenbeck (1992). If g_1 test scores obtained are smaller (or more negative) than the critical values, then the tree length distributions are said to be skewed and therefore exhibit phylogenetic signal. It should be noted that the critical values change little beyond 15 taxa being investigated so the values derived for 25 taxa can be used to test datasets with more than 25 taxa (Hillis & Huelsenbeck, 1992). Where the number of parsimony-informative sites or the number of taxa being investigated is not available in the table, then the next lower number should be used as a basis for deriving the critical value. Increasing the number of parsimony-informative sites or the number of taxa will increase the critical value (make it less negative), so if a g_1 score for a particular number of parsimony-informative sites and taxa is less than the critical value for a lower number of parsimony-informative sites or lower number of taxa, then there is phylogenetic signal for the dataset.

Table 2.10: Critical values for g_1 measure of skewness at $P=0.05$.

No. of parsimony-informative sites	No. of Taxa							
	5	6	7	8	9	10	15	25
10	-0.95	-0.70	-0.59	-0.51	-0.44	-0.34	-0.23	-0.16
50	-0.78	-0.58	-0.45	-0.37	-0.25	-0.28	-0.19	-0.12
100	-0.66	-0.56	-0.40	-0.31	-0.25	-0.30	-0.20	-0.10
250	-0.81	-0.43	-0.39	-0.26	-0.22	-0.20	-0.16	-0.08

Caution was taken, however, when interpreting strong skewness as this could be misleading. The presence of duplicated taxa or even very closely related taxa (i.e. those with very similar sequences) in a group of random taxa could lead to a very strong left skew as the tree that would show monophyly of these taxa would be considerably shorter than the other trees generated (Swofford *et al.*, 1996).

The g1 test statistic was performed in PAUP* and applied to all the gene datasets using the randtrees command with 10,000 replicates in PAUP*.

2.9.8. Tree Construction

Phylogenetic trees are graphical representations of the evolutionary relationships among taxa derived from morphological or molecular datasets. For molecular datasets, tree-construction methods either convert aligned sequences into a matrix of pairwise distances among the taxa (distance methods), or they use the positions in the sequences directly (character-state methods). Tree construction methods can use an algorithm to search for the single best tree estimate (algorithmic methods) or they define a criterion by which several alternative trees are compared and scored in order to identify the best tree (optimality criterion methods) (Graur & Li, 2000).

The maximum likelihood (ML) method (Cavalli-Sforza & Edwards, 1967; Felsenstein, 1981) is a character-state method that finds the best tree with the highest probability of fit with the data (measured as the likelihood score) using a specific model of DNA sequence evolution (Graur & Li, 2000). The objective of maximum likelihood is to find the tree with the best or largest likelihood score. However, phylogenetic analyses involving many taxa would entail evaluating a large number of trees (e.g. there are more than 34 million possible rooted trees for only 10 taxa) so that conducting an exhaustive search is impractical. Instead, a heuristic search is done in which an initial

start tree (obtained by stepwise addition, with a random sequence input order and 10 replicates) is successively improved by swapping branches until the most likely tree with the best likelihood score is obtained. Unlike an exhaustive search, a heuristic search only evaluates a subset of all the possible trees and provides the best guess of the optimal tree (Hall, 2008). ML was undertaken in PHYML Version 2.4.4 (Guindon *et al.*, 2005). The command lines in PHYML are listed in Appendix 2.3.1, pp. 368-369.

Bayesian inference (BI) is similar to maximum likelihood in that it also utilises likelihood values. Unlike maximum likelihood, Bayesian inference determines the best hypothesis that maximises the posterior probability, which is proportional to the likelihood value multiplied by the prior probability of the hypothesis. A prior probability is based on what is initially known about the dataset (Holder & Lewis, 2004). Computer programs that conduct Bayesian inference such as MrBayes make use of the Markov Chain Monte Carlo algorithm to approximate posterior probability values of randomly generated trees through a number of generations until the chain attains a stable likelihood value (Ronquist & Huelsenbeck, 2003; Hall, 2008). Four chains of a Markov Chain Monte Carlo algorithm in BI were used to compute the posterior probability values with an optimized number of generations and heating temperature (Temp). The number of generations was optimized when the values of the posterior probabilities across the four chains converged or did not vary significantly from each other. The heating temperature was optimized when the acceptance rates of the Metropolis-Hastings mcmc sampler ranged between 0.1 and 0.7. (See also Appendix 2.3.2, pp. 369-370.) Bayesian inference was conducted in MrBayes Version 3.1.2 package (Ronquist & Huelsenbeck, 2003), and the command lines for this package are listed in Appendix 2.3.2., pp. 369-370.

The neighbor-joining (NJ) method (Saitou & Nei, 1987) is an algorithmic distance method that uses a modified distance matrix that adjusts the separation of each pair of sequences based on the pair's average divergence from all the other sequences. This method identifies the shortest tree by sequentially finding neighbors that minimize the total length of the tree (Graur & Li, 2000). NJ trees were built in PAUP*, with the command lines summarized in Appendix 2.3.3, p. 371.

The maximum parsimony (MP) method is a non-model based method that identifies a topology that requires the least number of possible changes (Eck & Dayhoff, 1966; Fitch, 1977). As with ML, a heuristic search for the most parsimonious tree or a set of equally most parsimonious trees is conducted by starting with an initial tree (obtained by stepwise addition, with a random sequence input order and 10 replicates) and swapping branches until the optimal tree is obtained. MP analysis was undertaken in PAUP* with a heuristic search using the tree bisection and reconnection (TBR) swapping routine and 10 random input orders. The command lines are listed in Appendix 2.3.4, pp. 371-372.

Advantages and disadvantages of the four tree methods are summarized in Table 2.11.

Table 2.11: Comparison of methods (table taken from Holder & Lewis, 2004)

Tree method	Advantages	Disadvantages
Neighbor-joining	Fast	Information is lost in compressing sequences into distances; reliable estimates of pairwise distances can be hard to obtain for divergent sequences
Maximum likelihood	The likelihood fully captures what the data tell us about the phylogeny under a given model	Can be prohibitively slow
Bayesian inference	Has a strong connection to the maximum likelihood method; might be a faster way to assess support for trees rather than the	The prior distributions must be specified; it can be difficult to determine whether the Markov chain Monte Carlo (MCMC)

	maximum likelihood bootstrapping	approximation has run for long enough
Maximum parsimony	Fast enough for the analysis of hundreds of sequences; robust if branches are short (closely related sequences or dense sampling)	Can perform poorly if there is substantial variation in branch length

2.9.9. Estimating reliabilities of clades in phylogenetic trees

A phylogenetic tree represents the best estimate of the true tree based on the sequences available and the method of tree construction. Reliability of the tree estimates, which can be measured by how many times the members of a particular clade are recovered as members of that clade, can be done through the bootstrap method in the maximum likelihood, neighbor-joining and maximum parsimony methods and through measuring posterior probabilities in the Bayesian inference method (Hall, 2008). The bootstrap method is a resampling technique used to estimate the confidence level of hypotheses in phylogenetic estimation. A measure of support for the branches in the tree is provided by bootstrap values, which are based on the percentage of the number of times a particular branch is generated in the bootstrap datasets (generated by random sampling from the dataset with replacement) (Graur & Li, 2000). For instance, a particular dataset is resampled 1000 times with replacement to produce 1000 resampled datasets, with each dataset used to generate a tree. Out of the 1000 trees, the percentage of the trees in which a particular branch appears becomes its measure of support. Bootstrap resampling with 1000 replicates was undertaken in PHYML for ML (see Appendix 2.3.1, pp. 368-369) and in PAUP* for NJ and MP (see Appendix 2.3.3, p. 371 for NJ and 2.3.4, pp. 371-372 for MP). In Bayesian inference analysis, the fraction of times a clade occurred among the last 1000 trees surveyed as part of the MCMC process was directly computed as posterior probabilities. Unlike the bootstrap method, the

underlying data are not resampled (Hall, 2008). The posterior probabilities were computed in MrBayes (see Appendix 2.3.2, pp. 369-370).

2.9.10. Combining datasets

As well as undertaking phylogenetic analyses for each gene independently, where possible the individual gene datasets were combined in order to get a better estimate of the phylogeny. It has been recognized that a phylogeny based only on a single gene, known as a gene tree, cannot truly represent a species tree (Cummings *et al.*, 1995; Graur & Li, 2000) as organisms that participate in the evolutionary process are a collection of genes, each with its own evolutionary history, that interact with each other and with the environment (Morrison, 2006). Combining data from different genes tends to increase phylogenetic signal and disperse noise while at the same time uncovering the underlying signal found in the different data partitions (Baldauf *et al.*, 2000). Using genes in concert that have different evolutionary rates and modes of inheritance (i.e. the relatively slower evolving nuclear genes and the faster evolving, maternally inherited mitochondrial genes) can also provide information at different levels of phylogeny (Graybeal, 1994). Combining datasets are also known to reduce the phenomenon of long-branch attraction in parsimony analysis (Gontcharov *et al.*, 2004) in which long branches in a tree tend to cluster together (Graur & Li, 2000).

Concatenation is the combination of all datasets into a single data matrix for the purpose of phylogenetic analysis (Morrison, 2006). However, simply concatenating the datasets under consideration and analyzing them using only one evolutionary model could produce a phylogeny with the wrong groupings if the different datasets represent very divergent evolutionary histories (Cunningham, 1997). The partition homogeneity test (also known as the incongruence length test) based on maximum parsimony in

PAUP* was used in this study to distinguish between separate datasets that could be combined to give an improved phylogenetic accuracy (with P normally greater than 0.01) and individual datasets that give poorer accuracy when combined (with $P < 0.001$) due to their very divergent evolutionary histories (Cunningham, 1997). The command lines for the partition homogeneity test in PAUP* are shown in Appendix 2.4, p. 373. As the test makes use of maximum parsimony to determine if datasets can be combined, its utility is hampered if at least one of the genes being considered exhibit polytomies as this considerably slows down PAUP*. For such a scenario, the decision to combine would rely on whether or not the single gene phylogenies exhibited concordance with each other.

Two sets of concatenated sequence alignments were prepared for the combined gene analyses. The first included only taxa with complete sequences across all genes evaluated. In order not to lose any taxon, a second set of concatenated sequences was also prepared in which taxa with missing datasets were also included. Though including taxa with incomplete datasets could be potentially problematic in phylogenetic analyses, Wiens (2006) found that these taxa could still be included and their phylogenetic position accurately determined so long as enough characters have been surveyed; their inclusion could also remove misleading long branches and potentially increase the accuracy of the combined phylogeny. The trees generated from the second set of concatenated sequences were compared for any similarity with the individual gene phylogenies as well as those derived from the first set of concatenated sequences.

An alternative to combining datasets by concatenating sequences is to construct from multiple trees a consensus tree derived from several genes with the same set of taxa or a supertree if the taxa sampled are not identical but there are taxa overlapping

between trees (Morrison, 2006). Two strategies employed to construct consensus trees and supertrees are strict consensus (tree generated contains all the groups that occur on all trees considered) and majority consensus (tree generated includes groups that occur in at least 50% of all the trees considered) (Felsenstein, 2004). However, the effectiveness of consensus methods is limited by the following: 1) their tendency to produce spurious groups that do not appear in any of the trees; 2) the non-avoidance of pseudoreplication as a result of using overlapping data in some of the trees that increases the weight of these data; 3) the inappropriateness of using branch support methods such as bootstrapping (Morrison, 2006); and 4) their treatment of each tree, no matter how bad, as equally good (Wade, pers. comm.). The consensus methods were therefore not used.

2.9.11. Hypothesis testing

An optimal tree may not necessarily produce the expected topologies, as in the case where a supposedly monophyletic group based on taxonomy turned out not to be monophyletic in the optimal tree. A tree can be constrained to reflect the expected topology, and this tree can be compared for significant difference with the optimal tree. If the optimal tree turns out to be not significantly better than the constrained tree, then the hypothesis being put forward by the optimal tree is not a strong hypothesis. The Shimodaira-Hasegawa test (Shimodaira & Hasegawa, 1999; Goldman *et al.*, 2000) was applied on the datasets found in Chapters 3 and 4 to compare for significant difference between constrained and optimal NJ trees using likelihood. The command lines for the Shimodaira-Hasegawa test, which was carried out in PAUP*, are listed in Appendix 2.5, p. 374.

2.9.12. Inferring the presence of paralogous sequences in the dataset

The presence of paralogous sequences due to gene duplication could yield misleading trees. All genes were therefore evaluated for the presence of paralogs. Of the genes used in this study, actin is most susceptible to the problem of paralogy since several copies of the actin gene, which arose from gene duplication events, abound. Although the primers used to amplify the actin gene were known to be specific for the cytoplasmic actin (see Section 2.2.1.1.1.2, pp. 48 – 50), this was no guarantee that only orthologous sequences were amplified as duplication events could also have taken place within the cytoplasmic actin. To infer the presence of paralogous sequences, the PCR-direct sequences were checked for the presence of ambiguous sites; the presence of ambiguous sites (where more than one peak is obtained at the same position in a DNA sequence) implies that more than one copy of the gene is present in that individual. All

sequences obtained in this study were evaluated for the presence of ambiguities. Where evidence of multiple copies is found, cloning the PCR products prior to sequencing enables each copy of the gene to be examined individually. Cloning of the actin gene (Chapters 3 and 4) was undertaken for a limited number of clones from three taxa (time and cost constraints precluded a more detailed analysis).

2.9.13. Assessing the utility of the actin gene for phylogenetic analyses

The inherent problem of actin in relation to the presence of paralogous sequences could limit its utility in phylogenetic analyses. However, the potential of actin to provide some phylogenetic signal should not be underestimated. Several steps were undertaken to evaluate the utility of the actin dataset. First, the clones derived from representative taxa were checked for monophyly; if all clones derived from a taxon fall as a monophyletic group within the actin tree, then they will not mislead organismal phylogeny. Second, the number of ambiguous sites within a taxon was compared with the total number of nucleotide differences between the taxon and its closest relative based on the closest sequence identity of the actin sequence (i.e. the least number of nucleotide differences between any two taxa). In principle, the time of divergence between two taxa should be earlier than the evolution of the different actin genes within these taxa, such that the number of ambiguous sites should not exceed the number of differences between the two taxa being compared. However, it is possible for two taxa to have a higher number of ambiguous sites than nucleotide differences if these ambiguous sites are due to a large number of multiple copies, in which each contains only a small number of variable sites (when this variation is added together it may exceed the number of nucleotide differences). These sequences will still cluster together in a tree and will not mislead phylogeny. Third, a partition homogeneity test (see

Section 2.9.10, pp. 86-88) was undertaken to determine whether or not the sequences from the actin gene exhibited a significant difference in terms of phylogenetic signal with the other genes being evaluated. If no significant difference was observed, this would suggest that the actin gene could be utilized alongside the other genes in phylogenetic analyses and that the presence of multiple copies of the actin gene was not having a significant effect on phylogenetic estimation. Lastly, the phylogenetic tree obtained from the actin dataset was checked for concordance with the other gene phylogenies. Concordance would suggest that the presence of multiple copies of the actin gene was not misleading phylogeny. Phylogenetic analyses involving the actin gene were therefore approached with caution, with reference to the limitations of using this gene.

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CHAPTER 3 – Phylogeny of the Achatinoidea

3.1. Introduction

The Giant African Land Snail, *Achatina fulica*, and other members of the superfamily Achatinoidea belong to the suborder Stylommatophora, whose members possess a vascularised mantle cavity that serves as a lung and two pairs of caudal tentacles, the upper pair of which bears the eyes at their tips (Ruppert *et al.*, 2004; Barker, 2001). Using the Pilsbry-Baker system of classification, the Stylommatophora can be divided into four infraorders based on the structure of the excretory system (Pilsbry, 1900; Baker, 1955). *Achatina fulica* and other members of the Achatinoidea belong to the infraorder Sigmurethra, in which the excretory system does not have a distal ureteric pouch or orthureter but possesses a ureter that runs along the anterior portion of the nephridium and connects to the pallial cavity before terminating at the pneumostome. Recent molecular phylogenetic studies (Wade *et al.*, 2001, 2006) based on the nuclear ribosomal (r) RNA gene cluster (including part of the 5.8S and LSU genes) revealed a principal division of the Stylommatophora into two major clades. The ‘achatinoid’ clade comprises the superfamilies Streptaxoidea, represented by the single family Streptaxidae, and the Achatinoidea, represented by the families Achatinidae, Coeliacidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophorellidae. All other stylommatophoran families fall within the ‘non-achatinoid’ clade.

3.1.1. The Achatinoidea

The composition of the Achatinoidea varies according to author (Solem, 1978; Abbott, 1989; Tillier, 1989; Vaught, 1989; Schileyko, 1999; Bouchet & Rocroi, 2005; Table 1.1). Solem (1978) included the Achatinidae, Ferussaciidae, Megaspiridae,

Spiraxidae and Subulinidae (including the coeliacids and glessulids as the subfamilies Coeliacinae and Glessulinae, respectively), while Abbott (1989) regarded the Coeliacidae as a family in its own right and additionally included the monotypic Thyrophorellidae. Tillier (1989) classified land snails under the superfamily Achatinoidea based on certain morphological features such as a closed ureter in the excretory system and possession of symmetrical cerebro-pedal connectives, short commissure and contiguous left parietal and visceral ganglia in the nervous system. His Achatinoidea comprised the Achatinidae, Ferussaciidae, Subulinidae (including the subfamilies Coeliacinae and Glessulinae), Thyrophorellidae as well as the Streptaxidae, Succineidae and the Oleacinidae (Spiraxidae and Testacellidae). Vaught (1989) did not include the Streptaxidae, Succineidae, Spiraxidae or Testacellidae in her Achatinoidea but instead included the Thyrophorellidae and the Coeliacidae (elevated from a subfamily of subulinids) in addition to the Achatinidae, Ferussaciidae and Subulinidae. Schileyko (1999) placed only the Achatinidae in the Achatinoidea, whereas Bouchet & Rocroi (2005) included the Achatinidae, Ferussaciidae, Subulinidae (which also incorporated the coeliacids and glessulids as the subfamilies Coeliacinae and Glessulinae) and Micractaeonidae. Wade *et al.*'s (2001, 2006) molecular phylogenies based on a part of the rRNA gene cluster lend support to Vaught's Achatinoidea. In this study, I utilise an Achatinoidea that largely resembles Vaught's and comprising the Achatinidae, Coeliacidae, Ferussaciidae, Glessulidae (a subfamily of the Subulinidae in Vaught's classification), Subulinidae and Thyrophorellidae.

The family Achatinidae, which includes the Giant African Land Snail, *Achatina fulica*, has shells that are mostly dextral, are generally higher than wide and range in size from medium to very large (Schileyko, 1999). The reproductive structures of the achatinids are highly variable (Mead, 1991), but all members possess a penis sheath

(Schileyko, 1999). With the exception of *Achatina fulica*, which has been introduced by Man into new areas (Mead, 1979; Raut & Barker, 2002), all other achatinids are restricted to Africa and nearby islands (Schileyko, 1999).

The monotypic family Thyrophorellidae is represented by *Thyrophorella thomensis* from São Thomé. The shell is sinistral and is made up of three nearly flat whorls so that it appears low conic on top and bulging at the bottom (Schileyko, 2001). *Thyrophorella thomensis* is one of the few stylommatophorans that have secondarily acquired a structure similar in function to the prosobranch gastropod operculum; in the case of *T. thomensis*, a lobe-shaped outgrowth of the parietal wall of the shell's aperture is hinged on a flexible periostracal layer to serve as a covering of the aperture (Barker, 2001). Bouchet and Rocroi (2005) included the Thyrophorellidae in the superfamily Punctoidea, although recent molecular data from the LSU rRNA gene (Wade *et al.*, 2006) clearly places this family within the Achatinoidea.

Members of the Ferussaciidae have very small dextral shells that are elongate-ovate to nearly cylindrical and appear translucent or transparent (Schileyko, 1999). Ferussaciids are unique among the 'achatinoids' because they superficially resemble the 'non-achatinoid' Succineidae in possessing a short but transversely elongate kidney (Tillier, 1989) with a complete ureter (Schileyko, 1999). The ferussaciids are largely found in the Mediterranean, with several in tropical Africa, the Indian subcontinent and Indian Ocean islands, the Philippines, Hawaii, Mexico, the Caribbean, Brazil (Schileyko, 1999), Hawaii (introduced, Cowie, pers. comm.), New Caledonia (introduced, Schileyko, 1999) and Northwest Europe (Kerney & Cameron, 1979).

The family Subulinidae is a large group with a pantropical distribution (Tillier, 1989). With very few exceptions, its members have characteristic slender and turreted shells and are generally dextral. The internal anatomy of members of the group is not

well studied, and most taxa within the family are grouped together based largely on the similarity of their shells and their geographic distribution. Such an approach is exemplified by Zilch (1959). In revising the Subulinidae, Schileyko (1999) attempted to restructure their classification by including characters of the reproductive tract, primarily those proximal to the genital orifice. As the only comprehensive review of the group since Pilsbry (1906-1907; 1908-1910), it seems logical to use Shileyko (1999) as the primary source when reviewing the group as was done by Bouchet & Rocroi (2005). According to Schileyko (1999), eight subfamily groups can be recognized. These are the Subulininae (dextral shell with the last whorl lacking internal sculpture and with a reproductive tract that has a reduced or absent epiphallus, a muscular organ responsible for the formation of the spermatophores, and a short or absent flagellum, an extension of the epiphallus; distributed in tropical and subtropical regions of the world), Petriolinae (dextral or sinistral shell similar to Subulininae but with a prominent epiphallus; found in Africa and St. Helena), Rishetiinae (dextral shell with a simple columella and a reproductive organ with a long flagellum; found in the Hindustan Peninsula, Ceylon, and Southeast Asia), Rumininae (dextral shell decollated or entire and with a reproductive organ lacking an epiphallus and a flagellum; found in South Africa and the Mediterranean), Opeatinae (with dextral shell and an enlarged penis; found in subtropical regions of the Old and New World), Obeliscinae (dextral shell slender turreted to subfusiform and with a reproductive organ lacking an epiphallus and a flagellum; found in South America, the Caribbean and Southeast Asia), Tristaniinae (sinistral shell; found in Tristan da Cunha Islands), and Perrieriinae (shell sinistral and subcylindrical or pillar-shaped; found in New Guinea) (Schileyko, 1999). Other subulinid groups have been accorded by some authors with full taxonomic family status such as the Coeliacidae (Abbott, 1989; Vaught, 1989) and the Glessulidae (Schileyko,

1999). The coeliacids have slender dextral or sinistral shells and are distributed in South Africa (only one species), the Gulf Island of São Thomé in West Africa, the Caribbean, Colombia and Peru (Shileyko, 1999). The glessulids generally have glossy, oblong-conic dextral shells and are restricted to the Indian subcontinent and Sri Lanka (Shileyko, 1999).

However, closer examination of Shileyko's results raises doubts regarding his conclusions, and the taxonomy of the Subulinidae therefore warrants further scrutiny. For instance, rather than representing a subulinid subfamily, *Tristania* is a junior synonym of *Balea*, which is a member of the Clausiliidae (Preece & Gittenberger, 2003). *Rishetia* is anatomically close to *Glessula* (both have a shell with truncated columella and penis with epiphallus), and both are restricted to South Asia; therefore, there seems to be no basis for Shileyko's erection of the subfamily Rishetiinae. Shileyko's acceptance of Thiele's (1933) genus *Striosubulina* is supposedly justified by the reproductive anatomy described and figured by Shileyko. However, in describing the penis, Shileyko seems to have mistaken that part of the penis surrounded by the penial sheath for the complete penis, and most of what he describes as the penial retractor muscle is in fact the long thin penis almost identical to that of *Subulina octona*, the type species of *Subulina* (Naggs, pers. comm.). Thus, the arrangement of the Subulinidae into eight subfamilies by Shileyko (1999), subsequently accepted by Bouchet & Rocroi (2005), is unreliable and will need to be reassessed. Furthermore, some pieces of information on distributions of the subulinids based on published records were not included by Shileyko (1999). For example, in giving the range of *Glessula* as 'Hindustan peninsula and Ceylon (1999, page 541), Shileyko fails to mention that several species of *Glessula* occur in Myanmar, Yunnan (China), Thailand and east as far as Vietnam and southeast into the Indonesian peninsula (Pilsbry, 1908-

1910; Van Benthem Jutting, 1952, 1959; Hemmen & Hemmen, 2001; Vermeulen & Maassen, 2003). With the Rumininae, the geographical range for the group alters drastically if *Zootecus* is excluded (see Table 3.1) or included, when the range extends from circum-Mediterranean, Arabian and West African to encompass a large longitudinal range from the Cape Verde islands through Central and Northern India to Myanmar (Gude, 1914). It should be noted that Schileyko (1999) placed *Zootecus* under the Subulininae.

Table 3.1: Comparison of the composition and distribution of the Rumininae according to Zilch (1959) and Schileyko (1999).

	Rumininae sensu Zilch (1959)	Rumininae sensu Shileyko (1999)	Distribution according to Shileyko	Distribution according to Zilch
<i>Namibiella</i>	x	X	SW Africa	SW Africa
<i>Xerocerastus</i>	x	X	SW Africa	SW Africa
<i>Lubricetta</i>	x	X	SW Africa	SW Africa
<i>Krapfiella</i>		X	E. Africa	
<i>Riebeckia</i>		X	Sokotra	
<i>Balfouria</i>		X	Sokotra	
<i>Rumina</i>	x	X	Mediterranean	Europe and N. Africa
<i>Obeliscella</i>	x			Arabia
<i>Zootecus</i>	x			Cape Verde Islands, N Africa, N & C. India

3.1.2. Some questions about the Achatinoidea

Although the Achatinoidea is strongly supported in Wade *et al's* (2006) rRNA phylogeny with 93% NJ bootstraps (see Figure 3.1), their phylogeny was based on only 823 nucleotide sites from the rRNA gene cluster and incorporated only 15 achatinoid taxa. Clearly, the clade could be further refined by obtaining new sequence data (both

longer sequences of the rRNA as well as sequences from other new genes) as well as the inclusion of additional taxa.

For instance, the achatinids are a compact group based on the presence of a penis sheath (Schileyko, 1999) and their restricted geographic distribution to sub-Saharan Africa (Mead, 1979; Raut & Barker, 2002). Wade *et al.* (2006) used four taxa to represent the Achatinidae, but support for the monophyly of the group was equivocal (38% NJ bootstraps and $P=0.94$ BI) based on the small segment of the rRNA gene cluster used in their analyses.

The ferussaciid *Ferussacia folliculus* forms a distinct basal lineage in the Achatinoidea (Wade *et al.*, 2006). Whether this topology persists if other ferussaciids are surveyed remains to be seen.

The monophyly of the Subulinidae and its subfamilies also needs to be verified as, anatomically, this group is poorly studied (Schileyko, 1999), and both the coelioxids (Abbott, 1989; Vaught, 1989) and the glessulids (Abbott, 1989; Schileyko, 1999) have been regarded as separate families. In particular, the placement of *Zootecus* in the Subulininae according to Schileyko (1999) must be validated. The subulinids, coelioxids and glessulids were represented by nine taxa in the Wade *et al.* (2006) study, which formed several groups together with the thyrophorellid *Thyrophorella thomensis* but with no support apart from the *Riebeckia-Coeliaxis* group (92% NJ bootstraps and $P=1.0$ BI) and the *Pyrgina-Thyrophorella* group (100% NJ bootstraps and $P=1.0$ BI).

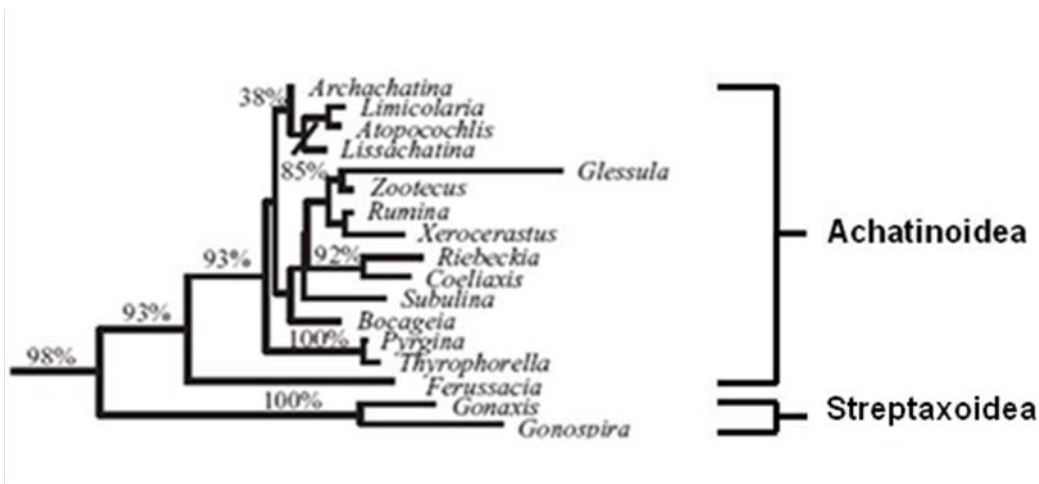


Figure 3.1: Phylogeny of the Achatinoidea based on 823 unambiguously aligned nucleotide sites of the rRNA gene cluster. (Taken from the stylommatophoran phylogeny of Wade *et al.*, 2006.)

3.1.3. Use of molecular markers to determine phylogeny

Various molecular markers are available for determining evolutionary relationships. These include the nuclear ribosomal RNA gene family (see Section 2.2.1.1.1 of Chapter 2, pp. 45-48). Already described is Wade *et al.*'s (2001, 2006) use of the nuclear ribosomal RNA gene cluster to infer phylogenetic relationships within the Stylommatophora. However, their results were derived from only a partial segment of the rRNA cluster. Wade *et al.* (2001, 2006) amplified an approximately 1460 nucleotide region of the rRNA (comprising approximately 80 nucleotides of the 5.8S gene, the complete internal transcribed spacer (ITS) 2 region and approximately 840 nucleotides of the LSU gene) of which 843 (2001 study) and 823 (2006 study) nucleotide sites could be aligned across all taxa and were therefore used in phylogeny reconstruction. Expanding the Wade *et al.* fragment to cover the entire LSU gene (approximately 4000 nucleotides) could provide better resolution of the Achatinoidea. The small subunit (SSU) rRNA gene has also been employed to elucidate evolutionary relationships among different gastropod groups. For example, Winnepenninckx *et al.*

(1998) applied the full-length SSU rRNA gene (approximately 1800 nucleotides), to reassess the groupings within the Gastropoda, demonstrating the basal position of the archaeogastropods. However, their results could not resolve whether the pulmonates or even the stylommatophorans were monophyletic.

In addition to the ribosomal genes, the nuclear actin and histone 3 genes have also been used to estimate phylogeny within the Mollusca. The actin gene encodes a protein that is involved in various functions such as muscle contraction, cell division and differentiation (Hernan, 1993, see also Section 2.2.1.1.2 of Chapter 2, pp. 48-50). However, several actin genes may exist in animals as a result of gene duplication and divergence (Hightower & Meagher, 1986); these genes code for protein isoforms that are broadly classified as either the muscle (α) type or the cytoplasmic (β) type (Vandekerckhove & Weber, 1984; Adema, 2002). Great care must therefore be taken to ensure that paralogous copies of the gene do not mislead the phylogenetic analysis. A 784 bp fragment of the actin gene has been successfully employed to show the monophyly of several groups within the coleoid cephalopods such as the Octopodiformes, the Decapodiformes, the Octopoda and the Incirrata (Carlini *et al*, 2000). The actin gene was also used together with the mitochondrial 16S and cytochrome c oxidase subunit I (COI) genes to resolve the phylogeny within the ancestral archaeogastropod monodontine topshells (family Trochidae) from the southern hemisphere, with three species of *Austrocochlea* being transferred to the genus *Chlorodiloma* (Donald *et al.*, 2005). The actin fragment (764 bp) used in this study was amplified using primers designed for the cytoplasmic actin fragment of the planorbid snail *Biomphalaria glabrata* previously sequenced by Adema (2002). To date, the cytoplasmic type actin gene has not been used in the Achatinoidea, and the utility of this

gene for phylogenetic analysis of the group is worth investigating, though the potential presence of paralogous sequences could limit its utility.

The histone 3 (H3) gene codes for two subunits of the histone core protein octamer that packs the eukaryotic chromatin into bead-like structures (Lewin, 2008) (see Section 2.2.1.1.1.3 of Chapter 2, pp. 50-51). Histone 3 was used, in conjunction with the nuclear SSU and LSU rRNA genes and the mitochondrial COI gene, to resolve the incongruence between molecular and morphological data for the gastropod phylogeny (Colgan *et al.*, 2003). The results demonstrated the monophyly of the Euthyneura, which include the stylommatophoran pulmonates. The H3 fragment (331 bp) used in this study corresponds to the 3' end of the gene in the bivalve *Spisula solidissima*. Like the actin gene, the histone 3 gene has not been used in the Achatinoidea; thus, its usefulness is worth investigating.

Using mitochondrial genes in concert with nuclear genes is desirable for constructing phylogenetic trees as they tend to improve phylogenetic accuracy (Lake & Moore, 1998; Steinke *et al.*, 2004). Nuclear and mitochondrial genes evolve at different rates and are not inherited in the same way; as such, they provide information at different levels of phylogeny (Graybeal, 1994). Mitochondrial genes generally evolve faster than nuclear genes; they are also maternally inherited and are therefore not subject to recombination (Brown, 1985; Avise, 1994). Two mitochondrial genes commonly used for phylogenetic analyses are the cytochrome c oxidase subunit I (COI) gene, which codes for an enzyme that accepts electrons from cytochrome c during the electron transport chain in the mitochondrion (Zubay *et al.*, 1995; see also Section 2.2.1.1.2.1 of Chapter 2, pp. 51-52), and the 16S rRNA gene, which transcribes a ribosomal rRNA that is incorporated in the mitochondrial ribosome (Lewin, 2008; see also Section 2.2.1.1.2.2 of Chapter 2, pp. 52-53). Steinke *et al.* (2004) used a combined

data set of nuclear rRNA genes (18S and ITS-1) and mitochondrial genes (16S rRNA and COI) to separate the Western Palaearctic helicoid stylommatophorans into two families: Helicidae and Hygromiidae. They reasoned that the faster evolving mitochondrial genes were able to resolve the terminal taxa while the more conserved nuclear rRNA genes separated the basal groups. The 16S rRNA fragment (approximately 400 bp) was amplified using Palumbi *et al.*'s (1991) universal primers while the COI fragment (approximately 500 bp) was amplified using Folmer *et al.*'s (1994) universal primers. As mentioned previously, these same fragments were also used by Carlini *et al.* (2000) alongside the actin gene for the coleoid cephalopod phylogeny. Furthermore, both 16S (Thollesson, 1999) and COI (Remigio & Hebert, 2003) were employed with some degree of success to evaluate higher level phylogenies within the Gastropoda even though both genes are susceptible to significant levels of saturation. Neither Palumbi *et al.*'s (1991) universal 16S primers nor Folmer *et al.*'s (1994) universal COI primers have been applied previously to phylogenetic analyses of the Achatinoidea.

Aside from the COI and the 16S rRNA genes, several other mitochondrial genes have also been used for inferring deep level phylogenies within the gastropods. Grande *et al.* (2004) employed several mitochondrial genes, in addition to the 16S and COI, such as the tRNA-valine, tRNA-arginine, tRNA-proline and the NADH dehydrogenase subunits 5 and 6 genes in the study of the Euthyneura (opisthobranchs and pulmonates) in which their molecular data rejected the monophyly of the pulmonates.

At present, no study has attempted to use nuclear and mitochondrial genes in concert to address the issues regarding the relationships within the Achatinoidea. Using several genes together approximates the species tree that traces the evolutionary relationships of the species under consideration, which is more desirable than

representing their relationships based on the history of individual genes, as shown by a gene tree (Grauer & Li, 2000). Combining these genes could also resolve the phylogeny of the Achatinoidea at all levels, with the faster evolving mitochondrial genes (COI and 16S) resolving the shallower nodes and the slower evolving nuclear genes (rRNA, actin and possibly histone 3) resolving the deeper nodes (Graybeal, 1994; Johnson & Clayton, 2000). Combining genes could also increase phylogenetic signal and disperse noise (Baldauf *et al.* 2000; Gontcharov *et al.*, 2004).

3.1.4. Objectives of this study

This study had the following objectives: (1) to identify the primary divisions within the superfamily Achatinoidea and to elucidate the relationship of the different families; (2) to establish if the Achatinidae are monophyletic and to identify their position relative to the other families within the Achatinoidea; (3) to determine whether the Ferussaciidae is monophyletic and to establish its phylogenetic position; (4) to settle the taxonomic designation of the Subulinidae, Coelioxidae and Glessulidae; and (5) to validate the different subfamilies of the Subulinidae. To address these objectives, the phylogeny of the Achatinoidea was inferred by examining representative taxa from all major groupings across the superfamily using molecular data from three nuclear genes (rRNA cluster, actin, and histone 3) and two mitochondrial genes (16S rRNA and COI). The genes were analyzed separately and in combination.

3.2. Materials and methods

3.2.1. Taxa used

Twenty-four taxa from six achatinoid families (Achatinidae, Coelioxidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophoprellidae) and three streptaxid outgroup taxa, two of which were used by Wade *et al.* (2006) and were found to fall at the base of the ‘achatinoid clade,’ were used to evaluate the phylogeny of the Achatinoidea (see Table 3.2). Four subulinid subfamilies following Schileyko’s (1999) classification (Petricolinae, Rishetiinae, Rumininae and Subulininae) were also represented in this study. Thirteen taxa were entirely new to this study while the remaining 14 taxa were used by Wade *et al.* (2006) in their phylogeny of the land snails based on a partial fragment of the rRNA cluster.

Table 3.2: Taxa used for the phylogenetic study of the Achatinoidea. Taxa marked with (*) are new to this study and were solicited through email by myself, C. Hudelot, F. Naggs and C. Wade from the collectors shown in the table. The DNA for the remaining taxa was provided by C. Wade from the collections obtained for Wade *et al.* (2001, 2006).

Family	Species	Collection/Location	Collector/Provider
Achatinidae	* <i>Achatina achatina</i> (Linnaeus, 1758)	Unknown (Zool. Soc. Lond. Colln.)	NHM**
	<i>Achatina fulica</i> Bowdich, 1822	Captive bred, unknown origin	NHM**
	* <i>Achatina stuhlmanni</i> von Martens, 1892	Semuliki National Park, Uganda	B. Rowson
	* <i>Cochlitoma ustulata</i> (Lamarck, 1822)	Western Cape Prov., South Africa	A. Moussalli & D. Stuart-Fox
Coeliacidae	<i>Coeliaxis blandii</i> (Pfeiffer, 1852)	New Bradford, South Africa	N. Smith
	<i>Pyrgina umbilicata</i> Greeff, 1882	São Thomé	A. Gascoigne
Ferussaciidae	* <i>Cecilioides gokweanus</i> (Boettger, 1870)	Cape Vida, Bhangazi Hill, Zululand, South Africa	D. Herbert
	<i>Ferussacia folliculus</i> (Gmelin, 1791)	Los Alcornales, Prov Cadiz, Spain	M. Seddon
Glessulidae	<i>Glessula ceylanica</i> (Pfeiffer, 1845)	Colombo, Sri Lanka	P. Karunaratne
Subulinidae			
S.f. Petriolinae	<i>Bocageia</i> sp.	São Thomé	A. Gascoigne
	* <i>Subulona</i> sp.	Ossen Forest, Tupen Hills, Kenya	M. Pickford
S.f. Rishetiinae	* <i>Eutomopeas layardi</i> (Benson, 1863)	Koralegama, Sri Lanka	?
	* <i>Tortaxis erectus</i> (Pilsbry, 1906)	Guilin, Guangxi Prov., China	R. Anderson
S.f. Rumininae	<i>Riebeckia</i> sp.	Samha, Sokotra Archipelago	E. Neubert
	<i>Rumina decollata</i> (Linnaeus, 1758)	Sicily	A. Davison
	<i>Xerocerastus</i> sp.	Otjiwarongo, Namibia	W. Sirgel
S.f. Subulininae	* <i>Allopeas clavulinum</i> (Potiez & Michaud, 1838)	University of Sao Paulo, Brazil	F. Florens & C. Baider
	* <i>Leptinaria lamellata</i> (Potiez & Michaud, 1838)	Botanical Garden, Rio de Janeiro, Brazil	F. Florens & C. Baider
	* <i>Paropeas achatinaceum</i> (L. Pfeiffer, 1846)	Agra Gajaba's Garden Sri Lanka	D. Raheem
	* <i>Subulina octona</i> (Bruguere, 1789)	Island of Pulo Anna, Sonsorol, Southwest Islands, Republic of Palau	R. Rundell & A.M. Gawel
	<i>Subulina striatella</i> (Rang, 1831)	Kew Gardens (introduced)	F. Naggs
	* <i>Subulina vitrea</i> (Mousson, 1887)	Gauss, Namibia	?
	<i>Zootecus insularis</i> (Ehrenberg, 1831)	Dubai, United Arab Emirates	S. Green
	<i>Thyrophorella thomensis</i> Greeff, 1882	Zampala, São Thomé, West Africa	A. Gascoigne
Thyrophorellidae			
Streptaxidae (outgroups)	* <i>Gibbulinella dewinteri</i> Bank, Groh & Ripken, 2002	Puntas Coloradas, La Gomera Island	M. Ibañez
	<i>Gonaxis quadrilateralis</i> Preston, 1910	Reunion	O. Griffiths
	<i>Gonospira</i> sp.	Mauritius	O. Griffiths

** NHM-Natural History Museum

3.2.2. DNA extraction, PCR amplification and sequencing

For all new specimens, tissue slices (approximately eight mm³) from the foot muscle of the snail were obtained and the DNA was extracted using the standard CTAB method of DNA extraction (Section 2.1.1 of Chapter 2, pp. 37-39). The DNA was

provided by C. M. Wade for those specimens utilized in the Wade *et al.* (2001, 2006) studies.

Amplification by PCR, gel migration and purification of PCR products from agarose gels were carried out as described in Sections 2.2-2.4 of Chapter 2, pp. 42-59. Nearly the entire fragment of the nuclear LSU rRNA gene, the internal transcribed spacer 2 (ITS2) and 80 bp of the 5.8S rRNA gene (for a total of approximately 4000 bp) were amplified using six overlapping primer pairs listed in Table 2.1 of Chapter 2, p. 47. Sequences of the Morgan (2002) nuclear cytoplasmic actin gene fragment (approximately 900 bp), the Colgan *et al.* (2003) nuclear histone 3 fragment (328 bp), the Folmer *et al.* (1994) mitochondrial COI fragment (approximately 650 bp) and the Palumbi *et al.* (1991) mitochondrial 16S fragment (420-500 bp) were amplified using the primers listed in Tables 2.2-2.5 of Chapter 2, pp. 50-53. For all fragments, both sense and anti-sense strands were sequenced directly using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 termination cycle sequencing chemistry (see Section 2.6 of Chapter 2, pp. 59-61 for details).

3.2.3. Sequence analysis

Sequences were assembled using the STADEN package version 1.5.3 (Staden *et al.*, 2000) and aligned manually within the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994). The rRNA and 16S rRNA sequences were aligned following alignments generated by C. Hudelot for the Stylommatophora based on the secondary structure of these genes. Ambiguous sites due to sequencing errors (for all genes) or the presence of multiple copies (for the actin gene) were assigned as described in Section 2.9.3 of Chapter 2, pp. 72-73. Cloned sequences of actin gene copies from three

representative taxa of the Achatinoidea were used as a guide to assign ambiguous sites in the actin gene (see Section 2.7 of Chapter 2, pp. 61-66).

For each gene fragment, the average base frequencies as well as the numbers of variable and parsimony-informative sites were determined in PAUP* Version 4.0b10 package (Swofford, 2002) (Section 2.9.4 of Chapter 2, p. 73). Corrected distances were computed after determination of the optimal model for DNA sequence evolution. This was carried out using likelihood by estimating the log likelihood scores in the PAUP* for the JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P (Kimura, 1980), HKY85 (Hasegawa *et al.*, 1985), TN93 (Tamura & Nei, 1993) and the GTR (Rodriguez *et al.*, 1990) models as well as their variant that incorporated gamma distributed rates (Γ) (Yang, 1993), after which the scores were compared for significant differences using the Likelihood Ratio Test (LRT) (see Section 2.9.5 of Chapter 2, pp. 73-77 for details of the LRT).

In order to examine the sequences for evidence of saturation, the following plots were generated: pairwise corrected versus pairwise uncorrected distances (Plot 1); pairwise uncorrected transition and transversion distances versus pairwise uncorrected total distances (Plot 2); and pairwise uncorrected transition distances versus pairwise uncorrected transversion distances (Plot 3) (see Section 2.9.6 of Chapter 2, pp. 77-81). Gene datasets that demonstrated saturation were excluded from subsequent phylogenetic analyses. To test for the presence of phylogenetic signal, tree length distribution using parsimony was also determined for a g1 measure of skewness for 10,000 tree length replicates (see Section 2.9.7 of Chapter 2, pp. 81-82). Due to the potential problems posed by the presence of paralogous sequences the actin gene was assessed for its utility in phylogenetic analyses using the procedures outlined in Section 2.9.13, pp. 90-91.

Phylogenetic trees were generated from the aligned datasets using the model based maximum likelihood (ML), Bayesian inference (BI) and neighbor-joining (NJ) methods and the non-model based maximum parsimony (MP) method (Section 2.9.8 of Chapter 2, pp. 82-85). The trees for NJ and MP were generated in PAUP* while that of ML was generated using PHYML Version 2.4.4 (Guindon *et al.*, 2005). Bootstrap resampling (Felsenstein, 1985) with 1000 replicates for MP and NJ (using PAUP*) and 1000 replicates for ML (using PHYML) were also carried out. Bayesian inference (BI) was performed using the MrBayes (Version 3.1.2) package (Ronquist & Huelsenbeck, 2003) using four chains of a Markov Chain Monte Carlo algorithm. The number of generations to explore the tree space and the heating temperature used for each gene were optimised as described in the BI segment of Section 2.9.8 of Chapter 2, pp. 83-84 and on Appendix 2.3.2, pp. 369-370. A consensus tree for each gene (where unsaturated) was constructed using the last 1000 trees.

To determine if the sequences from the genes that did not exhibit saturation could be combined and analyzed as a single dataset of concatenated sequences, a partition-homogeneity test (Swofford, 2002) was carried out within PAUP* (Section 2.9.10 of Chapter 2, p. 86-88). Two sets of concatenated sequences were prepared. The first one included only those taxa with complete sequences for all the gene fragments under consideration while the second set included all taxa. Studies show that taxa with incomplete datasets could still be used and their phylogenetic positions accurately determined provided that a lot of characters were surveyed from the other sequences (Wiens, 2006). Inclusion of such taxa could also benefit the combined gene analyses by removing misleading long branches and potentially increase the accuracy of the phylogeny (Wiens, 2006).

Where taxonomic groups expected to be monophyletic based on taxonomy did not cluster together in the optimal trees, their monophyly was tested using the Shimodaira-Hasegawa (1999) test in PAUP* (Section 2.9.11 of Chapter 2, p. 89).

3.3. Results

3.3.1. Molecular data

Twenty four achatinoid and three streptaxid outgroup taxa (*Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp.) were examined. Sequences were obtained for the nuclear rRNA cluster, actin and histone 3 genes as well as the mitochondrial 16S and cytochrome c oxidase I genes. Sequence alignments are presented in Appendices 3.1-3.5 on pp. 375-432. All sequences were scanned for the presence of ambiguous positions in the direct sequences that might be indicative of the presence of multiple gene copies. Ambiguities indicative of multiple gene copies were detected only for the actin gene (see Section 3.3.2.2, p. 155-156 for details). A summary of the molecular data for all the sequences is shown in Table 3.2. For the protein-coding genes (actin, histone 3 and COI), summary information was obtained separately for the entire gene (all codon positions), the combined 1st and 2nd codon positions and the 3rd codon position only. Uncorrected and corrected pairwise distances and their optimal models were determined for all five genes and are summarized in Table 3.3. The GTR+ Γ model was found to be the optimal model by LRT for the nuclear rRNA cluster, the mitochondrial 16S RNA and for both the full (all codon positions) and 3rd codon position datasets for the actin, histone 3 and COI genes. For the combined 1st + 2nd codon position datasets, TN93+ Γ was determined to be the

optimal model for the actin, TN93 for the histone 3, and GTR+ Γ for COI. (See Appendix 3.6, p. 433 for the summary of the LRT results).

Nearly the entire fragment of the LSU rRNA gene, the internal transcribed spacer 2 (ITS 2) and 80 nucleotides of the 5.8S rRNA gene was sequenced (approximately 4000 nucleotides) for the rRNA gene cluster, of which 3435 nucleotides could be aligned unambiguously. The ITS 2 region was too variable to align across all taxa and was therefore excluded from the analysis. A total of 260 (7.6%) variable sites were found, of which 151 were parsimony-informative. Pairwise distances across all taxa ranged from 0.001 to 0.034 (uncorrected) and 0.001 to 0.062 (corrected). The highest base frequency was obtained for G (0.318) followed by C (0.260), then by A (0.229) and finally by T (0.193). For the actin gene, 861 nucleotide positions were sequenced, of which all sites could be aligned unambiguously; of these, 288 (33.5%) were variable and 240 were parsimony-informative. Pairwise distances ranged from 0.009 to 0.173 (uncorrected) and 0.009 to 0.469 (corrected). Highest average base frequency was for A (0.274) followed by C (0.255) then T (0.250) and finally G (0.220). The majority of the variable sites (237) and parsimony-informative sites (210) were found at the 3rd codon position, while the combined 1st and 2nd codon positions only had 51 variable sites and 30 parsimony-informative sites. For the histone 3 gene, 328 nucleotides were sequenced and aligned unambiguously, with 103 (31.4%) variable positions and 71 parsimony-informative sites. Pairwise distances ranged from 0 to 0.183 (uncorrected) and 0 to 0.605 (corrected). Average base frequencies, in decreasing order, were as follows: C (0.321); G (0.271); A (0.242) and T (0.166). As with the actin gene, the 3rd codon position exhibited the most number of variable sites (93) as well as parsimony-informative sites (70) in comparison to the combined 1st and 2nd codon positions with only 10 and 1, respectively. Amplification of the histone 3 gene

fragment for *Cecilioides gokweanus* (Ferussaciidae), *Paropeas achatinaceum*, *Riebeckia* sp., and *Subulina vitrea* (Mousson) (Subulinidae) proved unsuccessful. Only a single streptaxid outgroup sequence (*Gibbulinella dewinteri*) was obtained for the histone 3 gene. For the COI gene, approximately 650 nucleotides were sequenced, with the length varying depending on the primers used. A total of 607 unambiguously aligned nucleotides were used; of which 279 (46.0%) were variable and 250 were parsimony-informative. Pairwise distances ranged from 0.160 to 0.265 (uncorrected) and 7.215 to 57.369 (corrected). Average base frequencies, in decreasing order, were as follows: T (0.395); A (0.249); G (0.194) and C (0.163), indicating a strong bias for T; this was heightened in the 3rd codon position where average frequency of T rose to 0.462. The 3rd codon position had 201 variable sites and 188 parsimony-informative sites, whereas the combined 1st and 2nd codon positions only had 78 and 62, respectively. Amplification of the COI gene for *Cecilioides gokweanus* was also unsuccessful. As with the histone 3 gene, only a single streptaxid outgroup sequence (*Gibbulinella dewinteri*) was obtained for the COI gene. The 16S rRNA gene product ranged from 420-500 nucleotides, with 294 that were unambiguously aligned, 139 (47.3%) that were variable, and 120 that were parsimony-informative. Pairwise distances ranged from 0.078 to 0.279 (uncorrected) and 0.107 and 1.082 (corrected). Base frequencies were as follows: T (0.306); A (0.296); G (0.217) and C (0.181), showing bias for A and T. Sequencing of the 16S rRNA gene fragment for *Eutomopeas layardi* and *Subulona* sp. (Subulinidae) was unsuccessful.

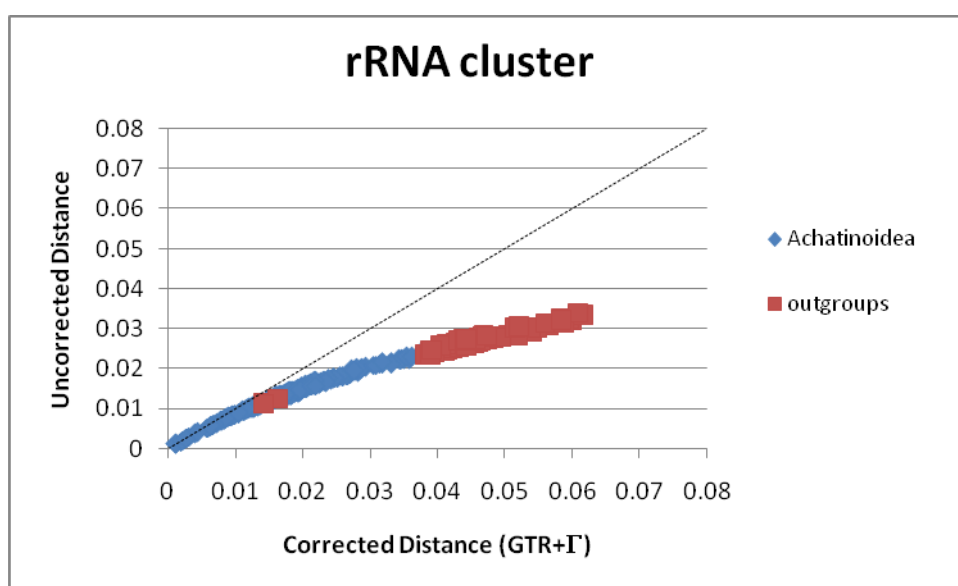
Table 3.3: Summary of molecular data across all genes used for the Achatinoidea and three streptaxid outgroup taxa (rRNA gene cluster, actin and 16S) and one streptaxid outgroup taxon (histone 3 and COI). A=Achatinoidea only; A+O=Achatinoidea and outgroup.

	Length (bp)	# of variable sites (%)		Range of distances (Uncorrected)		Optimal Model (α value for Γ distribution)	Range of distances (Corrected)		Average base frequencies								# of parsimony- informative sites	
									A		C		G		T			
		A	A+O	A	A+O		A	A+O	A	A+O	A	A+O	A	A+O	A	A+O	A	A+O
rRNA cluster	3435	212 (6.2)	260 (7.6)	0.001-0.029	0.001-0.034	GTR+ Γ (0.049)	0.001-0.048	0.001-0.062	0.229	0.229	0.260	0.260	0.319	0.318	0.192	0.193	113	151
Actin combined	861	283 (32.9)	288 (33.5)	0.009-0.173	0.009-0.173	GTR+ Γ (0.190)	0.009-0.469	0.009-0.469	0.276	0.274	0.254	0.255	0.219	0.220	0.251	0.250	230	240
Actin 1 st and 2 nd codon positions	574	50 (8.7)	51 (8.9)	0-0.046	0-0.046	TN93+ Γ (0.059)	0-0.105	0-0.105	0.316	0.316	0.228	0.228	0.228	0.228	0.229	0.229	29	30
Actin 3 rd codon position	287	233 (81.2)	237 (82.6)	0.025-0.444	0.025-0.444	GTR+ Γ (1.339)	0.023-1.056	0.023-1.056	0.192	0.189	0.307	0.312	0.203	0.206	0.298	0.294	201	210
Histone3 combined	328	97 (29.6)	103 (31.4)	0-0.156	0-0.183	GTR+ Γ (0.164)	0-0.491	0-0.605	0.241	0.242	0.322	0.321	0.272	0.271	0.165	0.166	66	71
Histone 3 1 st and 2 nd codon positions	218	8 (3.7)	10 (4.6)	0-0.023	0-0.028	TN93 (N/A)	0-0.024	0-0.029	0.287	0.287	0.285	0.285	0.261	0.261	0.167	0.167	1	1
Histone 3 3 rd codon positions	110	89 (80.9)	93 (84.5)	0-0.449	0-0.520	GTR+ Γ (1.811)	0-1.289	0-1.452	0.145	0.147	0.397	0.395	0.297	0.295	0.161	0.163	65	70
COI combined	607	276 (45.5)	279 (46.0)	0.160-0.265	0.160-0.265	GTR+ Γ (0.077)	7.215-57.369	7.215-57.369	0.249	0.249	0.163	0.163	0.194	0.194	0.394	0.395	250	250
COI 1 st and 2 nd codon positions	404	75 (18.6)	78 (19.3)	0.032-0.119	0.032-0.119	GTR+ Γ (0.086)	0.046-0.459	0.046-0.459	0.205	0.205	0.199	0.199	0.235	0.235	0.361	0.361	62	62
COI 3 rd codon positions	203	201 (99.0)	201 (99.0)	0.365-0.631	0.365-0.631	GTR+ Γ (0.386)	299.377-5420.470	299.377-5420.470	0.335	0.336	0.091	0.090	0.114	0.113	0.459	0.462	188	188
16S rRNA	294	137 (46.6)	139 (47.3)	0.078-0.279	0.078-0.279	GTR+ Γ (0.233)	0.107-1.082	0.107-1.082	0.292	0.296	0.184	0.181	0.220	0.217	0.304	0.306	113	120

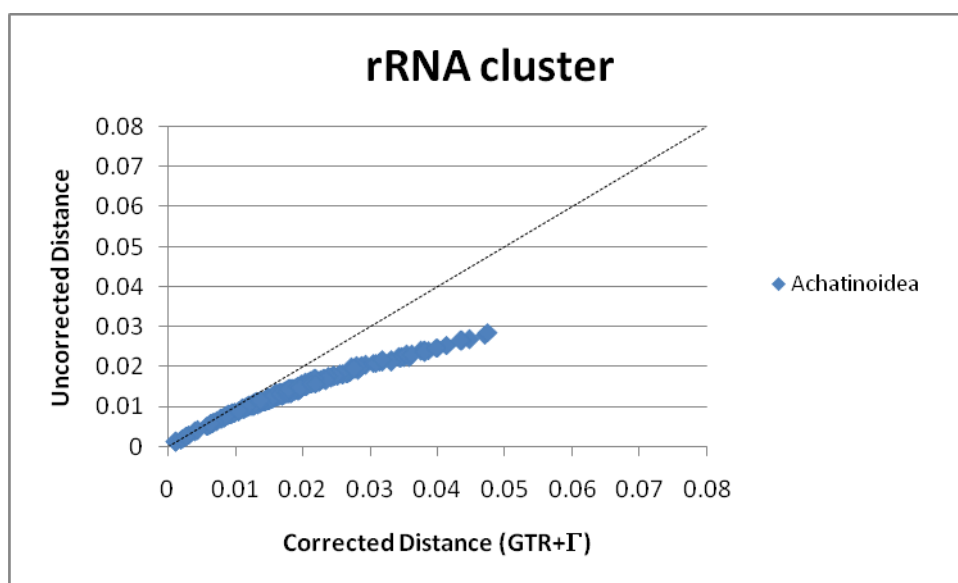
3.3.2. Sequence analyses

3.3.2.1. Evaluating for saturation and phylogenetic signal

For the rRNA cluster, the uncorrected versus corrected distances plots (Plot 1, Figure 3.2) demonstrated that the corrected distance based on the optimal GTR+ Γ model deviated from linearity starting at an uncorrected (p) distance of approximately 0.01 but no plateau was reached for the plots of either the Achatinoidea or the Achatinoidea plus outgroups. This suggested that the dataset for the rRNA cluster was far from being saturated and that the GTR+ Γ model was adequate at correcting the distances for multiple hits.



(A)



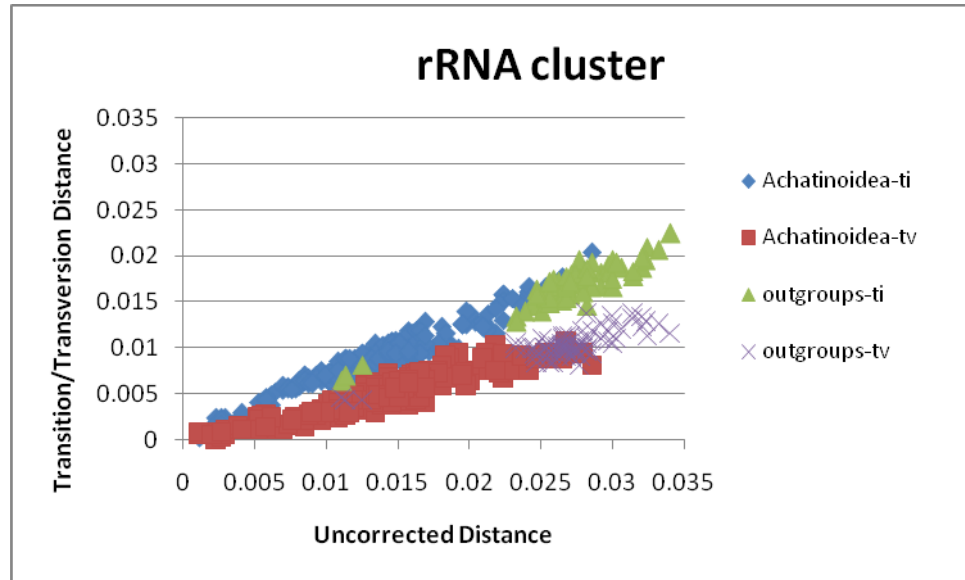
(B)

Figure 3.2: Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the rRNA cluster in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

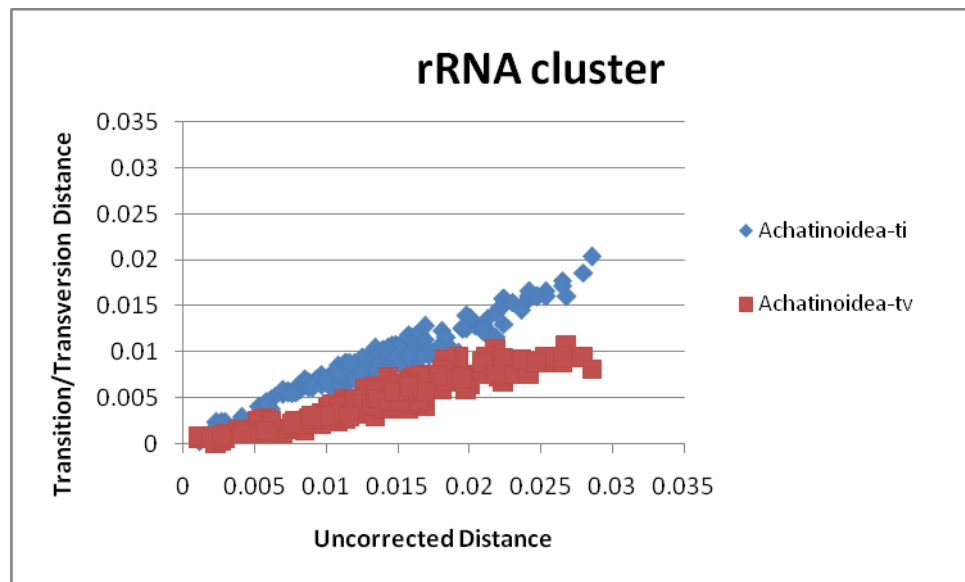
Plots for the transition and transversion rates versus uncorrected distances both with and without the outgroup taxa (Plot 2, Figure 3.3) showed transition distances that were higher than the transversion distances, with both types of substitutions increasing linearly. Plots for transitions against transversions both with and without the outgroup taxa (Plot 3, Figure 3.4) demonstrated that all transitions were higher than transversions. These suggested that neither transition nor transversion distances were saturated for the rRNA cluster.

A g_i value of -1.200 was obtained for the rRNA cluster with 10,000 replicates based on 27 taxa and 151 parsimony-informative characters. This value differed significantly from the critical g_i value of -0.1 at $P=0.05$ level of significance for 25 taxa and 100 parsimony-informative characters (Hillis & Huelsenbeck, 1992). This result was indicative of a strong phylogenetic signal.

The lack of substitution saturation and the presence of a strong phylogenetic signal suggested that the rRNA cluster was suitable for phylogenetic analyses of the Achatinoidea and outgroups.

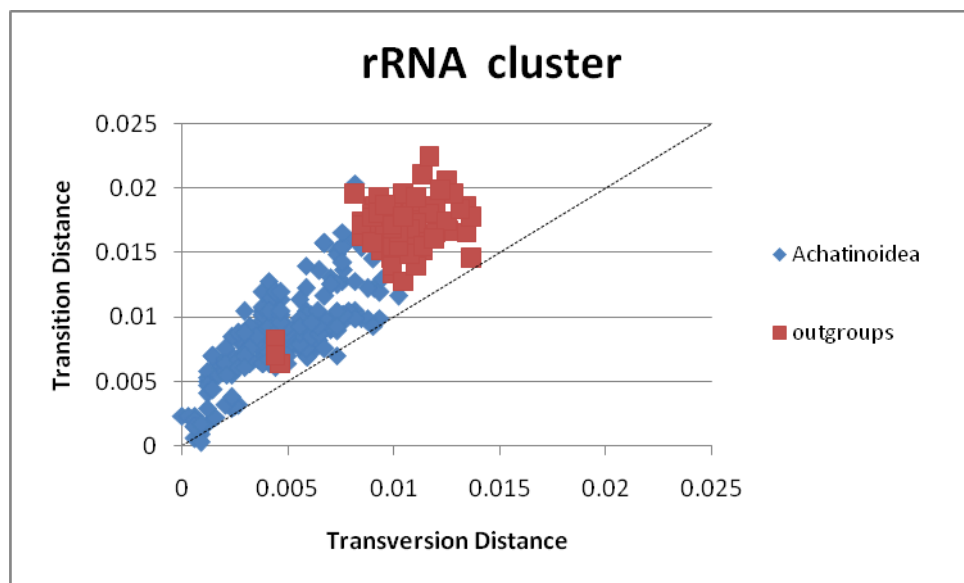


(A)

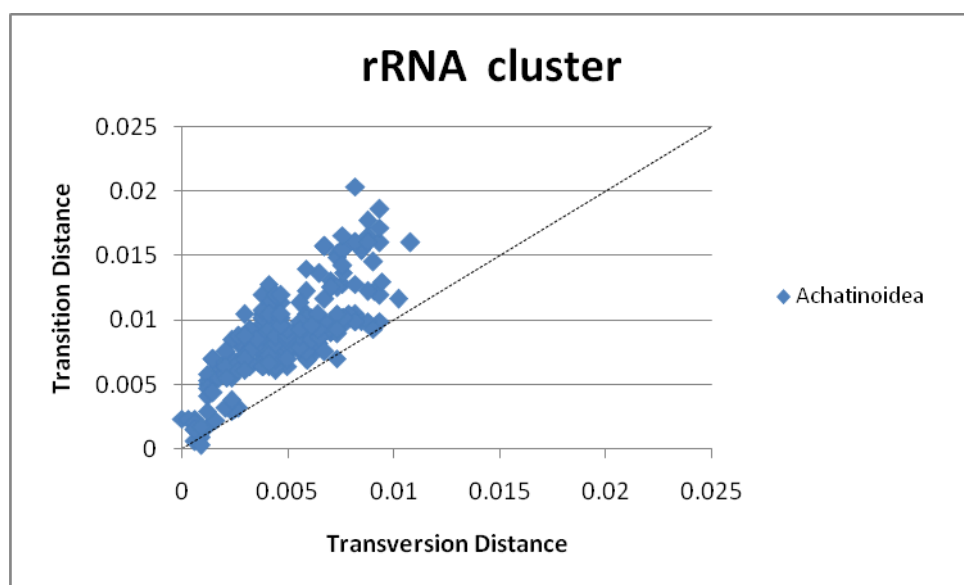


(B)

Figure 3.3: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the rRNA cluster in (A) the Achatinoidea and outgroup taxa and (B) the Achatinoidea only.



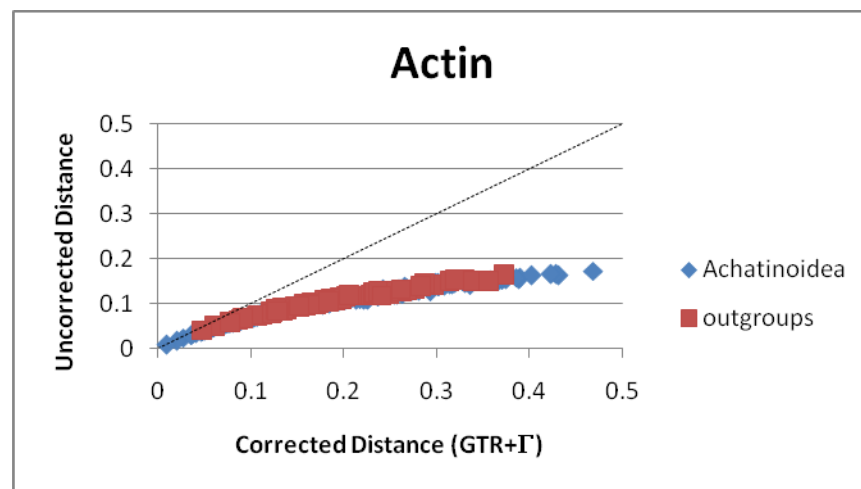
(A)



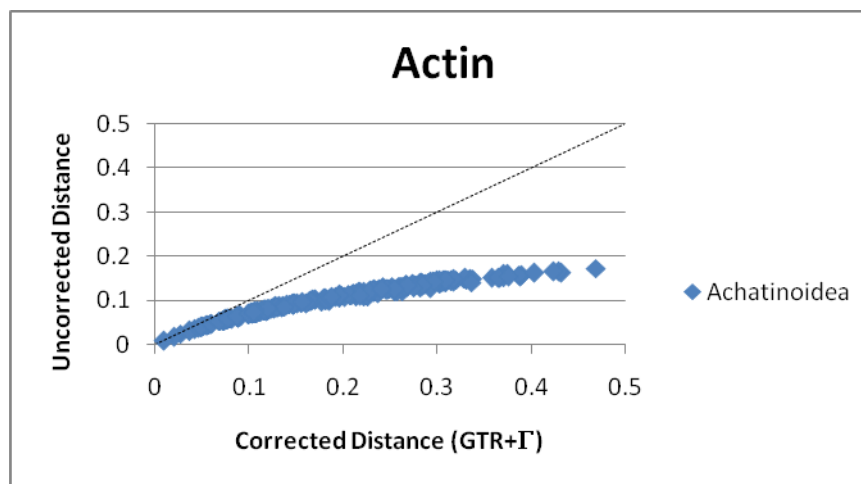
(B)

Figure 3.4: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the rRNA cluster in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

For the actin gene, Plot 1 (Figure 3.5) showed that the corrected distance, based on the optimal GTR+ Γ model, deviated from linearity starting at an uncorrected (p) distance of approximately 0.04 for both the Achatinoidea only and the Achatinoidea plus outgroup taxa. The plots were still increasing and had not reached a plateau, suggesting adequate correction of the optimal GTR+ Γ model and no saturation in the dataset.



(A)



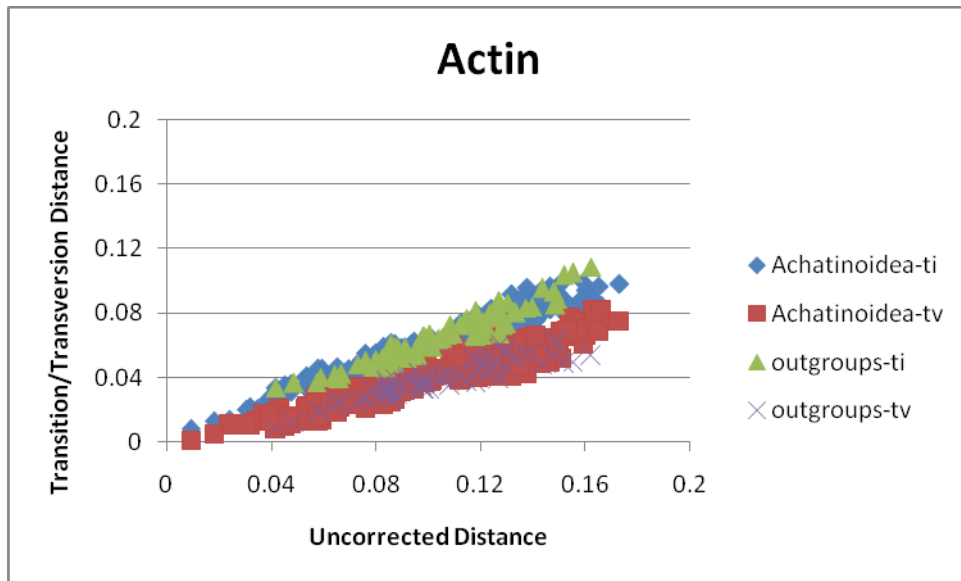
(B)

Figure 3.5: Plots of pairwise uncorrected distance against corrected distance for the actin gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

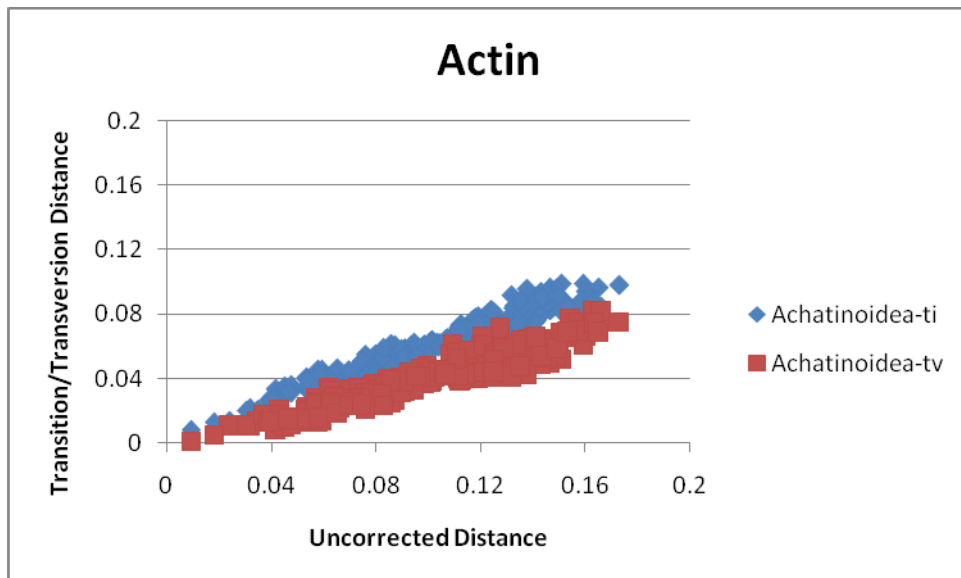
Plot 2 (Figure 3.6) for the actin gene, whether with or without outgroup taxa, exhibited a linear increase for both transitions and transversions. Plot 3 (Figure 3.7) showed that except for four points, all transitions were higher than transversions. These suggest that neither transition nor transversion distances were saturated for the actin gene.

A g_i value of -0.767 was computed for the actin gene based on 27 taxa and 245 parsimony-informative sites, much lower than the critical value of -0.1 at $p=0.05$ level of significance for 25 taxa and 100 parsimony-informative characters (Hillis & Huelsenbeck, 1992). The actin gene therefore exhibited strong phylogenetic signal.

The absence of substitution saturation and the presence of phylogenetic signal suggested that the actin gene was suitable for phylogenetic analyses of the Achatinoidea and outgroups.

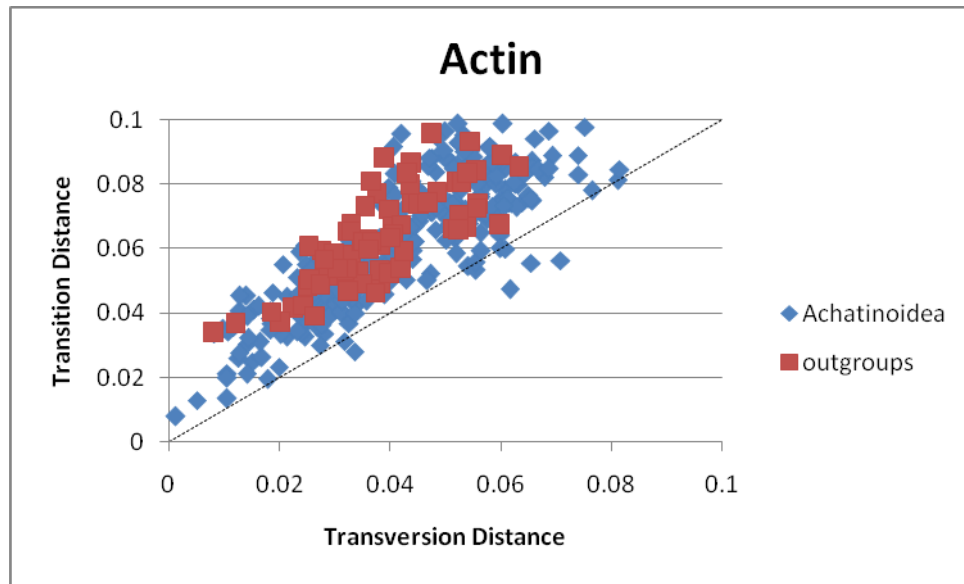


(A)

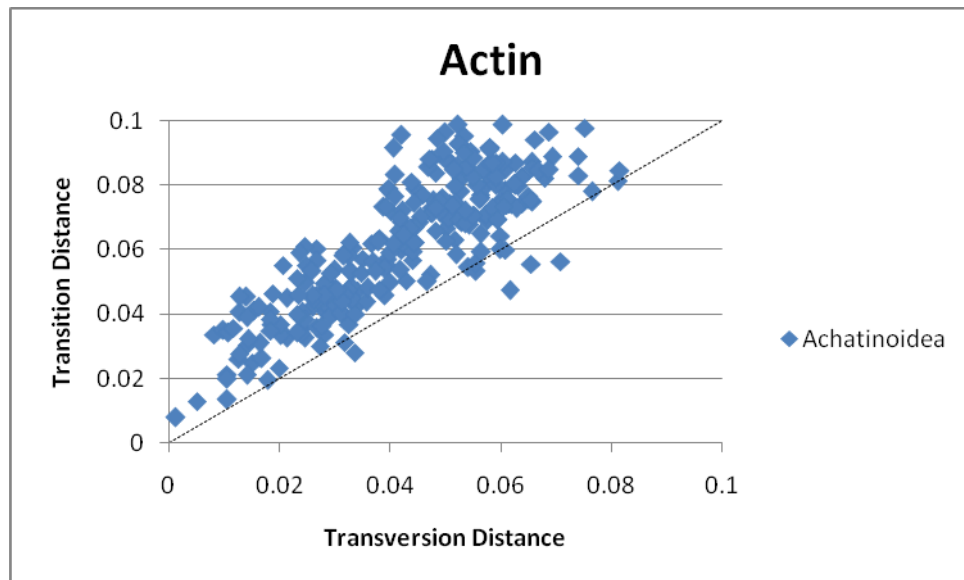


(B)

Figure 3.6: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the actin in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.



(A)



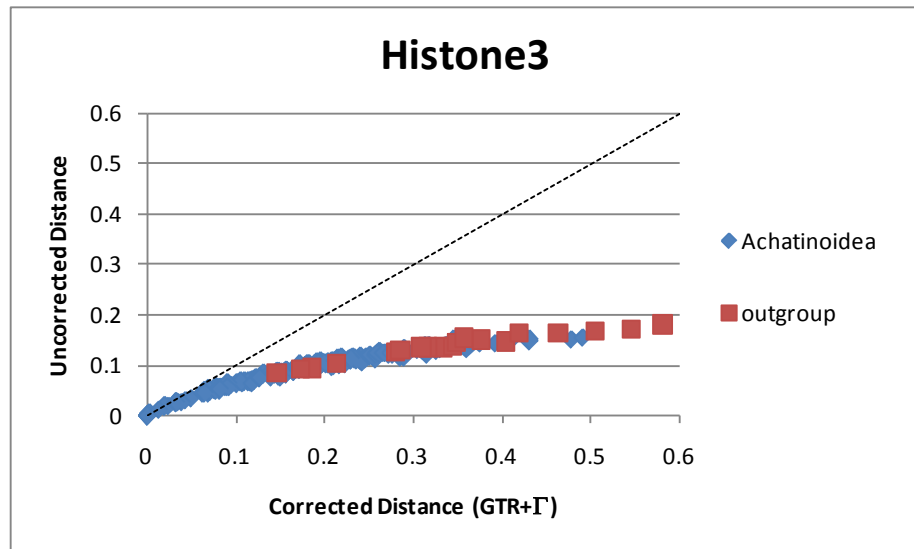
(B)

Figure 3.7: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the actin gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

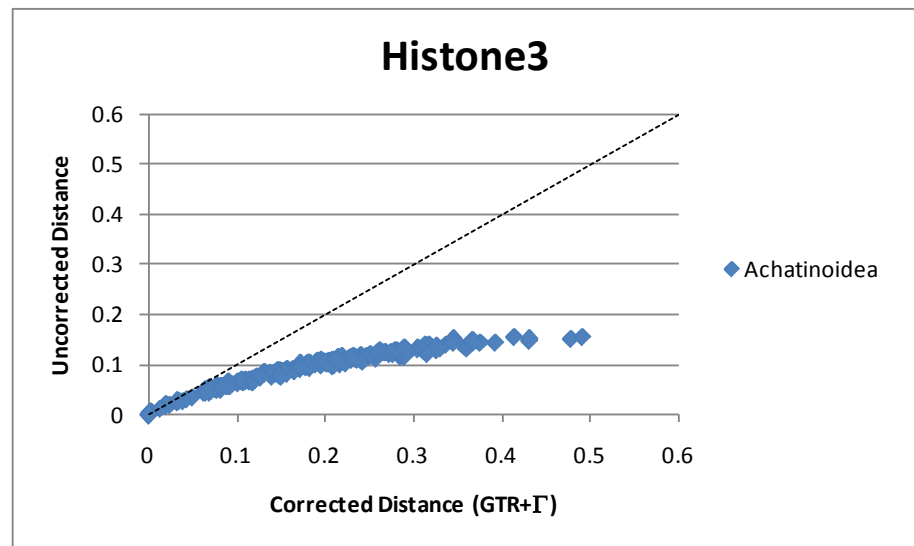
For the histone 3 gene, Plot 1 (Figure 3.8) revealed a curve for the corrected distances using the optimal GTR+ Γ model, with the deviation from a linear increase beginning at an uncorrected (p) distance of approximately 0.02 for both the

Achatinoidea only and including the streptaxid outgroup taxon *Gibbulinella dewinteri*.

The plots were still increasing and had not reached a plateau, implying that the histone 3 gene had not reached saturation.



(A)

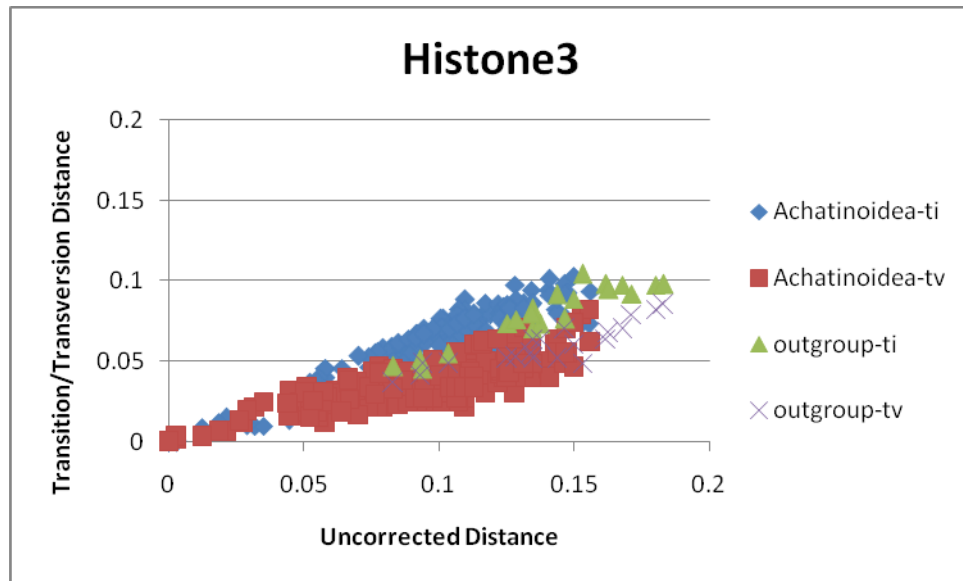


(B)

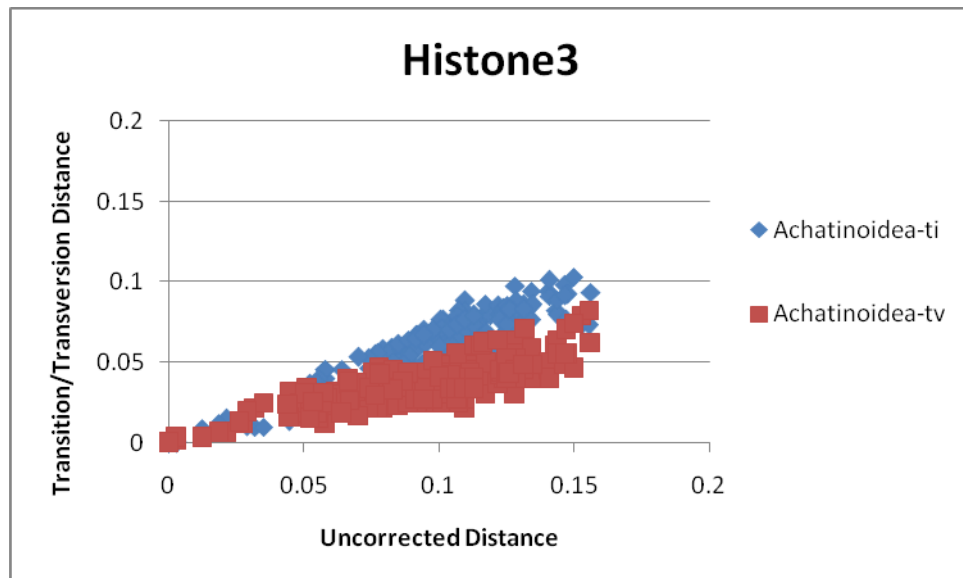
Figure 3.8: Plots of pairwise uncorrected distance versus corrected (GTR+Γ) distance for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.9) for histone 3 revealed a linear increase of both transitions and transversions for the Achatinoidea, but the transition line appeared to be beginning to curve when the outgroup was included. Although transitions were generally higher than transversions, some overlap was seen. Plot 3 (Figure 3.10) showed that the majority of transitions were higher than transversions. This suggested that the histone 3 dataset was just beginning to saturate, particularly if the outgroup taxon was included.

The g_i value for histone 3 based on 21 taxa and 71 parsimony-informative sites was -0.539, which was significantly smaller than the critical value of -0.16 based on 15 taxa and 50 parsimony-informative sites. The histone 3 gene therefore exhibited phylogenetic signal.

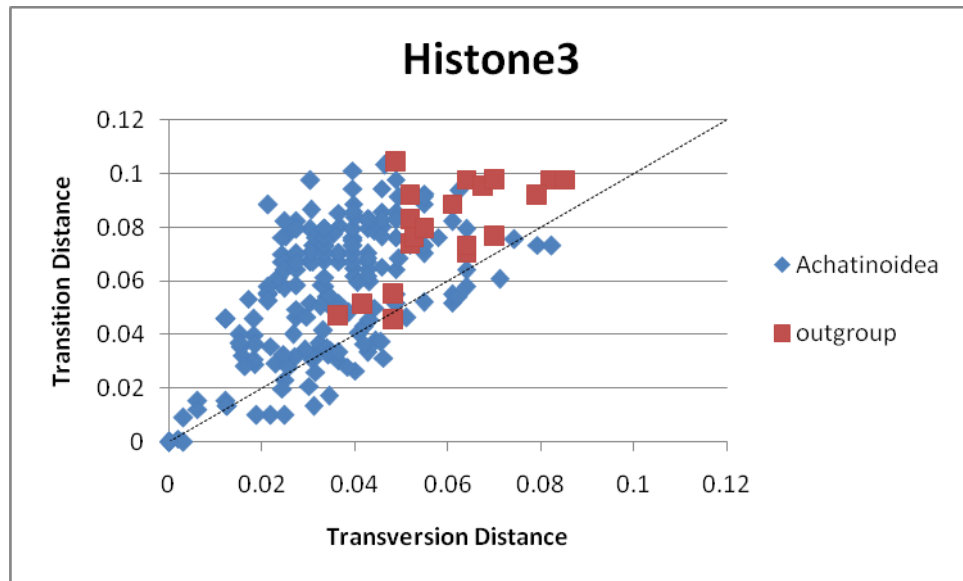


(A)

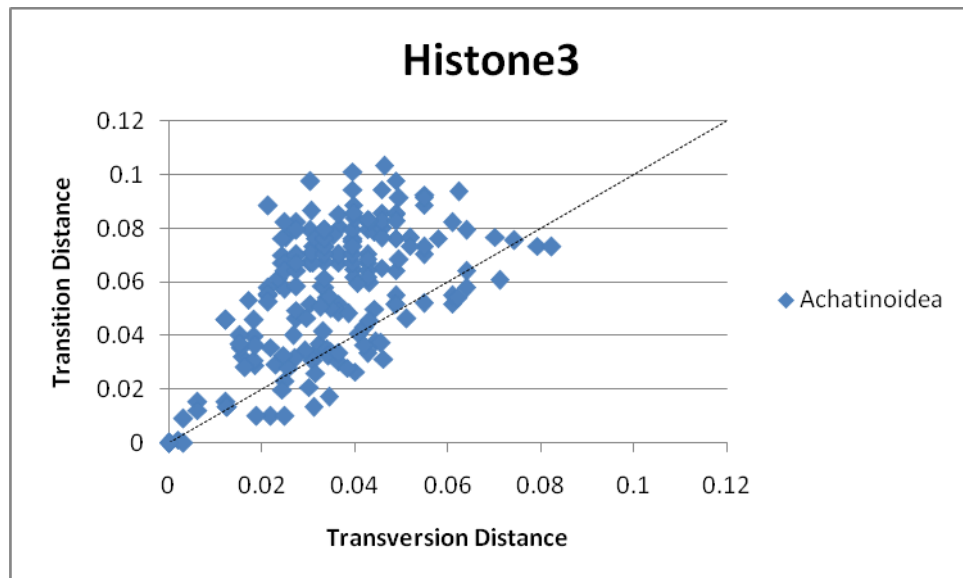


(B)

Figure 3.9: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

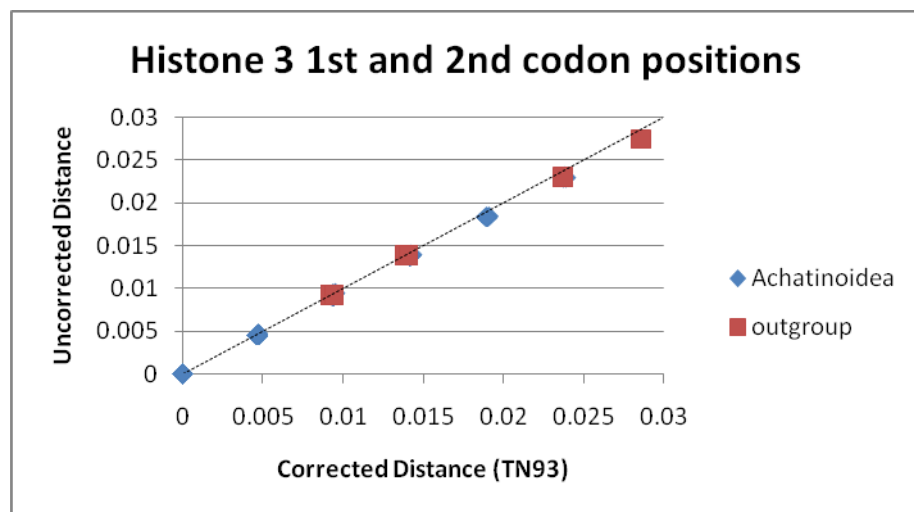


(B)

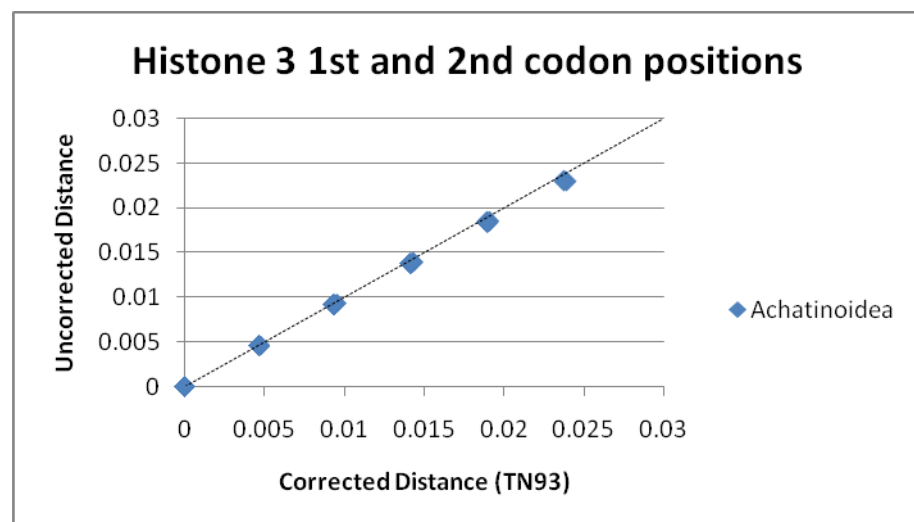
Figure 3.10: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

The analyses of the histone 3 gene as a whole (all codon positions) demonstrated that the dataset began to saturate when the outgroup taxon was included. The combined 1st and 2nd codon positions and the 3rd codon positions were therefore evaluated separately.

For the 1st and 2nd codon positions of the histone 3 gene, only a handful of sites varied (10 of 218 sites in the dataset that included the Achatinoidea and outgroup taxon). Plot 1 (Figure 3.11) showed a direct relationship between the corrected (based on the optimal TN93 model) and uncorrected distances, indicative of a highly conserved dataset with no saturation.



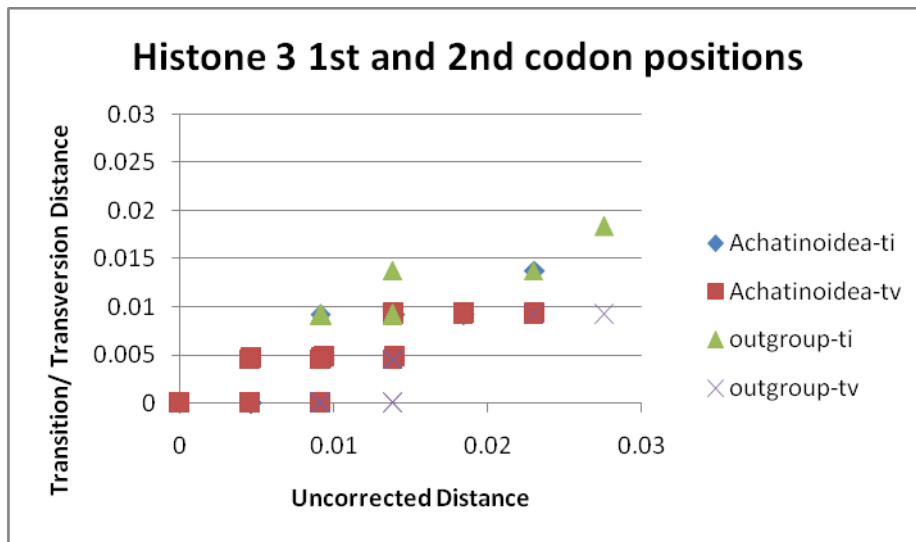
(A)



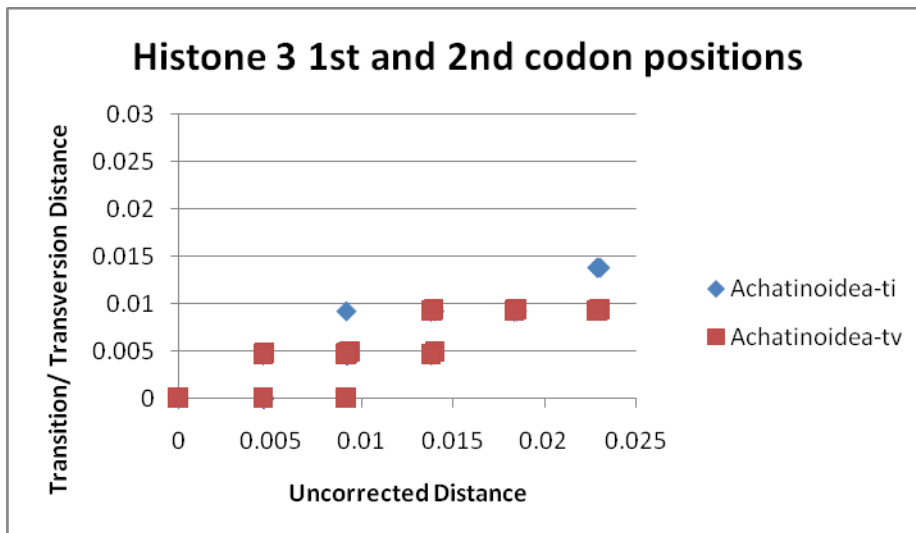
(B)

Figure 3.11: Plots of pairwise uncorrected distance versus corrected (TN93) distance for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plots 2 (Figure 3.12) and 3 (Figure 3.13) showed overlap between transitions and transversions, but curving due to saturation could not be assessed due to the small number of variable sites (10) evaluated.

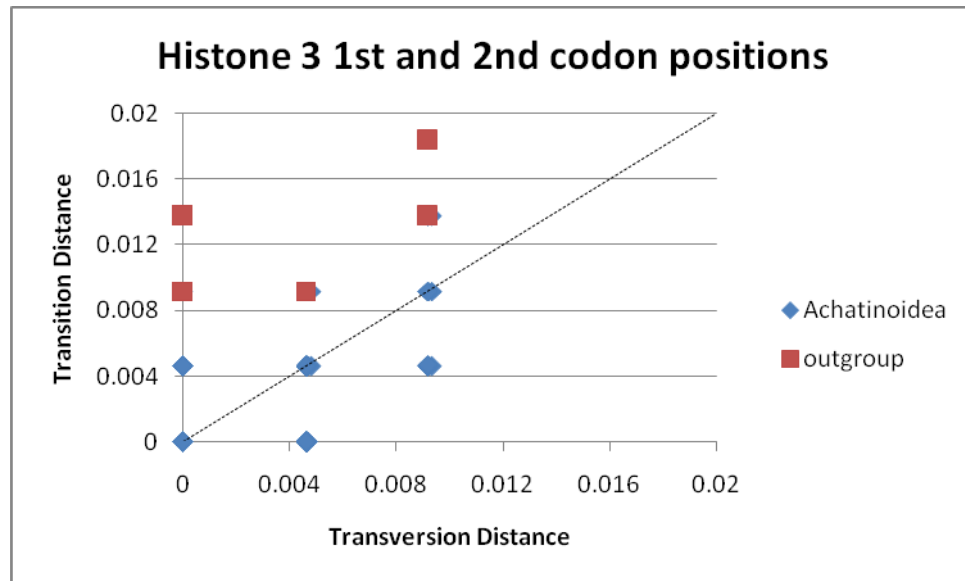


(A)

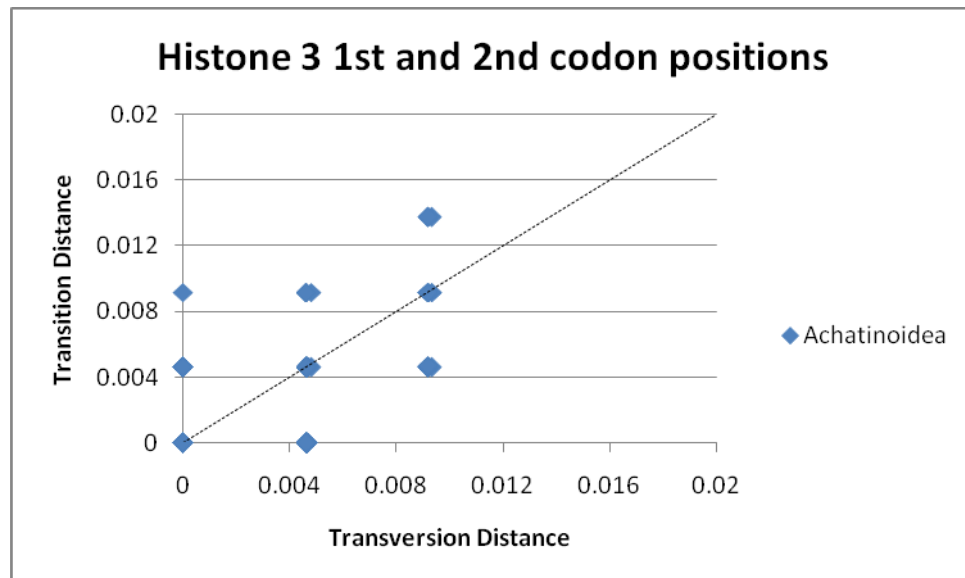


(B)

Figure 3.12: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only. Some transitions overlapped with transversions and were therefore not evident in the plots.



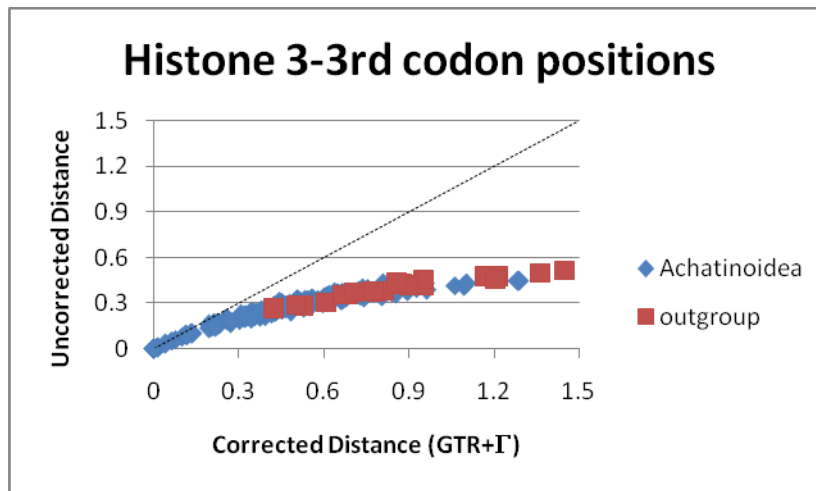
(A)



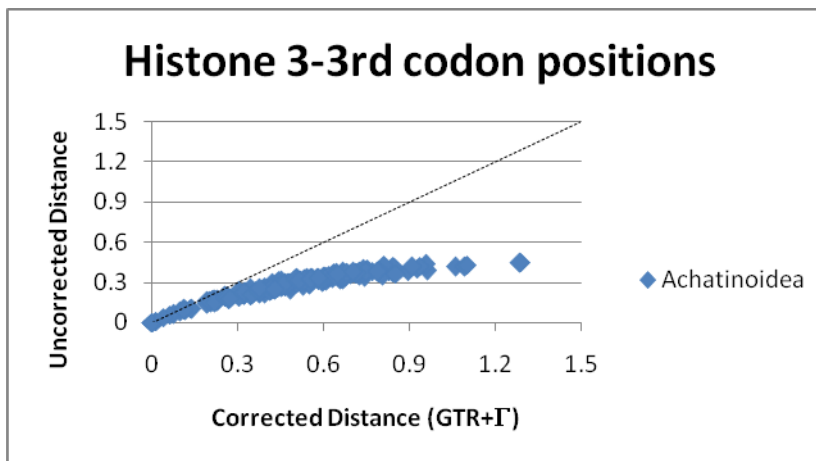
(B)

Figure 3.13: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

For the 3rd codon position, Plot 1 (Figure 3.14) revealed a curve for the corrected distances based on the optimal GTR+ Γ model that deviated from linearity at an uncorrected (p) distance of approximately 0.1. The corrected distances were still increasing and had not reached a plateau, implying that the dataset had not reached saturation.



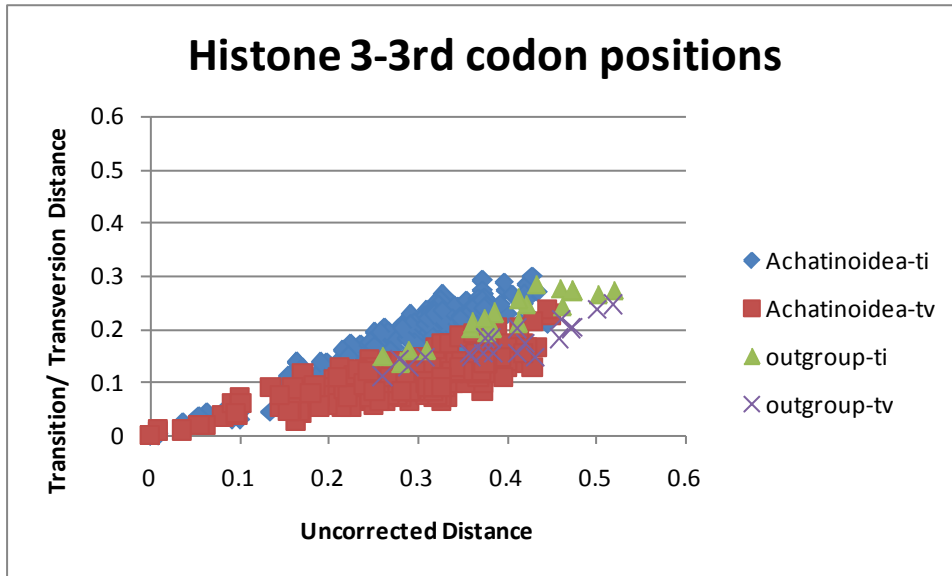
(A)



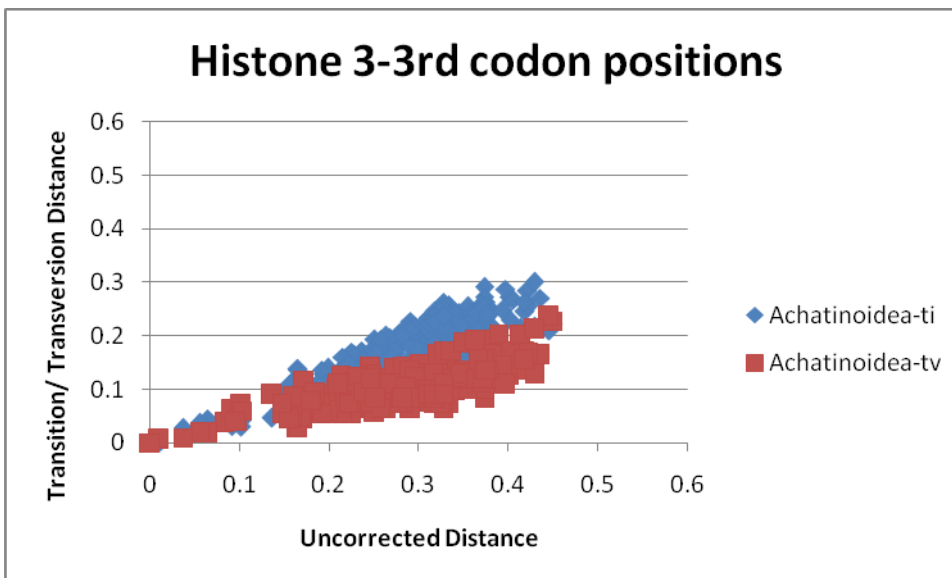
(B)

Figure 3.14: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 3rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.15) revealed a linear increase of both transitions and transversions, but the transition line appeared to be beginning to curve when the outgroup was included. Although transitions were generally higher than transversions, some overlap was seen. Plot 3 (Figure 3.16) demonstrated that majority of transitions were still higher than transversions. These results suggested that the 3rd codon positions of the histone 3 gene were just beginning to saturate, particularly as the outgroup taxon was included.

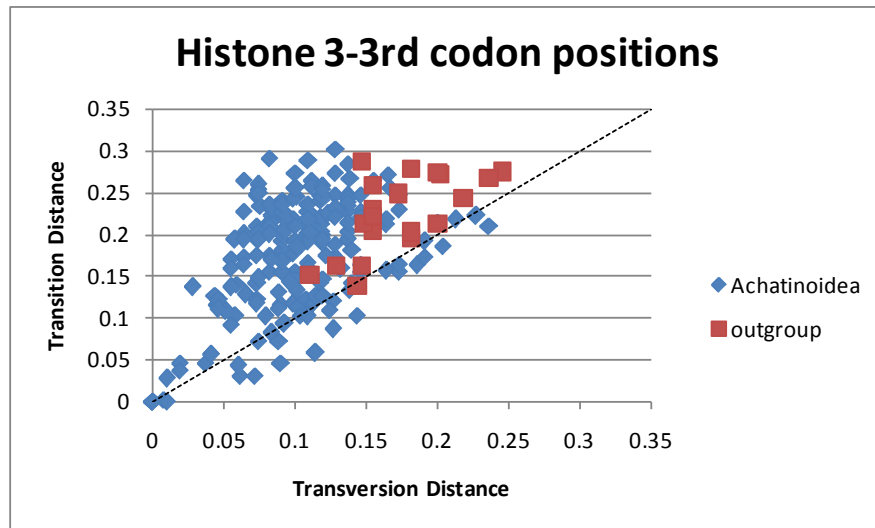


(A)

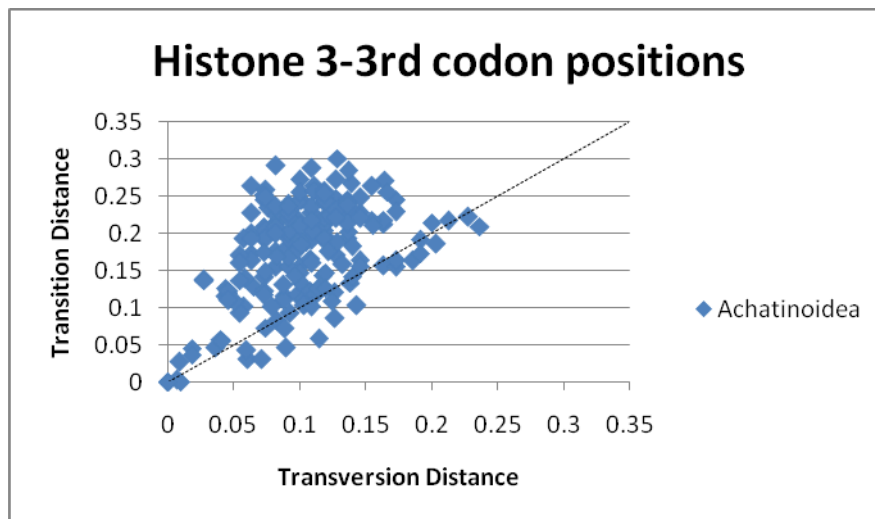


(B)

Figure 3.15: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

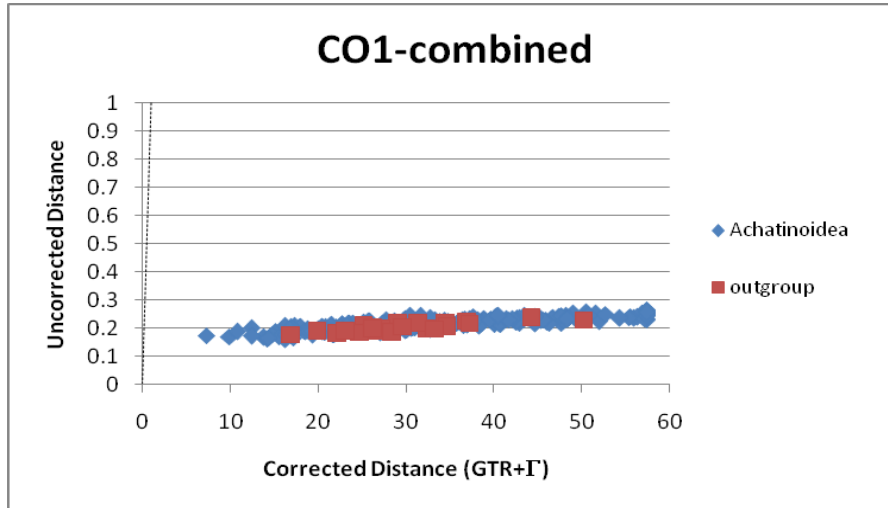


(B)

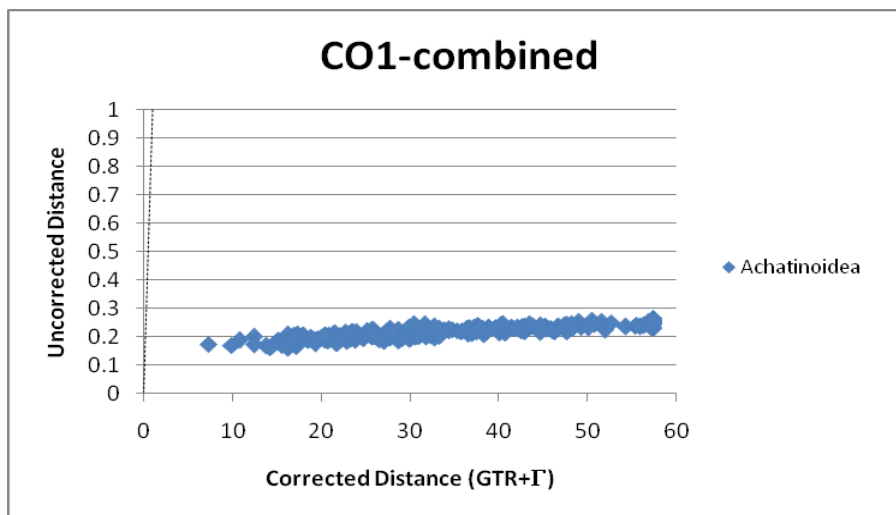
Figure 3.16: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

The findings suggested that the histone 3 dataset could be employed in its entirety in phylogenetic analyses of the Achatinoidea though with caution, particularly when the outgroup taxon was included. Moreover, particular care should be taken over the interpretation of the findings of the non-model based maximum parsimony method.

For the COI gene, Plot 1 (Figure 3.17) yielded an almost horizontal trend for all taxa and for the Achatinoidea only. This, along with corrected distances in excess of 50, indicated severe saturation for the COI gene and the optimal GTR+ Γ model was unable to correct the dataset for multiple hits.



(A)

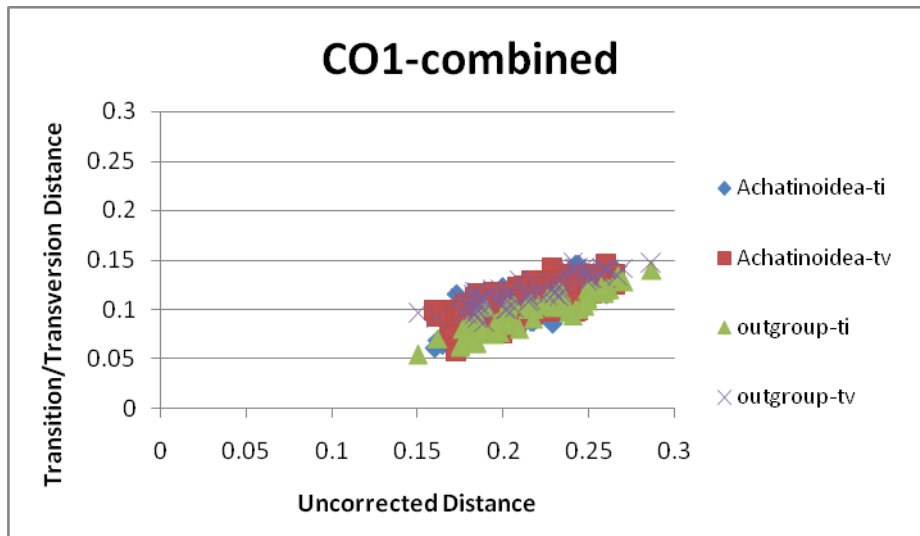


(B)

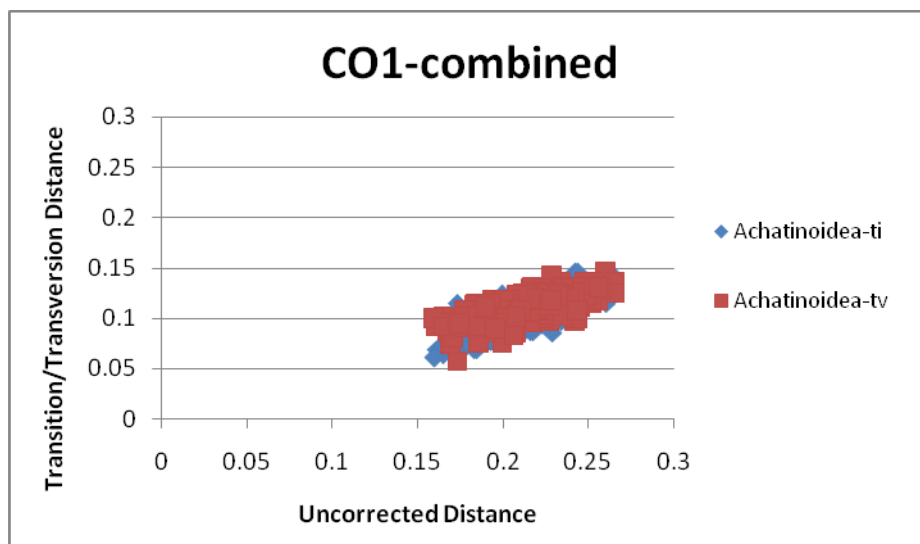
Figure 3.17: Plots of pairwise uncorrected distance against corrected distance for the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.18) revealed transversions overlapping with transitions in both the Achatinoidea only and the Achatinoidea plus outgroup datasets, while Plot 3 (Figure 3.19) revealed that the majority of the pairwise comparisons had higher rates of transversions than transitions.

The $g1$ test score for the COI gene based on 24 taxa and 250 parsimony-informative sites was -0.142, still significantly larger than the critical value of -0.16 for 15 taxa and 250 parsimony-informative sites. The $g1$ test therefore suggested that no phylogenetic information could be generated from the dataset.

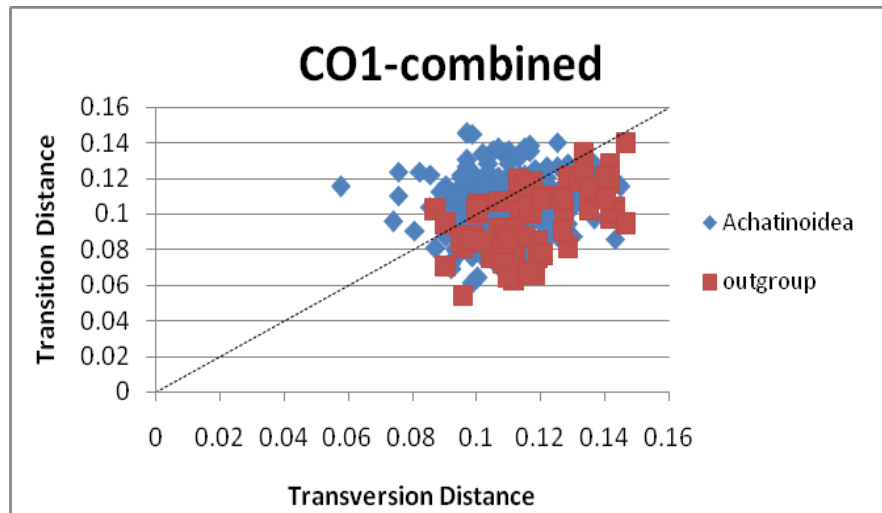


(A)

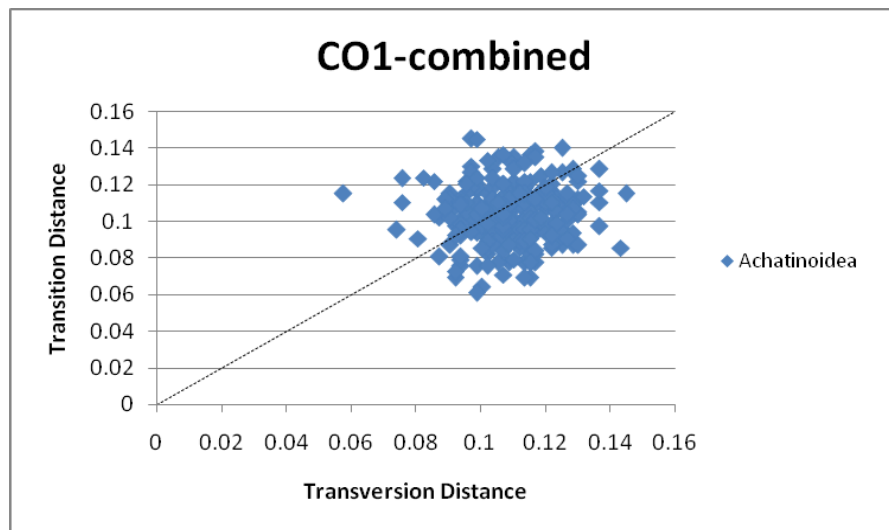


(B)

Figure 3.18: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

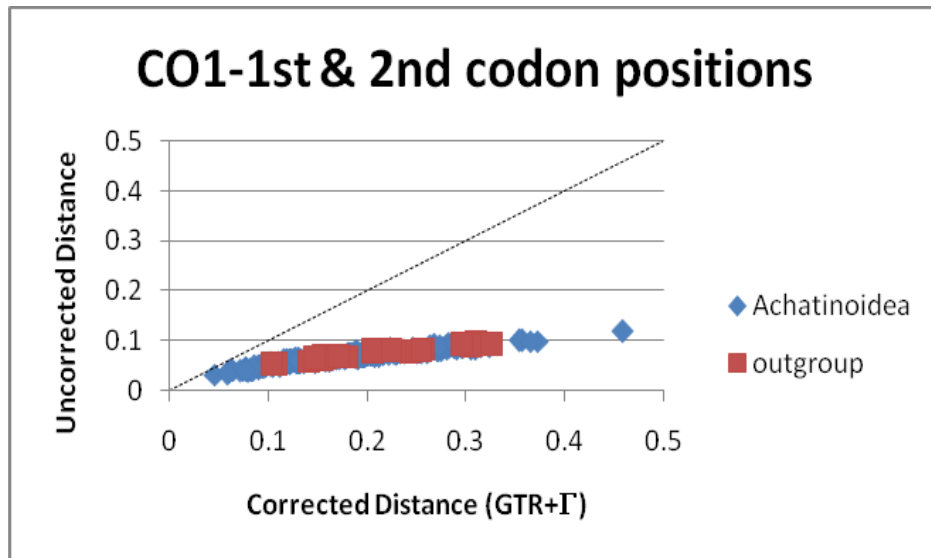


(B)

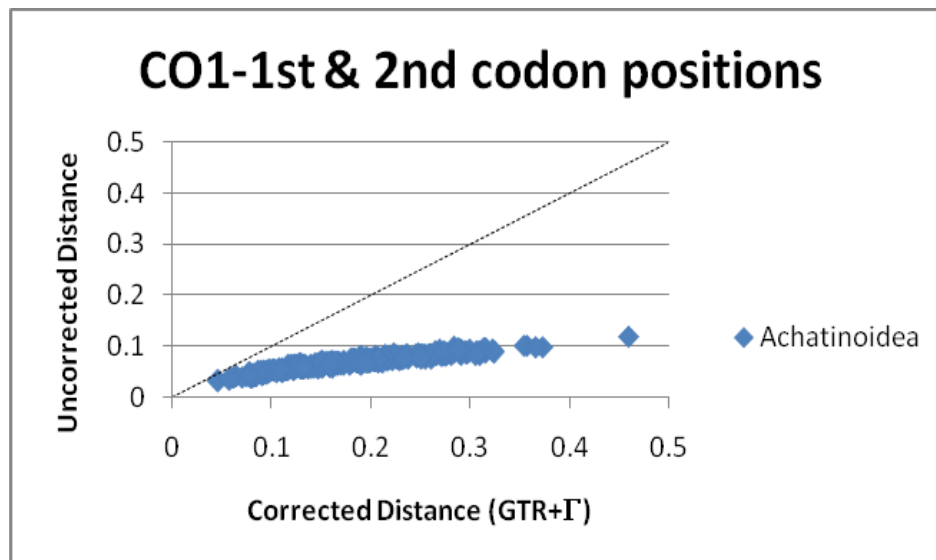
Figure 3.19: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the COI gene in **(A)** the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and **(B)** the Achatinoidea only.

The analyses of the COI gene as a whole (all codon positions) clearly demonstrated the existence of extensive saturation in the dataset, especially as shown by Plot 1. Its utility was further compromised by the absence of phylogenetic signal based on the g1 test. In order to evaluate whether this saturation was restricted to the 3rd codon position and whether some phylogenetic signal could be recovered from the 1st and 2nd codon positions, the combined 1st and 2nd codon positions and the 3rd codon position were evaluated separately.

For the 1st and 2nd codon positions of the COI gene, Plot 1 (Figure 3.20) revealed a curve for the corrected distances using the optimal GTR+ Γ model, with the deviation from linearity beginning at an uncorrected (p) distance of approximately 0.03 for both the Achatinoidea only and including the outgroup taxon. The plots were still increasing slightly and had not reached a plateau, implying that the 1st and 2nd codon positions of COI had not yet reached saturation.



(A)

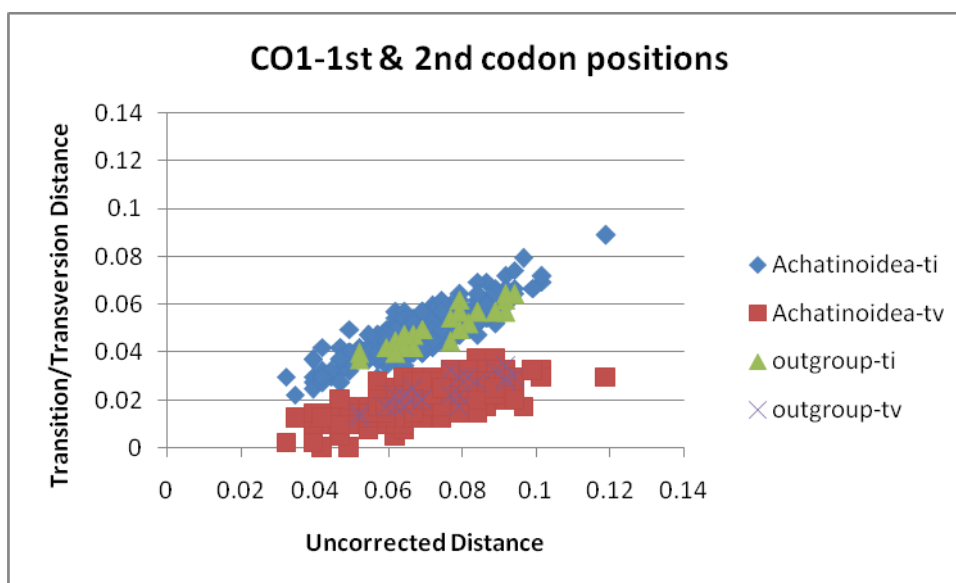


(B)

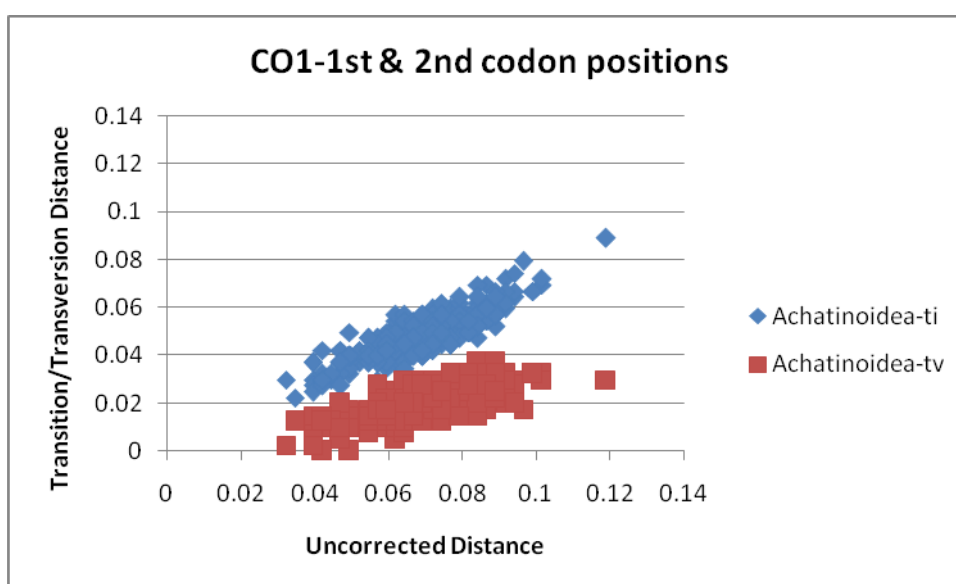
Figure 3.20: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 1st and 2nd codon positions of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.21) for the 1st and 2nd codon positions of the COI gene showed a linear increase of both transitions and transversions for the Achatinoidea and when the outgroup taxon was included, with the transitions being higher than the transversions. Plot 3 (Figure 3.22) also demonstrated the same conclusions as Plot 2. These findings suggested that the 1st and 2nd codon positions were not saturated.

The g1 value based on 24 taxa and 62 parsimony-informative sites was -0.283, which was significantly smaller than the critical value of -0.16 based on 15 taxa and 50 parsimony-informative sites. The 1st and 2nd codon positions of the COI gene therefore exhibited phylogenetic signal.

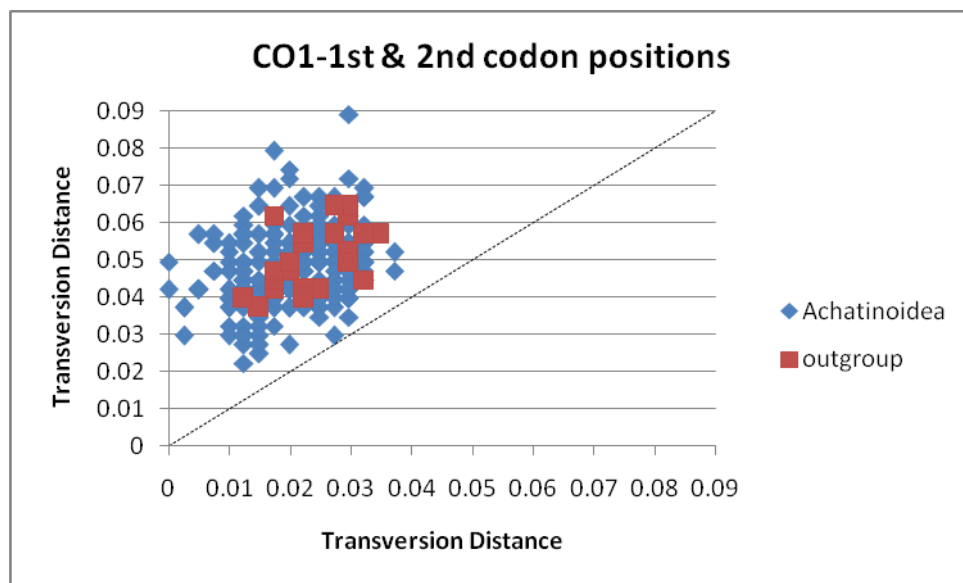


(A)

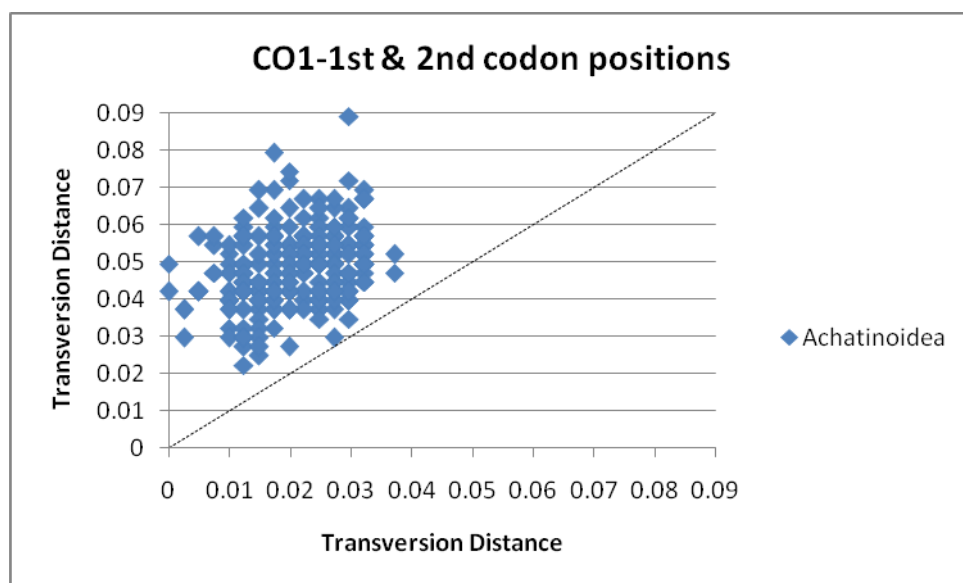


(B)

Figure 3.21: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1st and 2nd codon positions of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

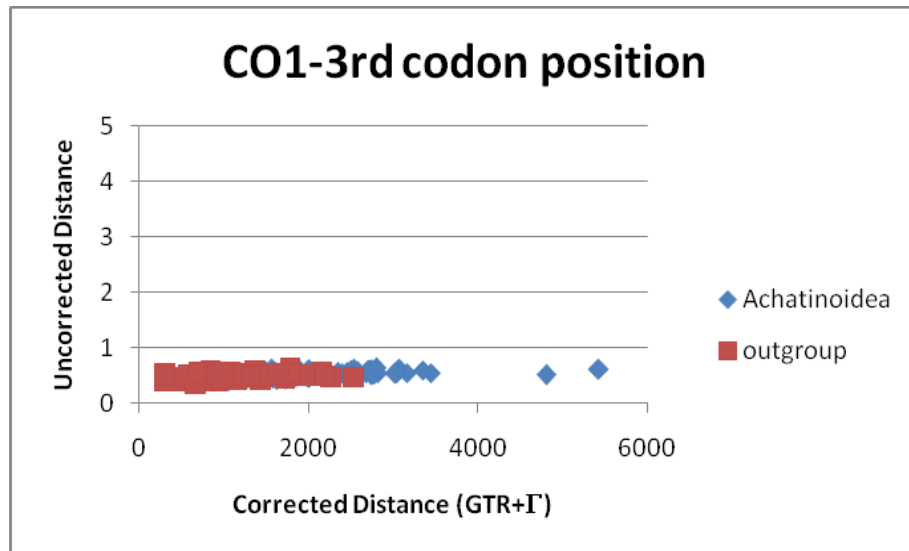


(B)

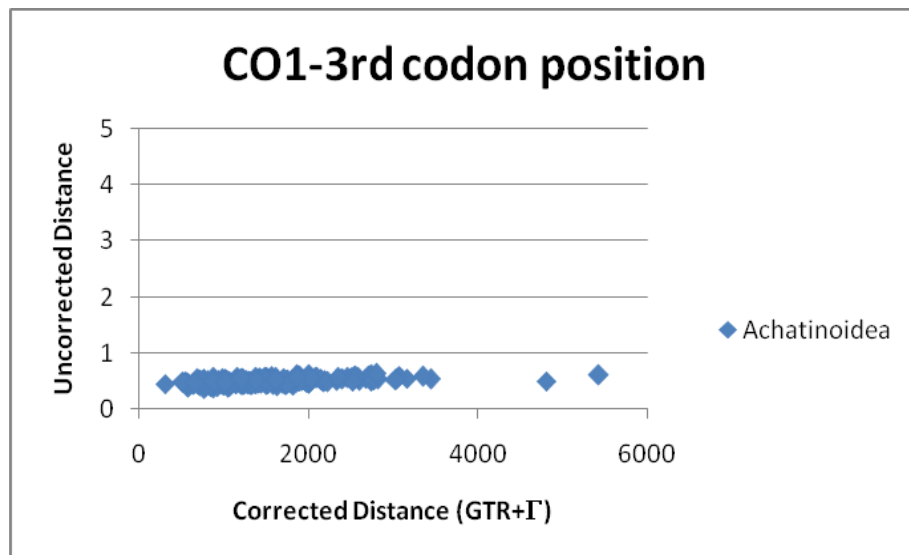
Figure 3.22: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1st and 2nd codon positions of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

For the 3rd codon position of the COI gene, Plot 1 (Figure 3.23) revealed that the optimal GTR+ Γ model was unable to correct the dataset for multiple hits as demonstrated by the nearly horizontal trend for both plots. The uncorrected distances

also ranged from 0.365 to 0.631 while corrected distances ranged from 299.377 to 5420.470 (see also Table 3.3). This indicated severe saturation for the 3rd codon position.



(A)

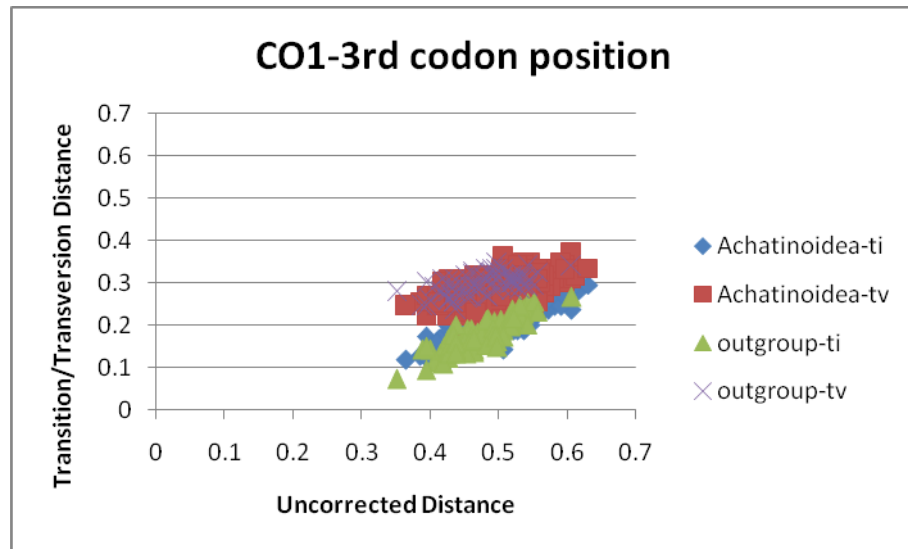


(B)

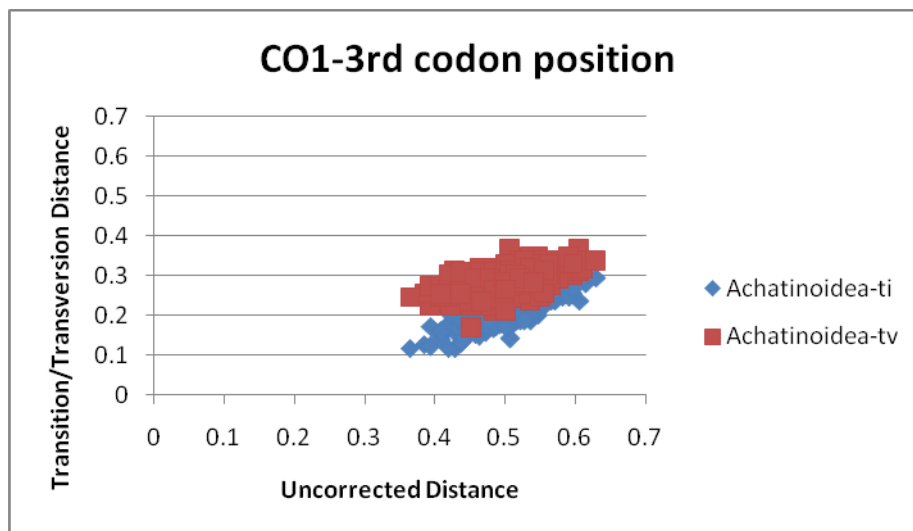
Figure 3.23: Plots of pairwise uncorrected distance against corrected distance for the 3rd codon position of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.24) for all taxa as well as for the Achatinoidea only showed transversions overlapping with transitions, with the transversions generally higher than the transitions. Plot 3 (Figure 3.25) demonstrated that most of the pairwise comparisons had higher rates of transversions than transitions.

The g1 test score for the 3rd codon position of the COI gene based on 24 taxa and 188 parsimony-informative sites was -0.050, which was significantly larger than the critical value of -0.16 for 15 taxa and 100 parsimony-informative sites. The 3rd codon position therefore did not exhibit any phylogenetic signal.

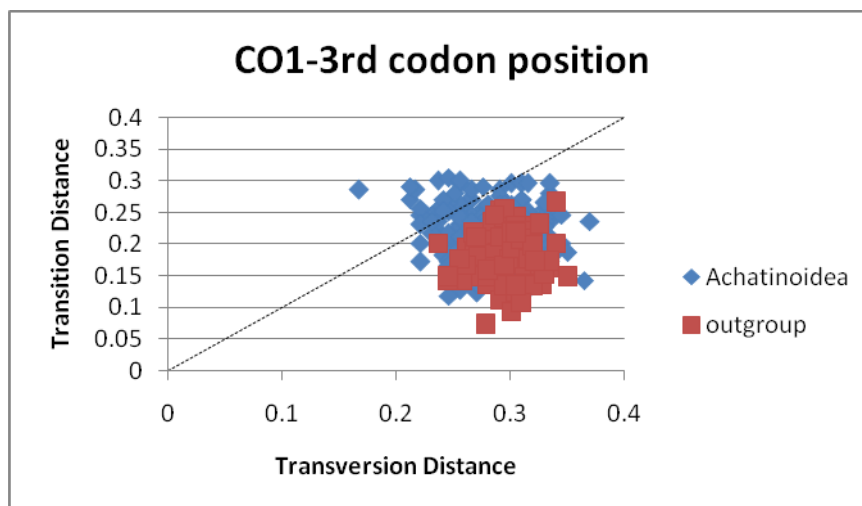


(A)

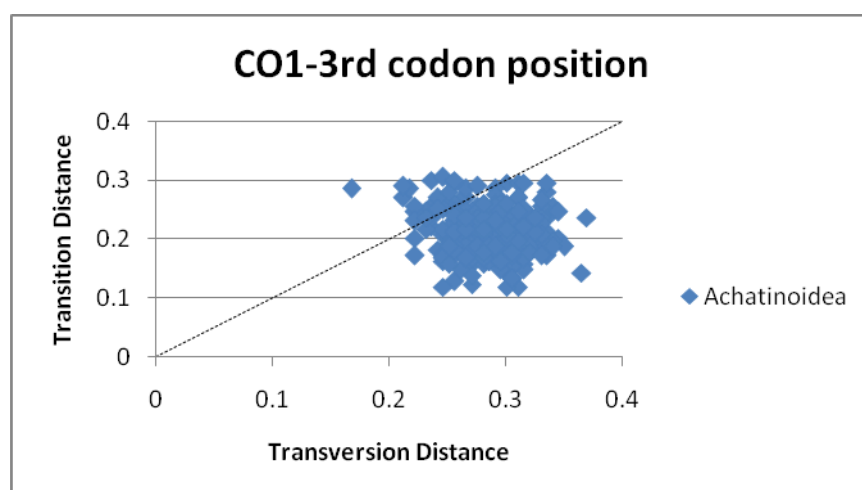


(B)

Figure 3.24: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3rd codon position of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

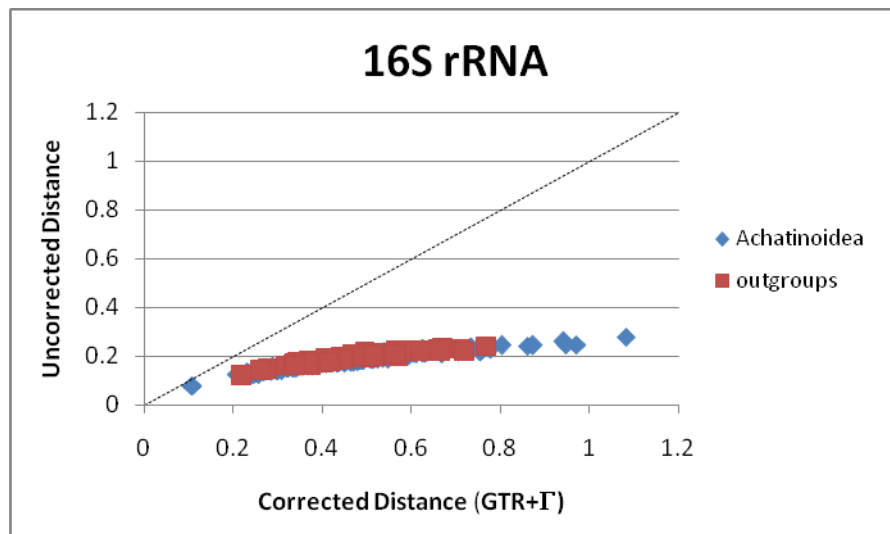


(B)

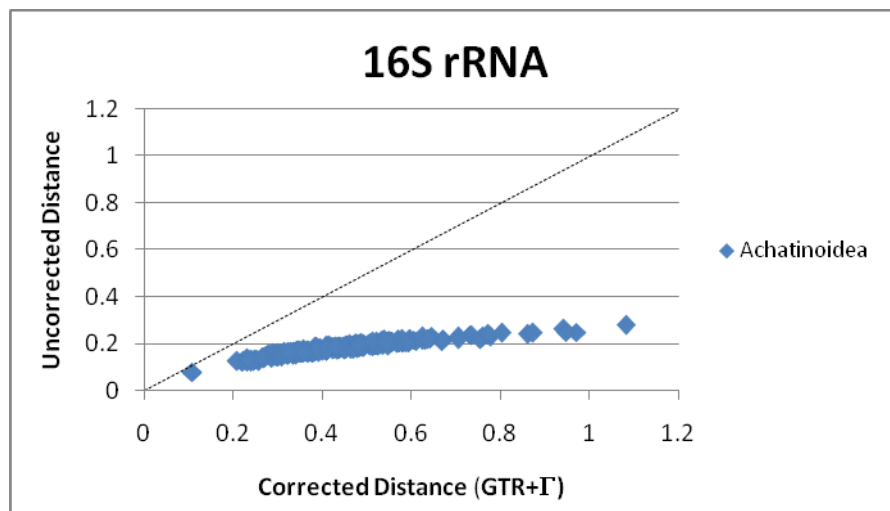
Figure 3.25: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3rd codon position of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

The lack of phylogenetic signal (as indicated by the g1 test) and the strong evidence for saturation of the entire COI gene suggested that the gene should not be used in its entirety for phylogenetic analyses. Further analysis revealed that the saturation is entirely due to the 3rd codon position. Phylogenetic analyses for the COI gene were therefore limited to the 1st and 2nd codon positions only.

For the 16S rRNA gene, Plot 1 (Figure 3.26) produced corrected distances based on the optimal GTR+ Γ model that curved from linearity at an uncorrected (p) distance of 0.08 or below and approached a plateau at an uncorrected distance of approximately 0.3. One corrected distance even exceeded the value of 1. These findings suggest the possibility of saturation in the 16S dataset.



(A)



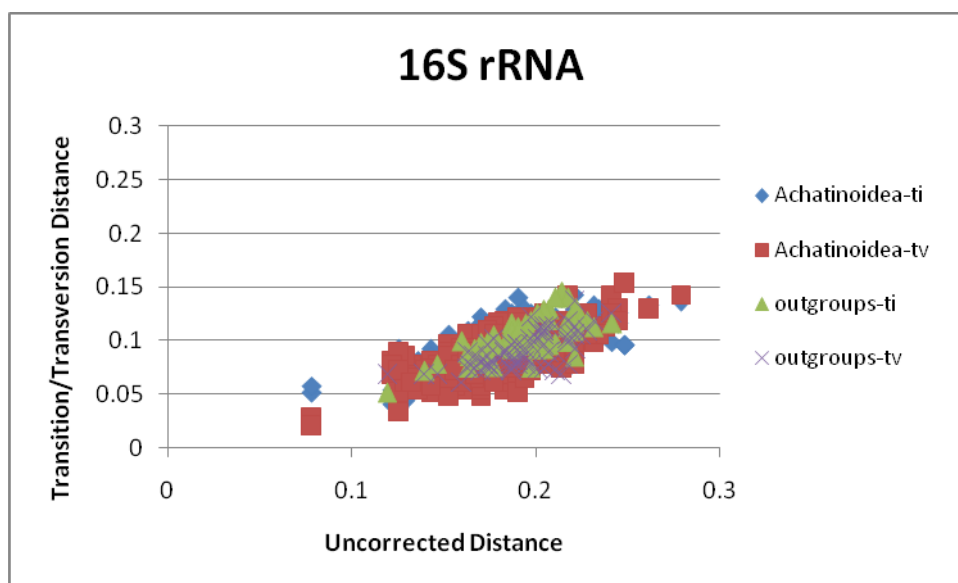
(B)

Figure 3.26: Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the 16S rRNA gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

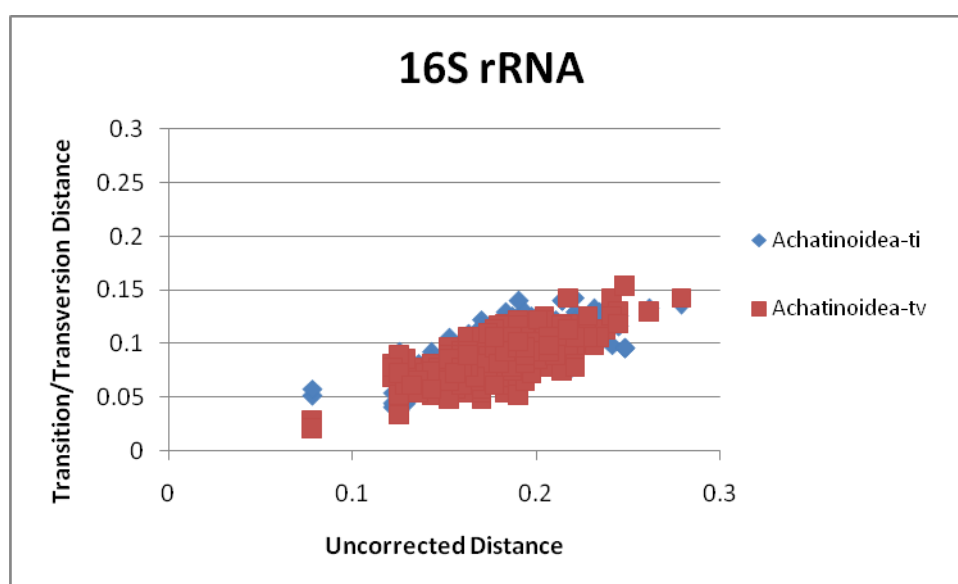
Plot 2 (Figure 3.27) yielded a transition line that began to curve at higher distances where it was overtaken by the transversion line that continued to increase, thus indicative of saturation. An unmistakable overlap was also seen between transitions and transversions. Plot 3 (Figure 3.28) revealed that approximately half of all pairwise comparisons had higher rates of transversions than transitions.

The $g1$ value based on 25 taxa and 120 parsimony-informative sites was -0.437, which was significantly smaller than the critical value of -0.1 at $p=0.05$ for 25 taxa and 100 parsimony-informative sites. Phylogenetic signal was therefore evident in the 16S rRNA gene.

Similarly to the COI gene, there was evidence of saturation in the 16S rRNA dataset. This suggested that the gene should not be used for phylogenetic analyses, despite apparently exhibiting some phylogenetic signal based on the $g1$ test.

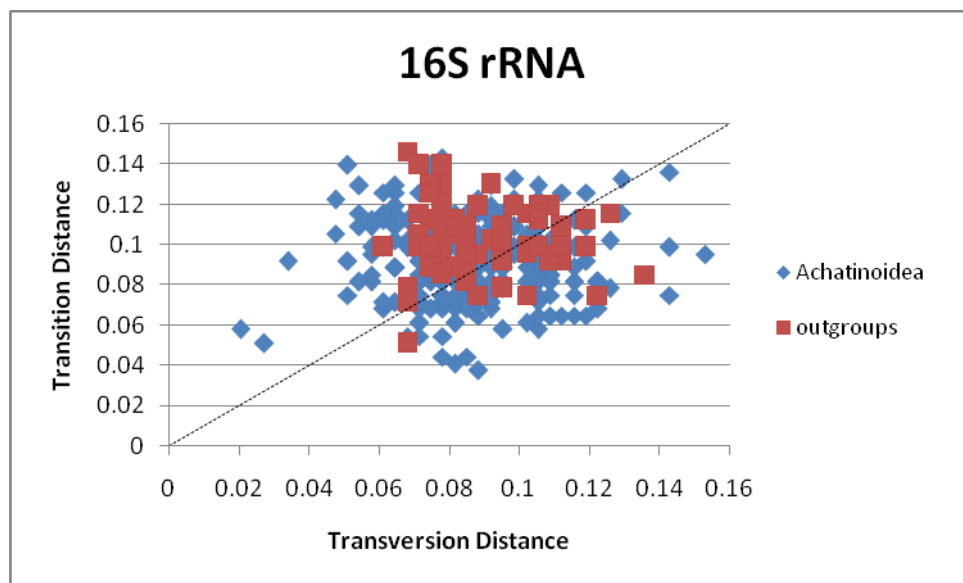


(A)

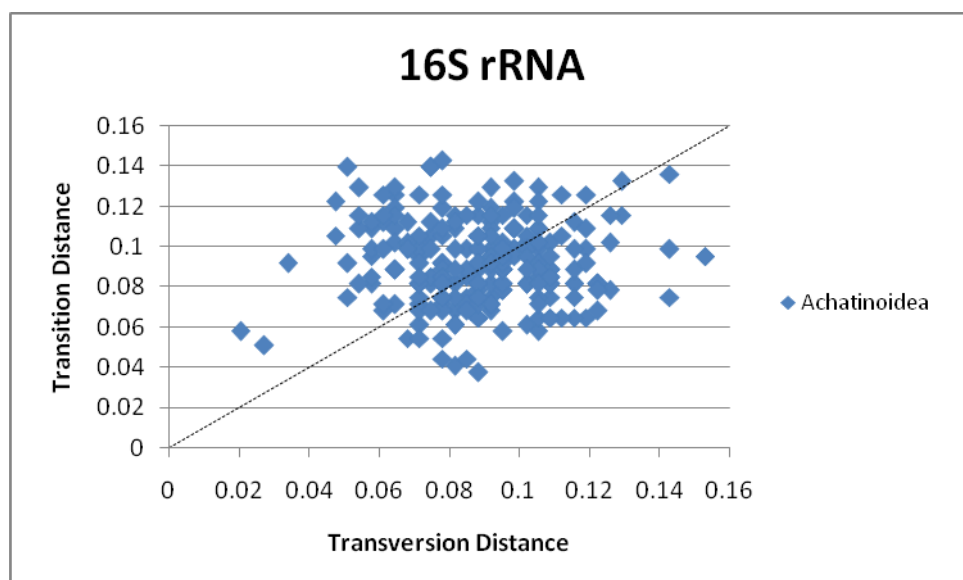


(B)

Figure 3.27: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 16S rRNA gene in **(A)** the Achatinoidea and the streptaxid outgroup taxa and **(B)** the Achatinoidea only.



(A)



(B)

Figure 3.28: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

3.3.2.2. Inferring the presence of paralogous sequences in the actin dataset and assessing its utility for phylogenetic analysis

To infer the presence of paralogous sequences, the actin sequences (obtained by direct sequencing of PCR products) were examined for the presence of ambiguous sites. Ambiguous sites were detected in 17 out of 24 achatinoid taxa and all three streptaxid outgroups (Appendix 3.7, p. 434), indicating the presence of more than one copy of the actin gene in the Achatinoidea and the Streptaxoidea.

Clones derived from three representative taxa, which had varied numbers of ambiguous sites in their direct sequences (*Coelioxys blandii*, 21 ambiguities; *Cochlitoma granulata*, 9 ambiguities; *Leptinaria lamellata*, 0 ambiguities), were examined to assess the ability of the PCR-direct sequences to detect variation among actin copies as ambiguities in the direct sequence. (See details of actin clones in Appendix 3.8, pp. 435-437). The three taxa were selected to represent those with many, few and no ambiguous sites, respectively and were chosen from a batch of actin sequences processed early on, with their cloned sequences used as guides to assign ambiguities in direct sequences. Other actin sequences obtained later yielded considerably more ambiguous sites after subsequent processing, as in the case of *Bocageia* sp. with 101 ambiguous sites. Although the sequences of the clones would be subject to Taq error, a reasonably close correlation would be expected between the amount of variation detected among clones and the amount of ambiguity detected in the direct sequences. However, for *C. blandii*, a total of 163 variable sites were detected among the 4 clones obtained, as opposed to only 21 ambiguities in the direct sequence (Appendix 3.8A, p. 435). Of these variable sites, 132 were attributed to a single clone (Clone 3) that was clearly not picked up in the direct sequence (see Appendix 3.8B-1, p. 436). Likewise for *C. granulata*, 72 variable sites were detected among the 7 clones obtained as

opposed to only 9 ambiguities in the direct sequence (Appendix 3.8A). The majority of these variable sites (55) were attributed to Clones 3 and 6 that were not picked up in the direct sequence (see Appendix 3.8B-2, p. 437). For *L. lamellata*, six clones were obtained, with three clones (Clones 1, 3 and 5) bearing sequences identical to the direct sequence (Appendix 3.8A). Clones 2 and 4 had one variable site each while Clone 6 had two; none of the four variable sites were picked up in the direct sequence (see Appendix 3.8B-3, p. 437). The very small number of variable sites in the *L. lamellata* clones and their absence in the direct sequence would suggest that these variable sites were probably due to Taq sequencing error. The cloned sequences therefore confirmed the presence of multiple actin genes in at least some achatinoid taxa, with some clones exhibiting highly divergent sequences when compared to other clones derived from the same taxon. Moreover, not all of the variation observed in the clones was detected in the PCR-direct sequences. The reason for the failure of the direct sequences to identify all of the variation among the clones is not clear. One explanation may lie in the fact that the PCR-direct sequence is effectively a consensus of the different actin copies within an individual in which rare copies of the gene might reasonably be expected to be averaged out.

To assess the utility of the actin gene for phylogenetic analysis of the Achatinoidea, four approaches were undertaken to determine whether the observed paralogy in actin was likely to mislead phylogeny. First, a neighbor-joining tree was constructed for all actin sequences for the Achatinoidea as well as all clones obtained for *C. blandii* (4 clones), *C. granulata* (7 clones) and *L. lamellata* (6 clones) in order to determine whether the clones for each species were monophyletic. Monophyly would suggest that the gene duplication event that led to the actin paralogs detected was recent relative to the date of species divergence. Moreover, if all copies of the gene were

monophyletic, the organismal phylogeny would not be misled. However, monophyly was not observed for the clones of either *C. blandii* or *C. granulata*, with the most divergent clones (Clone 3 for *C. blandii* and Clones 3 and 6 for *C. granulata*) falling separately from both the PCR direct sequence and the other clones from the species (see Appendix 3.9, p. 438). This suggested a high probability that the organismal phylogeny may be misled. Nevertheless, the most divergent sequences for *C. blandii* and *C. granulata* (that fell separately from the other *C. blandii* and *C. granulata* sequences in the neighbor-joining tree) were not represented as ambiguities in the PCR-direct sequences and would therefore not affect the phylogenetic signal of the direct sequences.

Second, the number of ambiguous sites within a taxon was compared with the total number of nucleotide differences between the taxon and its closest relative (based on actin sequence identity). The number of ambiguous sites within a taxon should not exceed the number of differences between this taxon and its closest relative if the time of divergence between the two taxa were to be earlier than the evolution of the different actin genes within these taxa. With the exception of *Pyrgina umbilicata* and *Thyrophorella thomensis*, a lower number of ambiguous sites was observed within each taxon when compared to the number of nucleotide differences between the taxon and its closest relative. Thus, based on the number of ambiguous sites observed from the direct sequences, in most cases the divergence among taxa was deeper than the divergence among the different copies of the actin gene, suggesting that some phylogenetic signal could be derived from actin at and above the genus level (see Table 3.4). However, the presence of divergent actin gene sequences falling deeper than the divergence among taxa should not be ruled out, as some divergent sequences, which were not picked up as ambiguities in the direct sequences, were detected by cloning; these sequences did not

cluster with the other clones or the direct sequence from that species (i.e. *Coelioxys blandii* and *Cochlitoma granulata*). It should also be noted that since the closest relative was identified based on actin sequence identity, such a relationship was not always found to be associative. For instance, *Achatina achatina* is most similar to *A. fulica* with 65 nucleotide differences, and yet the latter is most similar to *C. ustulata* with only 46 differences.

Third, a partition homogeneity test (see Section 2.9.10, p. 86-88) was undertaken to determine whether or not the sequences from the actin gene exhibited a significant difference in terms of phylogenetic signal compared to the other genes being evaluated (see results in Section 3.3.2.3, p. 161-165). The test revealed that the actin dataset was not too divergent in terms of its evolutionary history relative to the other datasets and that the presence of multiple copies of the actin gene was not having a significant effect on phylogeny.

Lastly, the phylogenetic tree obtained from the actin dataset was checked for concordance with the phylogeny obtained from the rRNA cluster. The actin phylogeny showed concordance with the rRNA phylogeny with respect to many well-supported groups (see results in Section 3.3.3, pp. 162-163 and 169-171), suggesting that the presence of multiple copies of the actin gene was not having a significant effect on phylogeny.

Thus, despite serious reservations over the utility of the actin gene in phylogenetic analysis of the Achatinoidea, it seems that some useful phylogenetic signal could be gleaned from the gene. Actin was therefore utilised in phylogenetic analyses of the Achatinoidea though its shortcomings suggest that its findings should be interpreted with extreme caution.

Table 3.4. Comparison of the actin ambiguous sites and nucleotide differences for each achatinoid and its closest relative. The closest relative for each taxon is determined based on the actin sequence identity (least number of nucleotide differences). Note that in scoring for the total differences, comparison between an ambiguous site and an unambiguous site is counted as different even if one of the possible nucleotides for the ambiguous site is the same as that found in the unambiguous site being compared (e.g. A vs. R, which is either A or G). See also Table 2.8 of p. 73 for assignment of ambiguous positions.

Species (total ambiguous sites)	Closest relative (total ambiguous sites)	Total number of nucleotide differences between each species and its closest relative
ACHATINIDAE		
<i>Achatina achatina</i> (45)	<i>Achatina fulica</i> (18)	65
<i>Achatina fulica</i> (18)	<i>Cochlitoma ustulata</i> (0)	46
<i>Achatina stuhlmanni</i> (2)	<i>Cochlitoma ustulata</i> (0)	28
<i>Cochlitoma ustulata</i> (0)	<i>Achatina stuhlmanni</i> (2)	28
SUBULINIDAE		
<i>Allopeas clavulinum</i> (0)	<i>Eutomoepas layardi</i> (0)	36
<i>Bocageia</i> sp. (101)	<i>Rumina decollata</i> (29)	126
<i>Eutomoepas layardi</i> (0)	<i>Allopeas clavulinum</i> (0)	36
<i>Leptimnaria lamellata</i> (0)	<i>Allopeas clavulinum</i> (0)	104
<i>Paropeas clavulinum</i> (1)	<i>Allopeas clavulinum</i> (0)	46
<i>Riebeckia</i> sp. (76)	<i>Coelioxys blandii</i> (21)	107
<i>Rumina decollata</i> (29)	<i>Zootecus insularis</i> (20)	55
<i>Subulina octona</i> (0)	<i>Subulina striatella</i> (26)	63
<i>Subulina striatella</i> (26)	<i>Subulina octona</i> (0)	63
<i>Subulina vitrea</i> (37)	<i>Coelioxys blandii</i> (21)	87
<i>Subulona</i> sp. (0)	<i>Achatina stuhlmanni</i> (2) <i>Cochlitoma ustulata</i> (0)	117
<i>Tortaxis erectus</i> (32)	<i>Rumina decollata</i> (29) <i>Zootecus insularis</i> (20)	110
<i>Xerocerastus</i> sp. (0)	<i>Zootecus insularis</i> (20)	60
<i>Zootecus insularis</i> (20)	<i>Rumina decollata</i> (29)	55
COELIAXIDAE		
<i>Coelioxys blandii</i> (21)	<i>Zootecus insularis</i> (20)	82
<i>Pyrgina umbilicata</i> (88)	<i>Thyrophorella thomensis</i> (62)	77
THYROPHORELLIDAE		
<i>Thyrophorella thomensis</i> (62)	<i>Pyrgina umbilicata</i> (88)	77
GLESSULIDAE		
<i>Glessula ceylanica</i> (53)	<i>Cochlitoma ustulata</i> (0)	149
FERUSSACIIDAE		
<i>Cecilioides gokweanus</i> (4)	<i>Xerocerastus</i> sp. (0)	81
<i>Ferussacia folliculus</i> (4)	<i>Xerocerastus</i> sp. (0)	99
STREPTAXIDAE (OUTGROUP)		
<i>Gibbulinella dewinteri</i> (6)	<i>Gonaxis quadrilateralis</i> (55)	94
<i>Gonaxis quadrilateralis</i> (55)	<i>Gibbulinella dewinteri</i> (6)	94
<i>Gonospora</i> sp. (77)	<i>Gonaxis quadrilateralis</i> (55)	117

3.3.2.3. Partition homogeneity test for combining datasets

A partition homogeneity test of the rRNA cluster, actin and histone 3 datasets and the 1st and 2nd codon positions of the COI gene (5028 nucleotides total) for all taxa

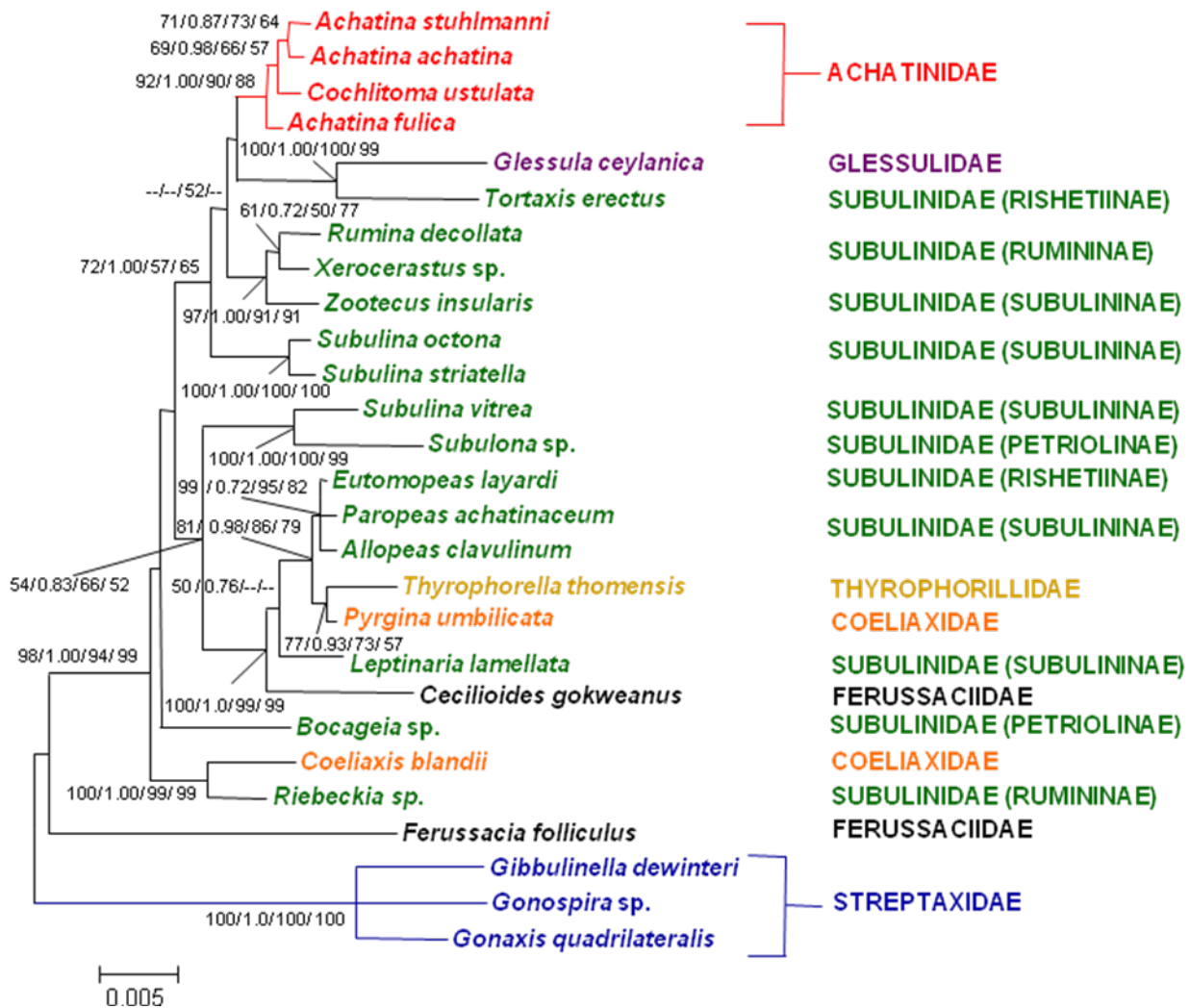
with complete DNA sequences yielded a *P* value of 0.01 (note that the 16S rRNA gene and the 3rd codon position of the COI gene were excluded due to saturation (see Section 3.3.2.1, pp. 138-154). This would indicate that the genes under consideration, including the actin, exhibited similar evolutionary histories (Cunningham, 1997). The datasets from the four genes could therefore be combined as a single dataset.

3.3.3. Molecular phylogeny

The maximum likelihood (ML) phylogenies for the rRNA cluster, actin, H3 and COI (1st and 2nd codon positions only) are shown in Figure 3.29 and those of the concatenated sequences from the four genes in Figure 3.30. Note that for the concatenated sequences, two sets of phylogenetic analyses were carried out. In the first, only taxa with sequences from all four genes were used in the phylogeny; thus, the achatinoids *Celilioides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp. were excluded (Figure 3.30A). In the second, all taxa were included in the analyses (Figure 3.30B), with missing data assigned in the analyses for taxa with incomplete datasets. Also shown in the phylogenies are the bootstrap support values for ML, neighbor-joining (NJ) and maximum parsimony (MP) analyses and the posterior probabilities for Bayesian inference (BI) analysis. The BI, NJ and MP phylogenies for the genes and the concatenated sequences are shown in Appendices 3.10-3.15, pp. 439-450.

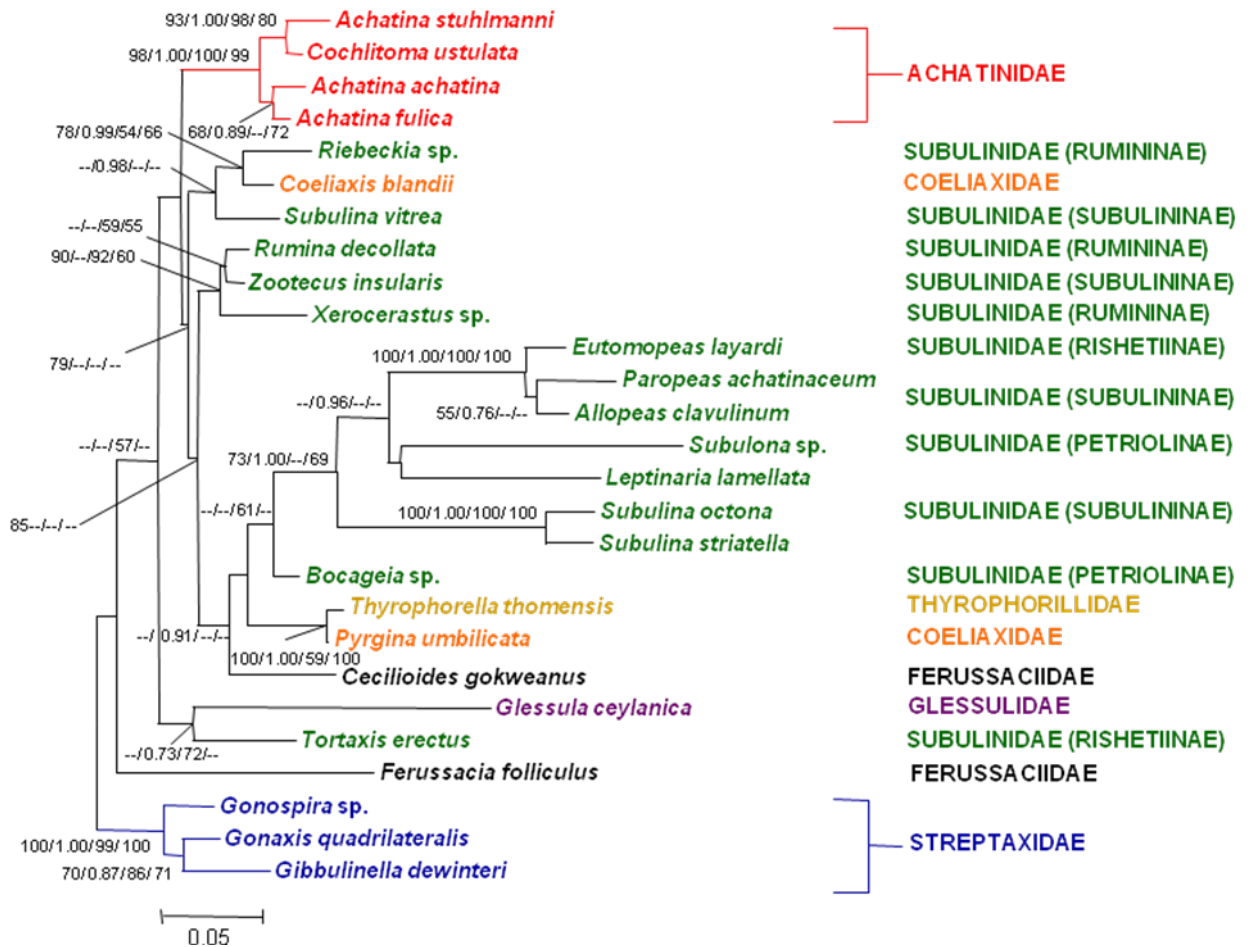
Figure 3.29: (See next four pages). Maximum likelihood phylogenetic trees of the Achatinoidea based on (A) the rRNA cluster (3435 nucleotides); (B) the actin gene (861 nucleotides); (C) the histone 3 gene (328 nucleotides) and (D) the 1st and 2nd codon positions of the COI gene (404 nucleotides). The phylogenies were rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp for the rRNA and actin datasets and *Gibulinella dewinteri* for the histone 3 and COI datasets. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap support (1000 replicates) for NJ and MP, respectively. Bootstrap support less than 50% and posterior probabilities less than 0.7 are not shown. For BI, the optimized number of generations to explore the tree space was 2,000,000 for all the genes; the optimized heating temperatures for each gene were as follows: rRNA cluster-0.1; actin-0.1; histone 3-0.1; 1st and 2nd codon positions of COI-0.075. The scale bar for (A) represents 5 substitutional changes per 1000 nucleotide positions; those of (B), (C) and (D) represent 5 substitutional changes per 100 nucleotide positions.

Figure 3.29 (contd.)



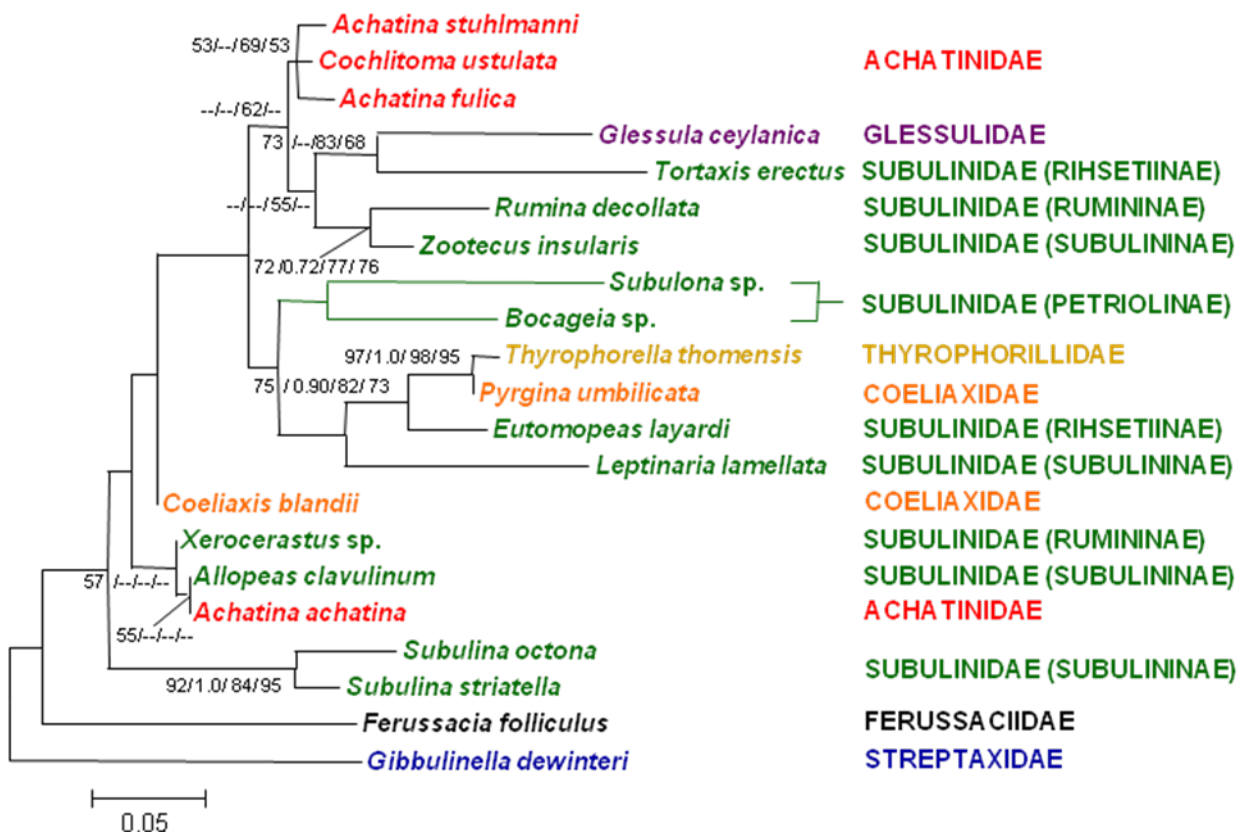
(A) – rRNA cluster

Figure 3.29 (contd.)



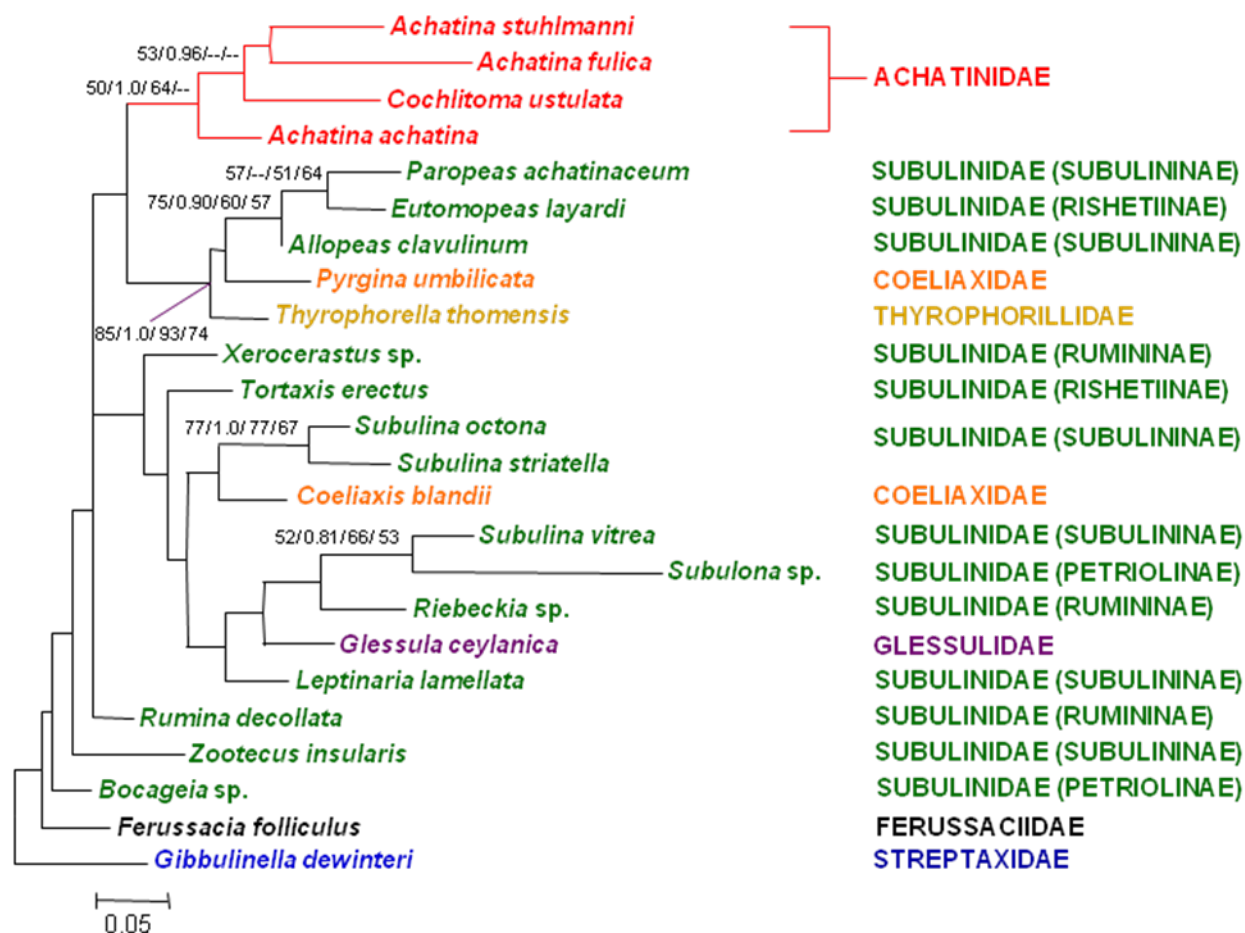
(B) – Actin

Figure 3.29 (contd.)



(C)–H3

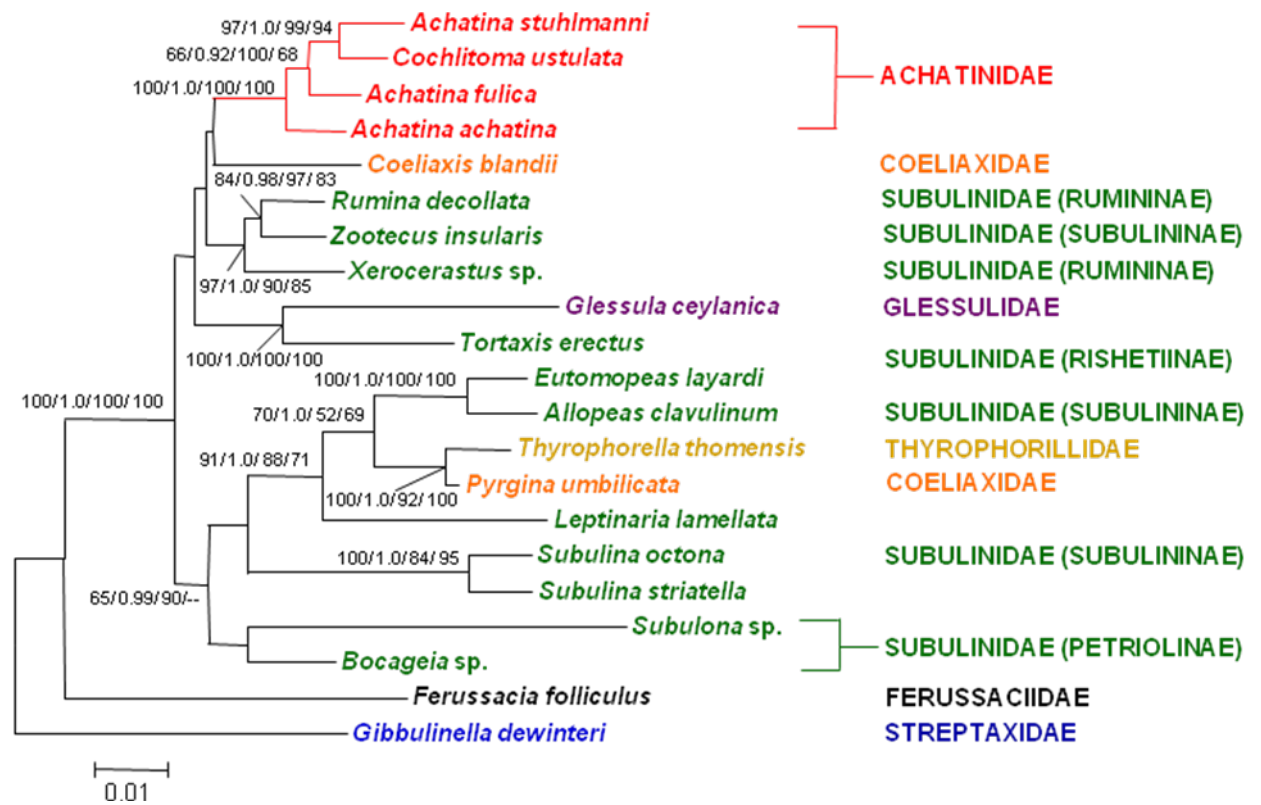
Figure 3.29 (contd.)



(D)– COI-1st and 2nd codon positions

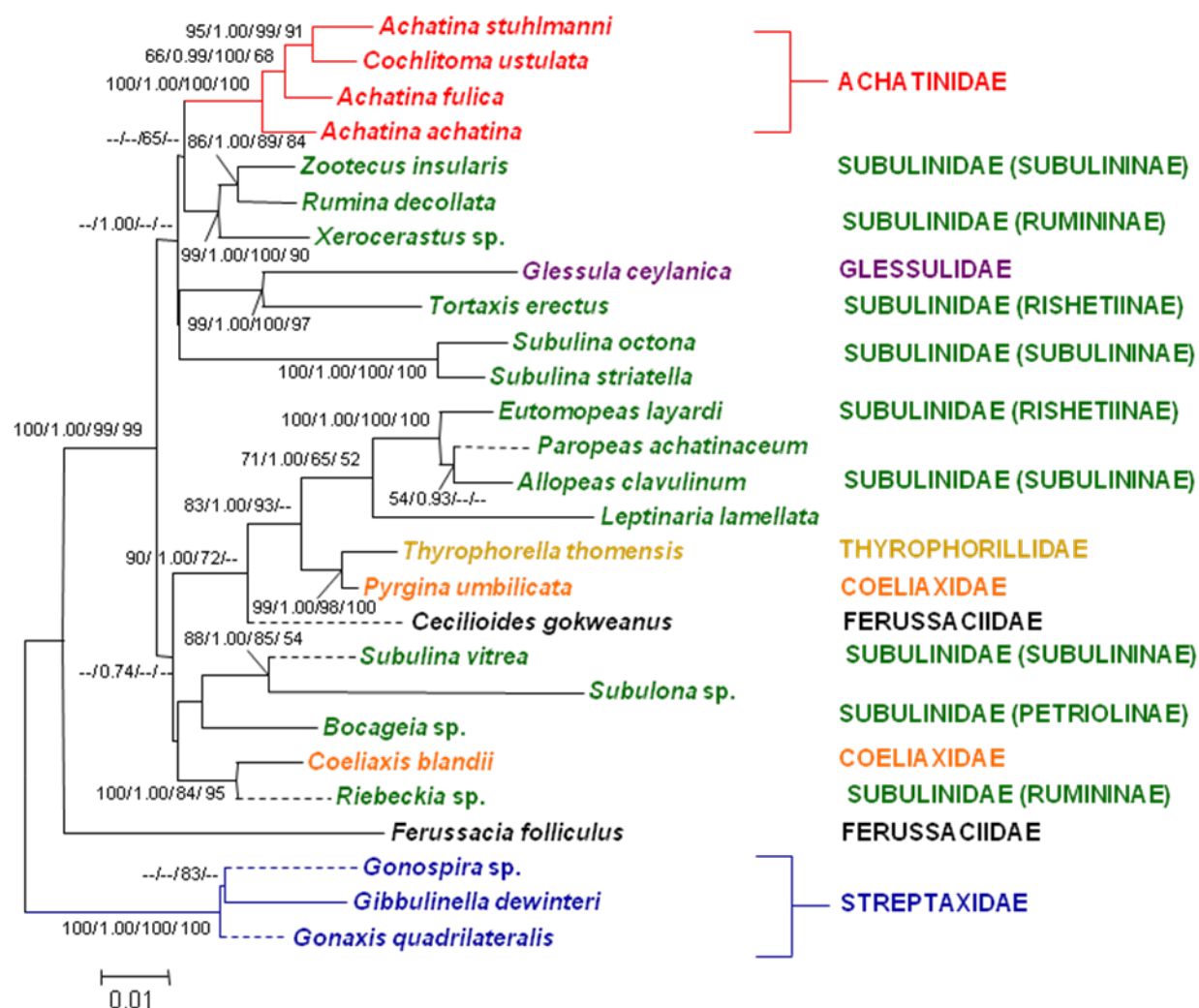
Figure 3.30: (See next two pages.) Maximum likelihood phylogenetic trees of the Achatinoidea based on a concatenated sequence of 5028 nucleotides derived from the combined dataset of the rRNA cluster, actin and H3 genes as well as the 1st and 2nd codon positions of the COI gene. The phylogeny in (A) includes only those taxa which have complete sequence data for all sequence regions while that in (B) includes all taxa including some with missing data. The following taxa had missing DNA sequence data: *Cecilioides gokweanus* (Histone 3 and COI), *Paropeas achatinaceum* (Histone 3), *Riebeckia* sp. (Histone 3), *Subulina vitrea* (Histone 3), *Gonaxis quadrilateralis* (Histone 3 and COI) and *Gonospira* sp. (Histone 3 and COI). The phylogeny in (A) is rooted on the streptaxid *Gibbulinella dewinteri* while that in (B) is rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap support (1000 replicates) for NJ and MP, respectively. Bootstrap support less than 50% and posterior probabilities less than 0.7 are not shown. For BI, the optimized number of generations to explore the tree space was 2,000,000 while the optimized heating temperature was 0.125 for both trees. The scale bars represent 1 substitutional change per 100 nucleotides. Branches for taxa with missing data are marked with dashes.

Figure 3.30 (contd.)



(A) – taxa with complete DNA sequence data

Figure 3.30 (contd.)



(B) - all taxa

3.3.3.1. Individual gene phylogenies

The phylogenies based on the four genes yielded several consistent groups. Even with the incorporation of new taxa and the addition of an extra 2612 nucleotide sites, the topology of the Achatinoidea based on the nuclear rRNA cluster (Figure 3.29A) remained similar to that obtained by Wade *et al.* (2006) based on a partial 823 bp fragment of the rRNA cluster. Several well-supported groups were recovered in the rRNA phylogeny. The Achatinidae, which included *Achatina fulica*, was recovered as monophyletic, with strong support with all tree methods (92% ML bootstraps, $P=1.0$ BI, 90% NJ bootstraps and 88% MP bootstraps). The other achatinoid families were not recovered in the tree as expected, but six groupings with strong support were resolved. The first group incorporated the coeliacid *Coeliaxis blandii* and the subulinid *Riebeckia* sp. (100 ML bootstraps, $P=1.00$, 99% NJ bootstraps and 99% MP bootstraps). The second group included the thyrophorellid *Thyrophorella thomensis*, the coeliacid *Pyrgina umbilicata*, the subulinids *Leptinaria lamellata*, *Eutomopeas layardi*, *Paropeas achatinaceum* and *Allopeas clavulinum* and the ferussaciid *Cecilioides gokweanus* (100% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 99% MP bootstraps). Within this group, the two São Thomé taxa, the ‘operculate’ *T. thomensis* and the coeliacid *P. umbilicata*, clustered together (77% ML bootstraps, $P=0.93$ BI, 73% NJ bootstraps and 57% MP bootstraps) while *E. layardi*, *P. achatinaceum* and *A. clavulinum* formed another cluster (99% ML bootstraps, $P=0.72$, 95% NJ bootstraps and 82% MP bootstraps). The third group included the subulinids *Subulina vitrea* and *Subulona* sp. (100% ML bootstraps, $P=1.0$, 100% NJ bootstraps and 99% MP bootstraps). The fourth group consisted of another pair of subulinids, *Subulina octona* and *Subulina striatella* (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). The fifth group incorporated the subulinids *Xerocerastus* sp., *Rumina decollata* and

Zootecus insularis (97% ML bootstraps, $P=1.0$ BI, 91% NJ bootstraps and 91% MP bootstraps). The sixth group clustered together the subulinid *Tortaxis erectus* and the glessulid *Glessula ceylanica* (100 ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 99% MP bootstraps). Finally, the ferussacid *Ferussaccia folliculus* fell at the base of the Achatinoidea and was separated from the other ferussacid in the study, *Cecilioides gokweanus*.

Despite the limitations of the nuclear actin gene for phylogenetic analysis due to the presence of multiple copies of the gene within an individual, the actin phylogeny was very similar to the rRNA tree, with many strongly supported groups in the actin tree also recovered in the rRNA tree. This suggests that the presence of multiple copies of the actin gene had a minimal effect on the achatinoid phylogeny. There was, however, less structure in the actin tree when compared to the rRNA tree as shown by the lower or equivocal support in many of the internal branches (Figure 3.29B). *Ferussaccia folliculus* again fell at the base of the tree; however, this was only moderately supported by NJ (57% bootstraps). Other groups that were consistent with the rRNA cluster phylogeny included the Achatinidae, which again formed a monophyletic group (98% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 99% MP bootstraps); the *C. blandii* and *Riebeckia* sp. group (78% ML bootstraps, $P=0.99$ BI, 54% NJ bootstraps and 66% bootstraps); the São Thomé taxa *T. thomensis* and *P. umbilicata* (100 ML bootstraps, $P=1.0$ BI, 59% NJ bootstraps and 100% MP bootstraps); the *E. layardi*, *P. achatinaceum* and *A. clavulinum* group (100 ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps); the *S. octona* and *S. striatella* group (100 ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps); the *Xerocerastus* sp., *R. decollata* and *Z. insularis* group (90% ML

bootstraps, $P < 0.7$ BI, 92% NJ bootstraps and 60% MP bootstraps); and the *T. erectus* and *G. ceylanica* group (with only $P = 0.73$ BI and 72% NJ bootstraps as support).

The phylogeny based on the nuclear histone 3 gene (Figure 3.29C), which excluded the achatinoids *Cecilioides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilaterlis* and *Gonospira* sp., revealed some groupings that were consistent with the rRNA phylogeny but showed less structure and a general reduction of support for most groups when compared to either the rRNA or the actin gene phylogenies (as shown in Figure 3.30C). For instance, *Achatina achatina* did not cluster with the rest of the Achatinidae and instead grouped with the subulinids *Xerocerastus* sp. and *A. clavulinum*, although it only received 57% ML bootstrap support. Some groups were recovered, however, such as *T. thomensis* and *P. umbilicata* (97% ML bootstraps, $P = 1.0$ BI, 98% NJ bootstraps and 95% MP bootstraps), *S. octona* and *S. striatella* (92% ML bootstraps, $P = 1.0$ BI, 84% NJ bootstraps and 95% MP bootstraps), and *T. erectus* and *G. ceylanica* (73% ML bootstraps, 83% NJ bootstraps and 68% MP bootstraps).

The mitochondrial COI gene phylogeny for codon positions 1 and 2 (Figure 3.29D) likewise revealed some groups that were consistent with the rRNA phylogeny but showed less structure and support (see Figure 3.30D). These were the Achatinidae (50% ML bootstraps, $P = 1.0$ BI and 64% NJ bootstraps), *S. vitrea* and *Subulona* sp. (52% ML bootstraps, $P = 0.81$ BI, 66% NJ bootstraps and 53% MP bootstraps), *S. octona* and *S. striatella* (77% ML bootstraps, $P = 1.0$ BI, 77% NJ bootstraps and 67% MP bootstraps), and *E. layardi*, *P. achatinaceum* and *A. clavulinum* (75% ML bootstraps, $P = 0.90$ BI, 60% NJ bootstraps and 57% MP bootstraps). It should be noted that *T. thomensis* and *P. umbilicata* also clustered with these three taxa with strong support

(85% ML bootstraps, $P=1.0$ BI, 93% NJ bootstraps and 74% MP bootstraps) as with the rRNA phylogeny.

3.3.3.2. Combined gene phylogenies

Combining datasets from genes with different evolutionary rates and mechanisms of inheritance provides information at different levels of phylogeny and therefore provides signals both for the internal branches and the terminal branches (Graybeal, 1994). The combined analysis for the rRNA cluster, actin, H3, and the 1st and 2nd codon positions of COI, but excluding the four taxa that did not yield either or both H3 and COI sequences, recovered most groups identified in the rRNA phylogeny and exhibited longer internal branches with similar or better support (Figure 3.30A). *Ferussacia folliculus* still fell at the base of the Achatinoidea while the other achatinoids clustered together but with improved support when compared to the rRNA tree (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). The monophyly of the Achatinidae was also recovered again with better support (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). Consistent with the rRNA tree was the group that consisted of *L. lamellata*, *T. thomensis*, *P. umbilicata*, *E. layardi* and *A. clavulinum* (91% ML bootstraps, $P=1.0$ BI, 88% NJ bootstraps and 71% MP bootstraps), with the São Thomé taxa *T. thomensis* and *P. umbilicata* again forming one pair and with increased support (100% ML bootstraps, $P=1.0$ BI, 92% NJ bootstraps and 100% MP bootstraps) and *E. layardi* and *A. clavulinum* forming a second pair likewise with increased support (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). Still other groups from the rRNA tree that were also recovered included *S. octona* and *S. striatella* (100% ML bootstraps, $P=1.0$ BI, 84% NJ bootstraps and 95% MP bootstraps), *Xerocerastus* sp., *R. decollata* and *Z.*

insularis (97% ML bootstraps, $P=1.0$ BI, 90% NJ bootstraps and 85% MP bootstraps) and *T. erectus* and *G. ceylanica* (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps).

When all taxa were included in the combined analysis (including those taxa for which there was missing data), all groups found in the rRNA phylogeny were recovered and in general received higher support (Figure 3.30B). *Ferussacia folliculus* fell at the base of the Achatinoidea while the remaining Achatinoidea formed a separate clade with increased support (100% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 99% MP bootstraps). Within this clade and consistent with the rRNA tree were the monophyletic Achatinidae with increased support (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps); *C. blandii* and *Riebeckia* sp. (100% ML bootstraps, $P=1.0$ BI, 84% NJ bootstraps and 95% MP bootstraps); *Subulina vitrea* and *Subulona* sp. (88% ML bootstraps, $P=1.0$ BI, 85% NJ bootstraps and 54% MP bootstraps); *S. octona* and *S. striatella* (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps); *G. ceylanica* and *T. erectus* (99% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 97% MP bootstraps); *Xeroxerastus* sp., *Rumina decollata* and *Zootecus insularis* with increased support (99% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 90% MP bootstraps); and *C. gokweanus*, *P. umbilicata*, *T. thomensis*, *L. lamellata*, *A. clavulinum*, *P. achatinaceum*, and *E. layardi* (90% ML: bootstraps, $P=1.0$ BI and 72% NJ bootstraps). Within the last group, *A. clavulinum*, *P. achatinaceum* and *E. layardi* clustered together with increased support (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps), and in turn clustered with *L. lamellata* (71% ML bootstraps, $P=1.0$ BI, 65% NJ bootstraps and 52% MP bootstraps) while the São Thomé group *T. thomensis* and *P. umbilicata* clustered

together with increased support (99% ML bootstraps, $P=1.0$ BI, 98% NJ bootstraps and 100% MP bootstraps).

3.3.3.3. Hypothesis testing of groups expected to be monophyletic based on taxonomy

Seven expected monophyletic groups (based on taxonomy) that were not recovered in either the single gene or combined gene phylogenies were subjected to hypothesis testing using the Shimodaira-Hasegawa (1999) test. These groups were the Coeliacidae, Ferussaciidae, and the Subulinidae; within the Subulinidae were the subfamilies Petriolinae, Rishetiinae, Rumininae and the Subulininae. The placement of *Zootecus insularis* in the Subulininae (according to Schileyko, 1999), in contrast to its placement in the Rumininae (according to Zilch, 1959), was also subjected to the SH test. A summary of the Shimodaira-Hasegawa tests are shown in Table 3.5. Constraining the NJ trees for the four single gene and combined gene phylogenies to show monophyly of the Coeliacidae produced likelihood scores that were significantly worse ($P<0.01$) than those of the optimal trees (see Table 3.5 for details). This confirmed the non-monophyly of the Coeliacidae. The monophyly of the Ferussaciidae was evaluated only for the rRNA cluster and actin gene phylogenies, for which both *Ferussacia folliculus* and *Cecilioides gokweanus* were successfully sequenced, as well as the combined gene phylogeny that included taxa with missing data. The constrained trees showing monophyly of the Ferussaciidae for the rRNA cluster and the combined genes that included all taxa were significantly worse than the optimal tree ($P<0.01$, Table 3.5), indicating that the family is not monophyletic. This was not the case for the actin gene where no significant difference was noted ($P=0.293$, Table 3.5). The monophyly of the Subulinidae was also rejected in the SH tests ($P<0.01$, Table 3.5) as

was the monophyly of two of its subfamilies, the Rishetiinae ($P < 0.03$, Table 3.5) and the Subulininae (with *Z. insularis*- $P < 0.01$; without *Z. insularis*- $P < 0.04$, Table 3.5). For the Petriolinae, the expected topology (Petriolinae monophyletic) was present in the actin NJ tree (see Appendix 3.10B, p. 440) while no significant difference was shown between the likelihood scores for the constrained trees in which the Petriolinae were monophyletic and the optimal NJ trees in the rRNA cluster ($P = 0.093$), histone 3 ($P = 0.184$), 1st and 2nd codon positions of COI ($P = 0.083$), combined genes but excluding taxa with missing data ($P = 0.433$) and combined genes with all taxa ($P = 0.150$). Support for the monophyly of the Petriolinae based on the molecular data is therefore equivocal. The constrained trees showing monophyly of the Rumininae yielded significantly worse likelihood scores than those of the optimal trees ($P \leq 0.001$ for both with and without *Z. insularis* for the rRNA cluster, actin and combined genes with all taxa, Table 3.5). *Riebeckia* sp. was not included in the datasets for histone 3, COI (1st and 2nd codon positions) and the combined genes that excluded taxa with missing data. Significant difference between the optimal tree and the constrained tree was observed in histone 3 ($P \leq 0.01$ for both with and without *Z. insularis*) but not in the 1st and 2nd codon positions of the COI ($P \leq 0.3$ for both with and without *Z. insularis*) and the combined genes but excluding taxa with missing data ($P = 0.370$ for without *Z. insularis* while expected NJ topology was generated when *Z. insularis* was included) (see Table 3.5). On the basis of the taxa examined, neither Shileyko's (1999) nor Zilch's (1959) Rumininae is monophyletic.

Table 3.5: Hypothesis testing for the monophyly of selected taxa of the Achatinoidea using the Shimodaira-Hasegawa test.

Gene	Likelihood Score		<i>P</i> for SH ^a Test (Conclusion ^b)
	Optimal NJ tree	Constrained NJ tree	
COELIAXIDAE			
rRNA cluster	7951.71731	8036.16984	<0.001 (Significant)
Actin	5881.25298	5972.47679	<0.001 (Significant)
Histone 3	1815.79561	1864.59737	<0.001 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1924.07354	0.006 (Significant)
Combined	16679.19409	16826.68130	<0.001 (Significant)
Combined-all taxa	18474.64048	18683.79407	<0.001 (Significant)
FERUSSACIIDAE			
rRNA cluster	7951.71731	7982.47034	0.010 (Significant)
Actin	5881.25298	5877.42426	0.293 (Not significant)
Histone 3	Not applicable; <i>Cecilioides gokweanus</i> not included		
COI (1 st and 2 nd codons)	Not applicable; <i>Cecilioides gokweanus</i> not included		
Combined	Not applicable; <i>Cecilioides gokweanus</i> not included		
Combined-all taxa	18474.64048	18518.72376	0.010 (Significant)
SUBULINIDAE			
rRNA cluster	7951.71731	8098.27402	<0.001 (Significant)
Actin	5881.25298	5922.88073	0.001 (Significant)
Histone 3	1815.79561	1874.36463	0.008 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1933.58362	0.005 (Significant)
Combined	16679.19409	16974.41302	0.005 (Significant)
Combined-all taxa	18474.64048	18638.18916	<0.001 (Significant)
PETRIOLINAE (SUBULINIDAE)			
rRNA cluster	7951.71731	7967.95035	0.093 (Not significant)
Actin	Not applicable; expected topology exists in optimal NJ tree		
Histone 3	1815.79561	1813.07222	0.184 (Not significant)
COI (1 st and 2 nd codons)	1885.86879	1911.13362	0.083 (Not significant)
Combined	16679.19409	16680.26472	0.433 (Not significant)
Combined-all taxa	18474.64048	18486.19798	0.150 (Not significant)
RISHETIINAE (SUBULINIDAE)			
rRNA cluster	7951.71731	8084.95708	<0.001 (Significant)
Actin	5881.25298	6059.01720	<0.001 (Significant)
Histone 3	1815.79561	1838.11663	0.026 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1924.68912	<0.001 (Significant)
Combined	16679.19409	16914.16222	<0.001 (Significant)
Combined-all taxa	18474.64048	18758.90087	<0.001 (Significant)
RUMININAE sensu Schileyko-without <i>Zootecus insularis</i> (SUBULINIDAE)			
rRNA cluster	7951.71731	8019.95480	<0.001 (Significant)
Actin	5881.25298	5911.57548	0.001 (Significant)
Histone 3	1815.79561	1837.83013	0.01 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1896.97810	0.226 (Not significant)
Combined	16679.19409	16681.37066	0.370 (Not significant)
Combined-all taxa	18474.64048	18578.49191	<0.001 (Significant)
RUMININAE sensu Zilch-with <i>Zootecus insularis</i> (SUBULINIDAE)			
rRNA cluster	7951.71731	8000.77016	0.001 (Significant)
Actin	5881.25298	6042.92852	<0.001 (Significant)
Histone 3	1815.79561	1835.88838	0.009 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1894.94276	0.244 (Not significant)
Combined	Not applicable; expected topology exists in optimal NJ tree		
Combined-all taxa	18474.64048	18542.21668	<0.001 (Significant)

SUBULININAE sensu Schileyko-with <i>Zootecus insularis</i> (SUBULINIDAE)			
rRNA cluster	7951.71731	8106.81200	<0.001 (Significant)
Actin	5881.25298	6067.38758	<0.001 (Significant)
Histone 3	1815.79561	1865.98883	0.002 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1940.19417	<0.003 (Significant)
Combined	16679.19409	16965.35856	<0.001 (Significant)
Combined-all taxa	18474.64048	18805.02183	<0.001 (Significant)
SUBULININAE sensu Zilch-without <i>Zootecus insularis</i> (SUBULINIDAE)			
rRNA cluster	7951.71731	8051.96505	<0.001 (Significant)
Actin	5881.25298	5898.27655	<0.028 (Significant)
Histone 3	1815.79561	1841.74798	0.034 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1933.96199	<0.001 (Significant)
Combined	16679.19409	16825.89704	<0.001 (Significant)
Combined-all taxa	18474.64048	18706.81385	<0.001 (Significant)

^a SH test – Shimodaira-Hasegawa Test

^b Significant at $P < 0.05$

3.4. Discussion

This phylogenetic study provides the first systematic coverage of the Achatinoidea with over 24 species from 20 genera and 6 families (Achatinidae, Coeliacidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophorellidae), including 4 subfamilies from the Subulinidae (Petriolinae, Rishetiinae, Rumininae and Subulininae).

3.4.1. Single versus combined gene phylogenies

Three nuclear genes and two mitochondrial genes were used for this study. Among these genes, the nuclear rRNA showed the least saturation, and its phylogeny provided the most resolved structure. Many of the basal nodes were well supported, most notably the splitting of the Achatinoidea into *Ferussacia folliculus* and the rest of the achatinoids. Many terminal nodes likewise received support. Expanding the coverage of the rRNA cluster to include nearly the entire LSU when compared to Wade *et al.*'s (2001, 2006) studies led to an increase in support for the Achatinoidea (99% NJ bootstraps and $P=1.0$ BI for this study versus 93% NJ bootstraps and $P=0.98$ BI in Wade *et al.*'s (2006) study). Support for the Achatinidae increased significantly from

38% NJ bootstraps and $P=0.94$ BI to 90% NJ bootstraps and $P=1.0$ BI, although a different set of achatinid taxa (apart from *A. fulica*) were used when compared to the Wade *et al.* (2006) study. There was also a slight increase in support for the *Coelioxys blandii* and *Riebeckia* sp. grouping (99% NJ bootstraps and $P=1.0$ BI for this study versus 92% NJ bootstraps and $P=1.0$ BI in Wade *et al.*'s (2006) study). However, a rather surprising decrease in support was noted for the *Pyrgina umbilicata* and *Thyrophorella thomensis* group (73% NJ bootstraps and $P=0.93$ BI for this study versus 100% NJ bootstraps and $P=1.0$ BI in Wade *et al.*'s (2006) study). The actin gene also recovered many strongly-supported groupings found in the rRNA cluster despite the presence of several copies of the actin gene within many taxa leading to ambiguous sites in the sequences. While many basal groups received less than 50% bootstrap support (ML, NJ and MP) and less than 0.7 posterior probabilities (BI), the splitting of the Achatinoidea into *F. thomensis* and the other achatinoids was still strongly supported with 100% bootstrap support (ML, NJ, MP) and a posterior probability of 1.0 (BI). Two terminal groups also received higher support in actin than in the rRNA cluster. For instance, the Achatinidae obtained 98% NJ bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 99% MP bootstraps. The second group, comprising *Pyrgina umbilicata* and *Thyrophorella thomensis*, received 100% ML bootstraps, $P=1.0$ BI, 59% NJ bootstraps and 100% MP bootstraps; however, *Pyrgina* exhibited more ambiguous sites (88) than the total number of nucleotide differences (77) between it and *Thyrophorella thomensis*. It is highly probable that the actin copies in *P. umbilicata* and *T. thomensis* diverged earlier than the two taxa, and interpretation of this grouping should be treated with caution. On the other hand, the histone 3 and COI (1st and 2nd codon position) gene phylogenies generally had less structure with little support for basal nodes in the tree. This could be attributed to the relatively smaller number of sites used, particularly in the

COI where the 3rd codon positions were excluded, as well as the possibility that both datasets were approaching saturation. In the COI, for example, the high levels of saturation of the 3rd codon position would imply the fast rate of evolution for this gene. Nevertheless, several groups in common with the rRNA and actin phylogenies were also recovered from the histone 3 and COI data. The 16S rRNA gene was excluded because it was found to be saturated and therefore not suitable for phylogenetic analyses at the level of the Achatinoidea.

The groups recovered in the rRNA tree also emerged in the phylogenies based on the concatenated sequence after the partition homogeneity test revealed that the individual datasets could be combined. The resolution within the internal branches and the statistical support for groupings in the tree clearly improved when the datasets were combined and analyzed under a single model (GTR+ Γ) for both the phylogeny that excluded taxa with missing datasets and for the phylogeny with all taxa. The combined phylogenies were dominated by the signal from the rRNA cluster as all groups from the rRNA phylogeny that did not appear in the other single gene phylogenies were recovered in the combined phylogenies.

3.4.2. Phylogenetic relationships within the Achatinoidea

3.4.2.1. The monophyly and phylogenetic position of the Achatinidae

The limited natural range of the Achatinidae to sub-Saharan Africa (Tillier, 1989, Raut & Barker, 2002) and the existence of fossils no earlier than the Pleistocene (0.01-1.8 MYA) (Zilch, 1959-1960; Solem, 1979) suggest that the achatinids are a relatively recent group (Tillier, 1989), although Raut & Barker (2002) believed them to be much older than the fossil record. The first molecular evidence for the monophyly of

the Achatinidae was provided by Wade *et al.* (2001) based on the analysis of a partial fragment of the rRNA cluster. Further molecular evidence for achatinid monophyly is provided in this study based on the analysis of an expanded rRNA dataset as well as the analysis of data from three other genes; actin, histone 3 and the 1st and 2nd codon positions of COI. High support for the monophyly of the family was also obtained when the genes were combined. The relative position of the Achatinidae within the Achatinoidea was not resolved.

3.4.2.2. The monophyly and phylogenetic position of the Ferussaciidae

This study included two representatives from the Ferussaciidae, *Ferussacia folliculus* and *Cecilioides gokweanus*, although the H3 and COI gene fragments failed to amplify in the latter. In the rRNA phylogeny, the two taxa did not cluster together as expected, and the early divergence of *F. folliculus* from the rest of the Achatinoidea, including *C. gokweanus*, was strongly supported (98% ML bootstraps, $P=1.00$ BI, 94% NJ bootstraps and 99% MP bootstraps). Furthermore, the Shimodaira-Hasegawa test conclusively rejected the monophyly of the Ferussaciidae. The actin gene also did not recover the Ferussaciidae as a monophyletic unit, though both bootstrap support/Bayesian posterior probabilities and the Shimodaira-Hasegawa test were equivocal. *Cecilioides gokweanus* has only been described conchologically; its internal anatomy has not been described, although that of *Cecilioides acicula* is already available. *Cecilioides acicula*, like *Ferussacia folliculus*, exhibits characteristics unique to Ferussaciidae such as the short but transversely elongated kidney (Watson, 1928; Tillier, 1989). However, the kidney of *C. acicula* is broader and curves gradually forward as it approaches the rectum whereas the kidney of *F. folliculus* bents abruptly where it meets the rectum. Furthermore, *C. acicula* has a simpler penis structure than *F.*

folliculus, as the former possesses an indistinct penis papilla (Watson, 1928). There is therefore no compelling evidence from morphological data to show that the Ferussaciidae are monophyletic, and the molecular data from this study confirm their polyphyly.

3.4.2.3. Evaluation of the taxonomic designation of the Subulinidae, Coeliacidae, Glessulidae and Thyrophorellidae

The single and combined phylogenies clearly showed that the Subulinidae (represented by eleven taxa in this study), are a polyphyletic group, with members of the Coeliacidae (two representatives), Glessulidae (one representative) and Thyrophorellidae (one representative) falling among the subulinids. Subulinid monophyly was also significantly rejected by the SH test. The phylogenies also revealed that the Coeliacidae, which are regarded as a subfamily, Coeliacinae, under the Subulinidae by Schileyko (1999), are likewise not monophyletic, as *Coeliaxis blandii* and *Pyrgina umbilicata* did not cluster together. Coeliacidae monophyly was also significantly rejected by the SH test.

The polyphyly of the subulinids is not surprising considering several taxa have not been described anatomically and are grouped together based only on their shell morphology and geographic distribution (Schileyko, 1999). However, some fascinating groupings have emerged. For instance, *Xerocerastus* sp. from southwest Africa (Schileyko, 1999), *Rumina decollata* from the Mediterranean (Schileyko, 1999) and *Zootecus insularis* from the Sahara, Arabia, India and Burma (Schileyko, 1999) formed one group despite having distinct geographic distributions. Most descriptions of *Xerocerastus* species are limited to the shell (Van Bruggen, 1970; Schileyko, 1999), but both *R. decollata* and *Z. insularis* have a very short to nearly absent oviduct (Schileyko,

1999). *Xerocerastus* should be evaluated anatomically to determine if it shares any morphological features with *R. decollata* and *Z. insularis*.

Two of the three *Subulina* species clustered together: *S. octona* from central and tropical South America (Schileyko, 1999) and *S. striatella* from tropical Africa (Jurickova, 2006). The third species, *S. vitrea*, which has a geographic distribution limited to within southwest Africa (Van Bruggen, 1970), clustered with the tropical African *Subulona* sp. The internal structures of *S. striatella* and *S. vitrea* have not been described, but *S. octona* and the type species of *Subulona*, *S. badia* (Martens), are distinct morphologically in that the former has a penis sheath while the latter does not (Schileyko, 1999). The reproductive tracts of *S. striatella* and *S. vitrea* are therefore worth investigating to determine if they are also distinct.

The subulinid *Allopeas clavulinum* (Subulininae), which has a New World distribution, clustered with another subulinid, *Eutomopeas layardi* (Rishetiinae), which is restricted to Sri Lanka and neighboring islands (Schileyko, 1999). Like most subulinid genera, which are classified according to their shell characteristics, both taxa can be distinguished from one another conchologically based on the shape of the apex, color pattern and the presence or absence of the umbilicus (Schileyko, 1999). However, *Allopeas* and *Eutomopeas* also share other shell characteristics, including shape, size and translucence (Schileyko, 1999). Cain (1977) observed that distinct taxa, such as the high spired *Cochlicella acuta* and *Cionella* (= *Cochlicopa*) *lubrica*, which live in separate but very similar environments and possibly exhibit similar feeding habits, may manifest strikingly similar shell morphology as a result of convergent evolution. Mead (1994) found the same observations for two species of the Achatinidae. This could also be the case for *A. clavulinum* and *E. layardi*. Only the internal anatomy of *A. clavulinum* has been described, so it is therefore worthwhile to note if the internal

structures of these taxa are also congruent with molecular data. If that is the case, then the shell characteristics used to distinguish the two taxa into separate subfamilies and even genera may not be valid.

The coeliacid *Coeliaxis blandii* from South Africa (Schileyko, 1999) grouped with the subulinid *Riebeckia* sp. found in Sokotra near the tip of Somalia (Schileyko, 1999). Only *C. blandii* has been described morphologically. Comparison of the two taxa based on internal structures is therefore not possible.

The other coeliacid, *Pyrgina umbilicata*, forms a group with the thyrophorellid *T. thomensis* that is consistent in all the single gene and combined gene phylogenies. This grouping is highly controversial in that, in contrast to *A. clavulinum* and *E. layardi*, these two taxa are very distinct from each other conchologically. Whereas *P. umbilicata* is dextral and turricate with 13 narrow whorls (Schileyko, 1999), *T. thomensis* is sinistral and low conic above and bulging below (Schileyko, 2001). Furthermore, the thyrophorellid bears a unique operculum-like structure (Schileyko, 2001). It is speculated that differences in shell morphology often reflect the differences of the feeding behaviour of the taxa being compared (Cain, 1977) and not necessarily their phylogenetic relationship. The only similarity exhibited by *P. umbilicata* and *T. thomensis* at present is their geographic distribution; both are restricted to São Thomé (Schileyko, 1999, 2001). It is not uncommon to have distinct morphological variation that arises between closely related taxa within a small geographic area as a result of adaptive radiation. A classic example of this is Darwin's finches in the Galapagos Islands, where different environmental conditions could have given rise to different feeding niches for these birds that could have led to the evolution of different beak forms (Whittaker & Fernandez-Palacios, 2007). Further examination of the internal

structures of *P. umbilicata* is imperative so that it can be compared with *T. thomensis*, which is already anatomically described (Schileyko, 2001).

Tortaxis erectus from Indochina and southern China (Schileyko, 1999) and the glessulid *Glessula ceylanica* from the Indian subcontinent and Sri Lanka (Schileyko, 1999) formed another group. Conchologically, the two species are somewhat similar in having glossy shells, although *T. erectus* has a turreted, cylindrical-turreted or subulate-turreted shell with 6-13 convex whorls while *Glessula* has an elongated-ovate to ovate-conic shell with 5-8 convex whorls (Schileyko, 1999). The anatomy of *T. erectus* is likewise unknown, so comparison of the two taxa based on internal structures is not possible. In addition to *G. ceylanica*, there are various other *Glessula* species that need to be characterized at the molecular level as they are very similar to each other conchologically (Beddome, 1906).

Based, therefore, on molecular data presented in this study, and in light of the fact that numerous subulinids are not described anatomically, revision of the Subulinidae is clearly needed as this no longer appears to be a valid group. The Coeliacidae must also be revised as its representatives in this study grouped with either another subulinid or a thyrophorellid. The designation of the monotypic Thyrophorellidae requires re-examination based on *Thyrophorella thomensis*' close association with the coeliacid *Pyrgina umbilicata*. Finally, the association of *Tortaxis erectus* with *Glessula ceylanica* raises the question as to whether the Glessulidae are really restricted to the Indian subcontinent.

3.4.2.4. Evaluation of the monophyly of the different subulinid subfamilies

Of the four subulinid subfamilies represented in this study, only the Petriolinae (represented by *Bocageia* sp. and *Riebeckia* sp.) were recovered in any of the gene

phylogenies, in this case the NJ tree based on the actin gene though bootstrap support was less than 50% (see Appendix 3.11B, p. 442). For the rRNA, H3, COI and combined gene phylogenies, the Shimodaira-Hasegawa test did not find any significant difference in the likelihood scores between the optimal NJ tree (in which the Petriolinae were not monophyletic) and the constrained tree showing monophyly of the Petriolinae. There is therefore no compelling evidence to rule out the monophyly of the Petriolinae but neither is there any convincing evidence in support of the subfamily. The Petriolinae is distinct from all the other subfamilies based on the presence of a well-developed epiphallus and a very large and fleshy stimulator in the penis. *Bocageia* sp. and *Subulona* sp. have not yet been described based on their reproductive structure and were only included in the subfamily based on their shell morphology (Schileyko, 1999). The monophyly of the other subulinid subfamilies, particularly Rumininae sensu Schileyko (without *Zootecus*) and Rumininae sensu Zilch (with *Zootecus*), were strongly rejected by the Shimodaira-Hasegawa Test, and a re-examination of their taxonomic position is warranted. As discussed in the previous section, many of these taxa that have been included in the subfamilies have not been described anatomically, particularly with regards to the reproductive structures. In fact, this situation applies to many subulinids in general, which leaves their taxonomic position questionable (Schileyko, 1999).

3.4.3. The next step

The monophyly of the Achatinidae using molecular data is based only on very limited taxa sampled to date. A comprehensive survey of taxa within the family will determine the extent of their monophyly and at the same time give a glimpse of the inter-relationships of the various groups in the family on the basis of molecular data. Mead (1991, 1995) has already described various groups within the Achatinidae using

the reproductive tract as a criterion, and it would be interesting to know if these groupings are supported by molecular data. The molecular data have revealed that the Ferusaciidae are not monophyletic; the inclusion of additional taxa from the family is therefore necessary in order to re-evaluate the relationships within the group. The Subulinidae are also determined to be non-monophyletic based on molecular data; this necessitates a re-examination of the different surprising groupings of these subulinids in the light of their association with the Coeliacidae (also non-monophyletic), Glessulidae and Thyrophorellidae by expanding the taxon coverage to include other members of the families mentioned. There is also a need to investigate the morphological features of many of the anatomically undescribed species to correlate the molecular data with morphological data. Within the Subulinidae, only the Petriolinae had equivocal support for its monophyly. Including other taxa from the Petriolinae could shed more light on its status as a valid group. Other subfamilies that were not represented in this study (Obeliscinae, Opeatinae, Perrieriinae and Tristaniinae) should likewise be surveyed to determine their validity in the light of the rejection of the monophyly of the Rishetiinae, Rumininae and Subulininae.

3.5. Summary

The study provides a phylogenetic analysis of the Achatinoidea based on 24 taxa representing six families using the nuclear rRNA, actin and histone 3 genes and the 1st and 2nd codon positions of the mitochondrial cytochrome c oxidase subunit I gene. The 16S rRNA gene and the 3rd codon position of the COI gene were severely saturated and were excluded from the phylogenetic analyses. Combined gene phylogenies recovered most of the groupings in the rRNA phylogeny with longer internal branches and similar or slightly better statistical support. The study provided further strong molecular support

for the monophyly of the Achatinidae. The Ferussaciidae did not form a monophyletic group within the molecular trees, with *Ferussacia folliculus* falling separately from *Cecilioides gokweanus* at the base of the Achatinoidea, thus warranting a re-evaluation of the Ferussaciidae as a valid taxonomic group. The Coeliacidae was not supported, with the two representatives sequenced to date falling separately in the molecular trees, thereby prompting a re-evaluation of the validity of the family. Finally, the Subulinidae were not supported in the molecular trees with members of the Coeliacidae, Glessulidae and Thyrophorellidae falling among the subulinids. As with the Ferussaciidae and the Coeliacidae, the molecular data prompt a re-evaluation of the Subulinidae as a valid taxonomic group.

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CHAPTER 4 - Phylogeny of the Achatinidae

4.1. Introduction

4.1.1. The Achatinidae

Other than as exotic alien species, the Achatinidae are confined to sub-Saharan Africa and the Atlantic Islands in the Gulf of Guinea, West Africa. *Achatina* (*Lissachatina*) *immaculata* Lamarck is considered to have been introduced into Madagascar (Raut & Barker, 2002), but the most notable colonizing achatinid is *Achatina* (*Lissachatina*) *fulica* Bowdich, which has been introduced by Man throughout much of the tropics where it is a major pest species (Mead, 1979a; Raut & Barker, 2002). Achatinids include some of the largest known terrestrial molluscs (Bequaert, 1950) and have been aptly called Giant African Land Snails. There are 13 genera and approximately 200 species (Raut & Barker, 2002) distributed across 3 subfamilies within the Achatinidae (Schileyko, 1999). With the exception of the sinistral *Columna columna* (Müller) and *Archachatina bicarinata* (Bruguière), the shells of the Achatinidae are dextral. In general, achatinid shells are higher than wide and are fusiform, ovoid or pillar-shaped (Schileyko, 1999). Although achatinids do not possess the wide range of anatomical accessory structures that are found in many other stylommatophoran groups, the morphology of the genital structures in achatinids is highly variable (Mead, 1991); all members of the Achatinidae possess a well-developed penis sheath (Schileyko, 1999).

4.1.2. Distribution and habitat of the Achatinidae

The Achatinidae are widely distributed in sub-Saharan Africa. They are present from Senegal to Guinea in the west, the Zaire Basin, the Lake District and Southern Sudan in the centre, Somalia and southern Ethiopia in the northeast, Kenya, Tanzania and Mozambique in the east, and to Zambia, Botswana, Zimbabwe, South Africa, Namibia and Angola in the south and southwest (Mead, 1991; Raut & Barker, 2002). The Atlantic Islands in the Gulf of Guinea, including São Thomé and Príncipe, also harbor certain achatinids such as *Columna columna* (Schileyko, 1999) and the monotypic *Atopocochlis exarata* (Müller) (Mead, 1991; Raut & Barker, 2002). Van Bruggen (1986) noted an unequal distribution of diversity of the Achatinidae within sub-Saharan Africa, with Western Africa exhibiting a high concentration of endemic species. Habitat preferences of the achatinids are also diverse. Members of the genus *Achatina* in Central Africa thrive in humid, tropical forest areas. Some achatinids such as *Achatina (Lissachatina) fulica* in Eastern Africa flourish in modified tropical forests and forest margins and even in highly disturbed areas such as gardens, intensively farmed crops and plantations. Others such as the lowland *Cochlitoma zebra* (Bruguière) and the montane *Cochlitoma omissa* (van Bruggen) live in temperate regions in Southern Africa. Still others can be found in less humid areas, for example *Achatina (Lissachatina) immaculata* Lamarck in savannah and *Achatina (Achatina) damarensis* (Pfeiffer) in deserts of south and southwest Africa (Raut & Barker, 2002).

4.1.3. The search for the phylogeny of the Achatinidae using morphological characters

Several studies have attempted to elucidate the phylogeny of the Achatinidae using morphological characters (Bequaert, 1950; Mead, 1950, 1979b, 1991, 1995).

Classification within the family is based mostly on shell form and structure as well as the anatomy of the genitalia. However, different forces of selection act to different extents on the two systems. Shell form is subjected to selection pressures associated with particular habitats and homoplasy in shell form is common (Cain, 1977; Mead, 1994); modifications to the genitalia are more likely to reflect evolutionary processes (Mead, 1994) partly in response to sexual selection and selection pressures associated with the need for compatibility of form within species and incompatibility of form acting as reproductive isolation mechanisms between species. It is therefore not unusual to have cases where there is no correspondence between shell structures and the genitalia (Mead, 1979b, 1991) where distinct taxa living in separate but very similar environments may undergo convergent evolution and manifest strikingly similar shell morphology (Mead, 1994). For instance, the western species *Achatina* (*Achatina*) *achatina* Linnaeus and the southern species *Cochlitoma zebra* (Bruguière) are very similar conchologically and were placed in the same genus, *Achatina*, by Bequaert (1950) when clearly they belong to different genera based on the differences in their reproductive organs (Mead, 1991).

A typical achatinid reproductive system based on *Archachatina* (*Calachatina*) *marginata* Swainson is shown in Figure 4.1. Achatinids, like all stylommatophorans, are hermaphroditic. Both the sperm and egg are produced in the ovotestis, though not at the same time, and pass through the hermaphrodite duct. During copulation, the sperm proceeds to the male portion of the common hermaphrodite duct and then to the vas deferens and the penis. The penis then protrudes through the genital atrium and the genital pore and inserts into the reproductive tract of the partner until it reaches the vagina where it injects the sperm into the spermatolytic sac through the spermatolytic duct. The penis then retracts through the penial retractor muscle. Most of the sperm injected

into the spermatolytic sac is digested by proteolytic enzymes. Sperm that survive go through the spermatolytic duct and pass through the oviduct into the female hermaphrodite duct (=uterus) before they are stored in the seminal receptacle (=carrefour). Fertilization also takes place in the seminal receptacle, after which the fertilized eggs or zygotes receive yolk from the albumin gland. The zygotes then pass through the female hermaphrodite duct where the yellow egg shells are added before the eggs are released through the genital pore (Segun, 1975; Egonmwan, 2007).

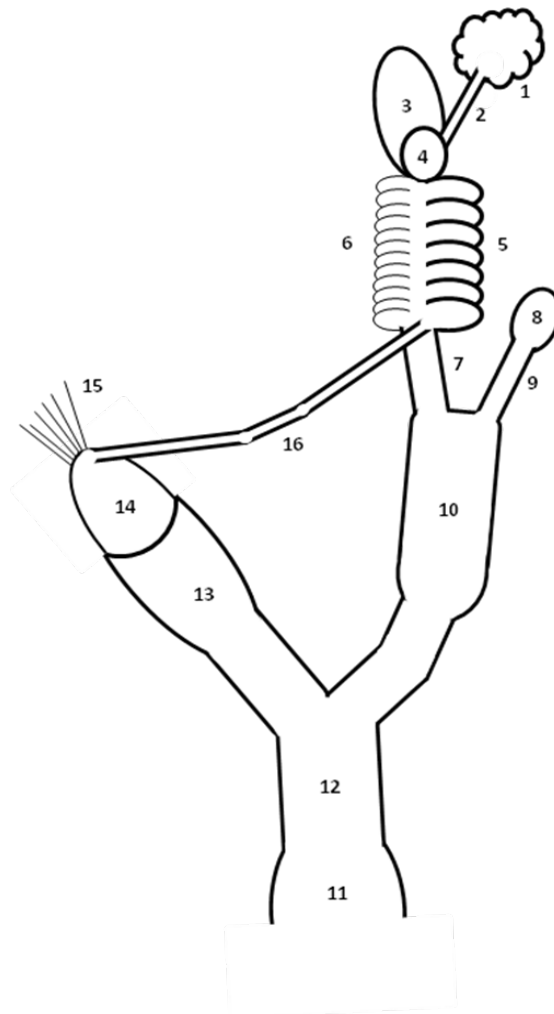


Figure 4.1: Generalized reproductive structure of achatinids. 1 – ovotestis; 2 – hermaphroditic duct; 3 – albumin gland; 4 – seminal receptacle; 5 - male part of common hermaphrodite duct; 6 – female part of common hermaphrodite duct (=uterus); 7 oviduct; 8 – spermatolytic sac (=spermatheca); 9 – spermatolytic duct; 10 – vagina; 11 – genital pore; 12 – genital atrium; 13 – penis sheath; 14 – penis; 15 – penial retractor muscle; 16 – vas deferens (Segun, 1975; Gomez, 2001; Egonmwan, 2007).

Morphological features of achatinid reproductive systems show a strong correlation with geographic distribution. For example, Mead (1991, 1994) hypothesized that members of the subfamily Callistoplepinae, which are found in Lower Guinea (Cameroon, Equatorial Guinea, Gabon and adjacent islands) in West Africa and include the genera *Callistoplepa* and *Leptocala*, probably gave rise to the other achatinids because they have the most 'primitive' genitalia in which the penis sheath does not enclose the vas deferens (Figure 4.2A). For the rest of the achatinids, the developing penis sheath envelopes the apical vas deferens to divide it into the apical and basal portions. Among these achatinids, the penis sheath either encloses the basal vas deferens but not the entire penis (microphallate type, Figure 4.2B) or it encloses both the basal part of the vas deferens as well as the entire penis (macrophallate type, Figure 4.2C). These two types can be found across different geographic regions of the Sub-Saharan Africa over several genera; for instance, the microphallate type is present in the West African *Achatina* (*Achatina*) *achatina*, *Archachatina marginata* and *Pseudachatina downesii* Sowerby and the Southern African *Metachatina kraussi* L. Pfeiffer while the macrophallate type is evident in the East African *Achatina* (*Lissachatina*) *fulica*, the Southern African *Cochlitoma* species and the East and West African *Limicolaria* species (Mead, 1979b; 1991). Apart from the penis sheath, other features also reflect groupings within the Achatinidae that have been thought to mirror their routes of dispersal. In the subfamily Limicolariinae, members are restricted to tropical regions of Africa and possess penial retractor muscles that originate from the diaphragm and are not enclosed by the penis sheath; a verge or penis papilla is also present at the base of the penis in this subfamily (Schileyko, 1999). Greater variation in the reproductive structure is exhibited in the subfamily Achatininae, members of which are distributed all over Sub-Saharan Africa (Schileyko, 1999), with some genera

possessing reproductive features that reflect their geographic distributions. For instance, extroversion muscles at the base of the penis sheath are very prominent in East African *Achatina* in the subgenus *Lissachatina* (Mead, 1979b, 1991), which include *A. (L.) fulica*, *A. (L.) immaculata*, *A. (L.) reticulata* Pfeiffer and *A. (L.) zanzibarica* Bourguignat. In Southern Africa, several species of *Archachatina* and *Achatina* exhibit a distinct folding of the penis, which prompted Mead (2004) to resurrect *Cochlitoma* Pilsbry. Based on these morphological features, it was proposed that the Achatinidae originated in Lower Guinea and dispersed through much of Sub-Saharan Africa (south of the Sahel) where they adapted to specific environmental conditions, from humid to arid, and tropical to temperate (Mead, 1991; Raut & Barker, 2002).

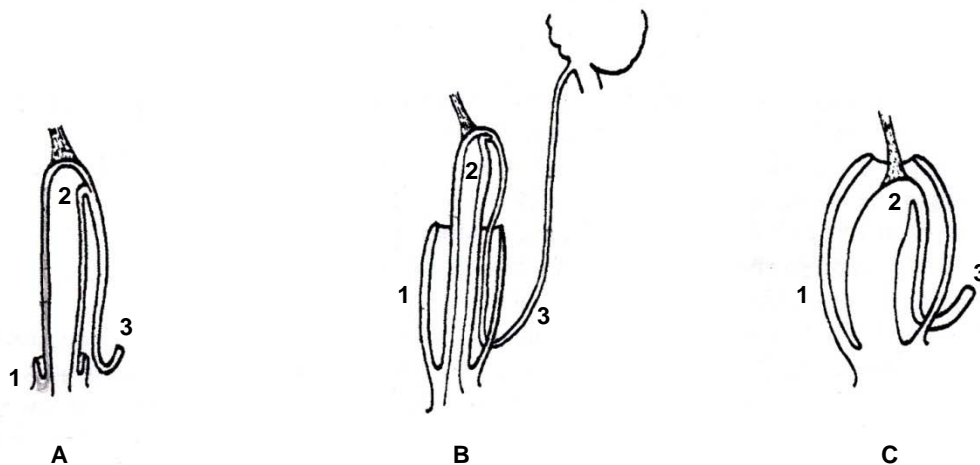


Figure 4.2: Schematic diagrams of how the penis sheath envelops certain parts of achatinid genitalia. 1 – penis sheath; 2 – penis; 3 – vas deferens. **A** – the penis sheath does not envelop the vas deferens, as in *Callistoplepa*. **B** – the penis sheath envelops the vas deferens and divides into the apical and basal portions, as in the West African *Archachatina*. **C** – the penis sheath envelops both the entire penis and the basal segment of the vas deferens, as in *Achatina (Lissachatina)*. (From Mead, 1991).

4.1.4. Taxonomic revision of the genus *Achatina* based on morphological characteristics

Bequaert (1950) used the appearance of the nepionic whorls of the shell to distinguish two subgenera of the genus *Achatina*, namely: (1) *Achatina* found in West and Central Africa; and (2) *Lissachatina* found in East Africa. He noted that the nepionic whorls in *Achatina* (*Achatina*) are sculptured whereas those of *Achatina* (*Lissachatina*) are either smooth or with faint vertical wrinkles. Mead (1995) further added that *Achatina* (*Achatina*) normally has a plateaued apex while *Achatina* (*Lissachatina*) exhibits a lopsided or half-dome apex. However, distinguishing these two subgenera using taxonomic characters based on the shell can be unreliable in the case of specimens with worn or weathered shells (Mead, 1995). The morphology of the reproductive organs is considered to provide more reliable characters for recognizing natural groups (Mead, 1991). For example, *Achatina* (*Achatina*) has a short spermolytic duct (=spermathecal duct *sensu* Mead), which causes the spermolytic sac (=spermatheca) to be located below the junction of the apical vas deferens and the uterine portion of the common hermaphrodite duct (=spermoviduct). In contrast, *Achatina* (*Lissachatina*) has a very long spermolytic duct, thus pushing the spermolytic sac way above the junction of the apical vas deferens and the uterine portion of the common hermaphrodite duct. Furthermore, *Achatina* (*Lissachatina*) tends to have a muscular, bulboid enlargement at the basal vagina (Fig. 4.3) in addition to the presence of extroversion muscles at the base of the penis sheath (Mead 1991; 1995).

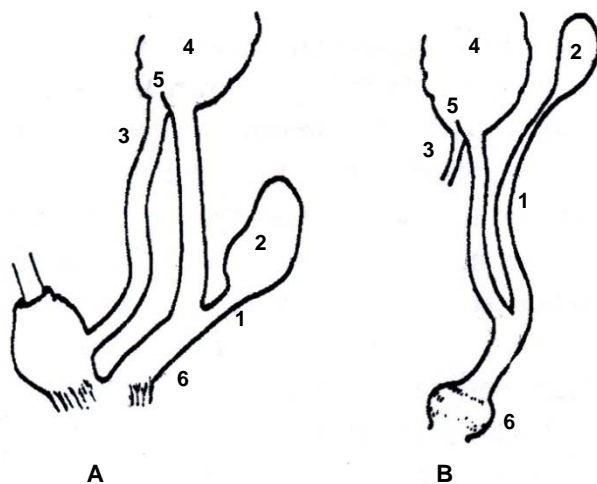


Figure 4.3: Schematic illustrations of the reproductive organs of the West and Central African *Achatina* (*Achatina*) (A) and the East African *Achatina* (*Lissachatina*) (B). 1 – spermolytic duct; 2 – spermolytic sac; 3 – apical vas deferens; 4 – uterine portion of the common hermaphrodite duct; 5 – junction of 3 and 4; 6 – basal vagina. Note that the spermolytic duct is longer in *Achatina* (*Lissachatina*), pushing the spermolytic sac above the junction of the vas deferens and the uterine portion of the common hermaphroditic duct. *Lissachatina* also tends to have a bulbous basal vagina. (From Mead, 1991)

4.1.5. Objective of the study

Results from Chapter 3 confirm the monophyly of the Achatinidae as represented by four taxa. However, no molecular based phylogenetic analyses have been carried out to determine the relationships within the Achatinidae. The current investigation aimed to address this issue by obtaining molecular data for the nuclear ribosomal rRNA cluster, actin and histone 3 genes as well as the mitochondrial cytochrome c oxidase subunit 1 and 16S rRNA genes to elucidate the phylogeny of the Achatinidae. The validity of the taxonomic designation of the East African *Lissachatina* and the West African *Achatina* was also evaluated using the molecular data.

4.2. Materials and methods

4.2.1. Taxa used

Twenty-nine taxa from the family Achatinidae were used to evaluate the phylogeny of the Achatinidae (see Table 4.1). Except for five taxa used by Wade *et al.* (2006), namely *Achatina* (*Lissachatina*) *fulica*, *Archachatina marginata*, *Atopocochlis exarata*, *Limicolaria kambeul* and the subulinid outgroup taxon *Rumina decollata*, all other taxa have not been evaluated in previous studies. *Rumina decollata* was chosen as the outgroup because it fell immediately outside of the Achatinidae in the phylogenetic studies found in Chapter 3, and it was the taxon that allowed incorporation of the greatest number of unambiguously aligned nucleotide sites across the Achatinidae for the ITS 2 region. Three subgenera of *Achatina* were represented in this study, namely *Achatina* (represented by *A. achatina*, *A. bisculpta*, *A. damarensis* and *A. stuhlmanni*), *Euaethiopina* (*A. loveridgei*) and *Lissachatina* (*A. fulica*, *A. immaculata*, *A. reticulata* and *A. zanzibarica*).

Table 4.1: Taxa used for the phylogenetic study of the Achatinidae. With the exception of *Achatina* (*Lissachatina*) *fulica*, *Archachatina marginata*, *Atopocochlis exarata*, *Limicolaria kambeul* and *Rumina decollata* from Wade *et al.* (2006), all other taxa were not used in previous studies and were solicited through email by myself, C. Hudelot, F. Naggs and C. Wade from the collectors shown in the table. The DNA for the five taxa used by Wade *et al.* (2006) was provided by C. Wade.

Family	Species	Collection/Location	Collector/Provider
Achatinidae	<i>Achatina</i> (<i>Achatina</i>) <i>achatina</i> (Linnaeus, 1758)	Unknown (Zool. Soc. Lond. Colln.)	NHM*
	<i>Achatina</i> (<i>Achatina</i>) <i>bisculpa</i> Smith, 1878	Gauteng Prov., South Africa	K. du Preez
	<i>Achatina</i> (<i>Achatina</i>) <i>damarensis</i> (Pfeiffer, 1870)	Damaraland, Namibia	D. Herbert
	<i>Achatina</i> (<i>Achatina</i>) <i>stuhlmanni</i> von Martens, 1892	Semuliki National Park, Uganda	B. Rowson
	<i>Achatina</i> (<i>Lissachatina</i>) <i>fulica</i> Bowdich, 1822	Captive bred, unknown origin	NHM
	<i>Achatina</i> (<i>Lissachatina</i>) <i>immaculata</i> Lamarck, 1822	Cato Ridge, South Africa	D. Herbert
	<i>Achatina</i> (<i>Lissachatina</i>) <i>reticuata</i> Pfeiffer, 1845	Mkungwe Forest reserve, Uluguru Mts, Tanzania	B. Rowson
	<i>Achatina</i> (<i>Lissachatina</i>) <i>zanzibarica</i> Bourguignat, 1879	Lulanda Forest Reserve, Udzungwa Scarp, Tanzania	B. Rowson
	<i>Achatina</i> (<i>Euaethiopina</i>) <i>loveridgei</i> (Clench & Archer, 1930)	Uluguru Forest Reserve, Tanzania	B. Rowson
	<i>Archachatina marginata</i> (Swainson, 1821)	Purchased in Lagos Market, Nigeria	NHM
	<i>Atopocochlis exarata</i> (Müller, 1774)	Nr. Belem, São Thomé	A. Gascoigne
	<i>Cochlitoma churchilliana</i> (Melvill & Ponsonby, 1895)	KwaZulu-Natal, South Africa	D. Herbert
	<i>Cochlitoma dimidiata</i> (Smith, 1878)	Mpumalanga, South Africa	A. Moussalli & D. Stuart-Fox
	<i>Cochlitoma granulata</i> (Krauss, 1848)	KwaZulu-Natal, South Africa	A. Moussalli & D. Stuart-Fox
	<i>Cochlitoma kilburni</i> Mead, 2004	Eastern Cape Prov., South Africa	L. Davis & M. Bursey
	<i>Cochlitoma marinae</i> (Sirgel, 1989)	Mossel Bay, Eastern Cape Prov., South Africa	K. Tolly
	<i>Cochlitoma montistempli</i> (van Bruggen, 1965)	Cathedral Peak area, Fern Forest, South Africa	D. Herbert
	<i>Cochlitoma omissa</i> (van Bruggen, 1965)	Monks Cowl area, South Africa	D. Herbert
	<i>Cochlitoma semidecussata</i> (Pfeiffer, 1846)	Durban, Pigeon Valley Park, South Africa	A. Moussalli & D. Stuart-Fox
	<i>Cochlitoma simplex</i> (Smith 1878)	Colenso area, Emaweni Game Ranch, near Brakfontein, South Africa	V. Albers
	<i>Cochlitoma</i> sp. cf. <i>vestita</i> (Pfeiffer, 1861)	KwaZulu-Natal, South Africa	D. Herbert
	<i>Cochlitoma ustulata</i> (Lamarck, 1822)	Western Cape Prov., South Africa	A. Moussalli & D. Stuart-Fox
	<i>Cochlitoma varicosa</i> (Pfeiffer, 1861)	Transkei, South Africa	J. Marais
	<i>Cochlitoma zebra</i> (Bruguère, 1789)	W. Cape, Witsand, South Africa	A. Moussalli & D. Stuart-Fox
	<i>Limicolaria kambeul</i> (Bruguère, 1792)	Somalia (NHM collection)	M. Leng
	<i>Limicolaria martensii</i> (Smith, 1880)	Bulyanbulu, Tanzania	A. Moussalli
	<i>Limicolariaopsis ruwenzoriensis</i> Pilsbry, 1919	Rwenzori Mts, National Park, Uganda	B. Rowson
	<i>Limicolariaopsis</i> sp.	Saimofores, Tupen Hills, Kenya	M. Pickford
	<i>Metachatina kraussi</i> Pilsbry, 1904	Zululand, South Africa	D. Herbert
Subulinidae (outgroup)	<i>Rumina decollata</i> (Linnaeus, 1758)	Sicily	A. Davison

*NHM-Natural History Museum

4.2.2. DNA extraction, PCR amplification and sequencing

The CTAB method of DNA extraction (Section 2.1.1 of Chapter 2, pp. 37 – 39) was used on tissue slices (approximately 8 mm³) derived from the foot muscle of each snail. The DNA for *Achatina (Lissachatina) fulica*, *Archachatina marginata*, *Atopocochlis exarata*, *Limicolaria kameul* and *Rumina decollata* was provided by C.M. Wade.

Amplification by PCR, gel migration and purification of PCR products from agarose gels were carried out as described in Sections 2.2 – 2.4 of Chapter 2, pp. 42 – 59. Approximately 4000 bp of the nuclear rRNA cluster (80 bp of the 5.8S rRNA, the full ITS2 and nearly the entire fragment of the LSU rRNA), 900 bp of the nuclear cytoplasmic actin gene, 328 bp of the nuclear histone 3 gene, 650 bp of the Folmer *et al.* (1994) mitochondrial COI fragment and 420-500 bp of the Palumbi *et al.* (1991) mitochondrial 16S (420-500 bp) fragment were amplified using the primers listed in Tables 2.1 – 2.5 of Chapter 2, pp. 47 – 53. Both sense and anti-sense strands were sequenced directly using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 terminator cycle sequencing chemistry (see Section 2.6 of Chapter 2, pp. 59 – 61 for details).

4.2.3. Sequence analysis

Sequences were assembled using the STADEN package version 1.5.3 (Staden *et al.*, 2000) and aligned manually within the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994). Ambiguous sites due to sequencing errors (for all genes) or the presence of multiple copies (for the actin gene) were assigned as described in Section 2.9.3 of Chapter 2, pp. 72-73 and using as a guide the three cloned sequences from representative taxa of the Achatinoidea in Chapter 3.

The average base frequencies and the numbers of variable and parsimony-informative sites for each gene fragment were determined (Section 2.9.4 of Chapter 2, p. 73). Corrected distances were computed after determination of the optimal model for DNA sequence evolution. Models evaluated by the Likelihood Ratio Test (LRT) were the JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P (Kimura, 1980), HKY85 (Hasegawa *et al.*, 1985), TN93 (Tamura & Nei, 1993) and the GTR (Rodriguez *et al.*, 1990) models as well as their variant that incorporated gamma distributed rates (Γ) (Yang, 1993) (Section 2.9.5 of Chapter 2, pp. 73-77). The sequences were also examined for evidence of saturation by plotting the pairwise corrected versus pairwise uncorrected distances (Plot 1), pairwise uncorrected transition and transversion distances versus pairwise uncorrected total distances (Plot 2), and pairwise uncorrected transition distances versus pairwise uncorrected transversion distances (Plot 3) (Section 2.9.6 of Chapter 2, pp. 77-81). Gene datasets that demonstrated saturation were excluded from subsequent phylogenetic analyses. The sequences were also subjected to the *g1* measure of skewness test based on 10,000 tree length replicates to determine if there is phylogenetic signal (Section 2.9.7 of Chapter 2, p. 81-82). The presence of paralogous sequences in actin that could affect phylogenetic analyses was detected (Section 2.9.12 of Chapter 2, pp. 89-90) and the utility for of the actin gene for phylogenetic studies assessed (Section 2.9.13 of Chapter 2, pp. 90-91).

Phylogenetic trees were generated from the aligned datasets using the model based maximum likelihood (ML), Bayesian inference (BI) and neighbor-joining (NJ) methods and the non-model based maximum parsimony (MP) method (Section 2.9.8 of Chapter 2, pp. 82-85). Bootstrap resampling (Felsenstein, 1985) with 1000 replicates for ML, MP and NJ were also carried out. Bayesian inference (BI) was performed using four chains of a Markov Chain Monte Carlo algorithm. The number of

generations to explore the tree space and the heating temperature used for each gene were optimised as described in the BI segment of Section 2.9.8 of Chapter 2, p. 83-84 and in Appendix 2.3.2, pp. 369-370. A consensus tree for each gene (where unsaturated) was constructed using the last 1000 trees.

A partition-homogeneity test (Swofford, 2002) was carried out to determine if the sequences from the different genes could be combined into one dataset (Section 2.9.10 of Chapter 2, pp. 86-88). Two sets of concatenated sequences were prepared; one set included only those taxa with complete sequences for all gene fragments while the other set included all taxa.

The Shimodaira-Hasegawa (1999) test was used to test the monophyly of taxonomic groups that were expected to be monophyletic based on taxonomy but did not cluster together in the optimal trees (Section 2.9.11 of Chapter 2, p. 89).

4.3. Results

4.3.1. Molecular data

Twenty nine achatinids and the subulinid *Rumina decollata* were examined for the nuclear rRNA cluster, actin and histone 3 genes as well as the mitochondrial 16S and cytochrome c oxidase I genes. Sequence alignments are presented in Appendices 4.1 – 4.5 on pp. 451-507. All sequences were checked for the presence of ambiguous positions in the direct sequences that might indicate the presence of multiple gene copies. (Ambiguities due to multiple gene copies were detected only in the actin gene; see Section 4.3.2.2, pp. 247-249 for details). A summary of the molecular data for all the sequences is shown in Table 4.1. For the protein-coding genes (actin, histone 3 and COI), information was obtained separately for the entire gene (all codon positions), the

combined 1st and 2nd codon positions and the 3rd codon position only. Uncorrected and corrected pairwise distances and their optimal models were determined for all five genes and are likewise summarized in Table 4.2. The following models were found to be the optimal models for the different datasets: GTR+ Γ for the 16S rRNA gene and the full (all codon positions) of the actin and COI genes as well as the 1st + 2nd codon position and the 3rd codon position datasets of COI and the 3rd codon position dataset of actin; TN93+ Γ for the actin 1st and 2nd codon positions; TN93 for the nuclear rRNA cluster; HKY for the histone 1st and 2nd codon positions; and HKY+ Γ for the histone 3 full and 3rd codon position datasets. (See Appendix 4.6, p. 508 for the summary of the LRT results).

Nearly the entire fragment of the LSU rRNA gene, the internal transcribed spacer 2 (ITS 2) and 80 nucleotides of the 5.8S rRNA gene were sequenced (approximately 4000 nucleotides) for the rRNA gene cluster, of which 3648 nucleotides (including 194 nucleotides of the ITS2) could be aligned unambiguously. A total of 105 (2.9%) variable sites was found, of which 40 were parsimony-informative. Pairwise distances across all taxa ranged from 0 to 0.014 (uncorrected) and 0 to 0.139 (corrected). *Cochlitoma montistempli* and *C. omissa* yielded identical sequences based on the 3648 unambiguously aligned sites, although variations were exhibited between these two taxa in the excluded nucleotides, particularly in the ITS2 region. The highest base frequency for the rRNA gene cluster dataset was obtained for G (0.320) followed by C (0.266), then by A (0.224) and finally by T (0.190). For the actin gene, 873 nucleotide positions were sequenced, of which all sites could be aligned unambiguously; of these, 185 (21.2%) were variable and 84 were parsimony-informative. Pairwise distances ranged from 0 to 0.147 (uncorrected) and 0 to 0.372 (corrected). Highest average base frequency for actin was computed for A (0.268)

followed by T (0.257) then by C (0.254) and finally by G (0.221). Looking at the different codon positions, the majority of the variable sites (161) and parsimony-informative sites (72) were at the 3rd codon position, while the combined 1st and 2nd codon positions only had 24 variable sites and 12 parsimony-informative sites. Amplification of the actin gene for *Limicolariopsis ruwenzoriensis* was unsuccessful. For the histone 3 gene, 328 nucleotides were sequenced and aligned unambiguously, with 61 (18.6%) variable positions and 27 parsimony-informative sites. Pairwise distances ranged from 0 to 0.090 (uncorrected) and 0 to 0.148 (corrected). Five sets of identical sequences were observed for the histone 3 gene; the first set included *Achatina* (*Lissachatina*) *fulica*, *A. (L.) reticulata* and *A. (L.) zanzibarica*; the second included *Achatina* (*Achatina*) *bisculpta*, *A. (A.) damarensis* and *A. (A.) stuhlmanni*; the third included *Limicolaria kambeul* and *Limicolaria martenssii*; the fourth included *Cochlitoma dimidiata*, *C. montistempli*, *C. omissa*, *C. semidecussata*, *C. simplex* and *C. varicosa*; and the fifth included *C. kilburni* and *C. sp. cf. vestita*. The presence of these sets of identical sequences and the low uncorrected distances demonstrate the conserved nature of the histone 3 gene fragment at the level of the Achatinidae. Average base frequencies for histone 3, in decreasing order, were as follows: C (0.320); G (0.275); A (0.243) and T (0.162). As with the actin gene, the 3rd codon position exhibited the most number of variable sites (57) as well as parsimony-informative sites (26) in comparison to the combined 1st and 2nd codon positions with only 4 and 1, respectively. Amplification of the histone 3 gene fragment for *Achatina* (*Euaethiopina*) *loveridgei* and *Limicolariopsis ruwenzoriensis* was unsuccessful. For the mitochondrial COI gene, approximately 650 nucleotides were sequenced, with the length varying depending on the primers used. A total of 641 unambiguously aligned nucleotides were used, of which 293 (45.7%) were variable and 276 were parsimony-informative. Pairwise

distances ranged from 0.067 to 0.264 (uncorrected) and 0.211 to 11.856 (corrected), making the COI gene fragment more variable than the nuclear genes above. No two sequences were found alike, and the smallest uncorrected distance is 0.067, way above the 0.03 sequence divergence threshold for any two species set by Hebert *et al.* (2003) for the COI.

Average base frequencies, in decreasing order, were as follows: T (0.376); A (0.239); G (0.204) and C (0.180), indicating a strong bias for T; this was heightened in the 3rd codon position where average frequency of T rose to 0.404. The 3rd codon position had 214 variable sites and 212 parsimony-informative sites, whereas the combined 1st and 2nd codon positions only had 79 and 64, respectively. Note that sequencing was unsuccessful for *A. (E.) loveridgei* and *Limicolariopsis ruwenzoriensis*. The mitochondrial 16S rRNA gene product ranged from 420-500 nucleotides, with 310 that were unambiguously aligned, 147 (47.4%) that were variable, and 110 that were parsimony-informative. Pairwise distances ranged from 0.019 to 0.242 (uncorrected) and 0.021 and 0.837 (corrected); thus, the gene is likewise more variable than the nuclear genes. As with the COI, no two sequences were found alike. Base frequencies were as follows: T (0.285); A (0.277); G (0.247) and C (0.191), showing bias for A and T. Sequencing of the 16S rRNA gene fragment for *L. ruwenzoriensis* was unsuccessful.

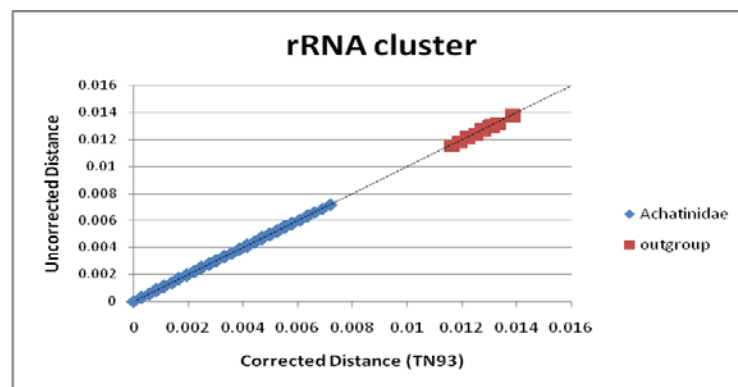
Table 4.2: Summary of molecular data across all genes used (rRNA gene cluster, actin, histone 3, COI and 16S) for the Achatinidae and the subulinid outgroup *Rumina decollata*. A=Achatinidae only; A+O=Achatinidae and outgroup.

	Length (bp)	# of variable sites (%)		Range of distances (Uncorrected)		Optimal Model (α value for Γ distribution)	Range of distances (Corrected)		Average base frequencies								# of parsimony- informative sites	
									A		C		G		T			
		A	A+O	A	A+O		A	A+O	A	A+O	A	A+O	A	A+O	A	A+O	A	A+O
rRNA cluster	3648	77 (2.1)	105 (2.9)	0-0.007	0-0.014	TN93 (N/A)	0-0.007	0-0.139	0.224	0.224	0.266	0.266	0.320	0.320	0.190	0.190	37	40
Actin combined	873	176 (20.2)	185 (21.2)	0-0.147	0-0.147	GTR+ Γ (0.175)	0-0.372	0-0.372	0.268	0.268	0.254	0.254	0.221	0.221	0.257	0.257	71	84
Actin 1 st and 2 nd codon positions	582	23 (4.0)	24 (4.1)	0-0.031	0-0.031	TN93+ Γ (0.043)	0-0.053	0-0.053	0.311	0.311	0.229	0.229	0.227	0.227	0.233	0.233	11	12
Actin 3 rd codon position	291	153 (52.6)	161 (55.3)	0-0.628	0-0.384	GTR+ Γ (0.686)	0-0.628	0-0.628	0.180	0.179	0.303	0.304	0.211	0.211	0.306	0.306	60	72
Histone3 combined	328	53 (16.2)	61 (18.6)	0-0.090	0-0.090	HKY85+ Γ (0.256)	0-0.148	0-0.148	0.243	0.243	0.320	0.320	0.275	0.275	0.162	0.162	22	27
Histone 3 1 st and 2 nd codon positions	218	4 (1.8)	4 (1.8)	0-0.019	0-0.019	HKY85 (N/A)	0-0.019	0-0.019	0.288	0.288	0.285	0.285	0.261	0.261	0.166	0.166	1	1
Histone 3 3 rd codon positions	110	49 (44.5)	57 (51.8)	0-0.280	0-0.280	HKY85+ Γ (2.967)	0-0.460	0-0.460	0.150	0.151	0.392	0.391	0.305	0.305	0.154	0.154	21	26
COI combined	641	292 (40.9)	293 (45.7)	0.067-0.264	0.067-0.264	GTR+ Γ (0.106)	0.211- 11.856	0.211-11.856	0.239	0.239	0.181	0.180	0.205	0.204	0.376	0.376	275	276
COI 1 st and 2 nd codon positions	427	78 (18.3)	79 (18.5)	0.007-0.098	0.007-0.098	GTR+ Γ (0.072)	0.008- 0.389	0.008- 0.389	0.197	0.197	0.204	0.204	0.236	0.236	0.362	0.362	63	64
COI 3 rd codon positions	214	214 (100)	214 (100)	0.164-0.625	0.164-0.625	GTR+ Γ (0.659)	1.304-120.141	1.304-120.141	0.321	0.323	0.134	0.133	0.142	0.140	0.403	0.404	212	212
16S rRNA	310	145 (46.8)	147 (47.4)	0.019-0.242	0.019-0.242	GTR+ Γ (0.250)	0.021- 0.837	0.021- 0.837	0.277	0.277	0.191	0.191	0.247	0.247	0.285	0.285	107	110

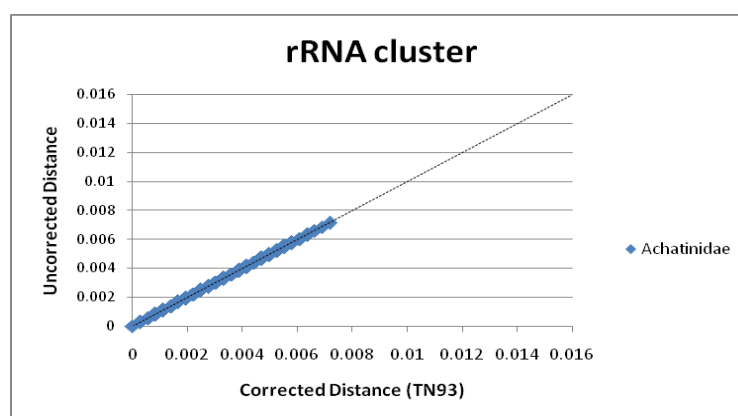
4.3.2. Sequence analyses

4.3.2.1. Evaluating for saturation and phylogenetic signal

For the rRNA cluster, the uncorrected versus corrected distances plots (Plot 1, Figure 4.4) demonstrated a linear relationship between the corrected distance (based on the optimal TN93 model) and the uncorrected (p) distance. This suggested that the dataset for the rRNA cluster was not saturated and that the TN93 model was adequate for correcting the distances for multiple hits.



(A)



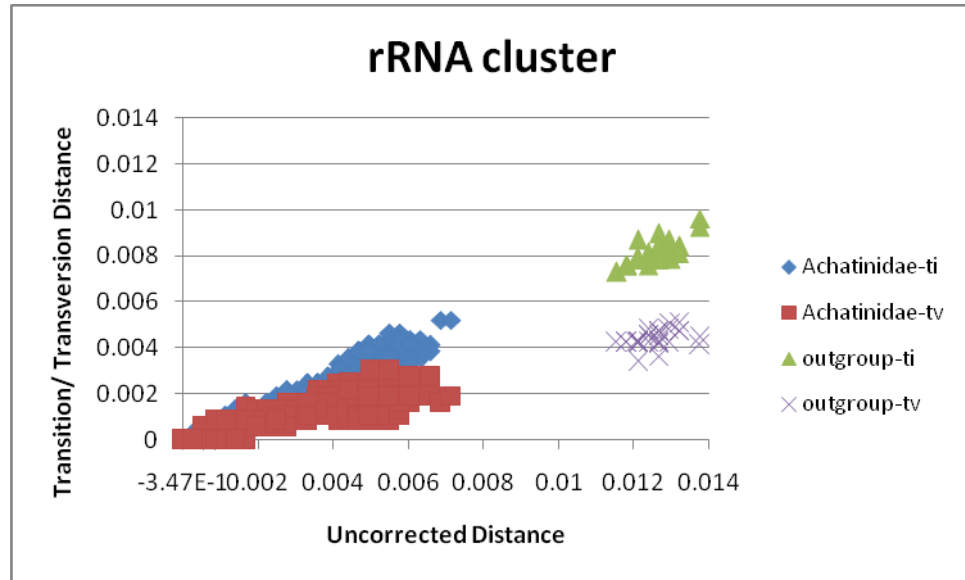
(B)

Figure 4.4: Plots of pairwise uncorrected distance against corrected (TN93) distance for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

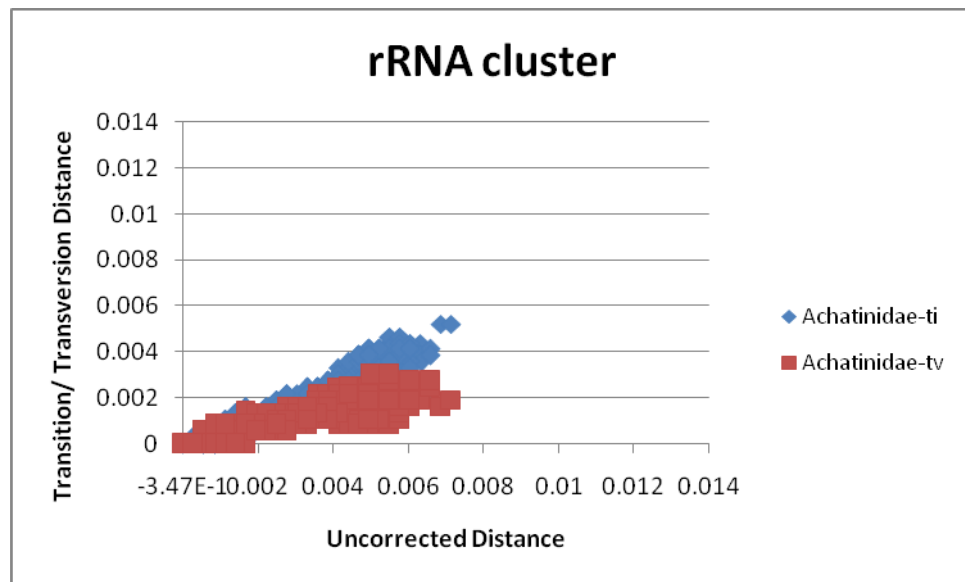
Plots for the transition and transversion rates versus uncorrected distances both with and without the outgroup taxon (Plot 2, Figure 4.5) showed that both transition and transversion substitutions were increasing linearly with transition distances higher than transversion distances. Plots for transitions against transversions both with and without the outgroup taxon (Plot 3, Figure 4.6) confirmed the results of Plot 2 where transition rates were higher than transversion rates. The results suggested that the rRNA cluster dataset was not saturated.

A g_i value of -0.596 was obtained for the rRNA cluster with 10,000 replicates based on 30 taxa and 40 parsimony-informative characters. This value differed significantly from the critical g_i value of -0.16 at $P=0.05$ level of significance for 25 taxa and 10 parsimony-informative characters (Hillis & Huelsenbeck, 1992). This result was indicative of a strong phylogenetic signal.

The absence of saturation and the presence of phylogenetic signal implied that the rRNA cluster was suitable for phylogenetic analyses of the Achatinidae.

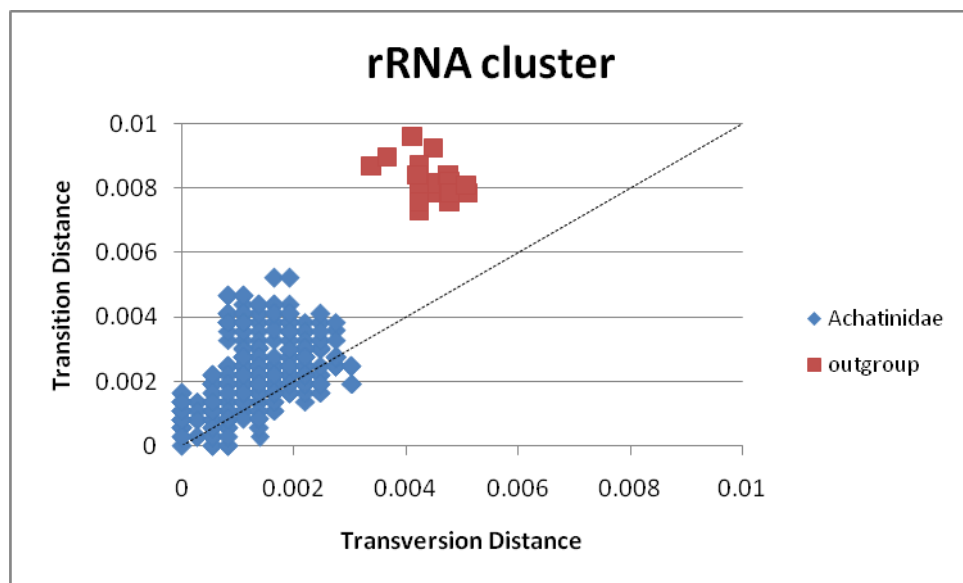


(A)

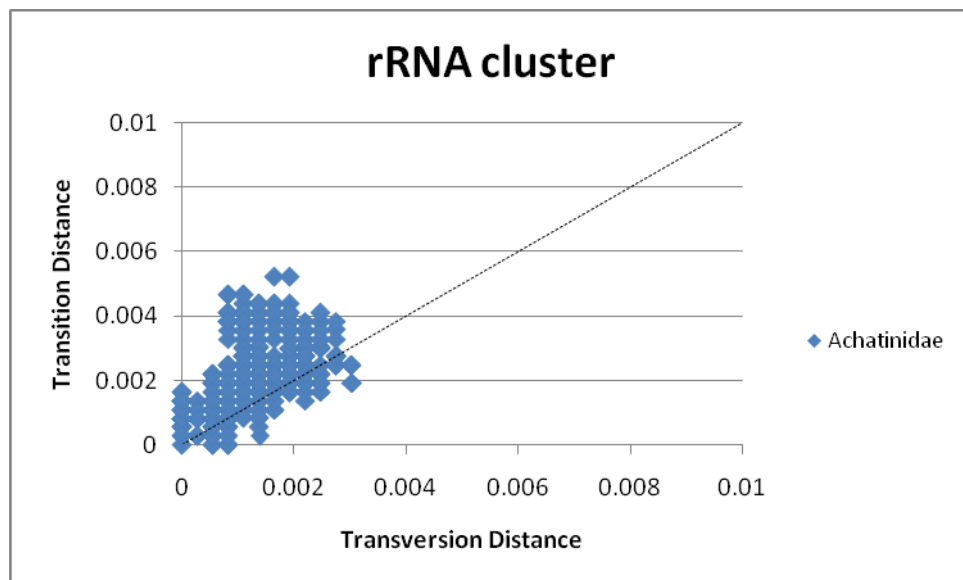


(B)

Figure 4.5: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



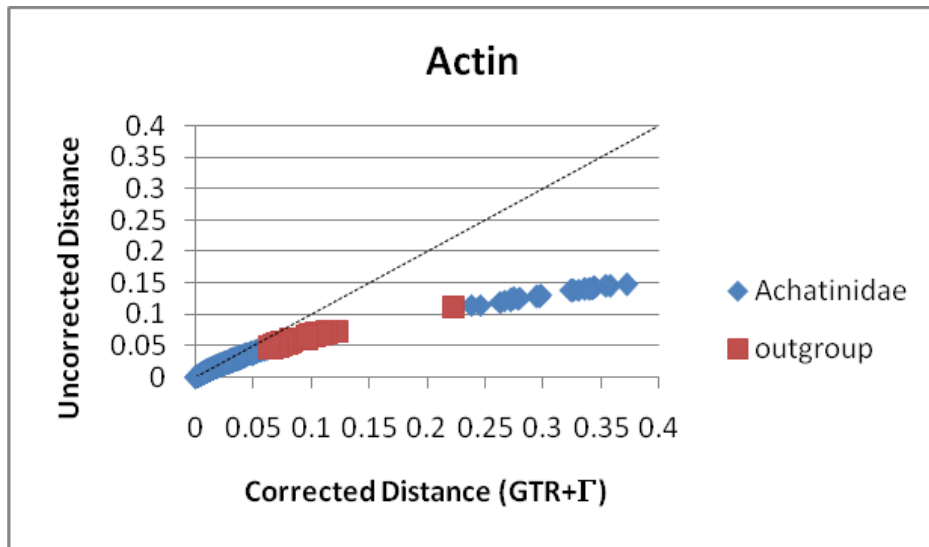
(A)



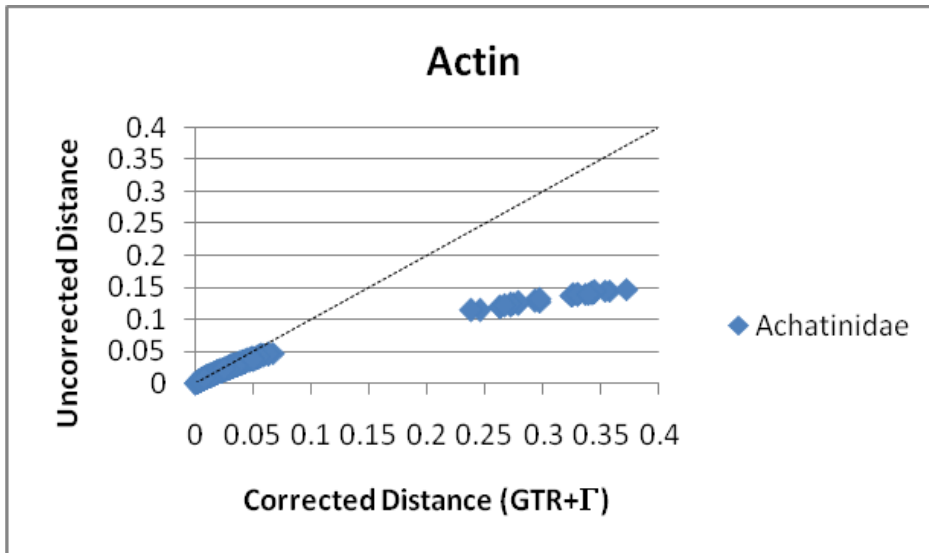
(B)

Figure 4.6: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

For the actin gene, Plot 1 (Figure 4.7) demonstrated that the corrected distance, based on the optimal GTR+ Γ model, deviated from linearity starting at an uncorrected (p) distance of approximately 0.05 for both the Achatinidae only and the Achatinidae plus outgroup taxon. A big gap was also shown in both plots; the points that came after the gap were attributed to the distance between the actin copy of *Cochlitoma marinae* and those of the other taxa. The presence of this highly divergent actin copy and its implications in the phylogenetic analysis of the Achatinidae using actin is discussed further in Section 4.3.2.2, pp. 247-249. Both plots were still increasing and had not reached a plateau, suggesting adequate correction of the optimal GTR+ Γ model and no saturation in the dataset.



(A)



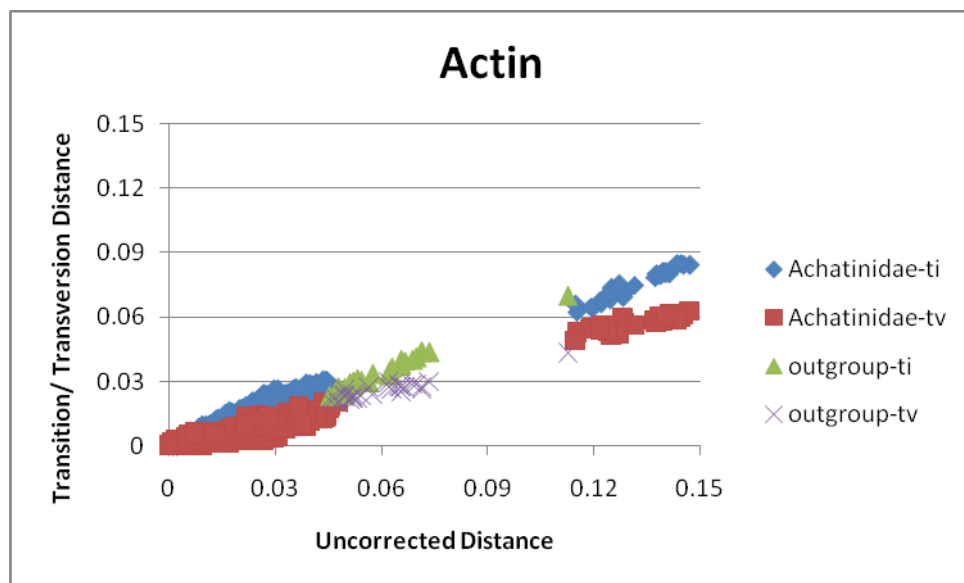
(B)

Figure 4.7: Plots of pairwise uncorrected distance against corrected distance for the actin gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only. Points after the gap were attributed to the divergent actin sequence of *Cochlitoma marinae*.

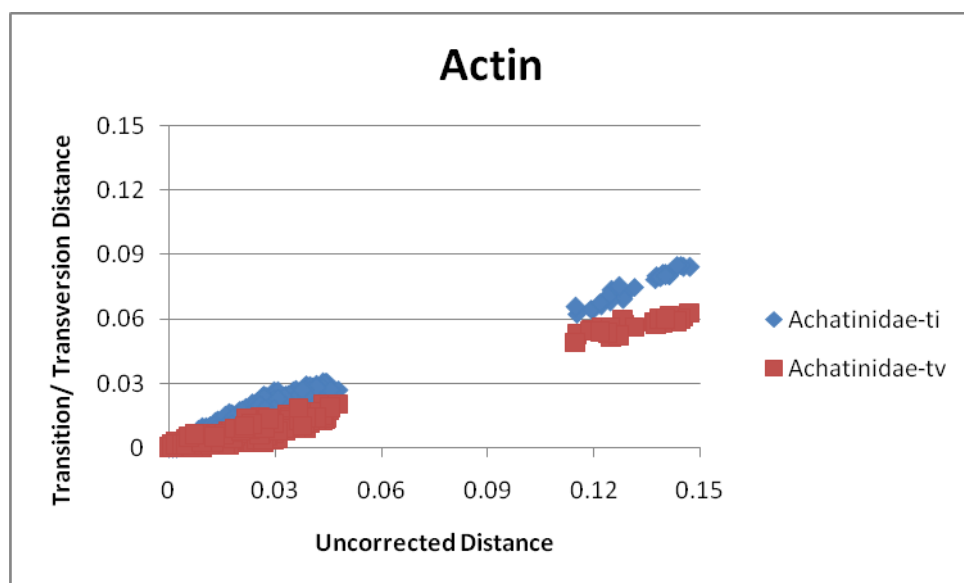
Plot 2 (Figure 4.8) for the actin gene, whether with or without outgroup taxa, exhibited a linear increase for both transitions and transversions, with transition distances higher than transversion distances. Plot 3 (Figure 4.9) confirmed the higher transition rates over the transversion rates. These results suggested that the actin gene was not saturated. Once again, a gap was observed in both plots due to the highly divergent *Cochlitoma marinae*.

A g_i value of -0.475 was computed for the actin gene based on 29 taxa and 84 parsimony-informative sites, much lower than the critical value of -0.12 at $p=0.05$ level of significance for 25 taxa and 50 parsimony-informative characters (Hillis & Huelsenbeck, 1992). The actin gene therefore exhibited strong phylogenetic signal.

Lack of saturation and the presence of phylogenetic signal implied that the actin gene was suitable for phylogenetic analyses of the Achatinidae and the outgroup taxon. However, the observed gap in the plots due to the highly divergent sequence of *Cochlitoma marinae* suggested caution must be exercised in using this gene in phylogenetic analyses. These issues are discussed in section 4.3.2.2, pp. 247-249.

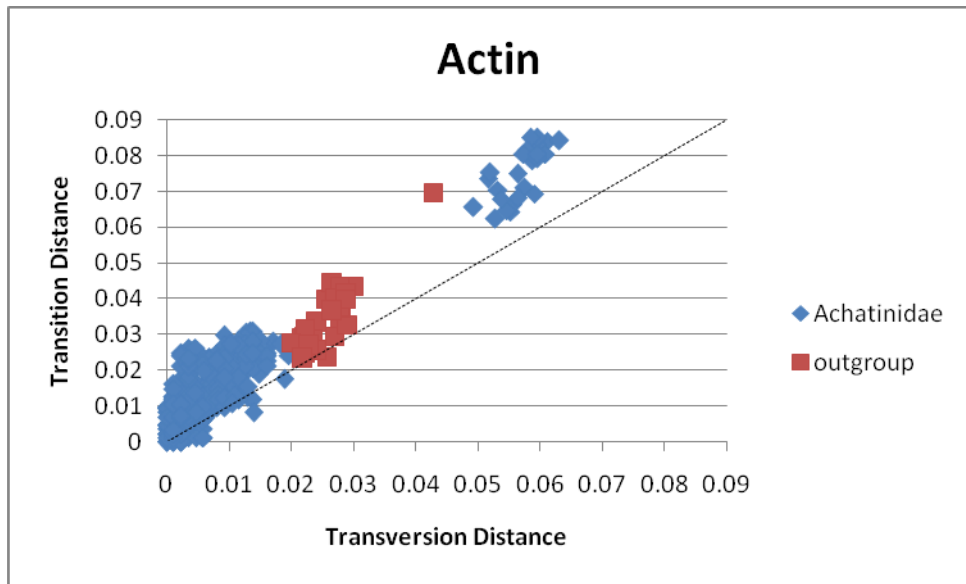


(A)

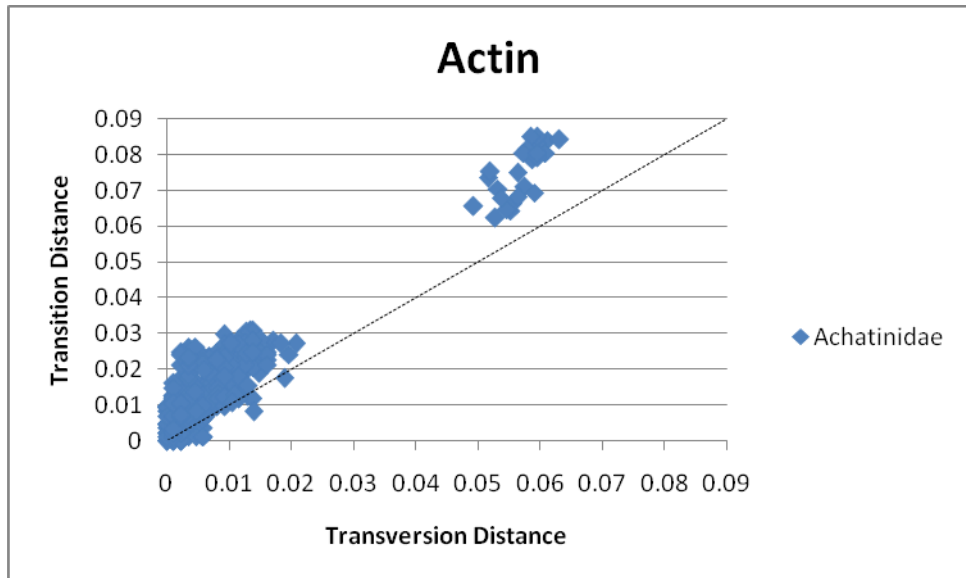


(B)

Figure 4.8: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the actin in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only. Points after the gap were attributed to the divergent actin sequence of *Cochlitoma marinae*.



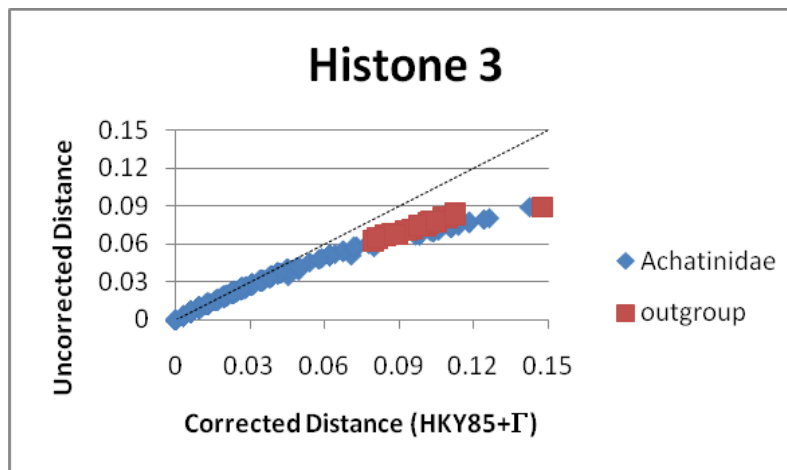
(A)



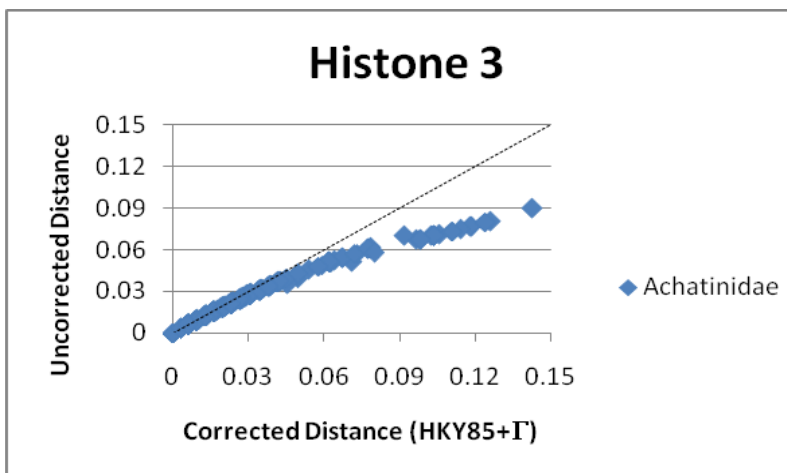
(B)

Figure 4.9: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the actin gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only. Points after the gap were attributed to the divergent actin sequence of *Cochlitoma marinae*.

For the histone 3 gene, Plot 1 (Figure 4.10) revealed a curve for the corrected distances using the optimal GTR+ Γ model, with the deviation from a linear increase beginning at an uncorrected (p) distance of approximately 0.03 for both the Achatinidae only and including the subulinid outgroup taxon. Both plots were still increasing and had not reached a plateau, implying that the histone 3 had not reached saturation.



(A)

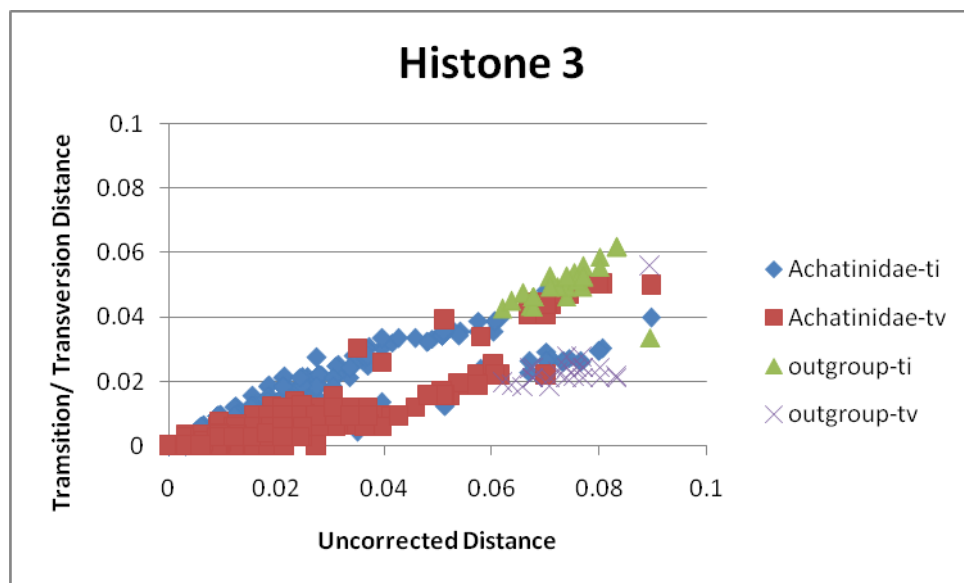


(B)

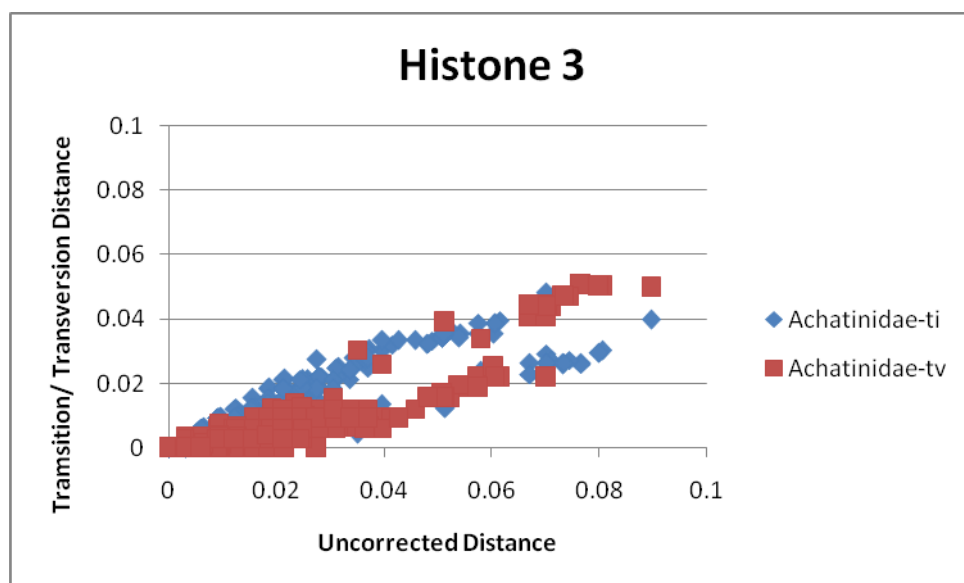
Figure 4.10: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

For plot 2, transitions and transversions were increasing and did not curve for both the Achatinidae only and including the outgroup taxon. Transitions were also generally higher than transversions. However, both the transition and transversion lines appeared to each form two separate lines beginning at approximately 0.035 uncorrected distance in both plots (Figure 4.11). Plot 3 (Figure 4.12) supported the observations from Plot 2, with several pairwise comparisons exhibiting higher transversions than transitions.

The g_i value for the histone 3 gene based on 28 taxa and 27 parsimony-informative sites was -0.900, significantly smaller than the critical value of -0.16 based on 25 taxa and 10 parsimony-informative sites. The histone 3 gene therefore exhibited phylogenetic signal.

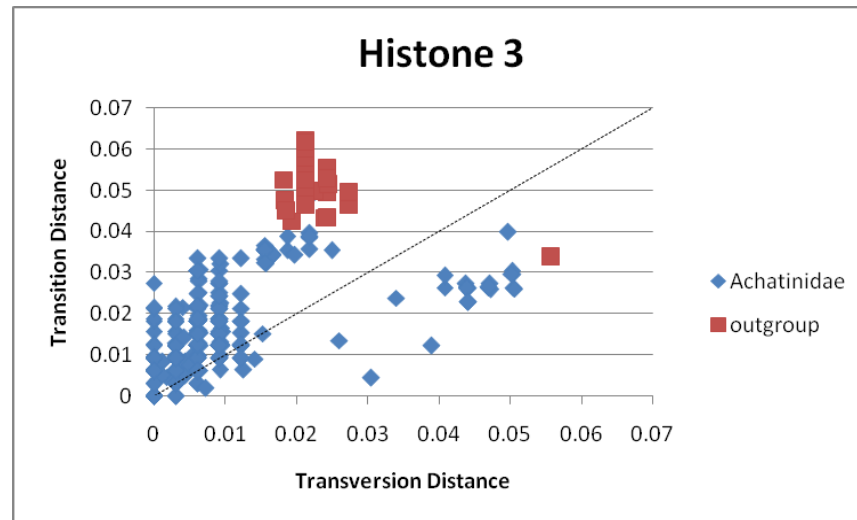


(A)

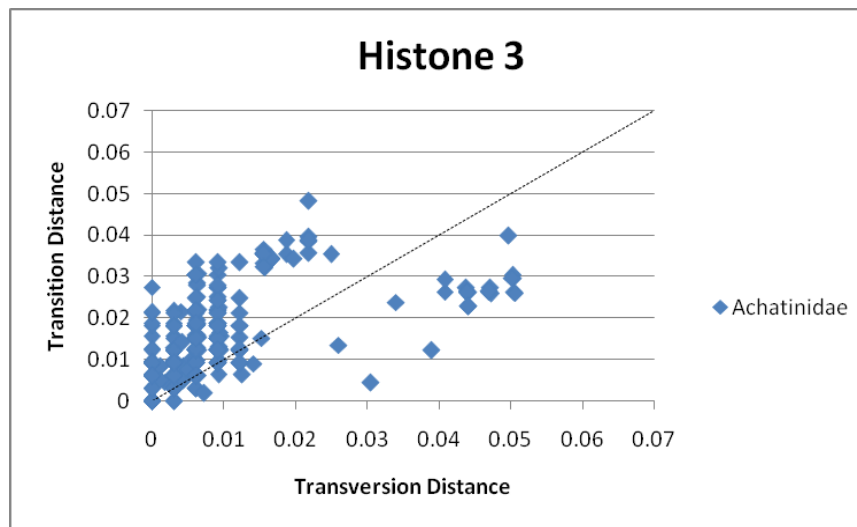


(B)

Figure 4.11: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



(A)

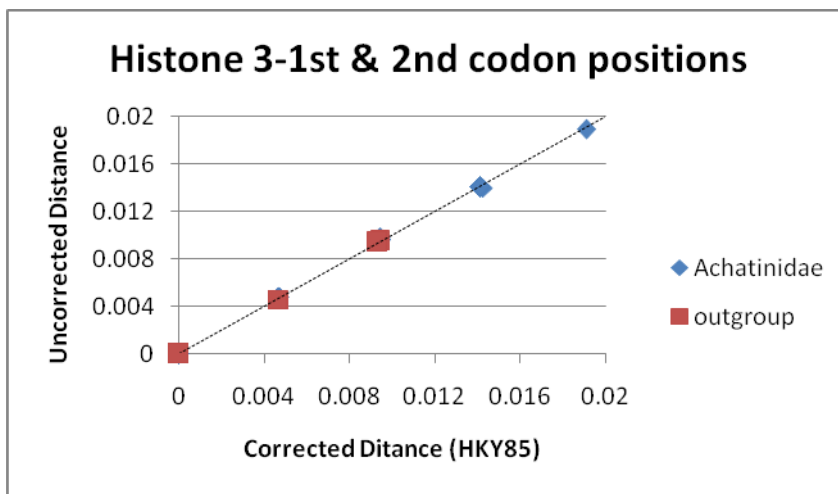


(B)

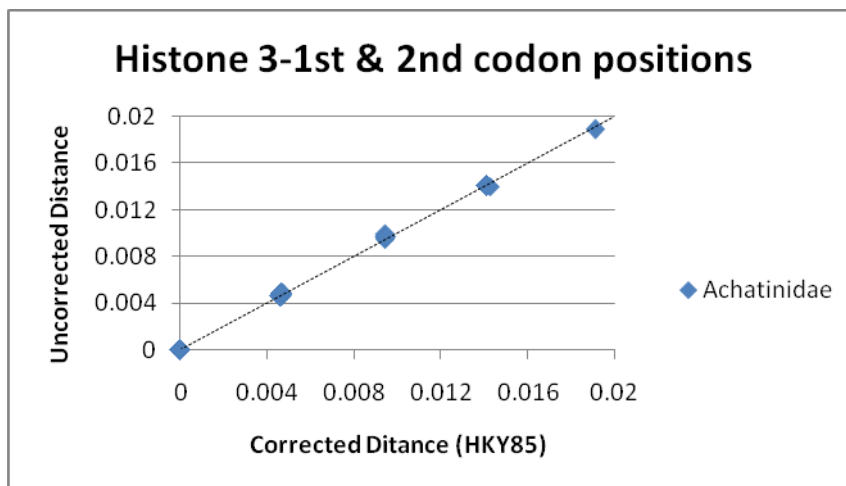
Figure 4.12: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

To account for the distinct lines that separated from the main transition and transversion lines observed in Figure 4.9, to determine if these observations were restricted to the 3rd codon positions, and whether such observations could be associated with saturation, separate plots were made for the combined 1st and 2nd codon positions as well as the 3rd codon positions.

For the 1st and 2nd codon positions, Plot 1 (Figure 4.13) showed a direct relationship between the corrected distances based on the optimal HKY85 model and the uncorrected distances, signifying adequate correction of the model and the absence of saturation. Note, however, that the distances were computed based only on four variable sites.



(A)

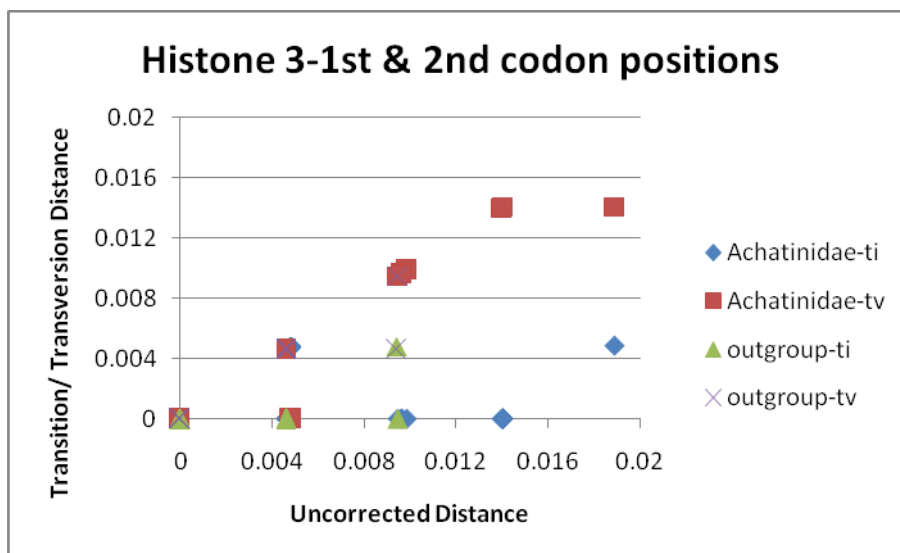


(B)

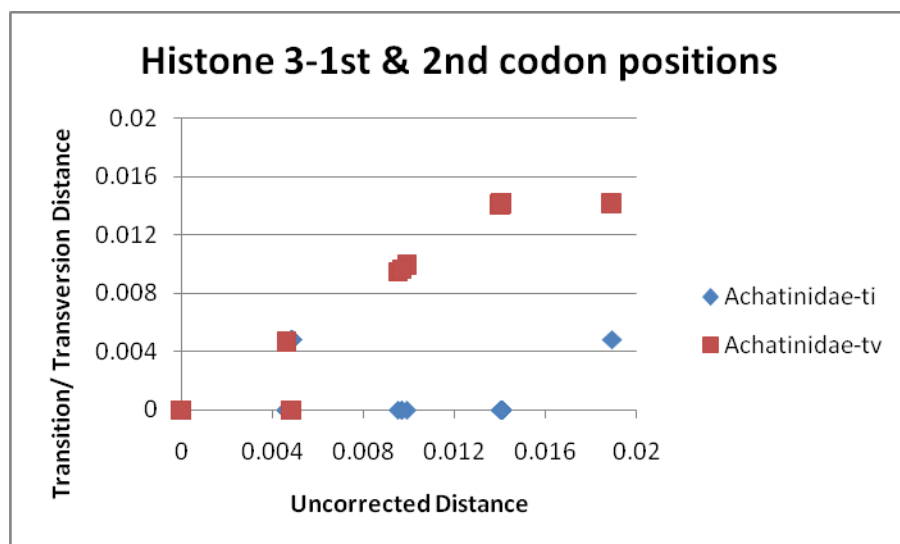
Figure 4.13: Plots of pairwise uncorrected distance versus corrected (HKY85) distance for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

For Plot 2 (Figure 4.14), transversion distances overtook the transition distances, and this was also evident in Plot 3 (Figure 4.15) where there were more transversions than transitions. However, the limited number of variable sites in the 1st and 2nd codon positions made it difficult to assess the presence of saturation through the curving of the plots.

The g1 value for the 1st and 2nd codon positions of histone 3 gene based on 28 taxa and one parsimony-informative site was -0.204. No critical value was available for 25 taxa and at least 2 parsimony-informative sites. The presence of phylogenetic signal could not be assessed based on one parsimony-informative site.

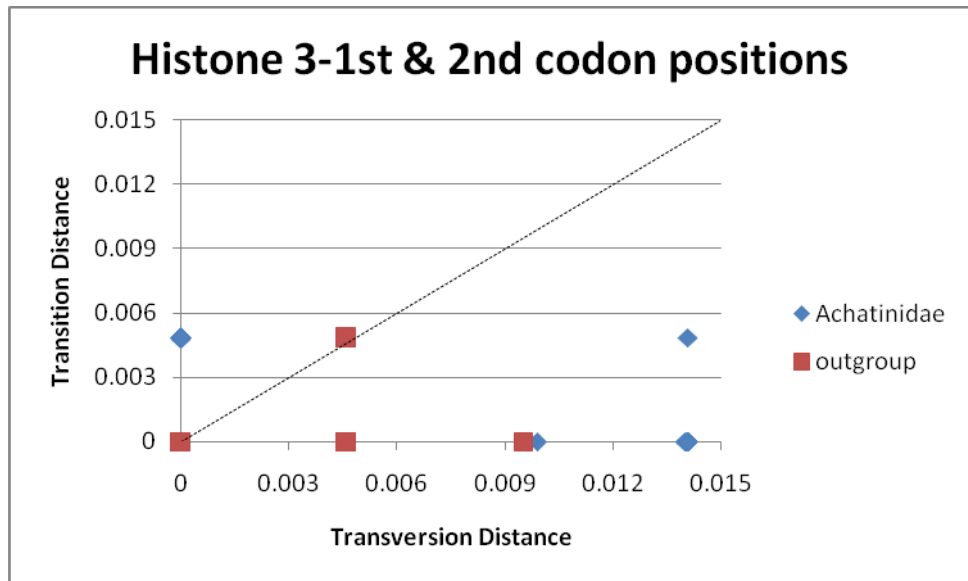


(A)

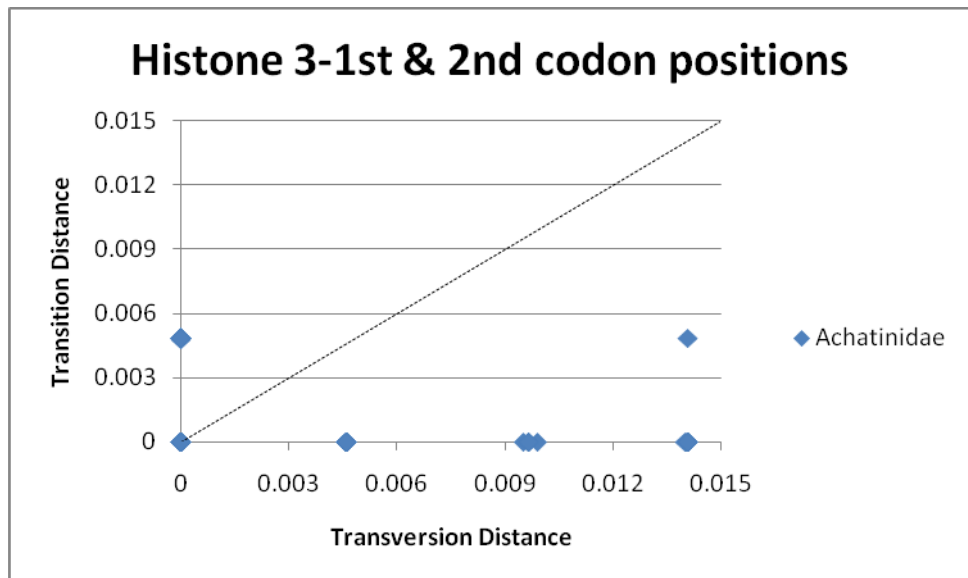


(B)

Figure 4.14: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



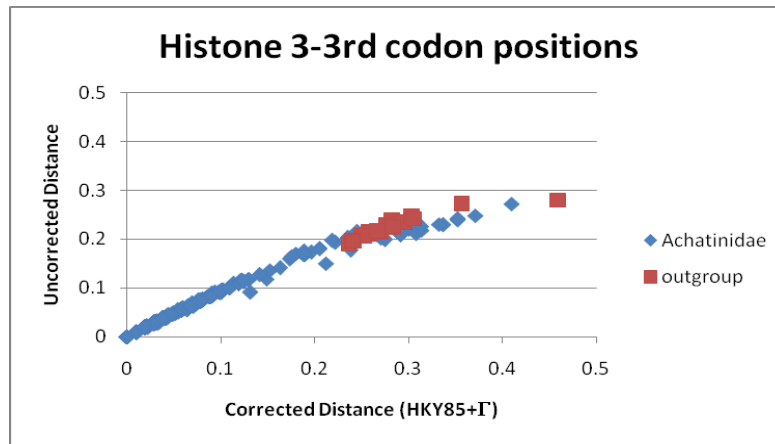
(A)



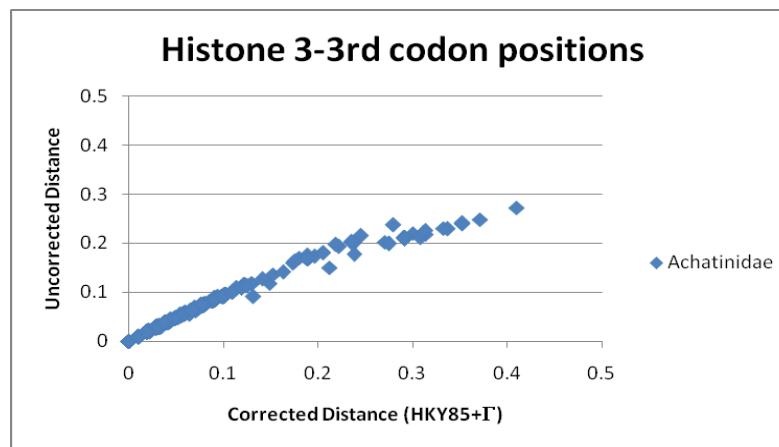
(B)

Figure 4.15: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

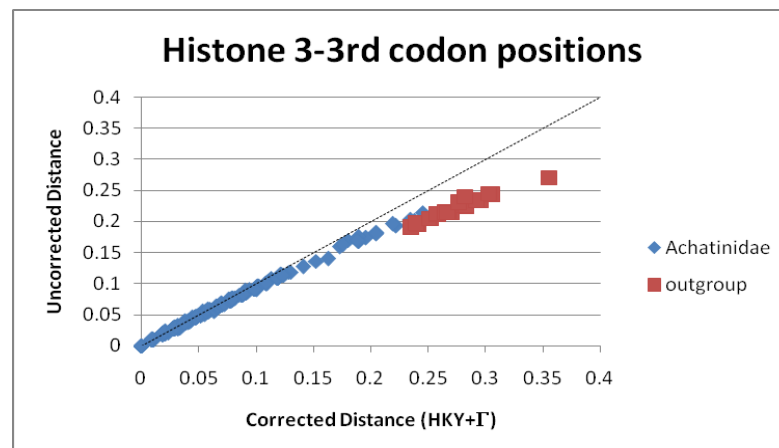
For the 3rd codon position of the histone 3 gene, Plot 1 (Figure 4.16) showed the corrected distances based on the optimal model HKY85+ Γ to be increasing but deviating from linearity at an uncorrected (p) distance of approximately 0.14. However, both plots showed an unmistakable second line starting at 0.13 corrected distance. This second line was attributed to the divergent sequence exhibited by *Archachatina marginata* as supported by the fact that the second line disappeared when *A. marginata* was removed from the plots (see Figure 4.16C). Nevertheless, no plateau was observed, suggesting that adequate correction was made by the optimal model and that the 3rd codon position was not saturated.



(A)



(B)

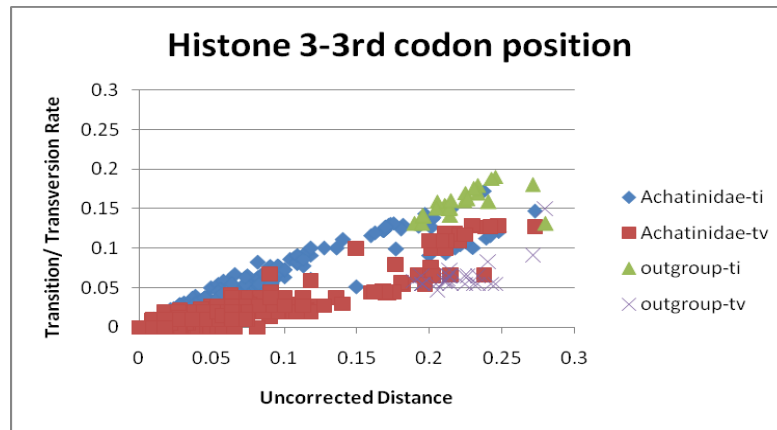


(C)

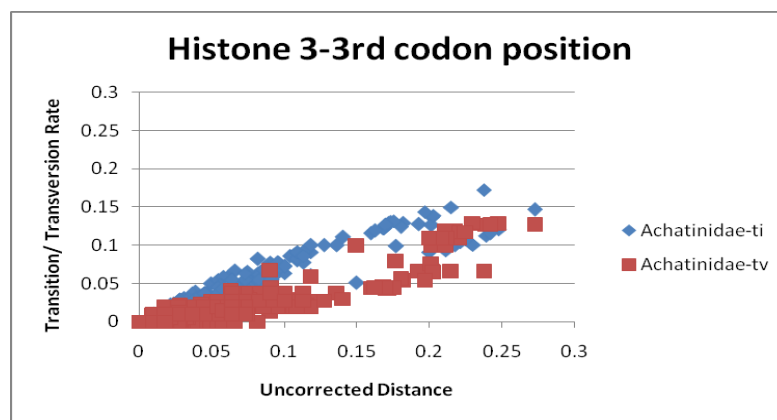
Figure 4.16: Plots of pairwise uncorrected distance versus corrected (HKY85+ Γ) distance for the 3rd codon position of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata*; (B) the Achatinidae only; and (C) Achatinidae and subulinid outgroup *Rumina decollata* but without *Archachatina marginata*.

For Plot 2 (Figure 4.17), the same observations were seen as in Figure 4.11 where both the transition and transversion lines each formed a distinct line beginning at approximately 0.09 uncorrected distance and resulting in a separate transversion line that was higher than the separate transition line in both the Achatinidae only and with the outgroup taxon. These separate lines were attributed to the divergent sequence of *Archachatina marginata*. When this sequence was excluded from the plots (see Figure 4.17C), it became clear that both the transition and transversion lines were increasing with no curving evident. Plot 3 (Figure 4.18) also showed majority of the pairwise comparisons with higher transitions than transversions. Those points with higher transversions than transitions involved *A. marginata* as these points disappeared when *A. marginata* was excluded from the plot (see Figure 4.18C). These results indicated that the 3rd codon position was not saturated.

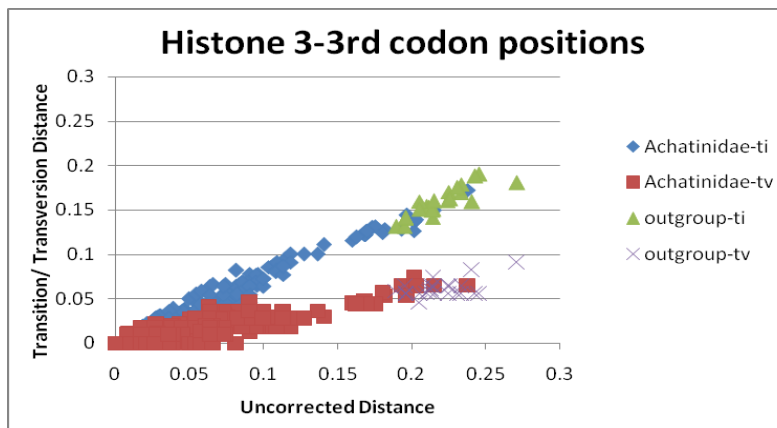
The g1 value for the 3rd codon position of the histone 3 gene based on 28 taxa and 26 parsimony-informative sites was -0.886, which was significantly smaller than the critical value of -0.16 based on 25 taxa and 10 parsimony-informative sites. The 3rd codon position of the histone 3 gene therefore exhibited phylogenetic signal.



(A)

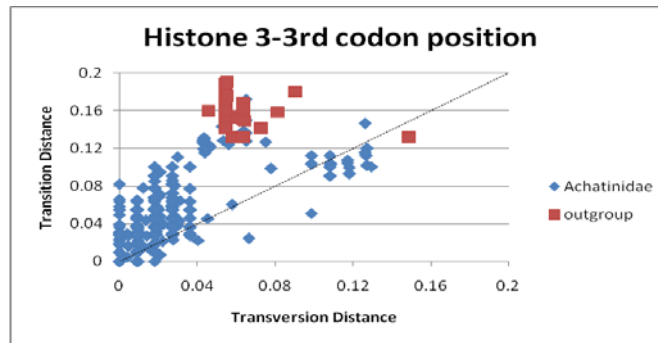


(B)

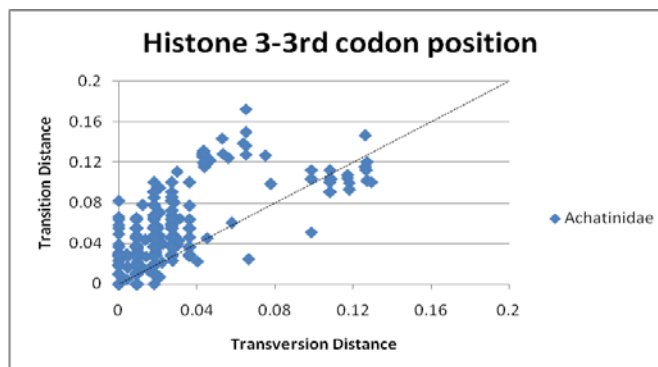


(C)

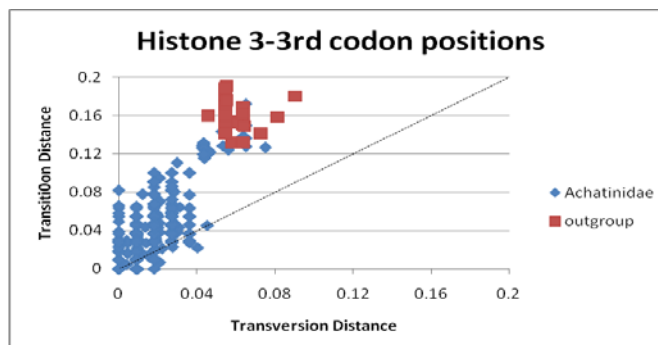
Figure 4.17: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3rd codon position of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata*; (B) the Achatinidae only; and (C) the Achatinidae and the subulinid outgroup *Rumina decollata* but excluding *Archachatina marginata*.



(A)



(B)

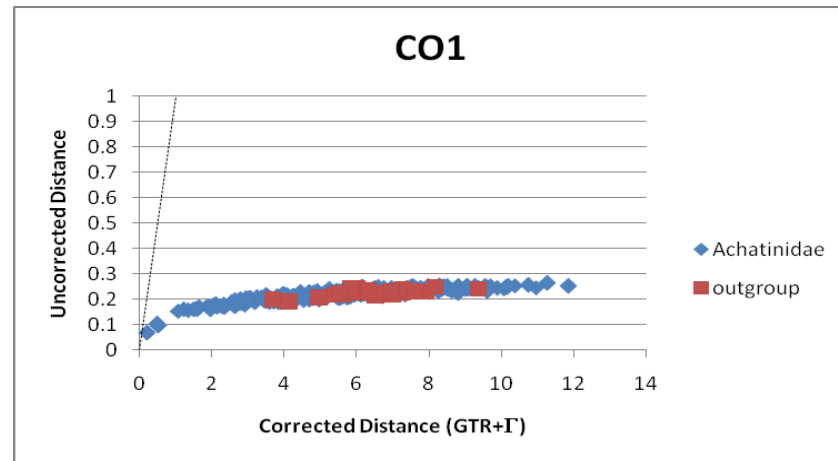


(C)

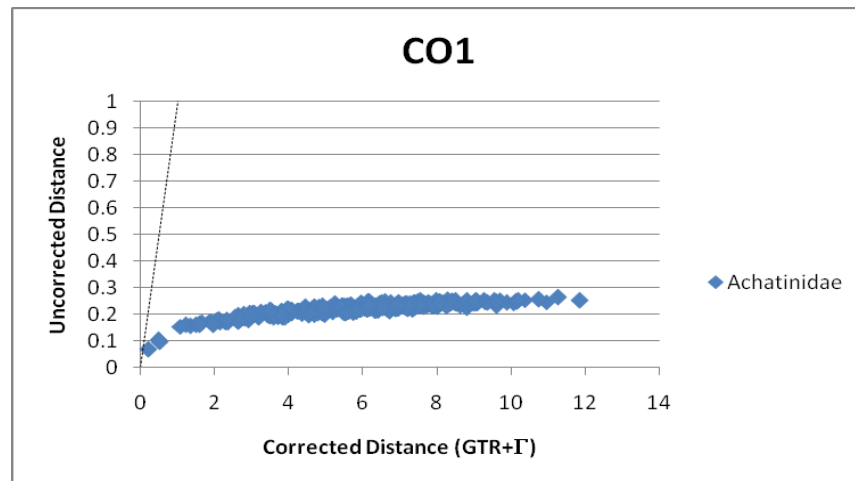
Figure 4.18: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3rd codon position of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata*; (B) the Achatinidae only; and (C) the Achatinidae and the subulinid outgroup *Rumina decollata* but excluding *Archachatina marginata*.

The findings suggested that the entire histone 3 dataset could be employed for the phylogenetic analyses of the Achatinidae, although the divergent sequence of *Archachatina marginata* should be taken into account when interpreting the trees.

For the COI gene, Plot 1 (Figure 4.19) yielded an almost horizontal trend for all taxa and for the Achatinidae only. This, along with corrected distances of up to 12, indicated severe saturation for the COI gene and the optimal GTR+ Γ model was unable to correct the dataset for multiple hits.



(A)

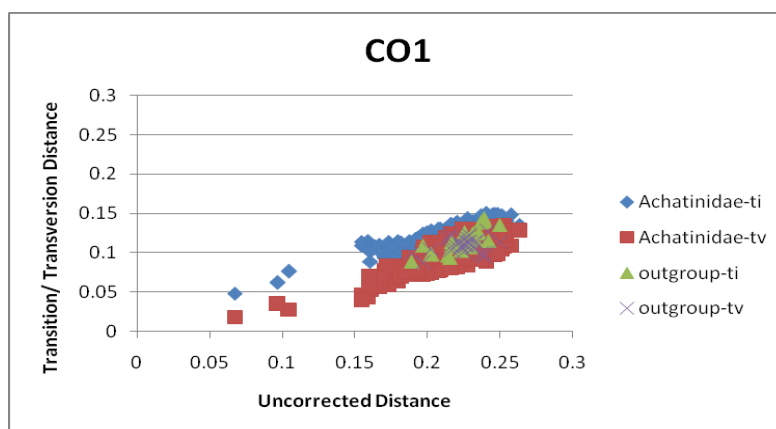


(B)

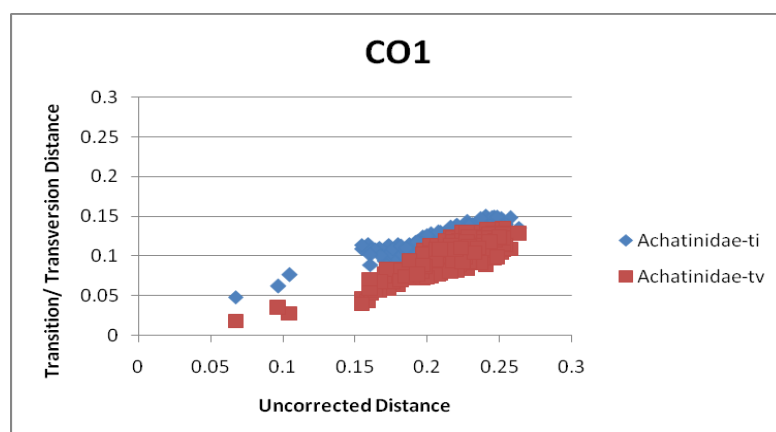
Figure 4.19: Plots of pairwise uncorrected distance against corrected distance for the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Plot 2 (Figure 4.20) for all taxa as well as the Achatinidae only revealed transversions overlapping with transitions. Plot 3 (Figure 4.21) showed approximately a quarter of all pairwise distances showing transversions that were higher than transitions.

The g1 test score for the COI gene based on 28 taxa and 276 parsimony-informative sites was -0.507, significantly smaller than the critical value of -0.08 for 25 taxa and 250 parsimony-informative sites. The g1 test suggested some phylogenetic information could be generated from the dataset.

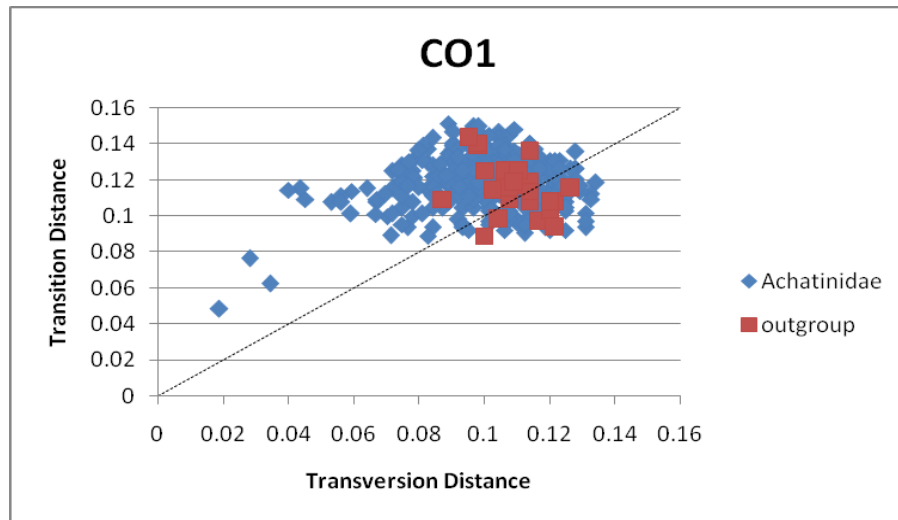


(A)

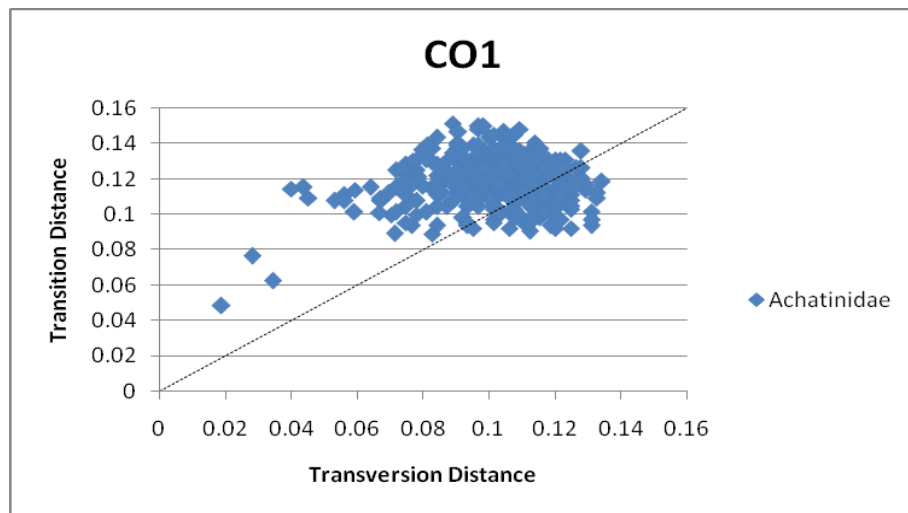


(B)

Figure 4.20: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



(A)

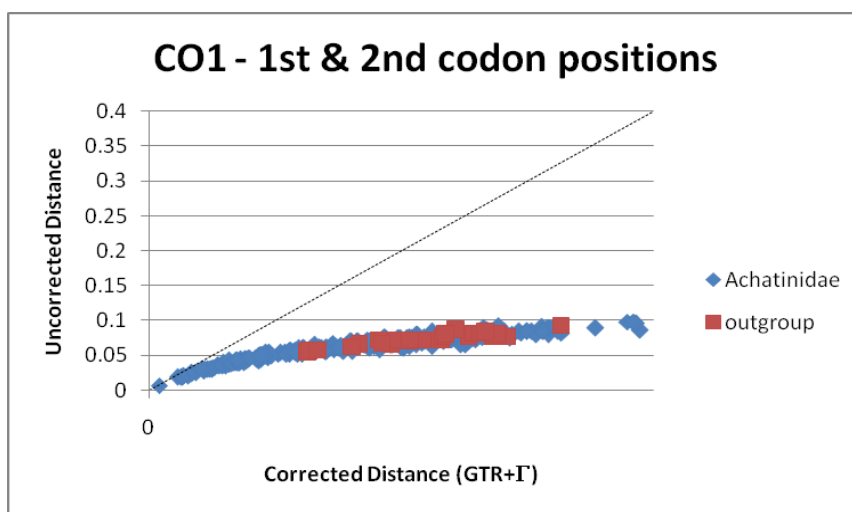


(B)

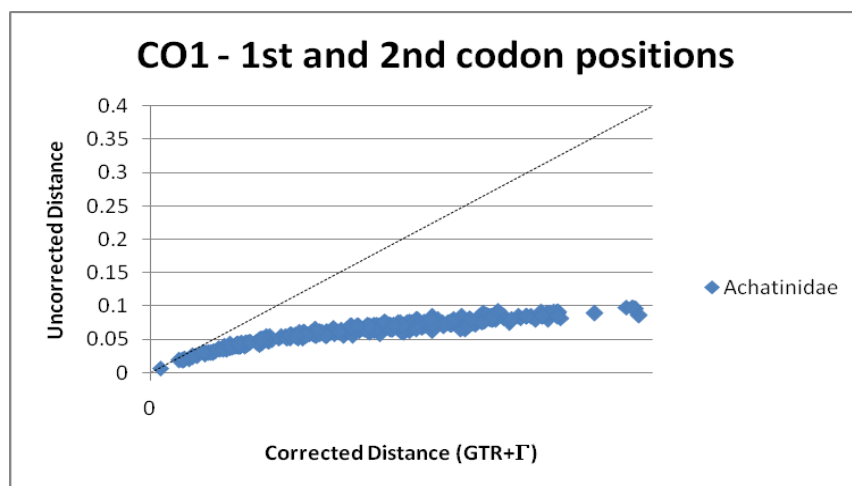
Figure 4.21: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Despite the presence of phylogenetic signal, the problem of saturation remained as shown in Plot 1. In order to evaluate whether saturation of the COI gene was restricted to 3rd codon positions, the combined 1st and 2nd codon positions and the 3rd codon positions were evaluated separately.

For the 1st and 2nd codon positions of the COI gene, Plot 1 (Figure 4.22) revealed a curve for the corrected distances using the optimal GTR+ Γ model, with the deviation from linearity beginning at an uncorrected (p) distance of approximately 0.03 for both the Achatinidae only and including the outgroup taxon. The plots were still increasing slightly and had not reached a plateau, implying that the 1st and 2nd codon positions of COI had not yet reached saturation.



(A)

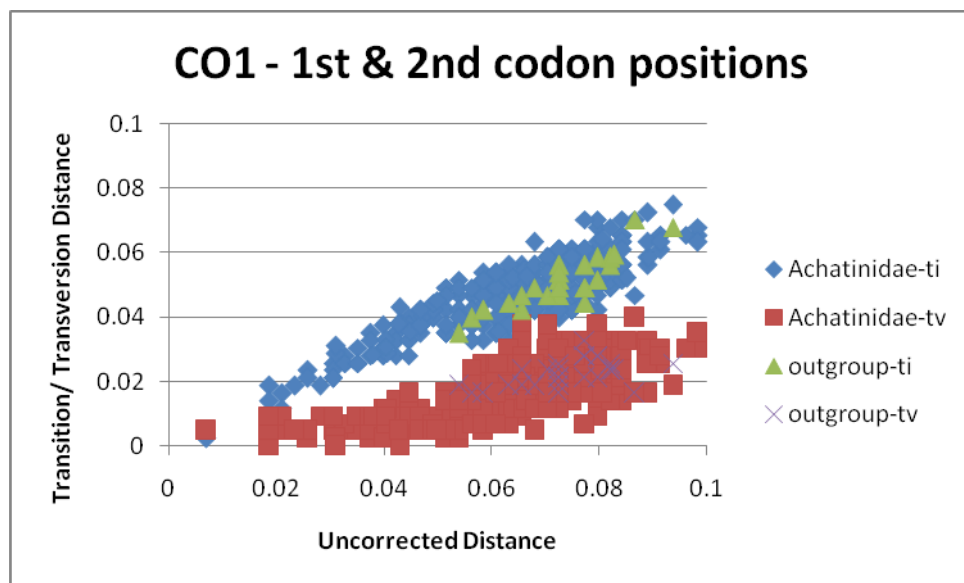


(B)

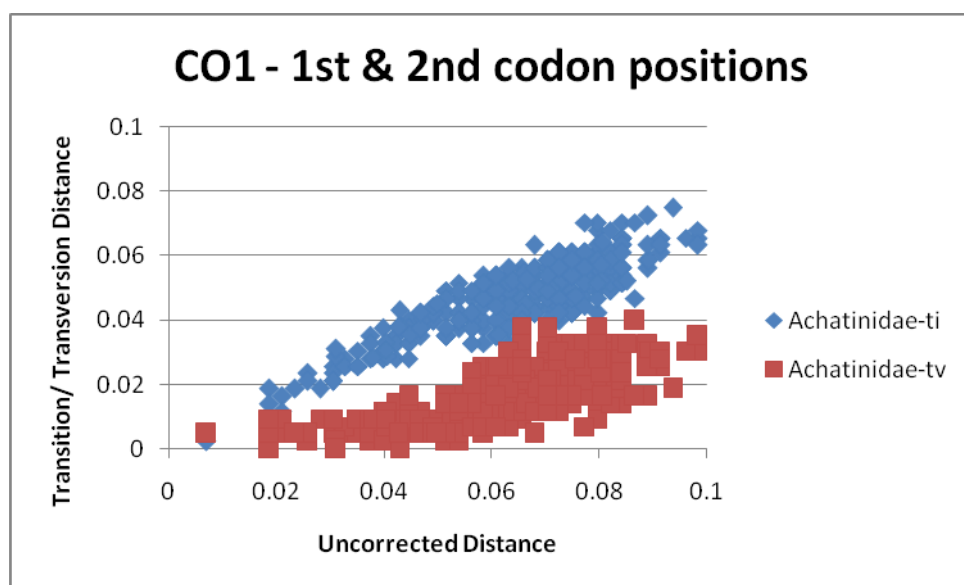
Figure 4.22: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 1st and 2nd codon positions of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Plot 2 (Figure 4.23) for the 1st and 2nd codon positions of the COI gene revealed a linear increase of both transitions and transversions for the Achatinidae and when the outgroup taxon was included, with the transitions being higher than the transversions, while plot 3 (Figure 4.24) revealed that all but three pairwise comparisons had higher rates of transversions than transitions. These findings suggested that the 1st and 2nd codon positions were not saturated.

A g1 value of -0.309 was obtained based on 28 taxa and 64 parsimony-informative sites, which was significantly smaller than the critical value of -0.12 based on 25 taxa and 50 parsimony-informative sites. The 1st and 2nd codon positions of the COI gene therefore exhibited phylogenetic signal.

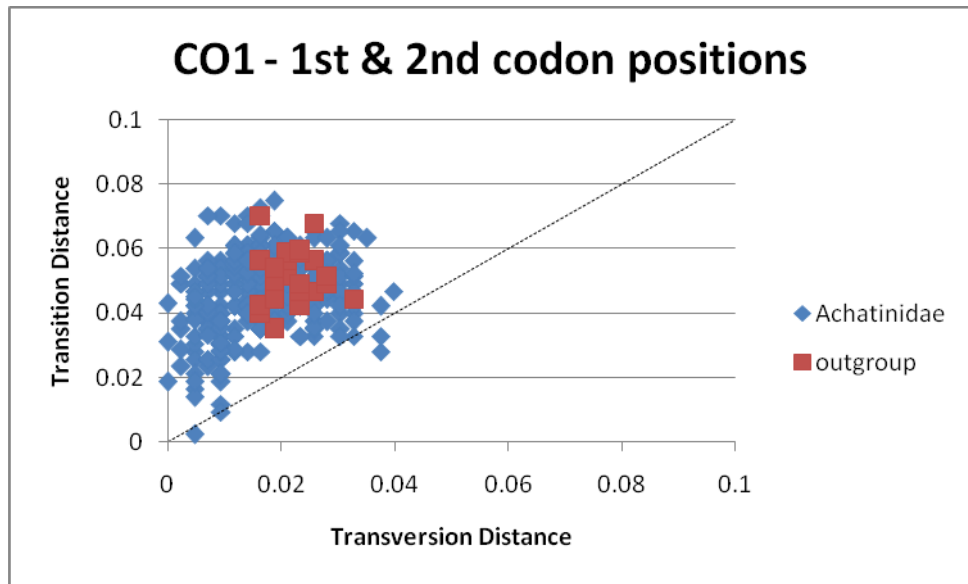


(A)

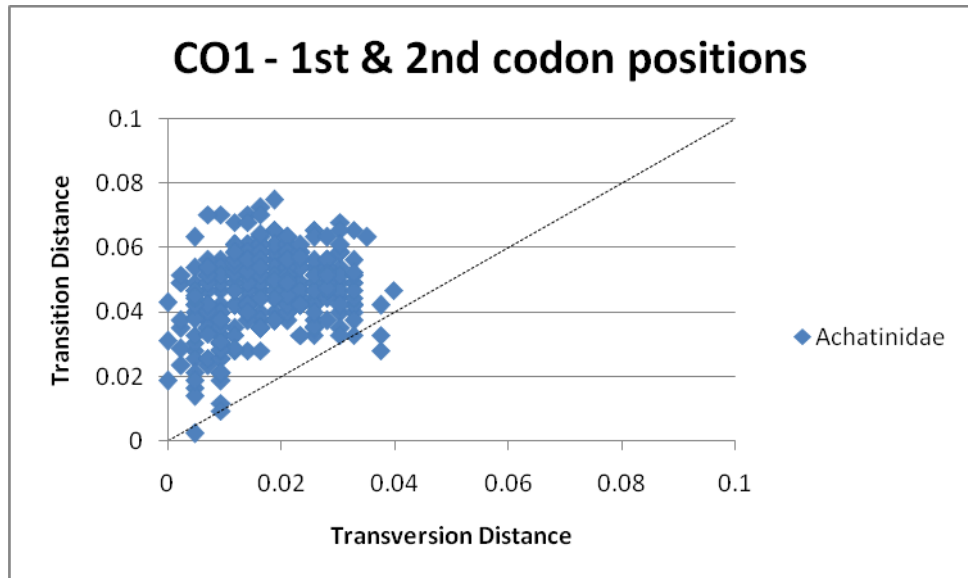


(B)

Figure 4.23: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1st and 2nd codon positions of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



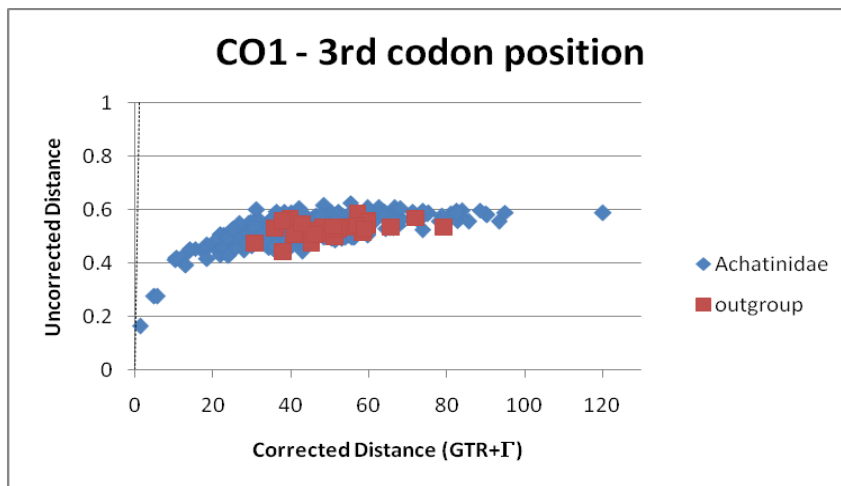
(A)



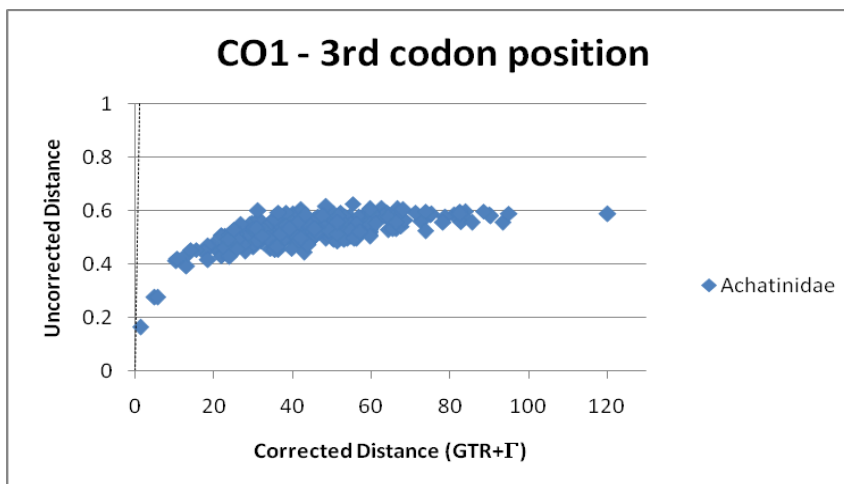
(B)

Figure 4.24: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1st and 2nd codon positions of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

For the 3rd codon position of the COI gene, Plot 1 (Figure 4.25) showed that the optimal GTR+ Γ model was unable to correct the dataset for multiple hits as demonstrated by the nearly horizontal trend for both plots. The uncorrected distances also ranged from 0.164 to 0.625 while corrected distances ranged from 1.303 to 120.141 (see also Table 4.1). This indicated severe saturation for the 3rd codon position.



(A)

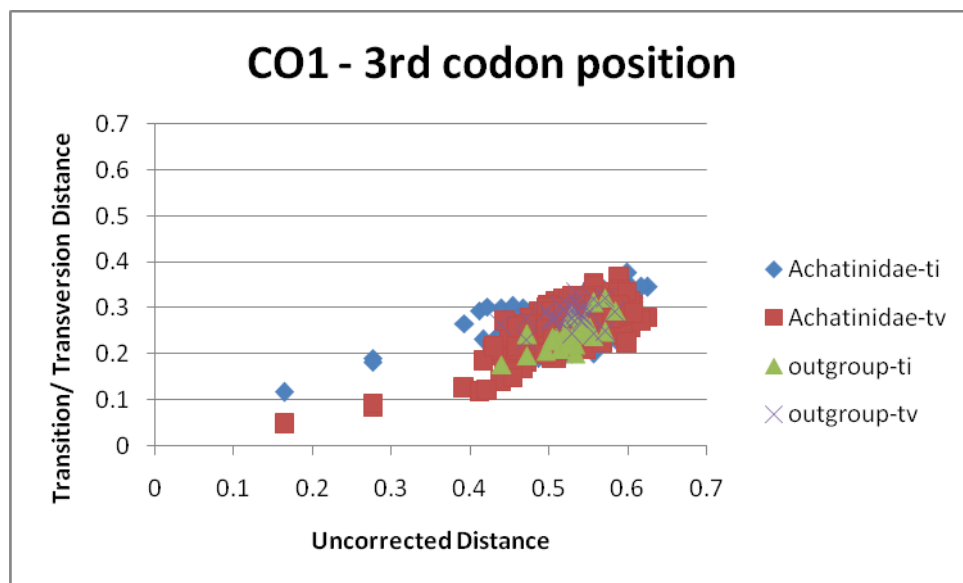


(B)

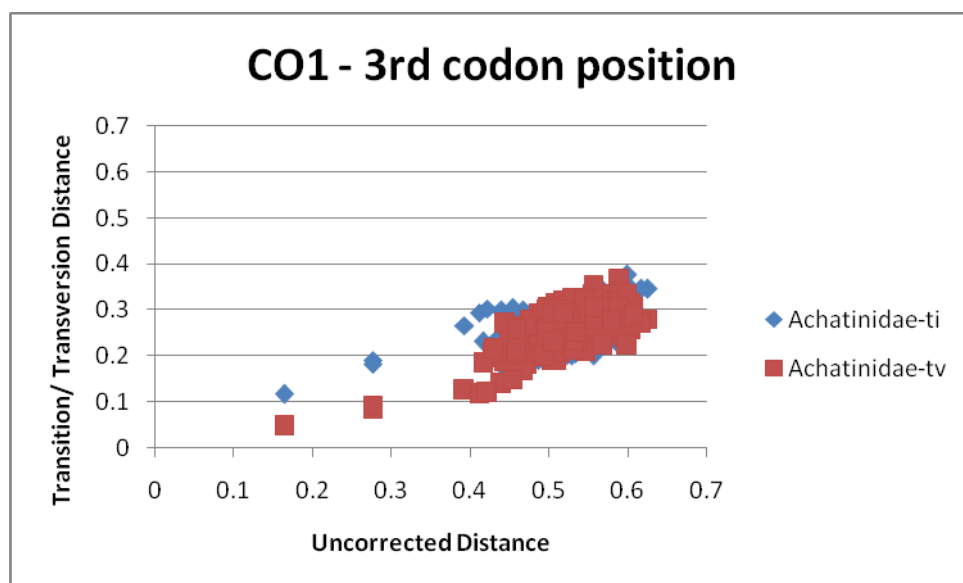
Figure 4.25: Plots of pairwise uncorrected distance against corrected distance for the 3rd codon position of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Plot 2 (Figure 4.26) for all taxa as well as for the Achatinidae only revealed transversions overlapping with transitions. Plot 3 (Figure 4.27) demonstrated that approximately half of the pairwise comparisons had higher rates of transversions than transitions.

The g_1 test score for the COI gene based on 28 taxa and 212 parsimony-informative sites was -0.457, significantly smaller than the critical value of -0.1 for 25 taxa and 100 parsimony-informative sites. The 3rd codon position therefore exhibited phylogenetic signal.

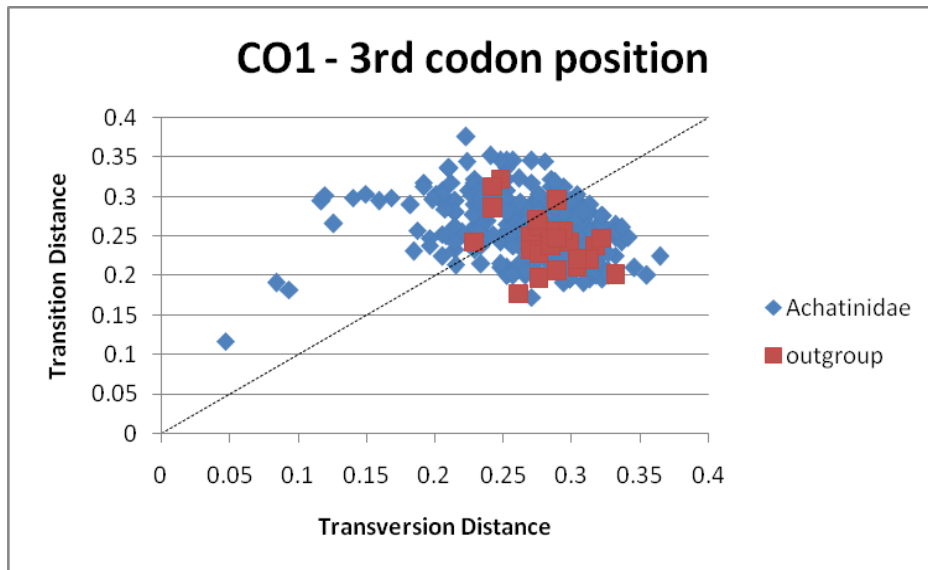


(A)

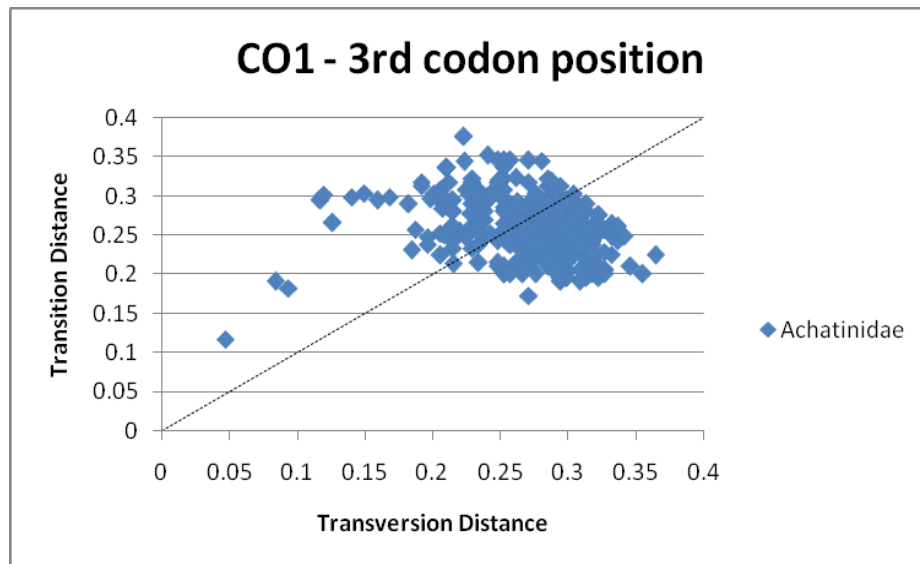


(B)

Figure 4.26: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3rd codon position of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



(A)

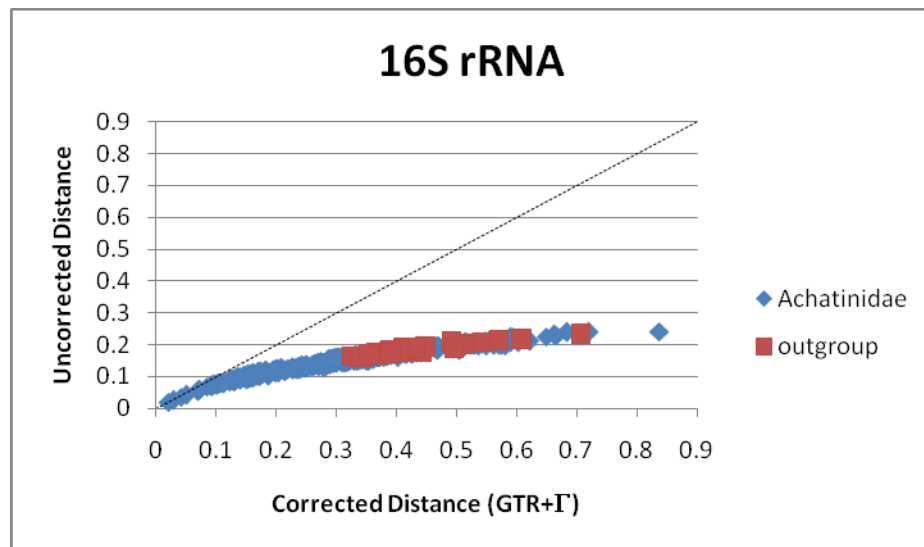


(B)

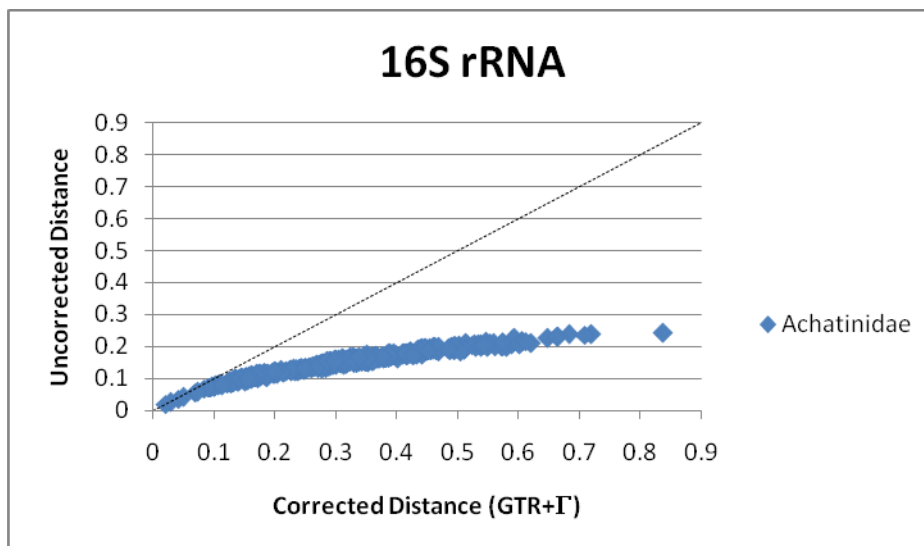
Figure 4.27: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3rd codon position of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Despite the presence of phylogenetic signal as indicated by the g1 test, the strong evidence for saturation of the entire COI gene suggested that the gene should not be used in its entirety for phylogenetic analyses. The results also revealed that saturation was restricted to the 3rd codon position despite showing phylogenetic signal based on the g1 test. These contradictory results from the saturation tests and the g1 test are most likely due to the presence of taxa with very similar sequences that led to a strong left skew of tree length distribution based on parsimony in the g1 test. In these instances, the saturation tests took precedence over the g1 test in deciding whether or not datasets should be included. Phylogenetic analyses for the COI gene were therefore limited to the 1st and 2nd codon positions only.

For the 16S rRNA gene, Plot 1 (Figure 4.28) produced corrected distances based on the optimal GTR+ Γ model that curved from linearity at an uncorrected (p) distance of 0.04 but was still increasing and had not reached a plateau. These findings did not indicate any saturation.



(A)



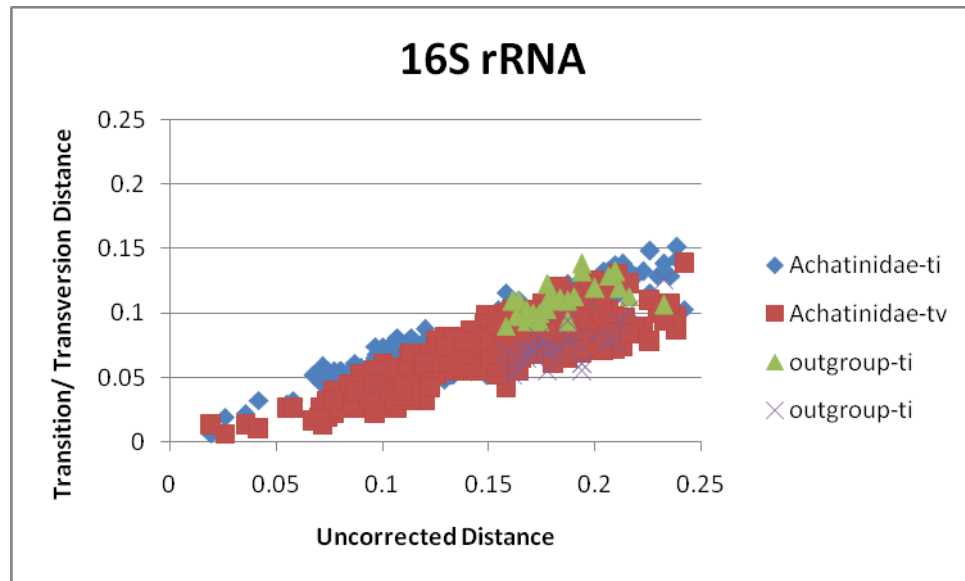
(B)

Figure 4.28: Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the 16S rRNA gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

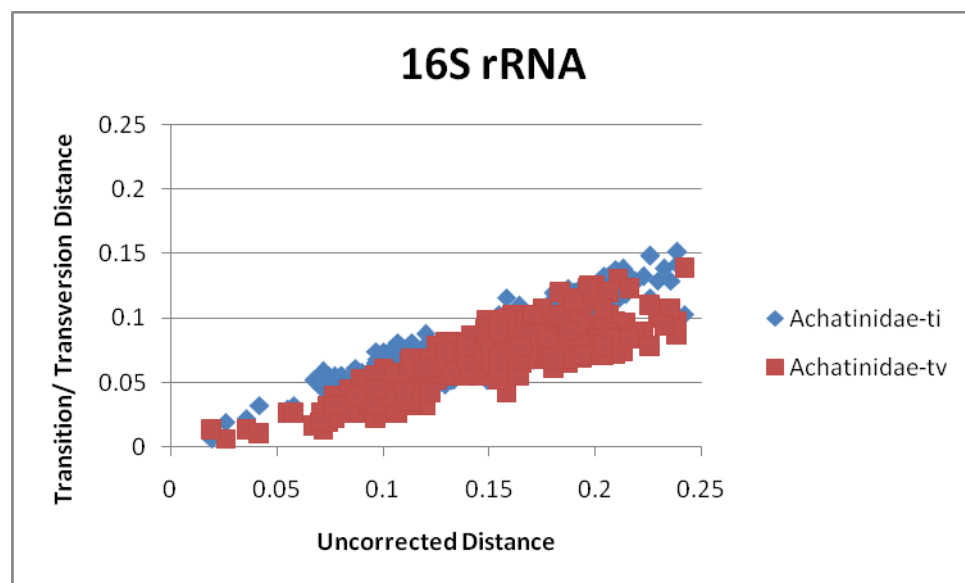
Plot 2 (Figure 4.29) showed an unmistakable overlap between transitions and transversions for the Achatinidae only and including the streptaxid outgroup. However, both plots were still increasing with no evidence of curving for either transitions or transversions. Plot 3 (Figure 4.30) revealed that many pairwise comparisons had higher rates of transversions than transitions, although more than half of the transitions were still higher than transversions.

A g_i value of -0.507 was obtained based on 29 taxa and 110 parsimony-informative sites, which was significantly smaller than the critical value of -0.1 at $p=0.05$ for 25 taxa and 100 parsimony-informative sites. Phylogenetic signal was therefore evident in the 16S rRNA gene.

The lack of evidence for saturation and the presence of strong phylogenetic signal implied that the 16S rRNA gene could be used to infer the phylogenetic relationship of the Achatinidae.

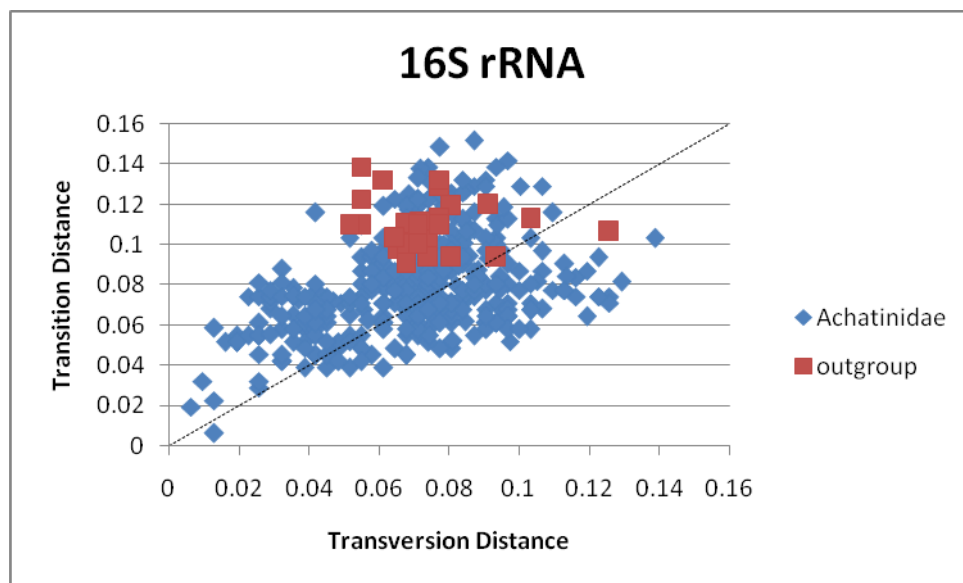


(A)

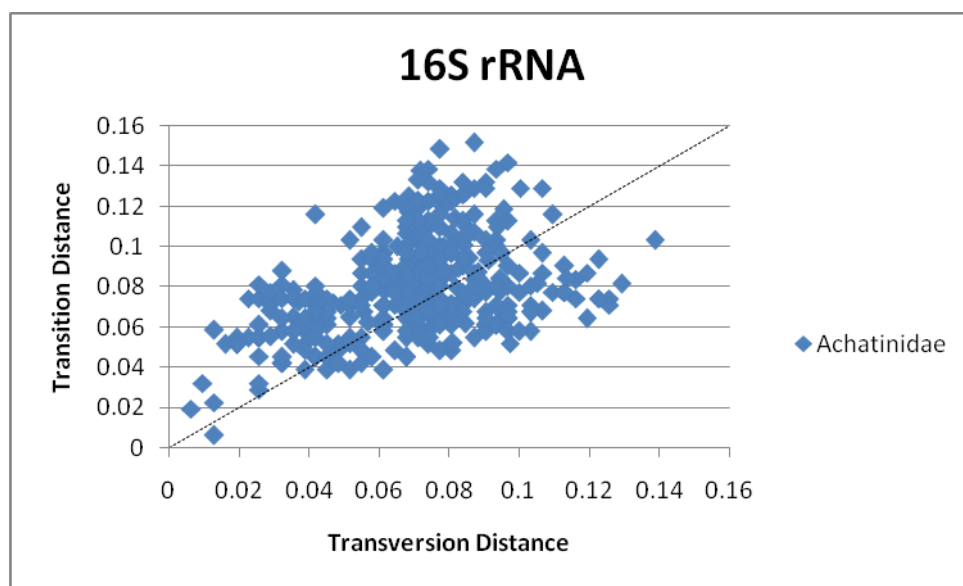


(B)

Figure 4.29: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 16S rRNA gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



(A)



(B)

Figure 4.30: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

4.3.2.2. Inferring the presence of paralogous sequences in the actin dataset and assessing its utility for phylogenetic analyses

To infer the presence of paralogous sequences, the actin sequences (obtained by direct sequencing of PCR products) were examined for ambiguous sites. Ambiguous sites were detected in 22 out of 28 achatinid taxa (Appendix 4.7, p. 509), which implied the presence of more than one copy of the actin gene in the Achatinidae. The presence of multiple actin copies was further supported when seven distinct cloned sequences were observed for *Cochlitoma granulata* (see Appendix 3.8, pp. 435-437; note that the clone data for *Cochlitoma granulata* is also discussed as part of the discussion on paralogy of the actin gene in the Achatinoidea, Section 3.3.2.2, Chapter 3, pp. 155-159). A total of 72 variable sites were detected among the seven clones as opposed to only 9 ambiguities in the direct sequence. The majority of these variable sites (55) were attributed to Clones 3 and 6 that were not picked up by the direct sequence. As discussed in chapter 3, the reason for the failure of the direct sequences to identify as ambiguous sites all of the variation among the clones is not clear, but as the PCR-direct sequence is effectively a consensus of the different actin copies within an individual, it is possible that rare copies of the gene might be averaged out.

Four approaches were carried out to determine whether the observed paralogy in actin was likely to mislead phylogeny. First, a neighbor-joining tree of all the actin sequences and the clones of *C. granulata* was generated to determine if the clones were monophyletic. As discussed previously in Section 3.3.2.2 of Chapter 3, the results revealed that some clones (Clones 1, 2, 4 and 5) formed a monophyletic group together with the direct sequence for *C. granulata* while others (Clones 3 and 6) fell elsewhere in the tree (see also Appendix 3.9, p. 438). Second, pairwise comparison between closely related taxa across the Achatinidae based on actin sequence identity (i.e. least number of

nucleotide differences) showed that for some pairwise comparisons, the number of ambiguous sites for at least one taxon in the pair were either very close to the number of total nucleotide differences (i.e. *Achatina* (*Achatina*) *achatina* and *Archachatina marginata*; *Achatina* (*Achatina*) *damarensis* and *Metachatina kraussi*; and *Cochlitoma montistempli* and *C. omissa*) or were larger than the total differences (i.e. *Achatina* (*Achatina*) *damarensis* and *Atopocochlis exarata*; *C. dimidiata* and *Atopocochlis exarata*; *C. varicosa* and *C. sp. cf. vestita*; *Limicolaria kambeul* and *L. martenssii*; and *Limicolariaopsis sp.* and *Achatina* (*Achatina*) *damarensis*). (See also Table 4.3.) These observations suggested that the divergence among the actin copies within some taxa were just as deep if not deeper than the divergence of the taxa themselves and that phylogenetic signal could be obscured at and above the genus level for these taxa. Third, a partition homogeneity test was supposed to evaluate whether or not the actin sequences exhibited significant difference in the phylogenetic signal with respect to the other genes. However, the test was not successfully carried out due to the polytomies exhibited by the actin gene using maximum parsimony. It was therefore not possible to assess the utility of the actin sequence for phylogenetic analyses in the Achatinidae using the partition homogeneity test. Last, the actin phylogeny was evaluated for concordance with the rRNA gene cluster phylogeny, and results showed little concordance. For example, *Cochlitoma marinae* had a very divergent sequence relative to the other achatinids, particularly the other Southern African *Cochlitoma* species; it fell at the base of the Achatinidae (see Appendix 4.8 on p. 510 for the actin phylogenies and Figure 4.31A on p. 252 for the rRNA gene phylogeny). Based on these results, the presence of multiple copies of the actin gene was having a significant effect on phylogeny; the actin gene was therefore excluded from further phylogenetic analyses.

Table 4.3. Comparison of the actin ambiguous sites and nucleotide differences for each achatinid and its closest relative. Closest relative for each taxon is determined based on the actin sequence identity (least number of nucleotide differences). Note that in scoring for the total differences, comparison between an ambiguous site and an unambiguous site is counted as different even if one of the possible nucleotides for the ambiguous site is the same as that found in the unambiguous site being compared (e.g. A vs. R, which is either A or G). See also Table 2.8 of p. 73 for assignment of ambiguous positions.

Species (total ambiguous sites)	Closest relative with least number of differences (total ambiguous sites)	Total number of nucleotide differences
WEST AFRICAN		
<i>Achatina (Achatina) achatina</i> (45)	<i>Archachatina marginata</i> (27)	48
<i>Archachatina marginata</i> (27)	<i>Achatina (Achatina) achatina</i> (45)	48
EAST AFRICAN		
<i>Achatina (Lissachatina) fulica</i> (18)	<i>Achatina (Lissachatina) immaculata</i> (15)	27
<i>Achatina (Lissachatina) immaculata</i> (15)	<i>Achatina (Lissachatina) fulica</i> (18)	27
<i>Achatina (Lissachatina) reticulata</i> (33)	<i>Achatina (Euaethiopina) loveridgei</i> (0)	39
<i>Achatina (Lissachatina) zanzibarica</i> (1)	<i>Achatina (Euaethiopina) loveridgei</i> (0)	13
<i>Achatina (Euaethiopina) loveridgei</i> (0)	<i>Achatina (Lissachatina) zanzibarica</i> (1)	13
OTHERS		
<i>Achatina (Achatina) bisculpta</i> (32)	<i>Limicolaria kambeul</i> (30)	49
<i>Achatina (Achatina) damarensis</i> (34)	<i>Atopocochlis exarata</i> (29)	33
<i>Achatina (Achatina) stuhlmanni</i> (2)	<i>Cochlitoma churchilliana</i> (0) <i>Cochlitoma montisptempli</i> (0) <i>Cochlitoma ustulata</i> (0) <i>Cochlitoma zebra</i> (0)	29
<i>Atopocochlis exarata</i> (29)	<i>Achatina (Achatina) damarensis</i> (34)	33
<i>Limicolaria kambeul</i> (30)	<i>Limicolaria martenssii</i> (33)	31
<i>Limicolaria martenssii</i> (33)	<i>Limicolaria kambeul</i> (30)	31
<i>Limicolariaopsis</i> sp. (40)	<i>Achatina (Achatina) damarensis</i> (34)	42
SOUTHERN		
<i>Cochlitoma varicosa</i> (27)	<i>Cochlitoma</i> sp. cf. <i>vestita</i> (30)	18
<i>Cochlitoma dimidiata</i> (31)	<i>Atopocochlis exarata</i> (29)	33
<i>Cochlitoma</i> sp. cf. <i>vestita</i> (30)	<i>Cochlitoma varicosa</i> (27)	18
<i>Cochlitoma marinae</i> (4)	<i>Cochlitoma semidecussata</i> (0)	124
<i>Cochlitoma churchilliana</i> (0)	<i>Cochlitoma zebra</i> (0)	8
<i>Cochlitoma kilburni</i> (1)	<i>Cochlitoma zebra</i> (0)	5
<i>Cochlitoma granulata</i> (9)	<i>Cochlitoma omissa</i> (6) <i>Cochlitoma semidecussata</i> (0) <i>Cochlitoma ustulata</i> (0)	15
<i>Cochlitoma simplex</i> (2)	<i>Cochlitoma zebra</i> (0)	10
<i>Cochlitoma montisptempli</i> (0)	<i>Cochlitoma omissa</i> (6)	5
<i>Cochlitoma omissa</i> (6)	<i>Cochlitoma montisptempli</i> (0)	5
<i>Cochlitoma semidecussata</i> (0)	<i>Cochlitoma ustulata</i> (0)	11
<i>Cochlitoma ustulata</i> (0)	<i>Cochlitoma churchilliana</i> (0) <i>Cochlitoma semidecussata</i> (0) <i>Cochlitoma zebra</i> (0)	11
<i>Cochlitoma zebra</i> (0)	<i>Cochlitoma kilburni</i> (1)	5
<i>Metachatina kraussi</i> (52)	<i>Achatina (Achatina) damarensis</i> (34)	59
SUBULINIDAE (OUTGROUP)		
<i>Rumina decollata</i> (29)	<i>Cochlitoma churchilliana</i> (0)	82

4.3.2.3. Partition homogeneity test for combining datasets

A partition homogeneity test was done separately for the dataset incorporating the rRNA cluster, histone 3, the 1st and 2nd codon positions of the COI, and the 16S rRNA (5586 nucleotides total) for all taxa with complete DNA sequences as well as the dataset that excluded actin (4713 total). However, PAUP* version 4.0b10 was unable to compute the *P* value for 1000 tree replicates using maximum parsimony. This was due to the presence of polytomies in the topology of the trees generated by maximum parsimony that severely slowed down PAUP. It was therefore not possible to assess whether the gene sequences could be combined as a single dataset using the partition homogeneity test. The decision to combine, therefore, relied on whether the single gene phylogenies exhibited some degree of concordance with each other, which they did for some groups of taxa based on the results in Section 4.3.3.1, pp. 259-261. On this basis, the rRNA cluster, histone 3, the 1st and 2nd codon positions of the COI, and the 16S rRNA were combined into a single dataset.

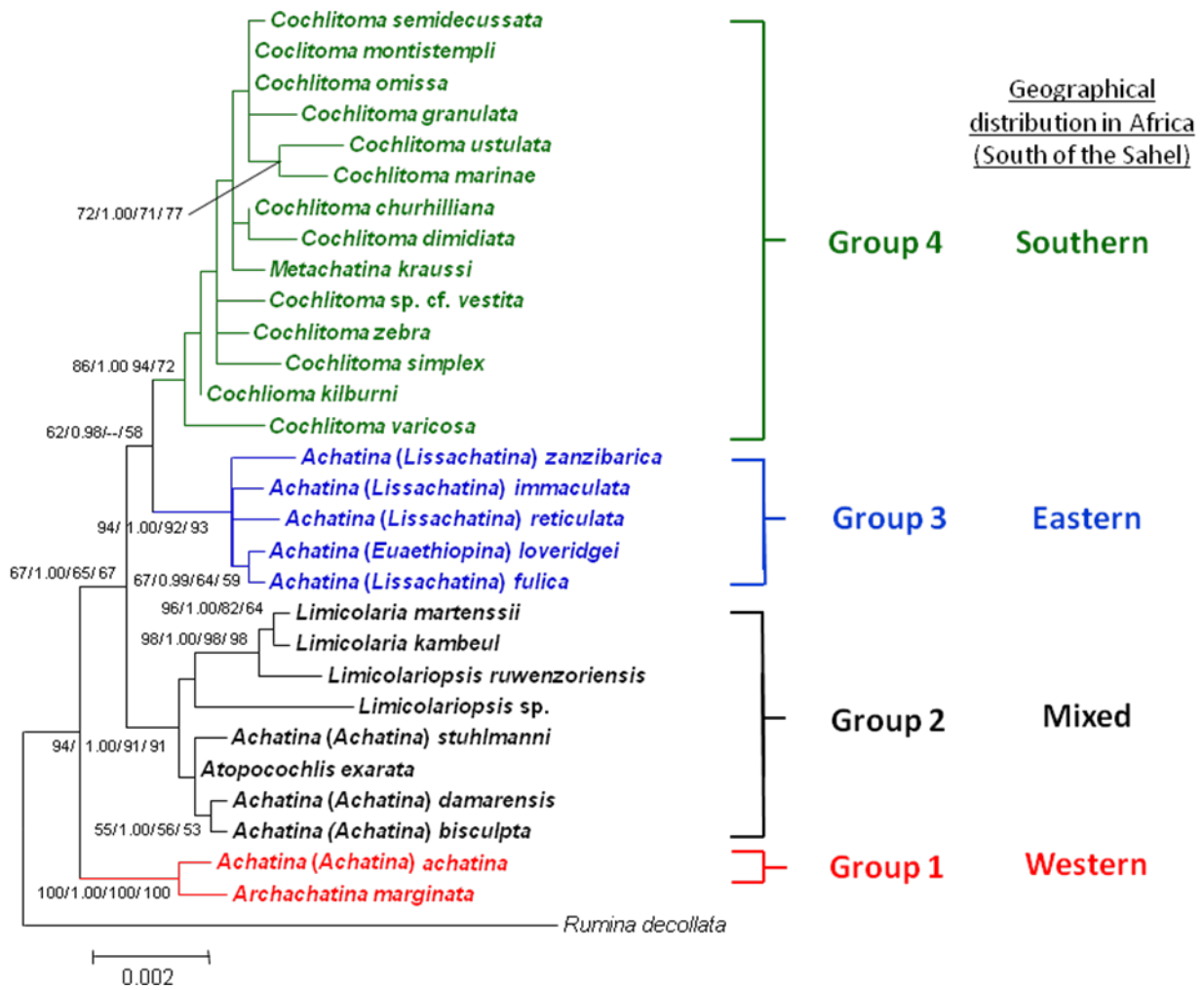
4.3.3. Molecular phylogeny

The maximum likelihood (ML) phylogenies for the rRNA cluster, histone 3, COI (1st and 2nd codon positions only) and 16S rRNA are shown in Figure 4.31 and those of the concatenated sequences from the four genes in Figure 4.32. For the concatenated sequences, two sets of phylogenetic analyses were done. In the first, only taxa with sequences from all four genes were used in the phylogeny; thus, *Achatina* (*Euaethiopina*) *loveridgei* (histone 3 and COI data missing) and *Limicolariopsis ruwenzoiensis* (histone 3, COI and 16S rRNA data missing) were excluded (Figure 4.30A). In the second, all taxa were included in the analyses (Figure 4.30B), with missing data assigned in the analyses for taxa with incomplete datasets. The bootstrap

support values for ML, neighbor-joining (NJ) and maximum parsimony (MP) analyses and the posterior probabilities for Bayesian inference (BI) analysis are also shown. For the BI, NJ and MP phylogenies for the genes and the concatenated sequences, see Appendices 4.9-4.14, pp. 511-522.

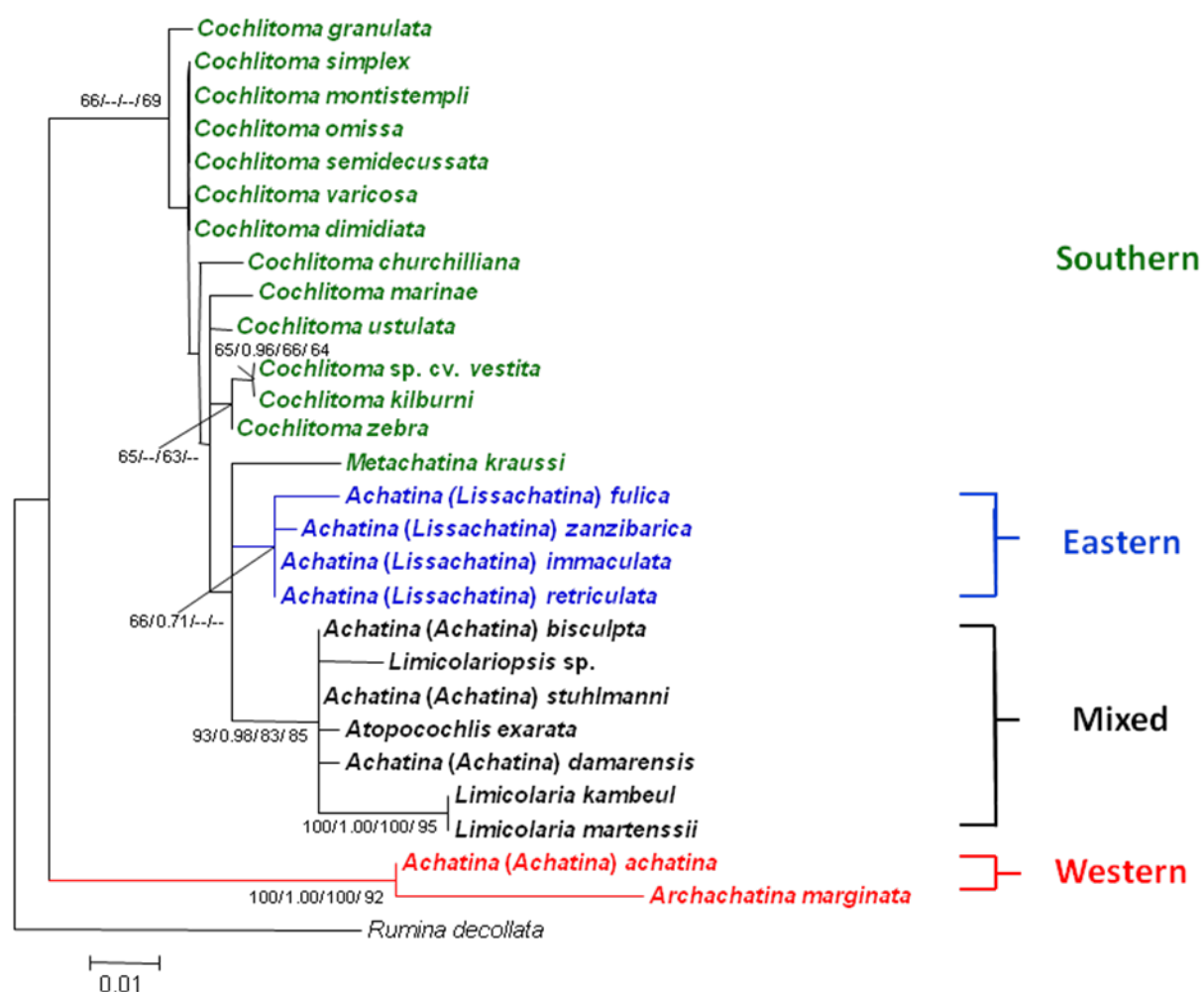
Figure 4.31: (See next four pages.) Maximum likelihood phylogenetic trees of the Achatinidae based on (A) the rRNA gene cluster (3648 nucleotides); (B) the histone 3 gene (328 nucleotides); (C) the COI gene (641 nucleotides) and (D) the 16S rRNA gene (310 nucleotides). The phylogenies were rooted on the subulinid *Rumina decollata*. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap support (1000 replicates) for NJ and MP, respectively. For BI, the optimized number of generations to explore the tree space was 2,000,000 for all the genes; the optimized heating temperatures for each gene were as follows: rRNA cluster-0.1; histone 3-0.075; 1st and 2nd codon positions of COI-0.075; 16S rRNA-0.1. The optimal model for the rRNA cluster was TN93; however, this model is not available in MrBayes, so the GTR model was instead used after determining that there was no significant difference between the TN93 and GTR models and that the GTR model was significantly better than HKY85 (see also Appendix 4.6, p. 508). The scale bar for (A) represents 2 substitutional changes per 1000 nucleotide positions; that of (B) represents 5 substitutional changes per 100 nucleotides; that of (C) represents 1 substitutional change per 100 nucleotides; and that of (D) represents 10 substitutional changes per 100 nucleotides.

Figure 4.31 (contd.)



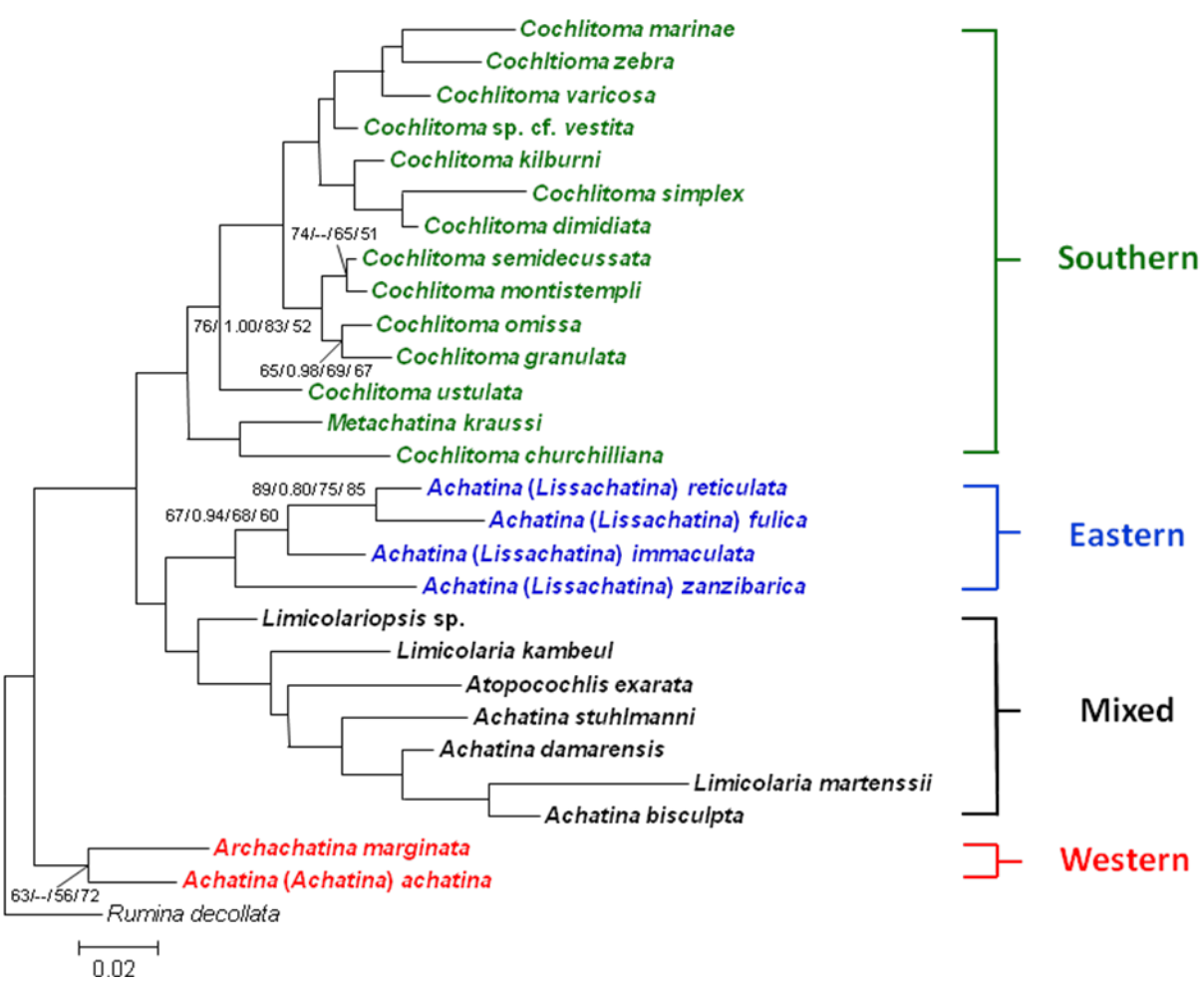
(A) rRNA gene cluster

Figure 4.31 (contd.)



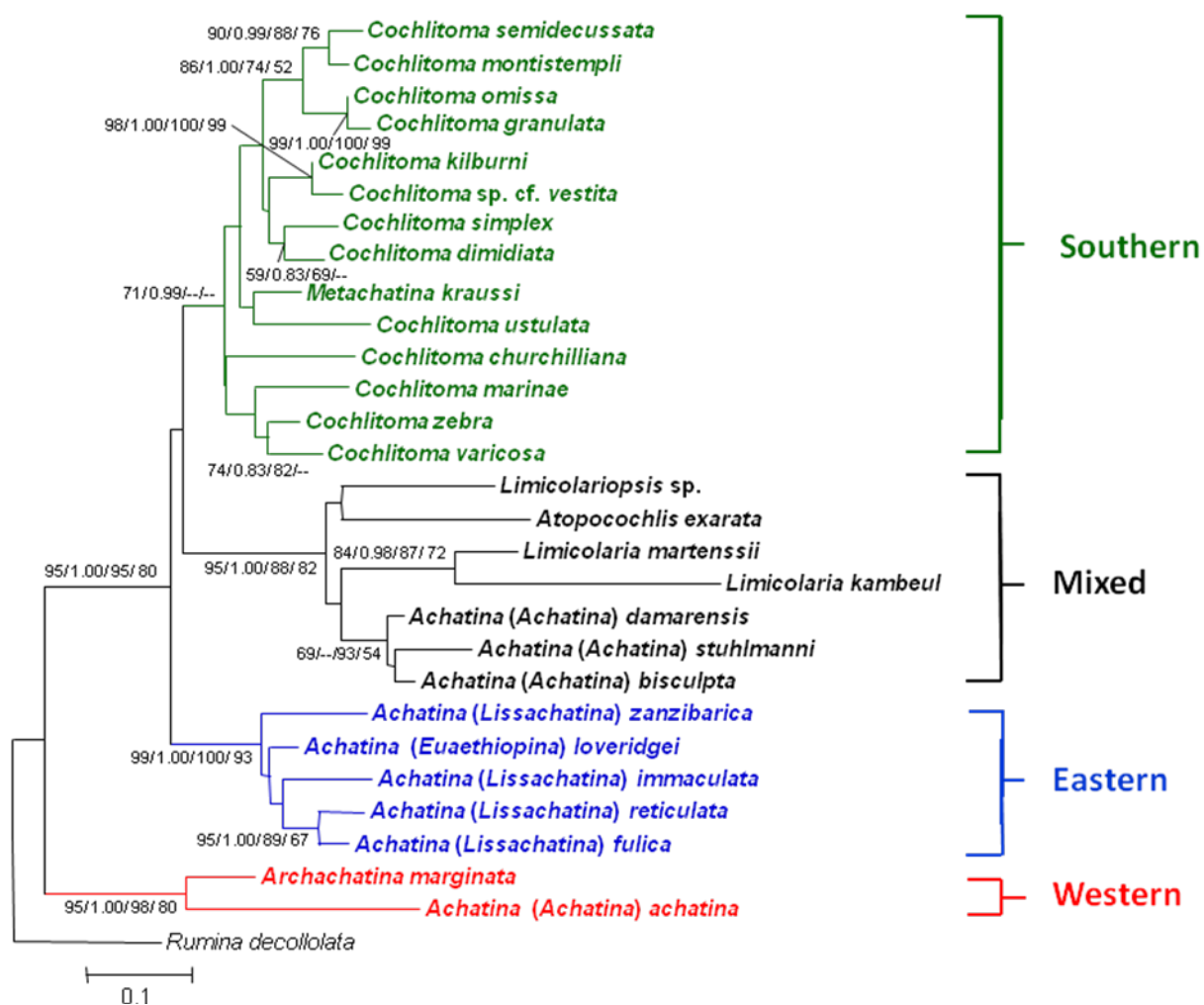
(B) Histone 3

Figure 4.31 (contd.)



(C) COI (1st and 2nd codon positions only)

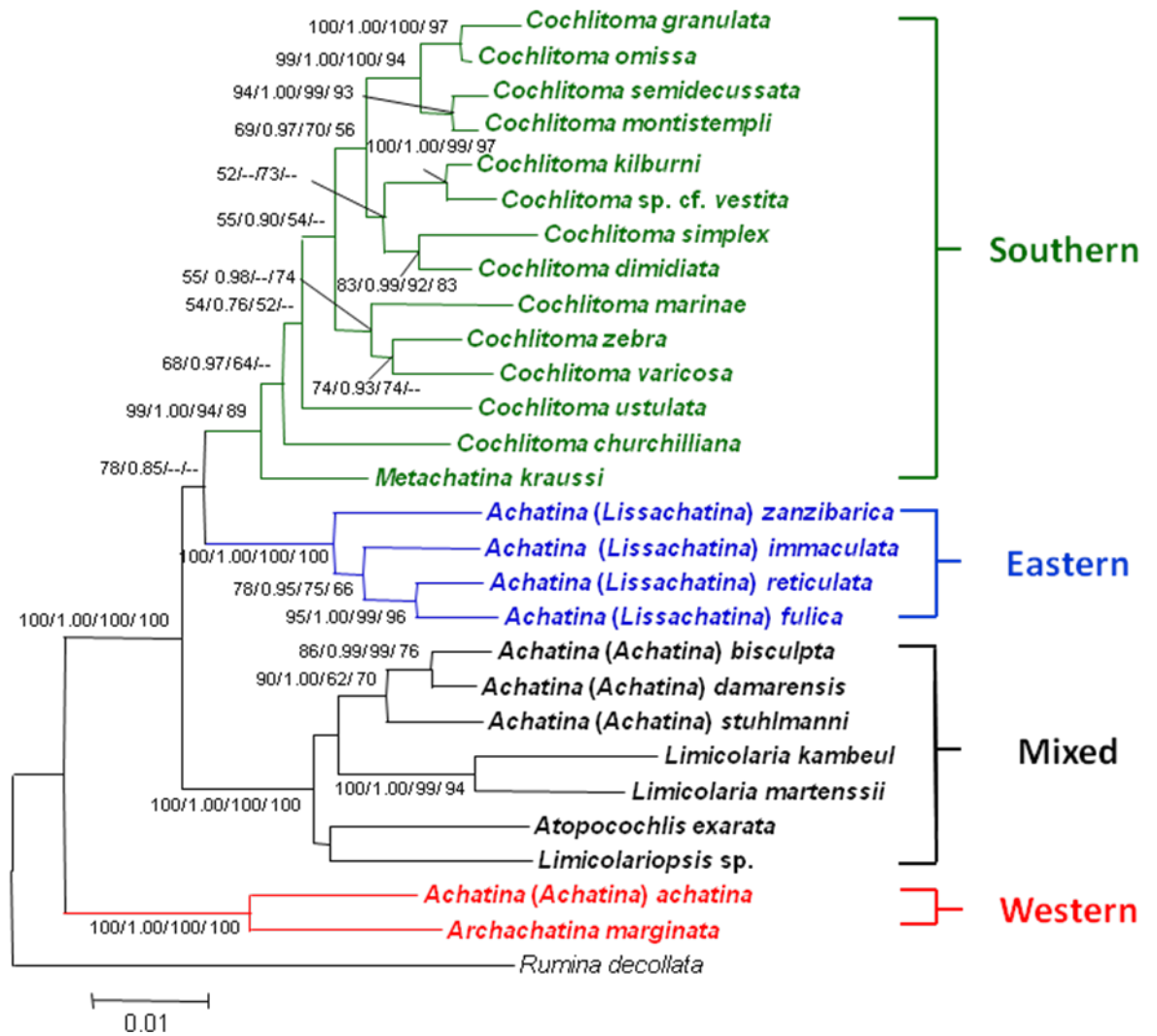
Figure 4.31 (contd.)



(D) 16S rRNA

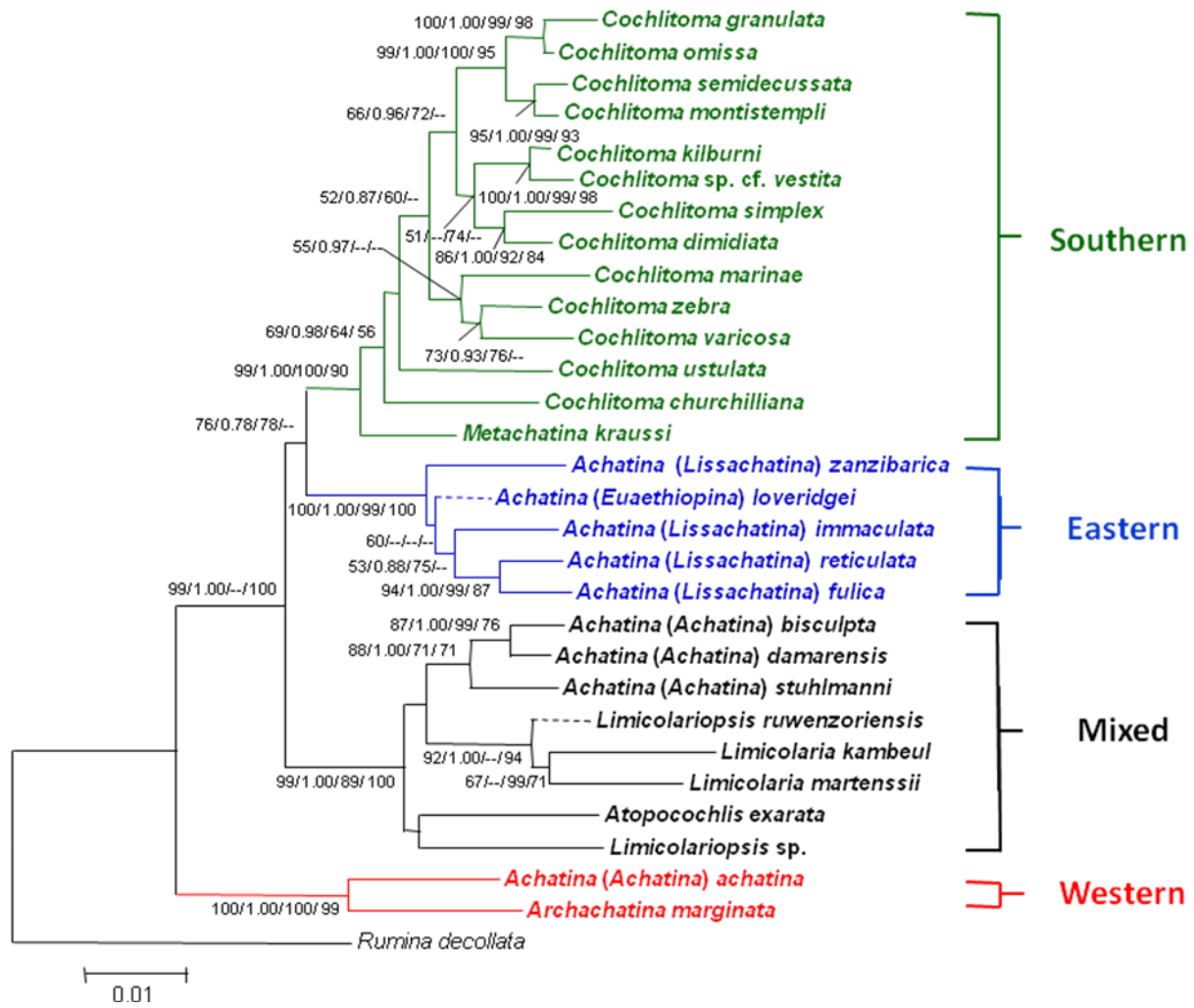
Figure 4.32: (See next two pages.) Maximum likelihood phylogenetic trees of the Achatinidae based on a concatenated sequence of 4403 nucleotides derived from the combined dataset of the rRNA cluster, H3 genes and the 1st and 2nd codon positions of the COI gene. The phylogeny in (A) includes only those taxa which have complete sequence data for all sequence regions while that in (B) includes all taxa including some with missing data. Both *Achatina* (*Euaethiopina*) *loveridgei* and *Limicolariopsis ruwenzoriensis* have missing datasets for the histone 3 and COI genes. Both phylogenies are rooted on the subulinid *Rumina decollata*. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap support (1000 replicates) for NJ and MP, respectively. Bootstrap support less than 50% and posterior probabilities less than 0.7 are not shown. For BI, the optimized number of generations to explore the tree space was 1,000,000 while the optimized heating temperature was 0.1 for both trees. The scale bars represent 1 substitutional change per 100 nucleotides. Branches for taxa with missing data are marked with dashes.

Figure 4.32 (contd.)



(A) Combined dataset (taxa with complete gene sequences only)

Figure 4.32 (contd.)



(B) combined dataset (all taxa)

4.3.3.1. Single gene phylogenies

For the rRNA gene cluster (Fig 4.31A), four major groups emerged that reflect certain geographical distributions. Group 1 at the base of the tree incorporated the two West African achatinids: *Achatina* (*Achatina*) *achatina* and *Archachatina marginata* (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). Group 2 incorporated several achatinids with mixed geographic distributions (94% ML bootstraps, $P=1.0$ BI, 91% NJ bootstraps and 91% MP bootstraps). These included the West African (São Thomé) *Atopocochlis exarata*, the Southern African *Achatina* (*Achatina*) *bisculpta*, the Southwestern African *Achatina* (*Achatina*) *damarensis*, the Central African *Achatina* (*Achatina*) *stuhlmanni* and *Limicolaria martenssii*, the tropical African (i.e. Uganda and Kenya) *Limicolariaopsis ruwenzoriensis* and *Limicolariaopsis* sp. and the widely distributed (from Senegal to Sudan and Somalia) *Limicolaria kambeul*. The other two groups formed sister clades. Group 3 incorporated the East African *Achatina* (*Lissachatina*) species, *A. (L.) fulica*, *A. (L.) immaculata*, *A. (L.) reticulata* and *A. (L.) zanzibarica*, as well as the East African *Achatina* (*Euaethiopina*) species, *A. (E.) loveridgei* (94% ML bootstraps, $P=1.00$ BI, 92% NJ bootstraps and 93% MP bootstraps). Group 4 included the Southern African *Cochlitoma* as well as *Metachatina kraussi* (86% ML bootstraps, $P=1.0$ BI, 94% NJ bootstraps and 72% MP bootstraps).

For the histone 3 gene (Figure 4.31B), three of the four groups identified in the rRNA gene phylogenies were recovered. These were the West African (Group 1) achatinids (100% ML bootstraps, $P=1.00$ BI, 100% NJ bootstraps and 92% MP bootstraps), the mixed group (Group 2) (93% ML bootstraps, $P=0.98$ BI, 83% NJ bootstraps and 85% MP bootstraps) and the East African group (Group 3), although support for this group was equivocal (66% ML bootstraps, $P=0.71$ BI and less than 50%

NJ and MP bootstraps). The Southern African group (group 4) resolved in the rRNA tree did not form a monophyletic unit in the H3 tree but instead was resolved as a paraphyletic group in that it included the Eastern (Group 3) and Mixed (Group 2) groups. In fact, *Metachatina kraussi*, a Southern African taxon, clustered with Groups 2 and 3, although this grouping is not well supported.

For the 1st and 2nd codon positions of the COI gene (Figure 4.31C), all four groups were recovered but there was generally less structure in the topology of the phylogeny and little support for groupings. Of the four groups, only the West African group (Group 1) received moderate support (63% ML bootstraps, $P < 0.7$ BI, 56% NJ bootstraps and 72% MP bootstraps).

For the 16S rRNA gene (Figure 4.31D), all four groups were recovered with good to moderate support: the West African group (Group 1) (95% ML bootstraps, $P = 1.00$ BI, 98% NJ bootstraps and 80% MP bootstraps); the mixed group (Group 2) (95% ML bootstraps, $P = 1.00$ BI, 88% NJ bootstraps and 82% MP bootstraps); the East African group (Group 3) (99% ML bootstraps, $P = 1.00$ BI, 100% NJ bootstraps and 93% MP bootstraps); and the Southern African group (Group 4) (71% ML bootstraps, $P = 0.99$ BI, and less than 50% NJ and MP bootstraps).

4.3.3.2. Combined gene phylogenies

The combined analysis for the rRNA cluster, H3, the 1st and 2nd codon positions of COI and 16S rRNA, but excluding *Achatina (Euaethiopina) loveridgei* and *Limicolariopsis ruwenzoriensis* as they had some gene sequences missing, recovered all four groups present in the rRNA phylogeny and exhibited longer internal branches with better support (Figure 4.32A). The West African group (Group 1) still fell at the base of the Achatinidae (100% ML bootstraps, $P = 1.00$ BI, 100% NJ and 100% MP bootstraps)

while the other achatinids clustered together (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). Within this cluster were the mixed group (Group 2) (100% ML bootstraps, $P=1.00$ BI, 100% NJ and 100% MP bootstraps), the East African group (Group 3) (100% ML bootstraps, $P=1.00$ BI, 100% NJ and 100% MP bootstraps) and the Southern African group (Group 4) (99% ML bootstraps, $P=1.00$ BI, 94% NJ and 89% MP bootstraps), with *Metachatina kraussi* falling at the base of the latter. Groups 3 and 4 clustered as sister clades within the tree.

When all taxa were included in the combined analysis (including the two taxa with missing data), all groups found in the rRNA phylogeny were recovered and in general received higher support (Figure 4.32B). The West African group (Group 1) again fell at the base of the Achatinidae (100% ML bootstraps, $P=1.0$ BI, 100% NJ and 99% MP bootstraps) while the remaining achatinids formed a separate clade with increased support in comparison to the rRNA cluster except for NJ (99% ML bootstraps, $P=1.0$ BI, less than 50% NJ bootstraps and 100% MP bootstraps). Within this clade and consistent with the rRNA tree were the mixed group (Group 2) with increased support (99% ML bootstraps, $P=1.0$ BI, 89% NJ and 100% MP bootstraps) and the clade incorporating the East African and Southern African groups with increased support except for MP (76% ML bootstraps, $P=0.78$ BI, 78% NJ and less than 50% MP bootstraps). The East African group (Group 3) also received increased support (100% ML bootstraps, $P=1.00$ BI, 99% NJ and 100% MP bootstraps) as did the Southern African group (Group 3) (99% ML bootstraps, $P=1.0$ BI, 100% NJ and 90% MP bootstraps). Some well-supported groupings were also observed within groups 2-4. Within the mixed group (Group 2), for instance, there was strong support for the clustering of *Achatina* (*Achatina*) *bisculpta*, *A. (A.) damarensis* and *A. (A.) stuhlmanni* (88% ML bootstraps, $P=1.0$ BI, 71% NJ bootstraps and 71% MP bootstraps) as well as

Limicolaria kambeul, *L. martenssii* and *Limicolariopsis ruwenzoriensis* (92% ML bootstraps, $P=1.0$ BI, less than 50% NJ bootstraps and 94% MP bootstraps). Within the East African group (Group 3), *Achatina (Lissachatina) fulica* clustered with *A. (L.) immaculata* (94% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 87% MP bootstraps). Within the Southern African group (Group 4), *Metachatina kraussi* again fell at the base. Very strong support was also shown for several groups. These were *Cochlitoma omissa* and *C. granulata* (100% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 98% MP bootstraps); *C. montistempli* and *C. semidecussata* (95% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 93% MP bootstraps); *C. kilburni* and *C. sp. cf. vestita* (100% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 98% MP bootstraps); *C. dimidiata* and *C. simplex* (86% ML bootstraps, $P=1.0$ BI, 92% NJ bootstraps and 84% MP bootstraps); and *C. varicosa* and *C. zebra* (73% ML bootstraps, $P=0.93$ BI, 76% NJ bootstraps and less than 50% MP bootstraps). The geographic distributions of the members of the four groups are shown in Fig. 4.33.

Table 4.4: Hypothesis testing for the monophyly of *Achatina* (*Achatina*) using the Shimodaira-Hasegawa test.

Gene	Likelihood Score		<i>P</i> for SH ^a Test (Conclusion ^b)
	Optimal NJ tree	Constrained NJ tree	
rRNA cluster	6146.48364	6240.31789	0.002 (Significant)
Histone 3	909.37626	949.45797	0.001 (Significant)
COI (1 st and 2 nd codons)	1995.68125	2004.66467	0.177 (Not significant)
16S rRNA	2876.32254	2906.36582	0.008 (Significant)
Combined	12509.66094	12643.04203	<0.001 (Significant)
Combined-all taxa	12640.55433	12756.19951	<0.001 (Significant)

^a SH test – Shimodaira-Hasegawa Test

^b Significant at $P < 0.05$

4.4. Discussion

4.4.1. Single versus combined gene phylogenies

Among the five genes used for the Achatinidae, the nuclear rRNA cluster provided the most resolved structure of the internal branches with the four major groups based on biogeography receiving good support. The terminal nodes, however, were not well supported, suggesting that the gene is too conserved for that level. The nuclear histone 3 gene yielded even less structure with only three groups recovered and very few terminal nodes received support. The gene was also too conserved as evidenced by the presence of five sets of identical sequences among 16 taxa. On the other hand, the two mitochondrial genes were more variable and were therefore evolving faster than the two nuclear genes. Of the two, the 16S rRNA gene gave support to most terminal nodes while at the same time recovering the four major groups. The COI had less structure than 16S because the groups were not well supported. Fewer sites were used for COI as the third codon positions were excluded due to their saturated condition, which is indicative of their faster rate of evolution. The fifth gene, the nuclear actin, was also not included due to significant levels of paralogy that disrupted phylogenetic signal.

When the four genes were combined, both the terminal branches and the internal nodes received support based on ML, NJ and MP bootstraps as well as BI posterior probabilities. Underlying signal was also detected in the combined phylogenies with very good support that was not evident in the single gene phylogenies (i.e. the basal position of *Metachatina kraussi* in the Southern African group).

4.4.2. Evolutionary history of the Achatinidae based on molecular data

The existence of no fossils earlier than the Pleistocene (0.01-1.8 MYA) (Zilch, 1959-1960; Solem, 1979) suggests that the Achatinidae are a relatively recent group (Tillier, 1989), although Raut & Barker (2002) believed them to be much older than the fossil record. Mead (1991) suggested that the primitive achatinids occurred in the region of the Lower Guinea in West Africa, and he postulated that the achatinids radiated from there to the other parts of Africa south of the Sahel. Tillier (1989) described the achatinids as giant subulinids, although the taxonomy of the latter is still in a state of flux, and Tillier could not identify with certainty from which subulinid group the achatinids arose.

This study presents the first attempt to elucidate the phylogeny of the Achatinidae using molecular data. Phylogenies derived from the combined nuclear rRNA cluster and histone 3 genes and the mitochondrial cytochrome c oxidase 1 and 16S rRNA genes demonstrated groupings that roughly corresponded to the geographical distributions of the Achatinidae. These groups were the West African group (Group 1) found at the base of the tree followed by the mixed group (Group 2), the East African (Group 3) group and Southern African (Group 4) group, the last two of which clustering as sister clades. Group 2 included species found in the other three regions as well as some that are present in Central Africa.

The West African *Achatina (Achatina) achatina* (found in the region of Upper Guinea of West Africa; Bequaert, 1950) and *Archachatina marginata* (found in the region of West Africa east of the Dahomey gap, Bequaert, 1950) both fell at the base of the tree, which supports Mead's (1991) hypothesis that the achatinids originated in the west, particularly in the Gabon-Cameroon region. Both species share a penis sheath that only covers half of the penis (Mead, 1950). In fact, *Archachatina* Albers was erected as a distinct genus from *Achatina* purely on conchological criteria, primarily on the form of the shell apices. The large obtuse apex of *Archachatina* is a result of the large embryonic whorls that form in their large eggs (about 19 by 14.5 mm); *Achatina* possesses an acute apex as a result of its small eggs (Bequaert, 1950).

The mixed group includes several species with varying distribution patterns, including the São Thomé endemic *Atopocochlis exarata*. Mead (1991) included *Atopocochlis exarata* as one of the most primitive achatinids together with *Callistoplepa*, *Leptocala*, *Tripachatina* and *Columna*, none of which was sampled in this study due to inability to obtain fresh material. The reproductive structures of *A. exarata* include a very short penis sheath similar to that of *Achatina (Achatina) achatina* and *Archachatina marginata* and an enlarged vagina (Mead, 1950); the vas deferens is also enclosed by the penis sheath, unlike in the other “primitive” achatinids like *Callistoplepa* and *Leptocala* where the vas deferens is free (Schileyko, 1999). The molecular data as well as the morphological features of the genitalia puts into question the position of *Atopocochlis exarata* among the “primitive” achatinids as suggested by Mead (1991). Another group within the mixed group with high support is the grouping of the two *Limicolaria* species. Both have a distinct penis papilla (Mead, 1979b; 1991). Also within the mixed group are *Achatina (Achatina) bisculpta* and *Achatina (Achatina) damarensis*, which clustered together with strong support. In both species, the basal vas

deferens and penis are either intimately and tightly connected to the penial retractor muscles, as in the case *A. (A.) bisculpta* (Sirgel, 2000), or cocooned by them, as in the case of *A. (A.) damarensis* (Mead, 1950, 1991; Van Bruggen, 1970). These two taxa clustered with *A. stuhlmanni*, and all three belong to the subgenus *Achatina*, though these taxa did not cluster with another member of the subgenus, *A. achatina*, which grouped with *Archachatina marginata*. The non-monophyly of this subgenus was further supported by the Shimodaira-Hasegawa test. The validity of this taxon therefore warrants re-examination.

The Southern African and East African groups together form a sister clade with the mixed group according to the combined gene phylogenies. Indeed, both *Achatina* (*Lissachatina*) from East Africa and *Cochlitoma* of Southern Africa share a “long spermathecal [=gametolytic] duct [that] places the spermatheca [=gametolytic sac] on the uterine portion of the spermoviduct, well above the junction of” the apical vas deferens and the uterine portion of the spermoviduct (Mead, 1991, p. 553). Such a characteristic is probably derived as it is not found in the Western group, which falls at the base of the Achatinidae based on the single and combined gene phylogenies.

The presence of the penial groove in *Cochlitoma* is most likely a derived character as it is not present in *Metachatina kraussi* found at the base of the southern African clade in the combined gene phylogenies, thus lending support to the assumption of Mead (1991) that *Cochlitoma* is an ‘advanced’ genus. This topology was not evident in the single gene phylogenies and was only revealed as an underlying signal when the genes were combined, as shown by the higher bootstrap and posterior probability supports. The clustering of *Cochlitoma* species within the Southern African group based on the molecular phylogenies supports Mead’s (2004) recognition of *Cochlitoma*

as a distinct genus rather than a subgenus of Western African *Archachatina* even if they share similar conchological features.

The members of the East African group under *Achatina* (*Lissachatina*) have extroversion muscles as well as a muscular, bulboid enlargement of the basal vagina (Mead, 1991, 1995). The results from the molecular data in this study support Mead's view that *Lissachatina* be distinguished from the West and central African *Achatina* species. Furthermore, judging from the topology of the East African group in relation to other *Achatina* species in the molecular trees, coupled with the distinct morphological characteristics of both the male and female conduits of the reproductive tract, it is clear that there is no basis for including *Lissachatina* within *Achatina* and *Lissachatina* should be recognized as a distinct genus as with *Cochlitoma*. Within the East African group, *Achatina* (*Lissachatina*) *fulica* clustered with *A. (L.) reticulata*. They differ morphologically in that *A. reticulata* has a short penis sheath that does not cover the entire penis (Mead, 1950). As with *Atopocochlis exarata*, the presence of a short penis sheath in *A. reticulata* is hypothesized to be a derived state and does not indicate plesiomorphic condition or 'primitive' state *sensu* Mead. It can be noted that the Southern African *Metachatina kraussi* (Mead, 1950) and *Cochlitoma granulata* (Mead, 2004) also possess a short penis sheath. The subgenus *Euaethiopina*, represented by *A. (E.) loveridgei*, has a shell that is more elongated ovoid than that of *Lissachatina*, which is more broadly ovate to obesely fusiform; the two taxa also have distinct coloration (Bequaert, 1950; Schileyko, 1999). However, *Achatina* (*Euaethiopina*) *loveridgei* is very similar to *Achatina* (*Lissachatina*) *zanzibarica* with respect to its reproductive anatomy (Mead, 1950). Its placement within *Lissachatina* based on molecular data, coupled with the evidence based on reproductive anatomy, would suggest that *A. loveridgei* should be moved to *Lissachatina*.

All these results support the hypothesis that the achatinids migrated to the central region from the west, then radiated to the east and south. As further support to this hypothesis, van Bruggen (1986) noted that terrestrial snails and slugs of the Afrotropical region (south of the Sahel) occur in four major centres of endemism: 1) Southern Africa; 2) East Africa; 3) North-East Africa; and 4) Central/West Africa. He also observed the highest taxon density for the Achatinidae in West Africa that extended to Central Africa, followed by East and North-East Africa and finally Southern Africa.

Aside from the Achatinidae, there are 33 other families of terrestrial gastropods in the Afrotropical region, of which eight are endemic. Of these, three families (Maizaniidae, Gymnarionidae and Urocyclidae) are found in all four major centres like the Achatinidae; one family (Aillyidae) is found only in West Africa and another (Thyrophorellidae) is restricted to São Thomé off the coast of West Africa. The remaining endemic families (Aperidae, Dorcasiidae and Prestonellidae) are distributed in Southern Africa (van Bruggen, 1986). Diamond and Hamilton (1980) also found that forest passerine birds roughly occur in the four endemic centres, either limited to one or a few of these centres or occurring in all. These centres most likely served as refugia for forest-dependent animals, such as terrestrial gastropods and passerine birds, when forests contracted due to changing rainfall patterns as a result of alternating glacial and interglacial periods during the Quaternary, particularly the Holocene (18,000 years ago), which in turn gave opportunities for vicariant speciation to occur (van Bruggen, 1986). However, Raut & Barker (2002) acknowledged that much of the speciation process, particularly for the terrestrial gastropods, could have predated the changing forest cover of the Holocene.

4.4.3. Limitations of the study and the next step

A number of achatinid groups were not available for this study. Among these are genera found in West Africa considered primitive by Mead (1991) in that the penis sheath is very short and does not enclose the vas deferens. These include *Leptocala* and *Callistoplepa*. If Mead's hypothesis is correct, then these genera should fall at the base of the tree. Furthermore, as West Africa harbors a high number of endemic species of achatinids (van Bruggen, 1986), it is desirable to include as many of these species as possible in future investigations. Of particular interest are the sinistral *Columna columna* and *Archachatina bicarinata*. Equally interesting is the relationship of the East African *Bequaertina* with *Lissachatina*.

4.5. Summary

This study is the first attempt to reconstruct the phylogeny of the Achatinidae using molecular data derived from the nuclear rRNA cluster, actin and histone 3 genes and the mitochondrial cytochrome c oxidase subunit 1 and 16S rRNA genes. Results demonstrated four distinct groups that to a large extent follow the geographical distribution of the sampled taxa and validate groupings previously erected on the basis of reproductive morphology. At the base of the trees are West African species followed by a group which includes taxa with varying geographical distributions ranging from the western, central, eastern and southern regions. Two other groups probably arose from the mixed group and form sister clades: one is the East African group that includes *Achatina* (*Lissachatina*) *fulica*; the other group encompasses most of the Southern African species. The results support Mead's recognition of Southern African *Cochlitoma* as a genus separate from the West African *Archachatina*. The results likewise support Mead's (1950) distinction of the Eastern subgenus *Lissachatina* from

the western and central *Achatina* and demonstrate that there is no basis for continuing to include *Lissachatina* as a subgenus of *Achatina*.

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CHAPTER 5 - Genetic variation in global populations of *Achatina fulica*

5.1. Introduction

The Giant African Land Snail, *Achatina (Lissachatina) fulica* Bowdich, is a tropical species with a widespread distribution across East Africa, the Indo-Pacific and the Caribbean. Its natural range is East Africa, but its current global distribution is primarily due to human intervention (Raut & Barker, 2002). It is a serious crop pest that can easily spread into new areas where it causes significant damage to vegetables and other food crops (Mead, 1979; Raut & Barker, 2002). The World Conservation Union (IUCN) has listed *A. fulica* in its 100 most invasive species (Lowe *et al.*, 2000), and among the land snails in the world, it is probably the most invasive (Raut & Barker, 2002). Its success as an invasive species is probably attributable to its high reproductive capacity and its generalist food behavior, feeding on a wide range of plants and detritus (Raut & Barker, 2002). The tendency for people to transport the snails and release them into the wild either intentionally or inadvertently also helps to spread them further (Thiengo *et al.*, 2007). The snail also serves as the intermediate host of the rat lungworm *Angiostrongylus cantonensis* that causes eosinophilic meningitis in humans (Marquardt *et al.*, 2000).

5.1.1. The Dispersal of *Achatina fulica* from East Africa

The ease with which *Achatina fulica* spreads into new areas is attributed to the availability of several pathways of dispersal for the snail. A pathway, which pertains to an activity that involves the introduction of a species, can be accidental or deliberate (Cowie & Robinson, 2003). *Achatina fulica* is deliberately introduced into new areas for the purposes of consumption, as pets, aesthetics or ornaments, religious symbols, or

for medicinal or research use (Mead, 1979; Kliks & Palumbo, 1992; Cowie & Robinson, 2003; Budha & Naggs, 2008). On the other hand, *A. fulica* is accidentally introduced when it “hitchhikes” with agricultural and horticultural products, the packaging or containers of any commodity, or even on vehicles such as cars and trucks (Cowie & Robinson, 2003). *Achatina fulica* can also escape from local farm operations where they are bred for their meat (Paiva, 1999).

Achatina fulica has a pan-tropical distribution brought about within the last 200 years by its human-aided dispersal (Fig. 5.1). The snail is indigenous to the coast of East Africa (probably originating in Kenya and Tanzania) but was introduced into the nearby islands of Madagascar, Mauritius, the Comoros, Mayotte and Reunion prior to 1800 (Bequaert, 1950). The snail was then brought to India and Sri Lanka by naturalists in the early 1900s (Naggs, 1997; Raut & Barker, 2002) and spread to Nepal to the north (Budha & Naggs, 2008) and to the Malay Peninsula to the southeast via sea routes in cargo vessels (Mead, 1961). Immediately before and during the Second World War, Japanese merchants and soldiers spread *A. fulica* further into Southeast Asia, Taiwan, and many islands of the Pacific as they were used for food and medicinal purposes and sometimes as pets (Kliks & Palumbo, 1992). By the late 1980's, *A. fulica* had been reported in the Caribbean, particularly in Guadeloupe and Martinique (Schotman, 1989) and later on in St. Lucia and Barbados by 2000 (Fields *et al.*, 2006) and in Antigua by 2008 (NAPPO-PAS, 2008). By the 1990's, *A. fulica* had already established itself in South America, particularly Brazil (Paiva, 1999; Carvalho de Vasconcellos & Pile, 2001; Thiengo *et al.*, 2007), Colombia, Ecuador and Peru (Borrero *et al.*, 2009). Recently, the snail has been observed in the Ivory Coast, Ghana and Morocco in West and Northwest Africa (Raut & Barker, 2002). Although *A. fulica* has reached Australia, Japan, and the United States mainland in the past, authorities in these countries have

successfully eradicated *A. fulica* for fear of its potential for causing damage to agriculture (Mead, 1961; Kliks & Palumbo, 1992). As an exotic pet (Kliks & Palumbo, 1992), *A. fulica* is now present in temperate countries such as the UK and France.

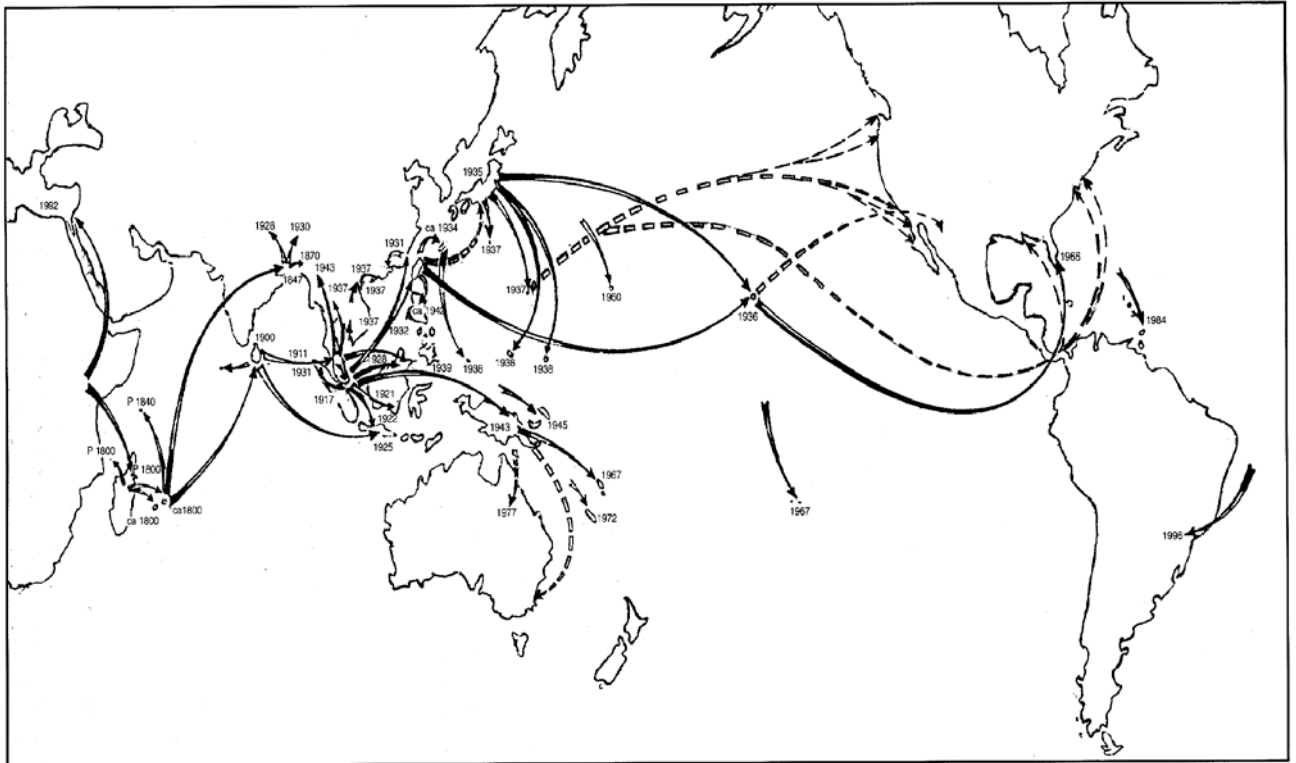


Figure 5.1: Dispersal route of *Achatina fulica* from East Africa. Years indicate approximate dates when the snail was first observed in each region. Solid lines denote successful introductions while broken lines denote introductions that were intercepted. (From Raut & Barker, 2002).

5.1.2. Genetic variation in introduced species

The genetic composition of the population of an introduced species in a new area may affect the success of that population. Invasion may involve founder events in which a population of a few individuals carries a reduced amount of genetic variation in comparison to the source population. Over time, this leads to a population bottleneck in which individuals suffer from a lack of genetic variation and where massive shifts in

allele frequencies are observed (Dlugosch & Parker, 2008). This becomes particularly problematic to the invasive species if the loss of variation includes adaptive traits that could have improved their chance of establishing in a new habitat, preventing extinction and increasing population growth (Kolbe *et al.*, 2007). Although some data do show lack of genetic diversity in introduced species, as seen in the apple snails *Pila conica* and *Pomacea canaliculata* in Hawaii (Tran Chuong *et al.*, 2008), other data on invasive species reveal that many populations do not exhibit such lack of diversity. One possible explanation is the role of multiple introductions in augmenting the genetic diversity of the invasive species. Multiple introductions may involve separate dispersal events over a long period of time in which new genes from recently introduced populations are spread among the individuals in the established population (Dlugosch & Parker, 2008). This was evident in the cheatgrass, *Bromus tectorum*, in which multiple introductions from Europe and Southwest Asia counteracted the effect of bottlenecking in introduced populations in western North America (Novack & Mack, 1993). Kolbe *et al.* (2007) also noted the same pattern in eight species of *Anolis* lizards introduced in Florida and the Dominican Republic from the northern Caribbean using the ND2, tRNA-Trp and tRNA-Ala genes of the mitochondrial DNA. Their results revealed that 80% of introductions were derived from multiple native sources. Furthermore, haplotypes that were found in distinct geographic locations in the native range occurred as variants within the introduced populations. They proposed a two-step process in introductions marked by the initial decrease in genetic variation as a result of founder events and bottlenecking followed by an increase in genetic variation if multiple introductions from different sources take place.

Despite the detailed historical and anecdotal information concerning the spread of *Achatina fulica*, no systematic evaluation of its genetic variation across global

populations has been undertaken. It is therefore not known whether multiple introductions of *A. fulica* from different sources have occurred in any part of its new range or whether the introduced populations are undergoing bottlenecking. In either case, these factors could have a profound impact on the success of *A. fulica* as an introduced species.

5.1.3. Mitochondrial molecular markers for evaluating genetic variation in snail populations and elucidating their population history

Data derived from molecular genetic variation in snail populations can yield useful information about the routes of introduction or dispersal and colonization of a particular snail species into new areas (Davison, 2000; Gittenberger *et al.*, 2004; Pinceel *et al.*, 2005; Rawlings *et al.*, 2007). Mitochondrial genes have been used as the markers of choice to assess variation within and between snail populations because they generally evolve faster than nuclear genes and are maternally inherited and therefore not subject to recombination (Brown, 1985; Avise *et al.*, 1987). For example, Davison (2000) analyzed a fragment of the mitochondrial 16S rRNA gene using single-stranded conformational polymorphism (SSCP) analysis and DNA sequencing to identify different haplotypes from populations of the banded grove snail *Cepaea nemoralis* from Britain and Ireland. The fragment he used for the SSCP analysis corresponded to the variable domain IV segment of the secondary structure of the post-transcribed 16S rRNA gene (Lydeard *et al.*, 2000) for which Palumbi (1996) designed universal primers. Davison's work revealed two mitochondrial lineages that had an east-west distribution, suggesting two possible routes of colonization of Britain after the last ice age. Pinceel *et al.* (2005) also used the same variable domain IV segment of the 16S rRNA gene as part of their population genetic survey of the terrestrial slug *Arion*

subfuscus in northeastern USA and Europe. They likewise used SSCP and DNA sequencing. Their results revealed the existence of two distinct 16S rRNA haplotypes in the USA representing at least two independent introductions from the British Isles on the one hand and mainland Europe on the other. In another study, Gittenberger *et al.* (2004) sequenced a fragment of the cytochrome c oxidase subunit I (COI) gene for several populations of the snail *Arianta arbustorum* and three other species of *Arianta* to elucidate the evolutionary history of the two shell forms of *A. arbustorum*.

5.1.5. Objective of the study

This study aimed to assess the genetic variation in global populations of *Achatina fulica*. Comparison of recently dispersed *A. fulica* populations from across the globe with those from East Africa and adjacent islands were undertaken to determine where the recently introduced global populations of *A. fulica* originated and whether they represent single or multiple introductions. To this effect, the genetic diversity of *A. fulica* populations from East Africa, the Indian subcontinent, Southeast Asia, the Pacific and the Caribbean were evaluated using single stranded conformational polymorphism (SSCP) analysis of the 16S ribosomal (r) RNA gene.

5.2. Materials and methods

5.2.1. Sample collection

In total, 382 individuals of *Achatina fulica* from 19 populations across the globe were sampled. Two populations were obtained from East Africa: Kampala, Uganda (5 snails) and Dar Es Salaam, Tanzania (38 snails). Three populations were from Indian Ocean islands near East Africa: Mayotte (50 snails) off the coast of Mozambique;

Souillac, Mauritius (45 snails) east of Madagascar; and Mahe, Seychelles (2 snails) northeast of Madagascar. Samples from Nepal (22 snails) represented the Indian subcontinent while those from Southeast Asia included Myanmar/ Burma (20 snails), Thailand (20 snails), the Philippines (2 population samples with 20 snails each), Malaysia (20 snails) and Singapore (20 snails). The Pacific was represented by Ogasawara/ Bonin (12 snails), New Caledonia (21 snails), Hawaii (20 snails), Moorea (10 snails) and Tahiti (5 snails). Two populations were obtained from the Caribbean: Martinique (20 snails) and Barbados (12 snails). The Martinique specimens were a captive bred population of F1 offspring bred from Martinique parents by F. Adnai. See also Table 5.1.

Table 5.1: Locality, collector and sample size of global populations of *Achatina fulica* used in the study

Locality	Collector	Sample size
Kampala, Uganda	B. Rowson	5
Dar Es Salaam, Tanzania	C. Ngereza	38
Mayotte	F. Barthelat	50
Souillac, Mauritius	O. Griffiths	45
Mahe, Seychelles	J. Gerlach	2
Bharatpur, Nepal	P. Budha	22
Yangon, Myanmar/ Burma	F. Naggs	20
Trok Nong Area, Chantaburi, Thailand	S. Panha & C. Sutcharit	20
Hahajima, Ogasawara/ Bonin Islands	A. Davison	12
University of the Philippines, Diliman, Quezon City, Philippines	I. Fontanilla	20
Los Baños, Philippines	M. Carandang	20
Singapore	M. Posa	20
Kota Kinabalu, Sabah, Malaysia	M. Schilthuizen & T. Liew	20
Noumea, New Caledonia	C. Wade	21
Moaroa Valley, Tahiti, French Polynesia	T. Coote	5
Haapiti Valley, Moorea, French Polynesia	T. Coote	10
Kaneohe, Oahu, Hawaii	K. Hayes	20
Martinique (captive bred F1 population, Nancy, Lorraine, France)	F. Adnai	20
Barbados	A. Norville	12

5.2.2. PCR and SSCP analysis using the 16S rRNA gene

Small tissue slices (approximately 8 mm³) were cut from the foot muscle of each snail and subjected to the NaOH direct lysis DNA extraction method for snails (Section 2.1.2 of Chapter 2, pp 39-40).

A 400 bp PCR fragment used for SSCP analysis can only detect 80% of single nucleotide differences whereas smaller fragments have an increased probability of the detection of differences (Sunnucks *et al.*, 2000). For the 16S rRNA gene, a primer pair, 16S1i and 16S_SSCP2i, was designed to produce a 293 bp fragment of domain IV of the post-transcribed 16S rRNA (Lydeard *et al.*, 2000). Note that primers 16S1i and 16S_SSCP2i bind to positions 781-802 and 1049-1068, respectively, of the 16S rRNA gene of *Cepaea nemoralis* (GenBank NC_001816). Details of the primer pair are shown in Table 2.5 of Chapter 2, p. 53.

The amplified PCR products were then subjected to SSCP analysis (Section 2.8 of Chapter 2, pp. 66-70). The bands in the native acrylamide gel were visualized using silver staining, after which haplotypes were identified and scored. Representative samples bearing unique haplotype gel profiles were checked for single nucleotide differences by re-amplifying the PCR product from the DNA sample and sequencing the amplified fragment. Both sense and anti-sense strands were sequenced directly using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 termination cycle sequencing chemistry (Section 2.6 of Chapter 2, pp. 59-61). The sequences were subsequently assembled in the STADEN package version 1.5.3 (Staden *et al.*, 2000) and aligned manually within GDE Version 2.2 (Smith *et al.*, 1994).

5.2.3. Phylogenetic and network analyses of the 16S rRNA haplotypes

To determine the evolutionary relationships of the different 16S haplotypes, the best model for sequence evolution with optimized parameters was determined and an NJ tree was generated in PAUP* Version 4.0b10 package (Swofford, 2002) (see Section 2.9.8 of Chapter 2, p. 84). *Achatina reticulata* was used as an outgroup as this species had the smallest uncorrected distance to *A. fulica* based on the same 16S rRNA SSCP fragment (see Appendix 5.1, p. 523). A median joining network of the haplotypes (Bandelt *et al.*, 1999) was also drawn using the Network Version 4.502 program (<http://www.fluxus-engineering.com>).

5.3. Results

5.3.1. PCR and SSCP analysis of the 16S rRNA gene

PCR and SSCP analysis carried out on 382 snails from 19 populations yielded 15 unique 16S rRNA haplotypes (Figure 5.2).

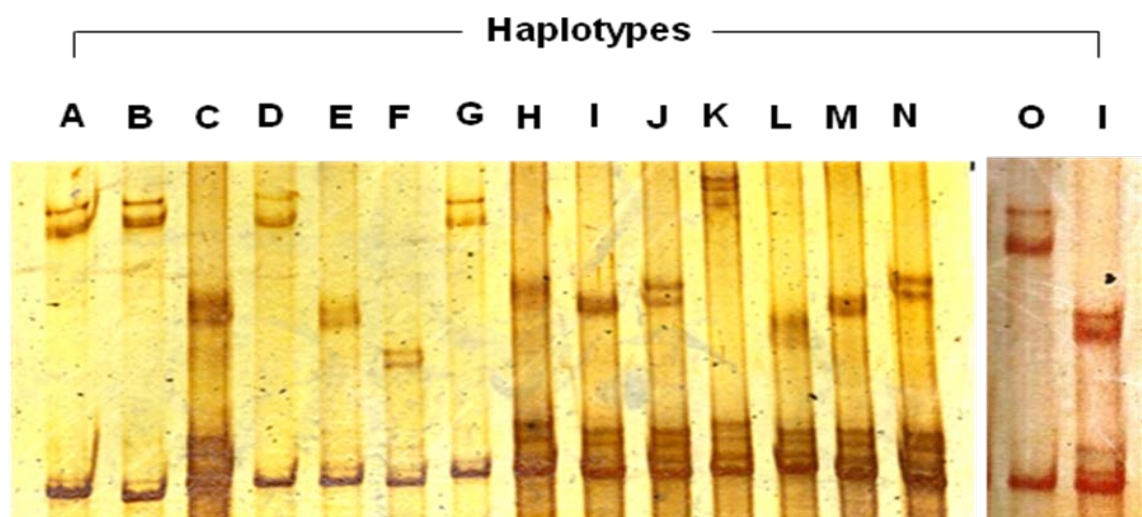


Figure 5.2: Gel profiles of 15 haplotypes (labeled A – O) based on a 293 bp fragment in domain IV of the 16S rRNA gene. (Haplotype O was run on a separate gel alongside haplotype I, which was used as basis of comparison.) Samples were run on a 50% native polyacrylamide gel at 180 V for 24 hours. The bands were visualized by silver staining.

PCR-direct sequencing of the haplotypes revealed 17 variable sites (Figure 5.3).

Two of these sites (pos. 285-286) were found to be indels.

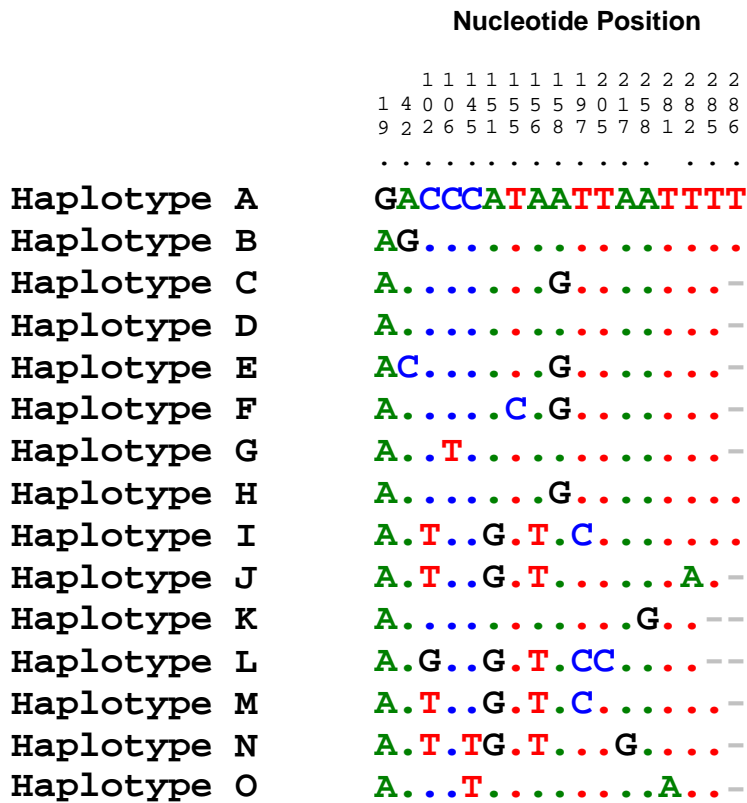


Figure 5.3: Seventeen variable sites across the 15 haplotypes. The number shown above each site refers to the variable site based on the 16S rRNA SSCP fragment.

A summary of the frequency and distribution of these haplotypes is shown in Table 5.2 and Figure 5.4. Seven haplotypes were found in the two East African populations: Uganda with one haplotype (O) and Tanzania with six haplotypes (I, J, K, L, M, and N). Among the Indian Ocean islands, Mayotte yielded the highest number of haplotypes with six (A, B, C, D, G and H), three of which were also found in Mauritius (C, D and H) and one in Seychelles (C). However, of the haplotypes found in Africa and on the Indian Ocean islands, only haplotype C was found in the populations surveyed outside East Africa and the Indian Ocean islands, thus making type C the only

pan-tropical haplotype. There was also one unique haplotype (E) detected in the Los Baños site in the Philippines where it was found in conjunction with haplotype C. Another unique haplotype (F) was found in New Caledonia in the Pacific and Barbados in the Caribbean where it was the only haplotype identified for both populations.

Table 5.2: Summary of the geographic distribution and frequency of the 16S rRNA haplotypes from *Achatina fulica* populations as evaluated by SSCP analysis.

Locality	Sample size (& no. of haplotypes)	Haplotype name (& no. of individuals)
Kampala, Uganda	5 (1)	O (5)
Dar Es Salaam, Tanzania	38 (6)	I (11) J (1) K (12) L (10) M (2) N (2)
Mayotte	50 (6)	A (1) B (1) C (6) D (40) G (1) H (1)
Souillac, Mauritius	45 (3)	C (34) D (9) H (2)
Mahe, Seychelles	2 (1)	C (2)
Bharatpur, Nepal	22 (1)	C (22)
Yangon, Myanmar/ Burma	20 (1)	C (20)
Trok Nong Area, Chantaburi, Thailand	20 (1)	C (20)
Hahasima, Ogasawara/ Bonin	12 (1)	C (12)
Quezon City, Philippines	20 (1)	C (20)
Los Baños, Philippines	20 (2)	C (17) E (3)
Singapore	20 (1)	C (20)
Kota Kinabalu, Sabah, Malaysia	20 (1)	C (20)
Noumea, New Caledonia	21 (1)	F (21)
Moaroa Valley, Tahiti, French Polynesia	5 (1)	C (5)
Haapiti Valley, Moorea, French Polynesia	10 (1)	C (10)
Kaneohe, Oahu, Hawaii	20 (1)	C (20)
Martinique (captive bred F1 population, Nancy, Lorraine, France)	20 (1)	C (20)
Barbados	12 (1)	F (12)
TOTAL	382 (15)	A (1) I (11) B (1) J (1) C (248) K (12) D (49) L (10) E (3) M (2) F (33) N (2) G (1) O (5) H (3)

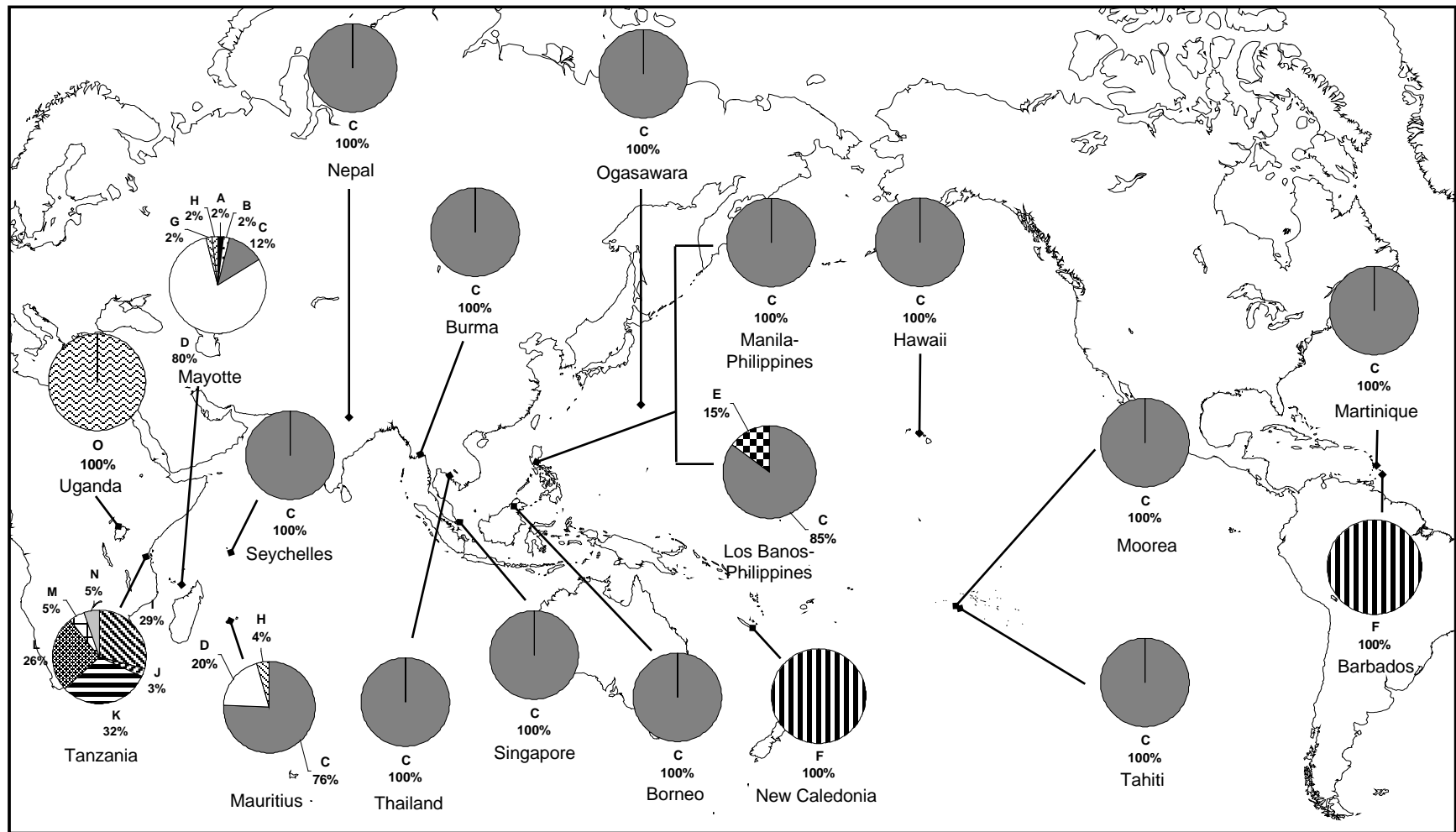


Figure 5.4: Distribution map of the 15 *Lissachatina fulica* populations and their 16S rRNA haplotypes.

5.3.2. Phylogenetic and network analyses of the 16S rRNA haplotypes

A neighbor-joining phylogenetic tree showing the interrelationships among the 15 haplotypes is shown in Figure 5.5 (see also alignment in Appendix 5.2, pp. 523-524). The tree was built using the optimal HKY85 model of DNA sequence evolution (see Appendix 5.3 for the LRT results, p. 525) and is rooted on *A. reticulata*. In general, there is very little phylogenetic structure within the tree and the interrelationships among the haplotypes remain largely unresolved. Nevertheless, two clades are apparent. The first comprises five of the six Tanzanian haplotypes (I, J, L, M and N) and is supported in 96% NJ bootstraps. The second comprises the pan-tropical (Mayotte) haplotype C, the East African (Mayotte) haplotype H, the Philippine haplotype E and the New Caledonian/Barbados haplotype F, albeit with only 54% NJ bootstrap support.

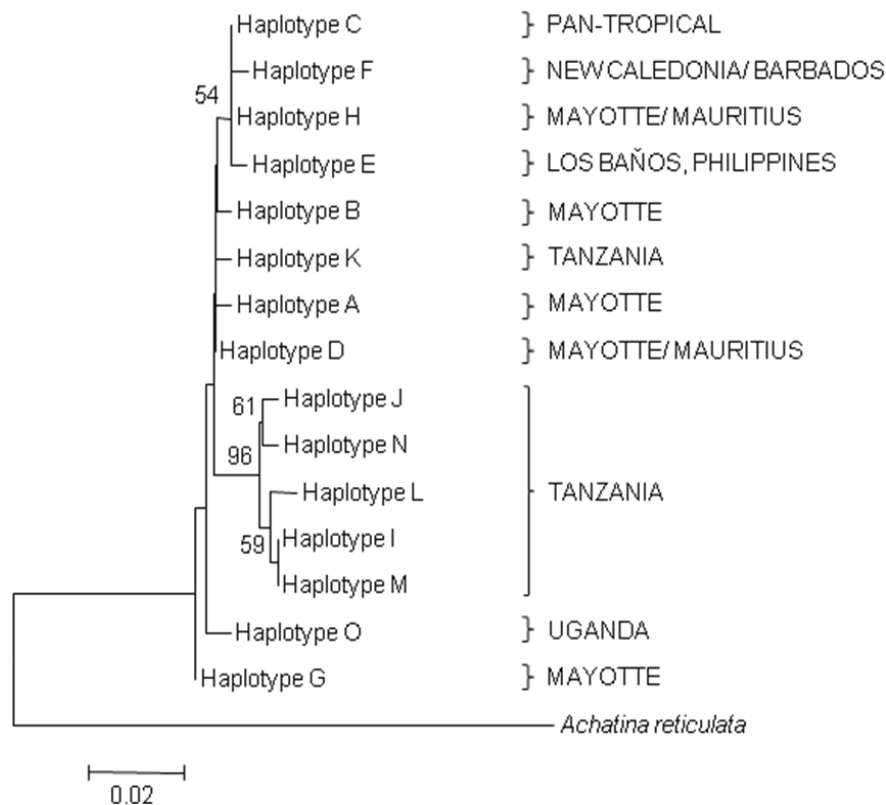


Figure 5.5: Neighbor-joining tree of the 15 *Achatina fulica* 16S rRNA haplotypes based on 293 unambiguously aligned nucleotide sites and using the optimal HKY model. The tree was rooted on the East African *Achatina reticulata*. Bootstrap values indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar represents two substitutional changes per 100 nucleotide positions.

Median network analysis (Figure 5.6) showed that the Tanzanian haplotypes (I, J, L, M and N) that formed a clade in the NJ tree were also grouped together in the network analysis where they were linked to the Mayotte haplotype D with three substitutional steps. The Tanzanian haplotype K was also linked to haplotype D by two substitutional steps but was separated from the other Tanzanian haplotypes. Other haplotypes linked to the Mayotte haplotype D were the Ugandan haplotype O, Mayotte haplotype G and the pantropical (Mayotte) haplotype C with one substitutional step each. Mayotte haplotype H, in turn, was linked to haplotype C by one substitution. Mayotte haplotypes A and B probably originated independently by one substitutional

step from a hypothetical haplotype, which, in turn, was probably only one substitutional step away from either D or H. The non-African haplotypes (E and F) arose through a single mutation each from the pantropical (Mayotte) haplotype C.

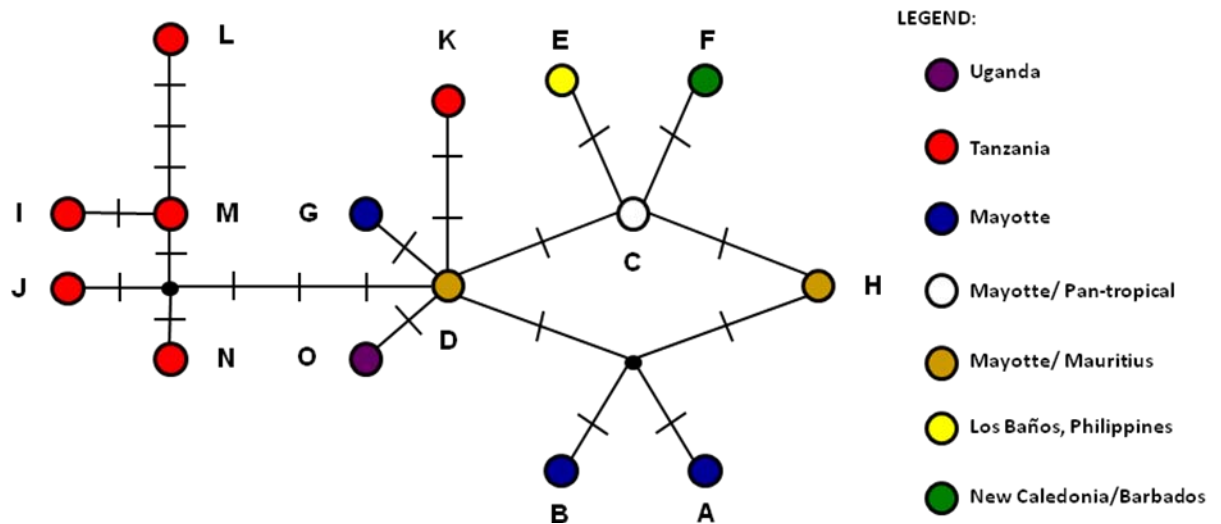


Figure 5.6: Median-joining network of the 15 *Achatina fulica* 16S haplotypes. Each mutation between haplotypes is represented by a bar. Unsourced putative haplotypes are represented by black dots.

5.4. Discussion

Results of the SSCP analysis, NJ tree and median-joining network indicated that all *Achatina fulica* populations outside East Africa and adjacent islands descended from haplotype C, presumably from a few individuals, as there is a complete absence of any other East African haplotypes among the non-African populations. Multiple introductions that could offset the lack of genetic variation as a consequence of founder events were therefore not detected. In India in particular, historical records suggest that the prevalence of *Achatina fulica* could be traced back to the introduction of two individuals from Mauritius to Calcutta by the malacologist William Benson in 1847 (Naggs, 1997). These snails were released by Benson's friend into his garden (Naggs,

1997) and then began to proliferate and slowly spread to the rest of India and later to Nepal (Raut & Barker, 2002; Budha & Naggs, 2008). Though samples were not available from India, particularly from Calcutta, they are expected to bear haplotype C because the population from Nepal also harbored this haplotype. Haplotype C was also the most common haplotype found in Mauritius, the source of Benson's *A. fulica*. The Giant African Land Snail was introduced separately to Sri Lanka from India and Mauritius by Oliver Collett sometime in 1900 (Mead, 1961; Naggs, 1997), and it would be worthwhile to investigate if *A. fulica* individuals in that island are also of the C haplotype or if they harbor other African haplotypes. It is highly likely, however, that the Sri Lankan *A. fulica* would carry the C haplotype as the snail populations from Peninsular Malaysia were derived from Sri Lanka, and Peninsular Malaysia, in turn, became the source of the snail for Burma, Singapore and Thailand (Mead, 1961), all of which bear the C haplotype; Singapore also became the source of the snail for Borneo (Jarrett, 1931), and the population there also carried the C haplotype. A similar case to India also took place in Hawaii where the snails descended from two individuals introduced there from Taiwan (Formosa) in 1936 (Pemberton, 1938), which in turn got the snail from Singapore after 1917 (Mead, 1961). As the *A. fulica* populations in Hawaii and Singapore are of the C haplotype, *A. fulica* populations in Taiwan would therefore also be expected to harbor the C haplotype.

The median-joining network demonstrated that the non-African haplotypes E (Philippines) and F (New Caledonia and Barbados) were probably recent mutations and were derived from haplotype C. To date, haplotype E has only been found in the Philippines where it is present alongside the pantropical haplotype C. It is therefore likely that haplotype E originated in the Philippines and since we know that *Achatina fulica* was introduced in the Philippines in 1931 (Muniappan *et al.*, 1986) this would

suggest that the single nucleotide substitution from C to E took place within the last 70 years or so.

The presence of F in areas that are far apart from each other such as New Caledonia in the Pacific and Barbados in the Caribbean would indicate that either both areas had the same source or that the snails from Barbados originated from New Caledonia as the snail was introduced earlier in New Caledonia (Lambert, 1974). A less likely possibility is that haplotype F arose independently by mutation in both places. Further sampling in New Caledonia would determine if the mutation from C to F took place there or elsewhere. To date, only haplotype F has been found in New Caledonia but so far sampling has been restricted to a single site in Noumea.

The relationship of the East African haplotypes from Tanzania and Uganda with Mayotte is less clear due to limited sampling in the region. It is believed that the Giant African Land Snail was introduced first in Madagascar from Kenya prior to 1800, and it is these Madagascan *A. fulica* snails that were eventually introduced to nearby islands (Bequaert, 1950; Raut & Barker, 2002) such as Mayotte. This raises some very important questions. Is haplotype C found in Madagascar and mainland East Africa? If so, then this would suggest that C was originally present in East Africa and was brought to nearby islands such as Mayotte. If C is *not* present in mainland East Africa, then this would indicate that this haplotype originated on islands off East Africa, possibly on Madagascar or indeed even on Mayotte itself. This question also applies to the other Mayotte haplotypes. Are they also found in Madagascar and mainland East Africa? Clearly, sampling from East Africa and Madagascar is necessary in order to answer these questions and get a clearer picture of the pattern of genetic variation in *A. fulica* populations in East Africa.

The use in this study of a single mitochondrial gene marker such as the 16S rRNA was sufficient to show that all the non-African *Achatina fulica* populations were very closely related and are undergoing bottlenecks as measured by the reduction or loss of genetic variation due to founder events. Whether or not this observation extends to other genes warrants further investigation. Using different markers to measure genetic variation can lead to conflicting results because these markers evolve independently of each other or may have varying degrees of sensitivity. This was proven to be the case when the effectiveness of various molecular markers on native and introduced North American house finches (*Carpodacus mexicanus*) were compared. Hawley *et al.* (2008) found that AFLP, mitochondrial DNA RFLP and allozyme markers did not show statistically significant difference in the degree of variability between introduced and native finches whereas 12 microsatellite loci and ND2 mitochondrial sequences did. They recommended that rapidly evolving single-locus haploid mitochondrial markers be used in conjunction with multi-locus variable nuclear microsatellite markers to detect the presence of recent bottlenecks.

Whether the lack of genetic variation has any effect on the overall fitness of the *A. fulica* populations is another matter altogether. Mead (1979) noted that the snail frequently undergoes three phases after establishment in a new area: (1) an exponential increase characterized by large individuals; (2) a stable phase of variable duration; and (3) decline exemplified by small individuals. Civeyrel and Simberloff (1996) also cited similar patterns in other introduced species; these include the cane toad (*Bufo marinus*) in Australia, the red-billed leiothrix (*Leiothrix lutea*) in Oahu, Hawaii and the weasel (*Mustela nivalis*) in New Zealand. Mead (1979) speculated that the decline in *A. fulica* could be attributed to the snail's susceptibility to the bacterium *Aeromonas hydrophila*, which causes leucodermic lesions. He based his theory on a study by Dean *et al.* (1970)

in which the bacterium, then called *Aeromonas liquefaciens* but was later synonymised with *Aeromonas hydrophila* by Ewing & Hugh (1974), was isolated from the snails and a positive correlation was detected between the presence of the bacteria and the manifestation of leucodermic lesions. What leads to the susceptibility is unknown, but Raut & Barker (2002) cited two possible explanations. One is the breakdown of natural resistance caused by stress in a densely populated environment (Mead, 1979) and the other is the increased chance of transmission of the bacterium due to crowding (Civeyrel & Simberloff, 1996). An alternative explanation is the depletion of food supply as a result of high population density, leading to starvation of the snails and making them more susceptible to bacterial infection (Civeyrel & Simberloff, 1996) or to any other disease-causing vector. Cowie (pers. comm.) offered yet another explanation in which he speculated that native and genetically diverse populations of *A. fulica* were naturally neither abundant nor large because they harboured the bacterium (or other disease agent). When the introduced populations were released from the disease agent pressure, it allowed them to become abundant and larger; eventually, however, the disease made its way to the new region and re-associated with the snails, reducing population density and snail size.

Whether or not the decline phenomenon observed in introduced *Achatina fulica* populations in many areas across the globe is a consequence of founder events is difficult to say. It should be noted that in general, the loss of genetic variation (i.e. heterozygosity and presence of rare alleles) may seriously affect the ability of future generations to adapt to changing environments and subsequently trigger a decline (Backeljau *et al.*, 2001). This was seen in the house finch, *Carpodacus mexicanus*, in which resistance to experimentally infected *Mycoplasma gallisepticum* is associated with increasing multilocus heterozygosity in 12 microsatellites (Hawley *et al.*, 2005). This

led Hawley *et al.* (2008) to speculate that the susceptibility of the house finch to the bacterium in its introduced range in eastern North America, the site of an epidemic in 1994, could be attributed to the reduced genetic variation of the introduced populations as a consequence of founder events. Indeed, their study (2008) revealed reduced genetic variability among the introduced populations of *C. mexicanus* as demonstrated by microsatellite and mitochondrial DNA data. A similar situation could also be taking place among global populations of *A. fulica* in which reduced heterozygosity or variability due to founder events could lead to susceptibility to bacteria or other disease-causing agents in the Giant African Land Snail.

As already mentioned in previously, this study is limited by the availability of sampling, and many crucial areas were not covered, particularly from East Africa (i.e. Kenya and Madagascar), the Indian subcontinent (Calcutta in India and Sri Lanka) and even in the Caribbean and South America (Brazil). The logical next step is to gather samples from these localities. Sampling from Kenya and Madagascar will most likely provide additional haplotypes not found in Uganda, Tanzania and Mayotte, which could give a clearer picture of the pattern of genetic variation in East Africa. Samples from Calcutta in India would determine if indeed the snails in India descended from two haplotype C individuals (if only C are found) or if multiple introductions took place (if other haplotypes are found). Likewise, samples from Sri Lanka would determine whether other haplotypes are present on the island, which could indicate more than one introduction. Sri Lanka is crucial as it served as a source of the Giant African Land Snail for some countries like Malaysia that ultimately became the source for many other countries. Additional sampling from the other Caribbean islands such as Guadeloupe could show if other areas also harbor haplotype F and if introduction occurred from different sources. The presence of *A. fulica* in Brazil is a result of recent introduction to

raise snails for food (Paiva, 1999; Carvalho de Vasconcellos & Pile, 2001; Thiengo *et al.*, 2007) and was probably derived from Indonesia (Thiengo *et al.*, 2007). Sampling from either Brazil or Indonesia will determine whether the pantropical haplotype C is also present in these areas or whether the populations in these areas comprise other haplotypes. Other South American countries such as Colombia, Ecuador and Peru are also worth investigating to track the spread of the snail in the continent.

5.5. Summary

The extent of genetic diversity in *Achatina fulica* populations was investigated using single-stranded conformational polymorphism (SSCP) analysis. A fragment of the 16S rRNA gene found within the variable domain IV was used as marker for the SSCP analysis. Fifteen haplotypes were identified from populations of *A. fulica*, but only haplotype C emerged to spread across the globe. Two non-African haplotypes (E from the Philippines and F from New Caledonia and Barbados) were recent mutations derived from haplotype C as shown by the neighbor-joining phylogenetic and median-joining network analyses. Non-East African populations of *A. fulica* therefore exhibited a lack of genetic variation based on the 16S rRNA marker, implying bottlenecking has taken place in these populations.

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CHAPTER 6 - The SSU rRNA gene as a genetic marker for identifying infective 3rd juvenile stage *Angiostrongylus cantonensis* from known gastropod intermediate hosts¹

6.1. Introduction

The rat lungworm, *Angiostrongylus cantonensis* (Chen), is a nematode parasite of murid rodents (Ubelaker, 1986; Marquardt *et al.*, 2000). Its life cycle requires invertebrate intermediate hosts such as snails and slugs (Roberts & Janovy, 2005). The lungworm can also accidentally infect humans where it causes eosinophilic meningoencephalitis (EME) or neurologic angiostrongyliasis, a disease of the central nervous system that leads to headache, vomiting, nausea, neck stiffness and photophobia (Kliks & Palumbo, 1992; Senanayake *et al.*, 2003; see also Section 1.8.5 of Chapter 1, pp. 26-28). Humans become infected with the 3rd juvenile stage *A. cantonensis* by handling infected snails (Wan & Weng, 2004) or by eating raw/undercooked infected snails or contaminated salad crops (Marquardt *et al.*, 2000). Reported cases of gastropod, rat and human infection reflect the wide geographic distribution of *A. cantonensis*, from East Africa and Madagascar to the Indian subcontinent, East and Southeast Asia, Australia, most islands in the Pacific (Kliks & Palumbo, 1992; Marquardt *et al.*, 2000) and the Caribbean (Raccurt *et al.*, 2003; Waugh *et al.*, 2005).

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6.1.1. Problems with identifying nematodes: the angiostrongylids as examples

Accurately identifying medically important nematodes such as *A. cantonensis* is crucial in diagnosing and controlling the diseases that they cause (Gasser & Newton, 2000). However, it is difficult to identify worms to species level due to a lack of suitable morphological characters in both the adult (Newton *et al.*, 1998a; Roberts & Janovy, 2005) and, more importantly, the infective juvenile stages (Newton *et al.*, 1998a). Some examples of these medically important nematodes include various species of the hookworm *Ancylostoma* as well as the other hookworm *Necator americanus*, in which the juveniles of these worms, which are difficult to distinguish from one another, infect humans either through the oral route or by penetration of the skin and cause hookworm disease. Other examples include different species of *Strongyloides*, most notably *S. stercoralis*, whose juveniles infect humans by contact through contaminated water or soil and cause strongyloidiasis, leading to damage of the skin, lungs and intestines; juveniles of *Strongyloides* are also difficult to detect and identify (Roberts & Janovy, 2005).

Identification of members of the order Strongylida such as the angiostrongylids relies heavily on expert knowledge of the morphological characteristics of the copulatory bursa found only in adult males, which it uses to wrap around the body of the female during copulation (Newton *et al.*, 1998a; Roberts & Janovy, 2005). For example, adult *A. cantonensis* can only be distinguished from the closely related species *A. vasorum* (Baillet) by differences in the copulatory bursa; in *A. cantonensis* the lateral rays arise from a single trunk whereas in *A. vasorum* the lateral rays arise separately (see Fig. 1.3 in Chapter 1, p. 23) (Ubelaker, 1986). Complicating the identification of nematodes is the ambiguity of morphological characters of the juvenile worms (Newton *et al.*, 1998a). For instance, the infective 3rd juvenile stages of *A. cantonensis* and *A.*

vasorum are differentiated only through the fine point termination of the tail of the former and the digitiform termination of the tail in the latter (Ash, 1970, see Fig. 6.1). Whether juveniles of other *Angiostrongylus* species can also be distinguished based on differences in the tail is not known, and some of these species may possibly exhibit similar tail terminations to those of either *A. cantonensis* or *A. vasorum*. In order to identify these juvenile angiostrongylids with certainty, they must be fed to laboratory mice until such time the adults can be observed (Wallace & Rosen, 1969). However, this process takes time, is tedious and does not guarantee that all the juveniles will mature into adults. A viable alternative is clearly needed to identify the 3rd juvenile stage *A. cantonensis*.

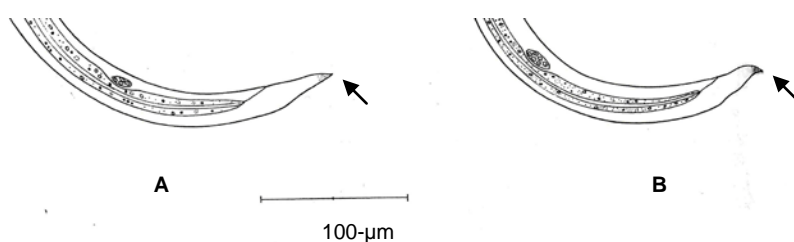


Figure 6.1: Posterior portion of the infective third juvenile stage of (A) *Angiostrongylus cantonensis* and (B) *Angiostrongylus vasorum*. Arrows indicate the type of termination of the tail for each nematode (e.g. fine point for *A. cantonensis* and digitiform, not fine point for *A. vasorum*). Taken from Ash (1970).

6.1.2. Identifying nematodes using molecular markers

In lieu of morphological expertise, the problem of identifying nematodes accurately and rapidly can be addressed by using various molecular genetic markers that are applicable at any stage of a nematode's development. Examples of these molecular markers are nuclear genes such as the ribosomal RNA genes and their internal transcribed spacers (ITS) (Newton *et al.*, 1998a; Gasser, 1999; Floyd *et al.*, 2002; Blaxter *et al.*, 2005) and mitochondrial genes like cytochrome c oxidase I (Caldeira *et*

al., 2003; Hebert *et al.*, 2003; Blaxter *et al.*, 2005). Using these markers requires PCR-based techniques that include polymerase chain reaction-restriction fragment length polymorphisms (PCR-RFLP) analysis and PCR-direct sequencing (Gasser, 1999).

In PCR-RFLP, PCR products from DNA target regions are digested by restriction enzymes. Depending on the sequence variations within the restriction sites that may result in either their recognition or non-recognition by restriction enzymes, fragments of varying lengths will be produced among different species. Through gel electrophoresis, each species can potentially generate a unique banding profile based on these fragments (Gasser, 1999). Newton *et al.* (1998a) employed this technique on the ITS-2 region between the 5.8S and the large subunit (LSU) ribosomal (r) RNA genes to characterize 24 previously described nematode species in the order Strongylida that parasitise livestock. The banding patterns they obtained from both adults and 3rd stage juveniles were unique for each of the species they surveyed except for *Cooperia oncophora* and *C. surnabada*. The ITS-2 can therefore be used to identify most of these parasites at specific developmental stages at which morphological characters are ambiguous. In another study, Caldeira *et al.* (2003) also used PCR-RFLP on the ITS2 as well as the mitochondrial cytochrome c oxidase subunit 1 to distinguish closely related taxa in the Angiostrongylidae such as *A. cantonensis*, *A. costaricensis* and *A. vasorum* using 3rd stage juveniles and adults. However, this technique cannot detect nucleotide substitutions outside the restriction site. Closely related species may share restriction sites but differ outside these restriction sites, thereby producing identical banding patterns after gel electrophoresis.

A more reliable molecular method of identifying nematodes is direct sequencing (Bhadury *et al.*, 2006). The ribosomal genes, which are advantageous for several reasons, have often been used for this purpose. First, they consist of highly variable and

phylogenetically informative regions interspersed with conserved regions that are suitable for primer-binding sites (Palumbi, 1996). Second, they are present as multiple copies of the same sequence (~50-100 copies in nematodes) arranged in tandem arrays across the genome, thus making them easier to PCR amplify (Floyd *et al.*, 2002). Finally, they show very little variation within a species due to concerted evolution (Gasser & Newton, 2000; Floyd *et al.*, 2002). When Newton *et al.* (1998a) generated the same banding profile for *Cooperia oncophora* and *C. surnabada* using PCR-RFLP of the ITS-2, they sequenced this region and discovered that these two taxa have identical sequences, which supports their hypothesis that they may in fact belong to the same species (Newton *et al.*, 1998b). In another example, Bhadury *et al.* (2006) employed a marker near the 5' end of the SSU rRNA gene to rapidly identify marine nematodes collected from southwest England. When they compared their sequences to public databases and conducted phylogenetic analyses, over 97% of their sequences corresponded with the correct species designation based on their morphological identification, and they concluded that amplifying and sequencing a specific region of the SSU rRNA gene could be utilized to rapidly identify nematodes. They accounted for the 3% misidentification either as DNA contamination or distortion of morphological features during ethanol preservation of the nematodes. In another study, Santos *et al.* (2006) successfully employed direct sequencing of the SSU rRNA to detect and identify nematode species in the family Anisakidae (i.e. *Anisakis simplex*, *Contraceacum maculatus*, *Pseudoterranova decipiens* and *Hysterothylacium aduncus*) that parasitise the commercially important sardine *Sardina pilchardus*. Humans get infected by the juvenile stages of these nematodes by eating raw or inadequately processed fish. Still other studies by Qvarnstrom *et al.* (2007) and Hollingsworth *et al.* (2007), which were done concurrently with this study, used a 1134 bp fragment of the

SSU to successfully detect by direct sequencing the presence of *A. cantonensis* from tissue and mucus secretions from gastropod intermediate hosts, primarily the semi-slug *Parmarion* cf. *martensi* in Hawaii. However, only *A. cantonensis* (Carreno & Nadler, 2003) and *A. vasorum* (Chilton *et al.*, 2006) SSU sequences were available to Qvarnstrom *et al.* (2007) and Hollingsworth *et al.* (2007), and their studies were therefore unable to exclude the possibility of detecting other closely related *Angiostrongylus* species. Moreover, sequencing the entire SSU rRNA gene to identify nematodes is not practical if a shorter fragment will suffice. Floyd *et al.* (2002) noted that 50% of the variations found in the SSU rRNA gene of nematodes occur in the first one third from the 5' end (approximately 450 bp), and sequences available for this region for many nematodes in GenBank could distinguish different species. They also considered individuals to belong to the same species if their sequences of the 5' end of the SSU rRNA gene are more than 99.5% identical.

6.1.3. Objectives of the study

The main objective of this study was to develop a quick and reliable molecular method for the identification of infective 3rd juvenile stage *Angiostrongylus cantonensis* based on PCR-direct sequencing a fragment of the small subunit (SSU) ribosomal (r) RNA gene. The SSU rRNA gene was chosen because it is well represented in nematodes in public databases (Floyd *et al.*, 2002) and is variable enough to permit differentiation of closely related nematode species (Gasser & Newton, 2000). The SSU sequence of *A. cantonensis* (voucher specimen no. 92610 of the US National Parasite Collection; identified using published descriptions) became available in Genbank during the course of my PhD studies (Accession AY295804, Carreno & Nadler, 2003) while that of *A. vasorum* (Accession AJ920365, Chilton *et al.*, 2006)

became available after I had already sequenced another individual from the same species (see below). However, there are no full length SSU sequences for many other closely related species that could be mistaken for *A. cantonensis*. To ascertain whether the SSU rRNA gene is indeed suitable to distinguish *A. cantonensis*, the full length SSU for several species of *Angiostrongylus* was sequenced and a suitable variable marker region within the gene determined. In particular, species closely related to *A. cantonensis* were chosen, such as those grouped together by Ubelaker (1986) under *Parastrongylus* based on the morphological characteristics of the male copulatory bursa and host specificity. To test the efficacy of the marker for identifying the 3rd juvenile stage *A. cantonensis*, worms were extracted and identified from gastropod intermediate hosts from a trial area, the Philippines, particularly in Manila, which is known for its prevalence of the parasite (De Leon & Saulog, 1965; Nishimura & Yogore, 1965) and its suburb Quezon City (Salazar & Cabrera, 1969). Furthermore, Salazar & Cabrera (1969) positively identified 3rd juvenile stage *A. cantonensis* in several gastropod intermediate hosts such as the widespread giant African land snail, *Achatina fulica* (with an infection rate of 4.52%), and the common black slug *Laevicaulis alte* (18.18% infected).

As this study resulted in new SSU sequences for several *Angiostrongylus* species, and with two other angiostrongylid SSU sequences already available in GenBank (Carreno & Nadler, 2003; Chilton *et al.*, 2006), a second objective was to conduct phylogenetic analyses of the Angiostrongylidae. These analyses had a dual purpose: to test the monophyly of the group as well as to test Ubelaker's (1986) hypothesis that some members of the genus *Angiostrongylus*, including *A. cantonensis*, are distinct and should be reclassified under the genus *Parastrongylus*.

6.2. Materials and methods

6.2.1. Sequence variation in angiostrongylid worms

Nearly the entire length of the SSU rRNA gene was obtained for four species of Angiostrongylidae: *Angiostrongylus costaricensis*, *A. dujardini*, *A. malaysiensis* and *A. vasorum*. These species were identified, based on published descriptions of the morphological characters of the adult and their definitive hosts, by the taxonomic experts listed in Table 6.1

Table 6.1: Detailed information of the angiostrongylid worms used in this study.

Specimen	Host	Location	Collector/Provider
<i>Angiostrongylus costaricensis</i>	<i>Sigmodon hispidus</i> (cotton rat) & <i>Biomphalaria glabrata</i> (blood fluke planorbid snail)	Maintained in the Department of Parasitology, Hamamatsu University School of Medicine, Japan	Akira Ishih
<i>Angiostrongylus dujardini</i>	<i>Clethrionomys glareolus</i> (red-backed mouse)	Prades, Pyrenees- Orientales, France	Serge Morand & Carlos Feliu
<i>Angiostrongylus malaysiensis</i>	<i>Rattus tiomanicus</i> (Malaysian field rat)	Endau-Rompin National Park, Johor, Malaysia	Siti Nursheena Mohd Zain
<i>Angiostrongylus vasorum</i>	<i>Vulpes vulpes</i> (common fox)	Swansea, Great Britain	Eric Morgan

With the exception of *A. vasorum*, the DNA of which was provided by Eric Morgan of the University of Bristol School of Biological Sciences, all other samples were subjected to DNA extraction following the NaOH direct lysis protocol of Floyd *et al.* (2002) (see Section 2.1.4 of Chapter 2, pp. 41-42).

For the PCR, the 4 overlapping primer sets (see Section 2.2.1.2 of Chapter 2, pp. 53-54) of Blaxter *et al.* (1998) were used to amplify nearly the entire fragment (approximately 1670 nucleotides) of the SSU rDNA gene. PCR amplification, gel

migration and purification of PCR products from agarose gels were done as previously described (see Sections 2.2.1.2 – 2.4 of Chapter 2, pp. 53-59). Both sense and anti-sense strands were sequenced directly using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 termination cycle sequencing chemistry (see Section 2.6 of Chapter 2, pp. 59-61).

Sequences were assembled using the STADEN package version 1.5.3 (Staden *et al.*, 2000) and aligned manually within the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994) together with the previously published SSU sequence of *Angiostrongylus cantonensis* (Carreno & Nadler, 2003). The primer set that amplified the most variable region that is also unique to *A. cantonensis* was chosen to identify the parasite.

6.2.2. Using the SSU rRNA gene to identify *Angiostrongylus cantonensis* in a trial population

The widely distributed Giant African Land Snail, *Achatina fulica*, and the black slug *Laevicaulis alte* were sampled for *A. cantonensis* from a grassy area within the College of Science complex at the University of the Philippines, Diliman campus in Quezon City, Philippines (samples collected by I. Fontanilla). *Angiostrongylus cantonensis* is common in Manila and its suburb Quezon City. Moreover, as rats frequent the Dilman collection site, snails and slugs were highly likely to be infected with *A. cantonensis*.

Snails and slugs were cut into small pieces and digested overnight in a Petri dish containing Ash's (1970) digestive fluid (0.7% pepsin in 0.5% HCl). Individual nematodes were collected under a dissecting microscope using a yellow-tipped pipette

(20-200- μ l capacity) and transferred into wells containing 100- μ l TE buffer in a 96-well microtitre plate. The worms were then stored in a -20 $^{\circ}$ C freezer until use.

DNA extraction, PCR amplification and direct sequencing were undertaken for each nematode using the same protocols described in Section 6.2.1 except that only the primer set that amplified the most variable region among the *Angiostrongylus* species was used (Primer Set A – see results). In cases in which the PCR amplifies products from both the nematode and the snail or slug host, the nematode band can be clearly distinguished as it is smaller (~480 bp) than the snail/slug host band (~510 bp). Once the purified PCR products were obtained, only the antisense strand (amplified by SSU_R09 of Set A) was directly sequenced.

Nematode sequences were processed using the STADEN package, after which distinct nematode sequences were identified and a BLAST search in GenBank was carried out to determine an exact match or identify the closest similarity for each sequence. To complement the BLAST results, a neighbor-joining (NJ) tree based on 376 unambiguously aligned nucleotide sites was generated in order to illustrate the phylogenetic position of the nematode sequences relative to all known nematode sequences in GenBank (GB Release No. 157, see Appendix 6.1, pp 526-536) as well as the new *Angiostrongylus* sequences obtained in this study. The phylogeny was rooted on 4 non-nematode taxa: *Chordodes morgani* (AF036639, Blaxter *et al.*, 1998) and *Gordius aquaticus* (X87985, Winnepenninckx *et al.*, 1995) of Phylum Nematomorpha, *Priapulius caudatus* (Z38009, Aleshin *et al.*, 1998) of Phylum Priapulida, and *Brachionus plicatis* (U49911, Aguinaldo *et al.*, 1997) of Phylum Rotifera. (See Appendix 6.2, pp 537-603 for the Nematoda alignment). The optimal model for DNA sequence evolution was determined using likelihood by estimating the log likelihood scores in PAUP* for the JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P

(Kimura, 1980), HKY85 (Hasegawa *et al.*, 1985), TN93 (Tamura & Nei, 1993) and the GTR (Rodriguez *et al.*, 1990) models as well as their variants that incorporated gamma distributed rates (Γ) (Yang, 1993) followed by comparing these scores for significant differences using the Likelihood Ratio Test (LRT) as described in Section 2.9.5 of Chapter 2, pp. 73-77). Bootstrap resampling (Felsenstein, 1985) with 1000 replicates was carried out. Once the phylogenetic positions of the nematode sequences were identified with specific clades, other nematode taxa that do not belong to these clades were removed in order to construct a smaller NJ tree based on the same fragment (376 sites) and using the best model with optimised parameters; however, a suitable outgroup [*Plectus acuminatus* (AF037628, Blaxter *et al.*, 1998), which falls immediately outside of the Rhabditida clade of Meldal *et al.* (2006) –see results] was retained.

6.2.3. Phylogeny of the Angiostrongylidae

In order to investigate relationships among the Angiostrongylidae, the full length SSU rRNA of all *Angiostrongylus* species that were not sequenced prior to this study as well as *A. cantonensis* from Carreno & Nadler (2003) were aligned with the rest of the sub-order Metastrongylina/ superfamily Metastrongyloidea and phylogenetic trees constructed. Two taxa from the Trichostrongylina, which falls immediately outside of the Metastrongylina based on phylogenetic studies of the Metastrongyloidea (Carreno & Nadler, 2003) and the Strongylida (Chilton *et al.*, 2006), were included in the alignment to serve as outgroups (Table 6.2). Phylogenies were constructed from the aligned dataset using the model based maximum likelihood (ML), Bayesian inference (BI) and neighbor-joining (NJ) methods as well as the non-model based maximum parsimony (MP) method (Section 2.9.8 of Chapter 2, pp. 82-85). A total of 1628 unambiguously aligned nucleotide sites were used for the phylogenies. The sequences were first

checked for base composition bias using the Chi square test in PAUP* (version 4.0b10) program (Swofford, 2002). The optimal model for DNA sequence evolution was then determined for the model-based methods. Bootstrap resampling (Felsenstein, 1985) with 1000 replicates for MP and NJ and 100 replicates for ML was also carried out. Bayesian inference (BI) was performed on the MrBayes (version 3.1.2) package (Ronquist & Huelsenbeck, 2003) using four chains of Markov Chain Monte Carlo algorithm to explore the tree space for 2 million generations with sampling at every 100 generations. The heating parameter was set to 0.125 to ensure adequate chain swapping. A consensus tree was constructed using the last 1000 trees (burnin=19001 samples).

Table 6.2: Taxa used for the sequence analysis of the angiostrongylid worms.

Sub-order	Family	Species	Reference	GenBank Accession #
Metastrongylina (Superfamily Metastrongyloidea)	Angiostrongylidae	<i>Angiostrongylus costaricensis</i> (Morera & Cespedes, 1971)	This study	EF514913
		<i>Angiostrongylus dujardini</i> (Drozdz & Doby, 1969)	This study	EF514915
		<i>Angiostrongylus malaysiensis</i> (Bhaibulaya & Cross, 1971)	This study	EF514914
		<i>Angiostrongylus cantonensis</i> (Chen, 1935)	Carreno & Nadler (2003)	AY295804
		<i>Angiostrongylus vasorum</i> (Baillet, 1866)	This study	EF514916
		<i>Aelurostrongylus abstrusus</i> (Railliet, 1898)	Chilton <i>et al.</i> (2006)	AJ920366
		<i>Didelphostrongylus hayesi</i> Prestwood, 1976	Carreno & Nadler (2003)	AY295806
	Crenosomatidae	<i>Crenosoma mephitidis</i> Hobmaier, 1941	Carreno & Nadler (2003)	AY295805
		<i>Otostrongylus circumlitus</i> (Railliet, 1899)	Carreno & Nadler (2003)	AY295813
		<i>Troglostrongylus wilsoni</i> (Stough, 1953)	Carreno & Nadler (2003)	AY295820
	Filaroididae	<i>Filaroides martis</i> (Werner, 1782)	Carreno & Nadler (2003)	AY295807
		<i>Oslerus osleri</i> (Cobbold, 1889)	Carreno & Nadler (2003)	AY295812
		<i>Parafilaroides decorus</i> Dougherty & Herman, 1947	Carreno & Nadler (2003)	AY295814
	Metastrongylidae	<i>Metastrongylus salmi</i> (Gedoelst, 1823)	Carreno & Nadler (2003)	AY295809
	Protostrongylidae	<i>Muellerius capillaris</i> (Müller, 1889)	Carreno & Nadler (2003)	AY295810
		<i>Parelaphostrongylus odocoilei</i> (Hobmaier & Hobmaier, 1934)	Carreno & Nadler (2003)	AY295815
	Pseudaliidae	<i>Halocercus invaginatus</i> (Quekett, 1841)	Carreno & Nadler (2003)	AY295808
		<i>Pseudalius inflexus</i> (Rudolphi, 1808)	Carreno & Nadler (2003)	AY295816
		<i>Stenurus minor</i> (Kühn, 1829)	Carreno & Nadler (2003)	AY295817
		<i>Torynurus convolutus</i> (Kühn, 1829)	Carreno & Nadler (2003)	AY295818
	Skrjabinylidae	<i>Skrjabinylus chitwoodrum</i> Hill, 1939	Carreno & Nadler (2003)	AY295819
Trichostrongylina (Superfamily Trichostrongylina) --OUTGROUP--	Molineidae	<i>Nematodirus battus</i> (Crofton and Thomas, 1951)	Zarlenga <i>et al.</i> (1994)	U01230
	Heligmonellidae	<i>Nippostrongylus brasiliensis</i> (Travassos, 1914)	Blaxter <i>et al.</i> (1998)	AF036597

6.3. Results

6.3.1. Sequence variation in the angiostrongylid worms

When the sequences of all five *Angiostrongylus* species available to date (including the four new *Angiostrongylus* sequences obtained in this study) were compared, a total of 31 variable sites were found, including an indel at position 1323 (Fig. 6.2). A pairwise comparison of the number of differences between any two *Angiostrongylus* species (Table 6.3) reveals that the least number of differences (9) was found between *A. cantonensis* and *A. malaysiensis* whereas the greatest number of differences (22) was seen between *A. malaysiensis* and *A. dujardini*. The sequences of the SSU rRNA clearly demonstrate that *A. cantonensis* has a distinct sequence that could be used to distinguish it from the other angiostrongylid worms in this study. Furthermore, 12 out of the 31 sites (or more than a third of the total) were within the first 480 bp from the 5' end. Moreover, *A. cantonensis* is distinct from all other *Angiostrongylus* species in this 480 bp 5' region. These findings demonstrate that the first 480 bp fragment of the SSU rRNA gene, which is amplified by primer Set A (see Section 2.2.1.2, pp. 53-54, and Table 2.6, p. 54 of Chapter 2), is a suitable marker to identify *A. cantonensis* and set it apart from other *Angiostrongylus* species sequenced to date, including three of its closest relatives, *A. costaricensis*, *A. dujardini* and *A. malaysiensis*, based on the morphology of the copulatory bursa (Ubelaker, 1986).

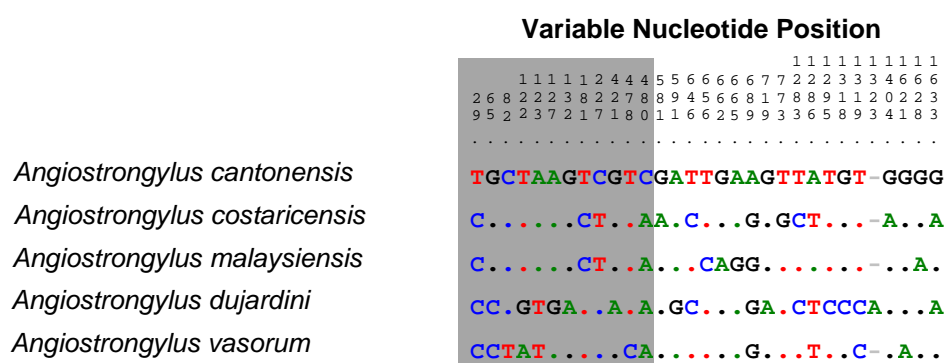


Figure 6.2: Variable sites across the SSU rDNA region for the previously sequenced *Angiostrongylus cantonensis* (Carreno & Nadler, 2003) and the four other angiostrongylid worms sequenced in this study. The number shown directly above each site pertains to the variable site relative to the SSU rRNA sequence of *A. cantonensis* (GenBank AY295804). The grey area corresponds to the 480 bp marker used for this study.

Table 6.3: Matrix showing the number of differences between any two *Angiostrongylus* species.

	<i>A. cantonensis</i>	<i>A. costaricensis</i>	<i>A. malaysiensis</i>	<i>A. dujardini</i>
<i>A. costaricensis</i>	12			
<i>A. malaysiensis</i>	9	11		
<i>A. dujardini</i>	19	16	22	
<i>A. vasorum</i>	11	15	14	16

6.3.2. Using the SSU rRNA gene to identify *Angiostrongylus cantonensis* in a trial population

In total, 27 nematodes were extracted from 15 *Achatina fulica* individuals (with 6 nematodes) and 19 *Laevicaulis alte* individuals (with 21 nematodes) from the University of the Philippines, Diliman campus in Quezon City. The 480 bp fragment at the 5' end of the SSU rRNA gene identified as a suitable marker for the discrimination of *A. cantonensis* (see Section 6.3.2, fragment amplified by Primer Set A) was then amplified for each nematode extracted. The results yielded two distinct nematode sequences (Philippine Sequences 1 and 2). Table 6.4 lists the distribution of these nematode sequences.

Table 6.4: Summary of the distribution of the two Philippine nematode sequences from the University of the Philippines, Diliman campus in Quezon City.

Nematode sequence	Gastropod host	Number of nematodes
Philippine Sequence 1	<i>Achatina fulica</i>	1
	<i>Laevicaulis alte</i>	20
Philippine Sequence 2	<i>Achatina fulica</i>	5
	<i>Laevicaulis alte</i>	1

The two Philippine nematode sequences were then subjected to a BLAST search in GenBank (GB Release No. 157, 22 December 2006), to identify an exact match or closest identity to a named nematode species. BLAST results showed Philippine Sequence 1 is identical to *Angiostrongylus cantonensis*. On the other hand, Philippine Sequence 2 did not match any named taxon in GenBank based on BLAST results but can only be assigned to its closest sequence identity to a named nematode species, which is *Oslerus osleri*, a dog lungworm, at 99% BLAST identity; since the value is less than 99.5% to be considered the same species as *O. osleri* based on Floyd *et al.* (2002), the exact species identification of Philippine Sequence 2 could therefore not be known as of GB Release No. 157.

To complement the BLAST results, a neighbor-joining tree of the Nematoda was generated in order to illustrate the phylogenetic position of the new nematode sequences relative to all 349 nematode taxa having at least the first 480 nucleotides of the 5' end of the SSU rRNA in GenBank (GB Release No. 157) as well as the new *Angiostrongylus* sequences obtained in this study. A GTR+ Γ model was used to correct distances for multiple hits after it was determined to be the best model (see Appendix 6.3A, p. 604). As this phylogeny is based on a small marker intended for barcoding purposes, it should not be expected to accurately show the deep-level evolutionary relationships amongst the nematodes (see Hajibabaei *et al.*, 2007 for a discussion of the limitations of “barcode” markers in phylogenetic studies). Nevertheless, the phylogeny

clearly demonstrates that the two Philippine sequences and all the *Angiostrongylus* taxa fall within the Rhabditida clade (as defined by Meldal *et al.*, 2006). Another NJ phylogeny of the same gene fragment (376 nucleotide sites) was then constructed (Fig. 6.3B) using 229 rhabditid taxa and rooted on the plectid nematode *Plectus acuminatus*. A GTR+ Γ model was determined to be the best model and was subsequently used for this analysis (see Appendix 6.3B, p. 604). The two Philippine nematode sequences and the *Angiostrongylus* taxa all fall within the Strongyloidea. Philippine Sequence 1 clusters with *Angiostrongylus cantonensis* (AY295804, Carreno & Nadler, 2003) with 100% BLAST identity. Philippine Sequence 2 clusters with *Oslerus osleri* (AY295812, Carreno & Nadler, 2003) with 99% BLAST identity.

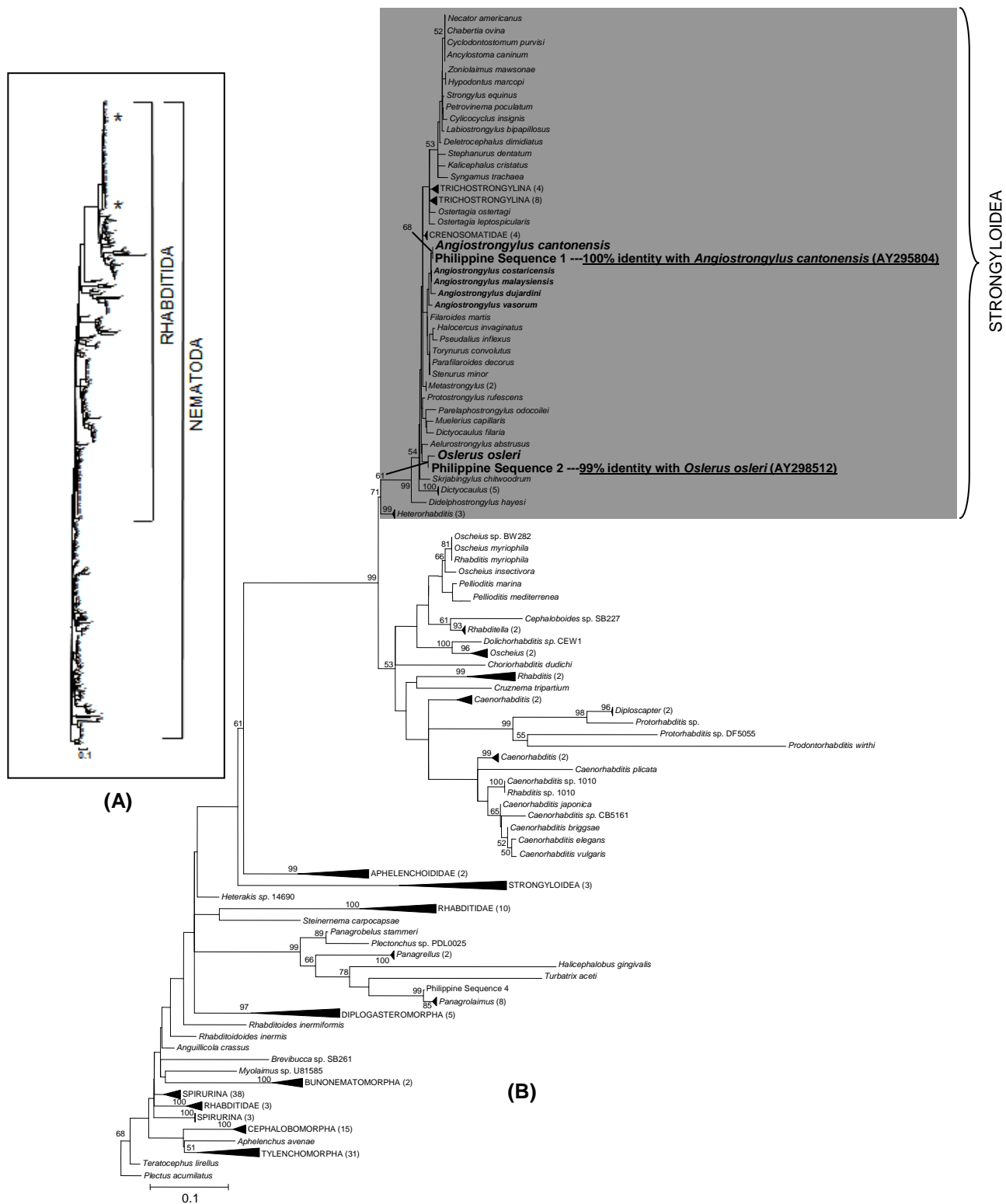


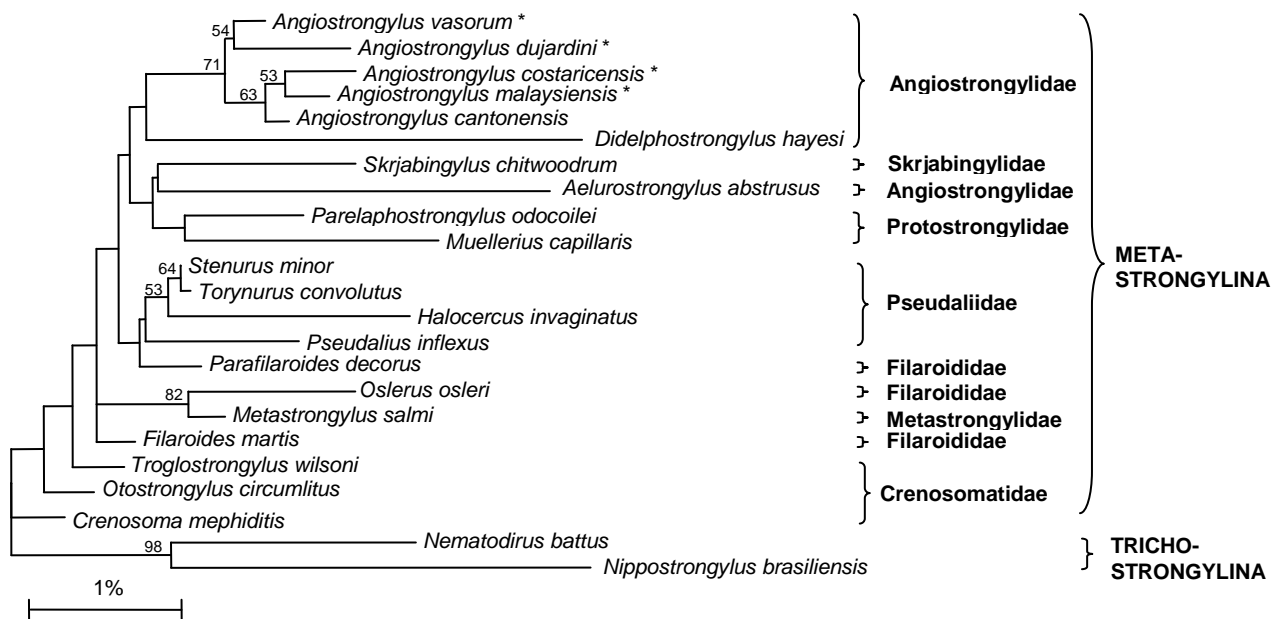
Figure 6.3: Neighbor-joining (NJ) phylogenetic trees based on 376 unambiguously aligned nucleotide sties of the SSU rRNA gene. (A) NJ tree of the Nematoda (349 taxa excluding the new nematode sequences from the Philippines) rooted on 4 non-nematode taxa: *Chordodes morgani* (Nematomorpha), *Gordius aquaticus* (Nematomorpha), *Priapulius caudatus* (Priapulida) and *Brachionus plicatis* (Rotifera). Both Philippine nematode sequences, as indicated by (*), fall within the Rhabditida clade. The scale bar shows one nucleotide change per ten nucleotide positions. (B) NJ tree of the Rhabditida (229 taxa excluding Philippine sequences) from (A), which is rooted on the nematode plectid worm *Plectus acuminatus*, and showing the positions (indicated by lines) of the 7 Philippine nematode sequences as well as their GenBank BLAST results from Table 6.6. Bootstrap values indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. Numbers in parentheses denote the number of species within a clade.

6.3.3. Phylogeny of the Angiostrongylidae

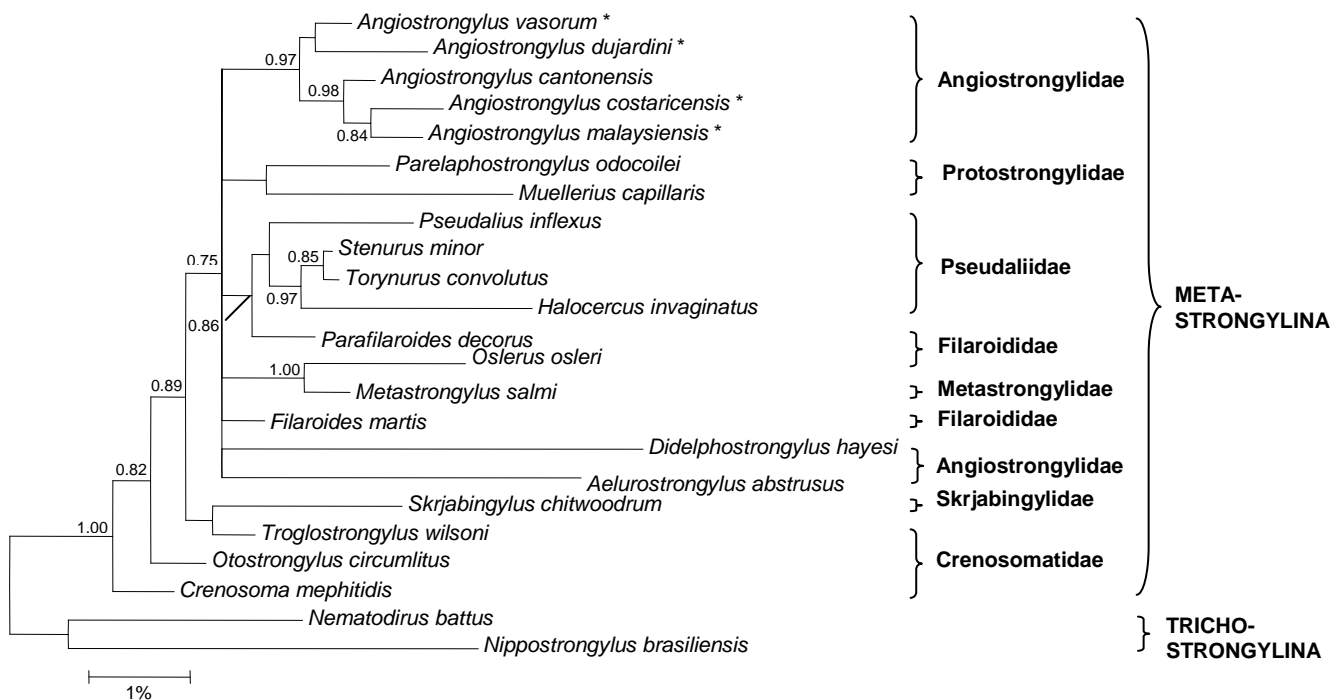
Phylogenetic analyses were undertaken to determine the relationships among the *Angiostrongylus* species sequenced to date (including the four angiostrongylid species sequenced for the first time in this study) and to determine the phylogenetic position of the Angiostrongylidae within the Sub-order Metastrongylina/ Superfamily Metastrongyloidea. (See alignment on Appendix 6.4, pp. 605-615.) Tests for base composition biases using the Chi square test in PAUP* showed no composition bias ($P=1.0$). GTR+ Γ was determined to be the best model (Appendix 6.5, p. 616) and was subsequently used for tree building in the model-based tree construction methods (ML, BI and NJ).

The maximum likelihood (ML), Bayesian inference (BI), neighbor-joining (NJ) and maximum parsimony (MP) phylogenies are shown in Figure 6.4. The trees were constructed using 1628 unambiguously aligned nucleotide sites and were rooted on the trichostrongylids *Nematodirus battus* (GenBank U01230, Zarlenga *et al.*, 1994) and *Nippostrongylus brasiliensis* (Genbank AF036597, Blaxter *et al.*, 1998).

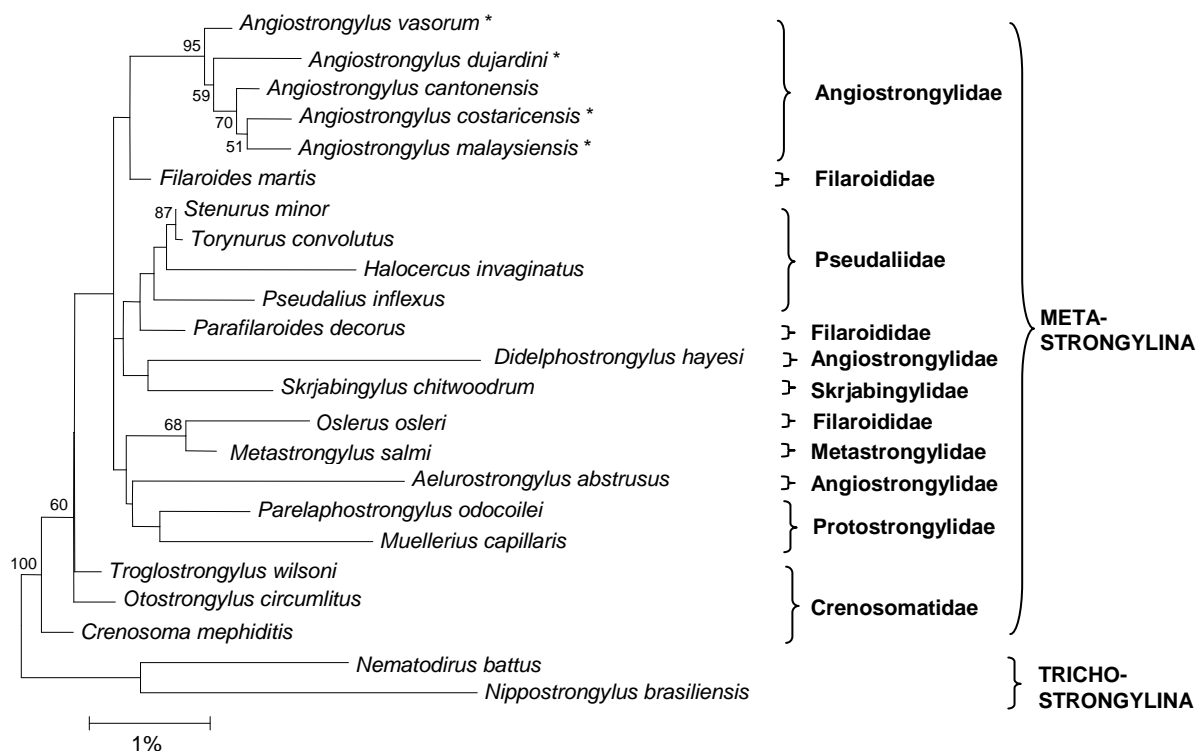
Figure 6.4: (See next two pages.) Phylogenetic trees of the angiostrongylids and worms in other families of the sub-order Metastrongylina based on the SSU rRNA gene using (A) maximum likelihood, (B) Bayesian analysis, (C) neighbor-joining and (D) maximum parsimony based on majority rule consensus of 3 equally parsimonious trees. The phylogenies are constructed from 1628 unambiguously aligned nucleotide sites and are rooted on the trichostrongylid worms *Nematodirus battus* (GenBank U01230) and *Nippostrongylus brasiliensis* (Genbank AF036597). Bootstrap values in (A) indicate the percentage support for individual branches based on 100 replicates. Bootstrap values lower than 50% are not shown. Bayesian posterior probabilities in (B) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (C) and (D) indicate the percentage support for individual branches based on 1000 replicates. The scale bars for (A), (B) and (C) represent one substitutional change per 100 nucleotide positions. *Angiostrongylus* species not sequenced in previous studies are highlighted with (*).



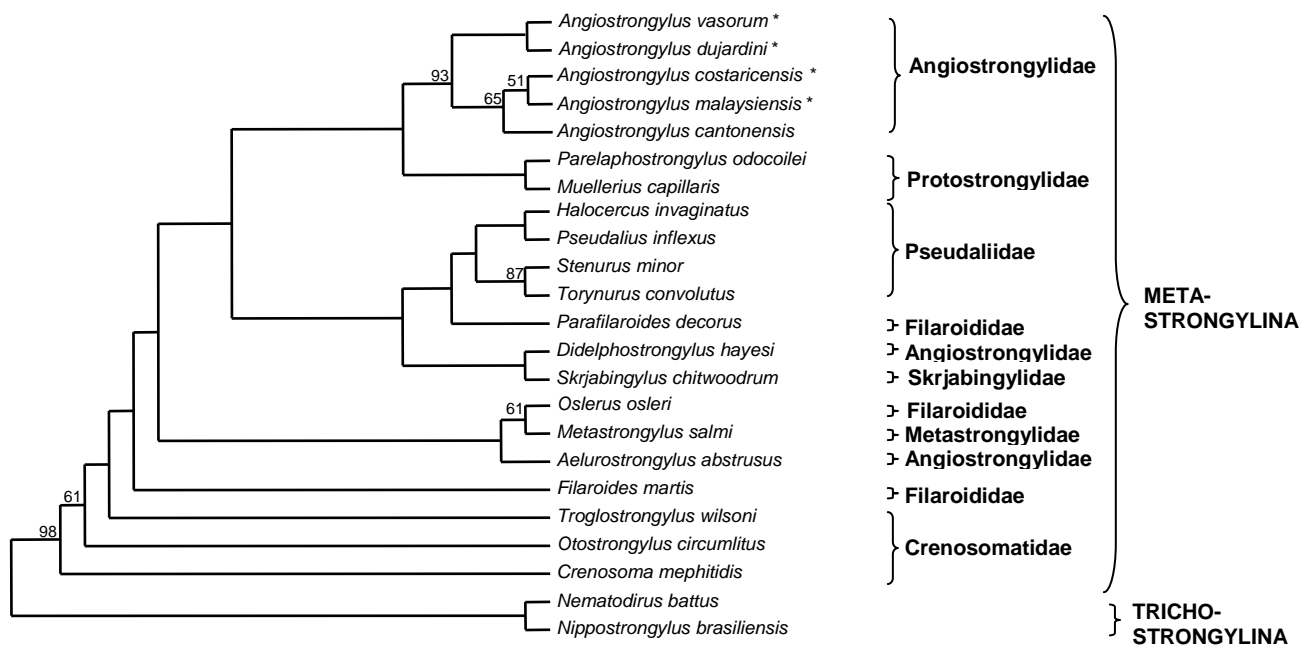
(A) - ML



(B) - BI



(C) - NJ



(D) - MP

The four phylogenies reveal several important points. First, monophyly of the Angiostrongylidae is neither conclusively supported nor rejected. For instance, in the ML tree (Fig. 6.4A) *Aelurostrongylus abstrusus* fell outside the main angiostrongylid group, while in Bayesian (Fig. 6.4B), NJ (Fig. 6.4C) and MP (Fig. 6.4D) analyses, both *Aelurostrongylus abstrusus* and *Didelphostrongylus hayesi* fell outside the main angiostrongylid group, though bootstrap support and posterior probabilities for these branches in the four trees were less than 50% and 0.7, respectively. Second, the phylogenies did not seem to support the distinction between *Angiostrongylus* and *Parastrongylus* as proposed by Ubelaker (1986) based on differences in morphology of the male bursa and the mammalian final host. The grouping of *A. vasorum*, [retained in the genus *Angiostrongylus* by Ubelaker (1986)] with *A. dujardini* [included in the genus *Parastrongylus* (Ubelaker, 1986)], was equivocal in these analyses ($P=0.67$ BI, 54% ML bootstraps, 30.2% NJ bootstraps and 43.9% MP bootstraps). Lastly, *Angiostrongylus cantonensis* was distinct from all other angiostrongylids and clustered with the *A. costaricensis* and *A. malaysiensis* group ($P=0.98$ BI, 68% ML bootstraps, 70% NJ bootstraps and 65% MP bootstraps).

6.4. Discussion

6.4.1. Suitability of the 5' end of the SSU rRNA gene as a molecular marker for *Angiostrongylus cantonensis*

A total of five *Angiostrongylus* species (including four from this study) have now been sequenced for almost the entire region (approximately 1670 nucleotides) of the SSU rRNA gene. All sequences are distinct and can therefore distinguish the *Angiostrongylus* species. However, this almost complete SSU rRNA gene region is too

long to be used as a molecular marker to rapidly and accurately identify specific taxa. When the first 480 bp of the 5' end of the gene was compared across the different *Angiostrongylus* taxa, 12 variable sites (more than a third of the total number of variations for the entire gene) were found. Furthermore, *A. cantonensis* exhibited a unique sequence for this region even when compared to its closest relatives *A. costaricensis*, *A. dujardini* and *A. malaysiensis* (based on morphology and host specificity). Furthermore, this segment of the SSU is available for the majority of the named nematodes in GenBank and encompasses the segment Bhadury *et al.* (2006) used to identify their marine nematodes. These factors make the first 480 bp region eminently suitable as a molecular marker to identify *A. cantonensis* in nematode surveys.

6.4.2. Molecular identification of nematodes from a trial population of *Achatina fulica* and *Laevicaulis alte* in the Philippines

Two distinct nematode sequences were detected by the SSU marker when applied to a trial population of intermediate hosts. Of these, Philippine Sequence 1 was identified as *A. cantonensis* on account of its 100% identity based on GenBank BLAST results of the variable region at the 5' end of the SSU rRNA gene. One hundred percent sequence identity could either mean 'the same taxon' or that 'there is insufficient variation to define taxa' (Floyd *et al.*, 2002). In the case of Philippine Sequence 1, its designation as *A. cantonensis* is valid as the current study already ascertained the uniqueness of the segment of the gene surveyed for this species in comparison to other closely related *Angiostrongylus* species (Figure 6.3).

This study clearly demonstrates the possibility of identifying infective 3rd juvenile stage *A. cantonensis* isolated from slug and snail intermediate hosts using the

5' end of the rRNA gene. This molecular approach to identification is a reliable alternative to morphological identification of nematode samples, especially in cases in which morphological characters are ambiguous in juvenile stages, thus rendering positive identification difficult.

6.4.3. Phylogenetic position of the Angiostrongylidae and the *Angiostrongylus* species within the Metastrongylina

Members of the Angiostrongylidae share certain features such as a posterior vulva but no buccal cavity or lips and the ability to infect a wide range of mammals as final hosts, including marsupials, insectivores, carnivores and rodents (Anderson, 2000; Roberts & Janovy, 2005). However, results from this study based on the SSU rRNA gene do not strongly support the monophyly of this group and show that only the *Angiostrongylus* species form a distinct group. Carreno and Nadler (2003) have already hinted that there is wide morphological diversity within the Angiostrongylidae. A further examination of the Angiostrongylidae that includes a broader taxonomic coverage of the group is required to determine its validity.

Ubelaker (1986) moved certain *Angiostrongylus* species, such as *A. cantonensis*, *A. costaricensis*, *A. dujardini*, *A. mackerrrasae*, *A. malaysiensis*, *A. petrowi*, *A. ryjikovi*, *A. sandarsae*, *A. schmidt*i, and *A. siamensis* to the genus *Parastrongylus*, which was first described by Baylis (1928) based on *Parastrongylus tateronae* as the type species. This is to distinguish *Parastrongylus* from *Angiostrongylus*, first described by Baillet (1866) based on *A. vasorum* as the type species and includes *A. chabaudi* and *A. raillet*i. This distinction was based on differences of the morphology of the male bursa and the mammalian final host. However, this distinction is not widely accepted, and

molecular data from this study based on five *Angiostrongylus* species neither conclusively support nor reject this distinction.

6.5. Conclusion

An alternative approach to morphological identification of infective 3rd juvenile stage *Angiostrongylus cantonensis* was developed and successfully employed using the 5' end of the SSU rRNA gene (~480 bp). Sequences of other closely related *Angiostrongylus* species were obtained for this region to ascertain the uniqueness of the marker for *A. cantonensis*. When the SSU rRNA marker was used on nematode populations extracted from the giant African land snail, *Achatina fulica*, and the black slug, *Laevicaulis alte*, from the Philippines, the infective 3rd juvenile stage *A. cantonensis* was detected without difficulty. The technique developed in this study therefore enables the rapid and accurate identification of medically important nematode taxa when morphological analysis proves difficult or inadequate.

Phylogenetic analyses based on the full length SSU rRNA of the Metastrongylina with 5 *Angiostrongylus* species and 2 other angiostrongylids do not support the monophyly of the Angiostrongylidae. The same data neither support nor reject Ubelaker's distinction of *Parastrongylus* from *Angiostrongylus*.

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CHAPTER 7 - Survey of *Angiostrongylus cantonensis* (and other nematodes) from global populations of the giant African snail, *Achatina fulica* (and other snails)

7.1. Introduction

Angiostrongylus cantonensis, like one of its gastropod intermediate hosts *Achatina fulica*, has a global distribution (Marquardt *et al.*, 2000) and is a major concern from a human health perspective as it causes the food-borne disease eosinophilic meningoencephalitis (EME) or angiostrongyliasis (Kliks & Palumbo, 1992; Prociv *et al.*, 2000). The presence of *A. cantonensis* in rats in New Orleans, USA (Campbell & Little, 1988) and the recent outbreak of EME among medical students visiting Jamaica (Waugh *et al.*, 2005) illustrate the expanding distribution of the parasite. The life cycle of *A. cantonensis* involves a gastropod intermediate host and a murid rodent definitive host (Fig. 1.4, Chapter 1, p. 25); interestingly, the parasite is not highly specific to either its intermediate or its definitive host, which could help to explain its global distribution (Prociv *et al.*, 2000). The route for human infection by *A. cantonensis* is mostly through the gastropod intermediate host. The consumption of contaminated salad crops or raw/undercooked snails (Marquardt *et al.*, 2000) either as an alternative food source during famine (Kliks & Palumbo, 1992) or as delicacies (Chau *et al.*, 2003), their use as medicine (Kliks & Palumbo, 1992) and the predilection of humans to keep gastropods as pets (Wan & Weng, 2004) has led to sudden outbreaks or isolated cases of eosinophilic meningoencephalitis due to 3rd juvenile stage *A. cantonensis* infection (Kliks & Palumbo, 1992). Some of these outbreaks were attributed to the Giant African Land Snail, *Achatina fulica* (Kliks & Palumbo, 1992), which Alicata (1966) suggested as primarily responsible for the spread of the parasite to Southeast Asia and the Pacific.

7.1.1. Dispersal of *Angiostrongylus cantonensis* from East Africa: Alicata's hypothesis

First reported in rats in Canton, China in 1933 (Chen, 1935), *Angiostrongylus cantonensis* has a wide distribution and infects a range of intermediate and final hosts (Alicata, 1966, Fig. 7.1). The parasite was thought to have spread from East Africa towards the Indian subcontinent until it reached the Malay Peninsula (Alicata, 1966; Marquardt *et al.*, 2000). From there it migrated northwards to Indochina, southern China, Taiwan, and Okinawa, and eastwards to the Philippines, New Guinea, Australia, New Zealand, and several islands and island groups of the Pacific including Palau, Guam, Pohnpei, Pingelap, Moen, Ponape, Mariana, Wallis, Solomon, New Hebrides, New Guinea, Fiji, Loyalty, New Caledonia, Tahiti, Tonga, Rarotonga, Raiatea, Samoa, Tokelau and Hawaii (Kliks & Palumbo, 1992). *Angiostrongylus cantonensis* has also been reported in Western Africa (Ivory Coast, Liberia and Nigeria) (Kliks & Palumbo, 1992; Marquardt *et al.*, 2000), the Middle East (Egypt) (Marquardt *et al.*, 2000), North America (Campbell & Little, 1988; New *et al.*, 1995), and the Caribbean (Cuba, Puerto Rico, Jamaica and Haiti) (Pascual *et al.*, 1981; Andersen *et al.*, 1986; Lindo *et al.*, 2002; Raccurt *et al.*, 2003). Prociv *et al.* (2000) observed a general pattern in the recognition of *A. cantonensis* in new locations that begins with the initial detection in rats and occasionally in captive primates, followed by reported human cases of EME, and finally by the recovery of worms from some patients. Subsequent surveys of gastropod intermediate hosts in these new locations oftentimes lead to the detection of the parasite following confirmation by infecting the juveniles in laboratory rats, as in the study of Wallace & Rosen (1969).

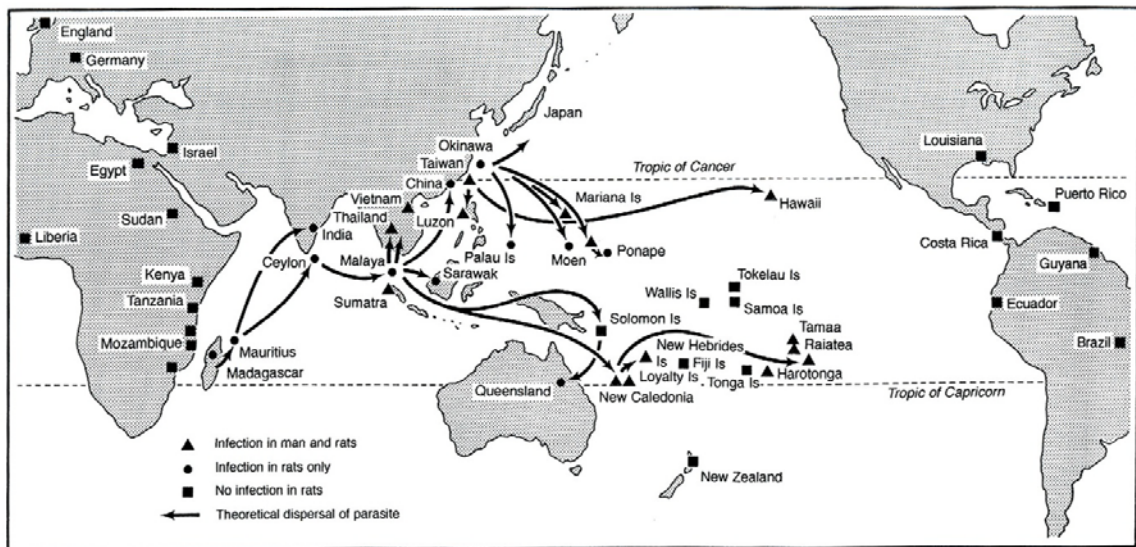


Figure 7.1: Probable dispersal route of *Angiostrongylus cantonensis*. (From Marquardt *et al.*, 2000.)

Alicata's (1966) hypothesis that the worm most probably originated from East Africa and migrated to East Asia and the Pacific is based on three observations. First, *A. cantonensis* is not found in all major Pacific islands despite the prevailing similar environmental conditions. Second, reported cases of eosinophilic meningoencephalitis in the Pacific are relatively recent, from just after the Second World War. Third, the parasite was only reported during the latter half of the 20th century in East Asia, the Pacific and Australia. This hypothesis was further supported by the total absence of the worm in 1967 in Central America (Costa Rica and Puerto Rico) and South America (Brazil, Ecuador and Guyana) based on a survey of 1810 rats (Alicata, 1967) only to appear later on in Cuba in 1973 (Pascual *et al.*, 1981), Puerto Rico in 1984 (Andersen *et al.*, 1986), Jamaica in 2000 (Waugh *et al.*, 2005) and Haiti in 2002 (Raccurt *et al.*, 2003; Marquardt *et al.*, 2000). The parasite was also reported for the first time in North America in New Orleans in 1986 (Campbell & Little, 1988).

It is not yet clear how *A. cantonensis* spread from the Pacific Basin to the Caribbean and beyond, although Kliks and Palumbo (1992) pointed to rats as culprits.

They suspected that infected rats from Indochina were accidentally brought to Cuba between 1966 and 1975 during the Vietnam War, which led to cases of EME beginning in 1973. From there it spread to Puerto Rico where the worms were detected in adult rats in 1981. How the worms got to West Africa, particularly the Ivory Coast and Nigeria is less clear. The first and only case of eosinophilic meningoencephalitis in the Ivory Coast was reported in 1979 (Nozais *et al.*, 1980). In Nigeria, adult worms were also recovered from rats in Port Harcourt but not in Ibadan (Kliks & Palumbo, 1992). Nematodes were also found in Giant African Land Snails in Port Harcourt, reported by Kliks and Palumbo as *Achatina fulica*, though they are more likely to be *Achatina achatina* based on the prevalence of this snail in that area and the fact that *A. fulica* has not yet been officially recorded there.

7.1.2. Perceived role of *Achatina fulica* in the spread of *A. cantonensis*

Alicata (1966) pinned his East Africa origin for *A. cantonensis* on the nearly parallel distribution of the parasite with one of its intermediate hosts, *Achatina fulica*, particularly in Southeast Asia and the Pacific. He based his hypothesis on the observations he made when he surveyed 8 areas from East Africa to the Western Pacific, in which he checked for the presence of *A. cantonensis* in adult rats. He found the parasite in Madagascar, Mauritius, Sri Lanka and Borneo but not in Mozambique, Kenya, Tanzania and India. By combining his findings with what was known at that time regarding the dispersal route of *A. fulica* and the reported cases of rat and human infections of *A. cantonensis*, he concluded that the rapid spread of the snail led to the spread of the parasite based on the following reasons. First, he observed *A. cantonensis* along the dispersal route of the snail. Second, he found the parasite in rats in places where *A. fulica* was introduced. Last, he also noted the close time relationship between

the introduction of the snail and the first reported case of EME in several Pacific islands. In the case of West Africa, however, the presence of *A. cantonensis* there can not be pinned on *A. fulica*. Although Kliks and Palumbo (1992) reported the presence of the parasite in *A. fulica*, this was probably a result of a misidentification of another giant African snail, probably *Achatina achatina*.

7.1.3. Dispersal of *Angiostrongylus cantonensis* from South and Southeast Asia: Drozd et al.'s alternative hypothesis

An alternative hypothesis for the dispersal of *A. cantonensis* was proposed by Drozd *et al.* (1975) in which they postulated that the parasite originated in South and Southeast Asia rather than East Africa and that the murid rodent definitive hosts and not *Achatina fulica* were responsible for the current distribution of *A. cantonensis*. First, they argued that the occurrence within close geographical regions (Indo-Malayan region and Australia) of closely related *Angiostrongylus* species and their respective *Rattus* final hosts would indicate a very long host-parasite association that probably encompassed the radiation of murid rodents in Asia. Such is probably the case for *A. cantonensis* (China, Southeast Asia) in *R. rattus* and *R. norvegicus*, *A. malaysiensis* (Malaysia) in *R. jalorensis*, and *A. mackerrasae* (Australia) in *R. fuscipes*. Second, they believed that the adaptation of rat hosts such as *Rattus rattus* and *R. norvegicus*, which originated in South and Southeast Asia, to survive high dosage infections (150 parasites per individual) would have taken a long time to evolve, which could not have happened had the parasite originated in East Africa and spread to Asia in a short period of time as postulated by Alicata (1966). They therefore concluded that *Achatina fulica* is a recent introduction in the parasite's life cycle. As further support of this hypothesis, *A. cantonensis* has so far only been found in Madagascar in *Rattus rattus*, a recent human

introduction to the island (Drodz *et al.*, 1975; Prociv *et al.*, 2000); no other endemic murid rodent is found in Madagascar (Drodz *et al.*, 1975). Prociv *et al.* (2000) believed that the role of *A. fulica* in the dispersal of *A. cantonensis* was over-emphasised as the parasite was more likely to have been introduced into new areas by rats, owing to their cosmopolitan distribution, and that local snails and slugs had just as easily assumed the role of intermediate hosts. Furthermore, Civeyrel & Simberloff (1996) pointed the possibility of other introduced snails, such as *Euglandina*, *Edentulina* and *Gonaxis*, in spreading the parasite.

7.1.4. Other nematodes in *Achatina fulica*

In addition to *Angiostrongylus cantonensis*, other nematodes may also be present in *Achatina fulica*. Indeed, land snails and slugs in general have often been used by nematodes either as intermediate hosts or as definitive hosts (Grewal *et al.*, 2003). There are 61 known nematode species that utilise molluscs as intermediate hosts, with 49 belonging to the superfamily Metastrongyloidea; 47 known nematode species use molluscs as definitive hosts, 33 of which belong to the order Rhabditida (Grewal *et al.*, 2003). Both groups are included in the Rhabditida clade by Meldal *et al.* (2006) based on phylogenetic analyses using the small subunit (SSU) ribosomal (r) RNA gene. In the case of *Achatina fulica*, previous studies have already demonstrated the presence of nematodes. For instance, the metastrongyloid lungworm *Oslerus ostratus* (= *Anafilaroides rostratus*), which parasitises cats as the final host and is closely related to the dog lungworm *Oslerus osleri*, can also infect *Achatina fulica* as an intermediate host (Grewal *et al.*, 2003). Another nematode, *Rhabditis* sp., was found in *A. fulica* populations in Thailand in which the parasite utilised the snail as a definitive host but without causing any noticeable damage (Viyada, 2005). Juvenile nematodes were also

noted in the mucus of *A. fulica* in Brazil though their identities were not reported (Fonseca & Nascimento, 2004).

7.1.5. Objectives of the study

Alicata's (1966) hypothesis states that the Giant African Land Snail, *Achatina fulica*, caused the spread of *Angiostrongylus cantonensis*, particularly in Southeast Asia and the Pacific, whereas Drozd et al.'s (1975) hypothesis points to murid rodents. The objectives of this study were to (1) survey global populations of *Achatina fulica* for the presence of *Angiostrongylus cantonensis* using the technique developed in Chapter 6 that employed the 5' end of the small subunit rRNA gene for rapid identification of the parasite, and (2) investigate the role of *A. fulica* in the dispersal of *A. cantonensis* (as proposed by Alicata, 1966) by comparing the movement of the snail through population genetic analysis with the movement of the parasite also through population genetic analysis using a suitable molecular marker. Both objectives, however, were dependent on the availability of sufficient population samples of both the parasite and the snail. While the focus was on *A. cantonensis* infection in *A. fulica*, where samples of other snails and slugs were obtained within the distribution range of the parasite (in this case two other species, *Laevicaulis alte* and *Achatina* sp.), they were also surveyed for *A. cantonensis* infection. This is the first systematic attempt to survey *A. cantonensis* from global populations of a known intermediate host using molecular identification. If Alicata's hypothesis is correct, then the nematode parasite should be present within the dispersal route of the snail intermediate host, and the dispersal patterns of the snail and the parasite based on population genetic analysis should also be similar.

7.2. Materials and methods

7.2.1. Samples and localities

Ten global populations of *Achatina fulica*, two West African populations of *Achatina* sp. and two Philippine populations of *Laevicaulis alte* were sampled for *Angiostrongylus cantonensis*. The *Achatina fulica* and *Laevicaulis alte* sample populations from UP Diliman used in Chapter 6 were also included in this survey. For the Ivory Coast *Achatina* sp. samples, the exact location of the sampling site was not identified as the snails were confiscated by customs officials at Waterloo Station in London from a man trying to smuggle the snails in October 2005. Details of the samples, their sampling localities and collectors are provided in Table 7.1.

Table 7.1: Snail samples, collectors and localities

Locality	Species	Collector
Bharatpur, Nepal	<i>Achatina fulica</i>	P. Budha
Yangon, Myanmar	<i>Achatina fulica</i>	F. Naggs
Trok Nong Area, near Phlieu Waterfall National Park, Chantaburi, Eastern Thailand near Cambodia	<i>Achatina fulica</i>	S. Panha & C. Sutcharit
Hahajima, Ogasawara, Japan	<i>Achatina fulica</i>	A. Davison
UP Diliman Campus, Quezon City, Philippines	<i>Achatina fulica</i>	I. Fontanilla
Batasan Hills, Quezon City, Philippines	<i>Achatina fulica</i>	E. Fontanilla & I. Fontanilla
Subic Bay Forest Reserve, Zambales, Philippines	<i>Achatina fulica</i>	M.R. Posa
Singapore	<i>Achatina fulica</i>	M.R. Posa
Haapiti Valley, Moorea, French Polynesia	<i>Achatina fulica</i>	T. Coote
Moaroa Valley, Tahiti, French Polynesia	<i>Achatina fulica</i>	T. Coote
Ivory Coast	<i>Achatina</i> sp.	Intercepted by customs officials at Waterloo Station in London from a man from the Ivory Coast (October 2005)
Ibadan, Nigeria	<i>Achatina</i> sp.	F. Opeyemi
UP Diliman Campus, Quezon City, Philippines	<i>Laevicaulis alte</i>	I. Fontanilla
Batasan Hills, Quezon City, Philippines	<i>Laevicaulis alte</i>	E. Fontanilla & I. Fontanilla

7.2.2. DNA extraction, PCR amplification and sequencing

The protocols in this section follow those developed in Chapter 6. The snails were cut into small pieces and digested overnight in Ash's digestive fluid, after which the worms were individually collected using yellow-tipped pipettes and stored at -20°C in TE buffer until use (see Section 6.2.2 of Chapter 6, pp. 305-307). DNA extraction, PCR amplification, gel extraction and purification were carried out for each nematode using the protocols in Chapter 2, Section 2.1.4, pp. 41-42 and Sections 2.2 - 2.4, pp. 42-59, except that only Primer Set A (SSU_F07 and SSU_R09) for the 5' end of the SSU rRNA was used to amplify an approximately 480 bp PCR product. The purified PCR products were subsequently sequenced using the SSU_R09 primer and the peaks visualised using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 termination cycle sequencing chemistry (Section 2.6 of Chapter 2, pp. 59-61). PCR amplification and sequencing of a second molecular marker for the population genetic analyses of the snail and parasite samples (second objective) were no longer carried out due to the limited and patchy sampling of the parasite and the difficulty of obtaining live snail intermediate host (see results and discussion).

7.2.3. Sequence analyses

Individual nematode sequences were processed using the STADEN package (Staden *et al.*, 2000) and the presence of *Angiostrongylus cantonensis* was determined by BLAST search in GenBank based on the 5' end of the SSU rRNA gene. Where possible, the non-angiostrongylid nematodes were also identified based on the closest sequence match to a named nematode species. If a 100% BLAST sequence identity to a named nematode species was obtained, it could mean the same taxon or that there is insufficient variation to distinguish taxa (Floyd *et al.*, 2002). In the case of

Angiostrongylus, results in Chapter 6 have already shown that the five closely related species of *Angiostrongylus*, which include *A. cantonensis*, can be distinguished from each other based on the 5' end of the SSU rRNA gene. To complement the BLAST results, a neighbor-joining phylogeny was then constructed to visualise the relationship of the nematode sequences against all known nematode taxa in Genbank as well as the *Angiostrongylus* species obtained in Chapter 6. The phylogeny was based on 376 unambiguously aligned nucleotide sites and rooted on 4 non-nematode taxa: *Chordodes morgani* (AF036639, Blaxter *et al.*, 1998) and *Gordius aquaticus* (X87985, Winnepenninckx *et al.*, 1995) of Phylum Nematomorpha, *Priapulus caudatus* (Z38009, Aleshin *et al.*, 1998) of Phylum Priapulida, and *Brachionus plicatis* (U49911, Aguinaldo *et al.*, 1997) of Phylum Rotifera (for alignment, see Appendix 6.2, pp. 537-603). The sequences were checked for base composition bias using the Chi square test in PAUP* (version 4.0b10) (Swofford, 2002), after which the optimal model for DNA sequence evolution was determined by comparing, through the Likelihood Ratio Test (LRT), the likelihood scores of the following: JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P (Kimura, 1980), HKY85 (Hasegawa *et al.*, 1985), TN93 (Tamura & Nei, 1993) and the GTR (Rodriguez *et al.*, 1990) models as well as their variants that incorporated gamma distributed rates (Γ) (Yang, 1993). Bootstrap resampling (Felsenstein, 1985) with 1000 replicates was then carried out (see Section 2.9.9 of Chapter 2, pp. 85-86). Once the clades to which the nematode sequences belong were identified, a second NJ tree based on the same gene fragment (376 sites) with optimised parameters was constructed but this time excluding the taxa that do not belong to these clades.

7.3. Results

A total of 191 *Achatina fulica* from 10 global populations, 25 *Achatina* sp. from 2 West African populations and 39 *Laevicaulis alte* from 2 Philippine populations were surveyed for *A. cantonensis* infections. Of the 10 global populations of *Achatina fulica*, 6 yielded nematodes; these were Thailand (20 snails sampled), Ogasawara (12 snails), UP Diliman-Philippines (15 snails), Batasan Hills-Philippines (50 snails), Singapore (33 snails) and Tahiti (5 snails). Population samples from Ogasawara and Tahiti had a 100% infection rate. Population samples from Myanmar (20 snails), Nepal (22 snails), Zambales-Philippines (4 snails) and Moorea (10 snails) had a 0% infection rate. The two *Achatina* sp. populations from West Africa, namely the Ivory Coast (22 snails) and Nigeria (3 snails), as well as the two Philippine populations of *Laevicaulis alte*, UP Diliman (19 slugs) and Batasan Hills (20 slugs), also manifested nematode infection, with the population sample from Nigeria exhibiting a 100% infection rate. See also Table 7.2.

Table 7.2: Population sizes and percentage of nematode infection

Locality	Species	Sample Size	% Infected
Bharatpur, Nepal	<i>Achatina fulica</i>	22	0
Yangon, Myanmar	<i>Achatina fulica</i>	20	0
Chantaburi, Eastern Thailand	<i>Achatina fulica</i>	20	5.0%
Hahajima, Ogasawara, Japan	<i>Achatina fulica</i>	12	100.0%
UP Diliman Campus, Philippines	<i>Achatina fulica</i>	15	6.7%
Batasan Hills, Philippines	<i>Achatina fulica</i>	50	14.0%
Subic Bay Forest Reserve, Philippines	<i>Achatina fulica</i>	4	0
Singapore	<i>Achatina fulica</i>	33	15.2%
Haapiti Valley, Moorea	<i>Achatina fulica</i>	10	0
Moaroa Valley, Tahiti	<i>Achatina fulica</i>	5	100.0%
Ivory Coast	<i>Achatina</i> sp.	22	18.2%
Ibadan, Nigeria	<i>Achatina</i> sp.	3	100.0%
UP Diliman Campus, Philippines	<i>Laevicaulis alte</i>	19	21.1%
Batasan Hills, Philippines	<i>Laevicaulis alte</i>	20	30.0%

A total of 25 distinct nematode sequences were identified from the snail and slug populations. For the *Achatina fulica* populations, a single unique nematode sequence was found in the Thailand population; two nematode sequences were each found in the UP Diliman-Philippines, Ogasawara and Tahiti populations; three were found in the Batasan-Philippines population, and four in the Singapore population. For the West African *Achatina* sp. populations, three unique nematode sequences were found in Nigeria and seven in the Ivory Coast. For the Philippine *Laevicaulis alte* populations, the same two nematode sequences found in *A. fulica* were also found in the UP Diliman *L. alte* population, whereas two unique sequences were found in the Batasan Hills population in addition to a nematode sequence that it shares with the *A. fulica* population from that site. One of the Tahiti sequences (Tahiti Sequence 1) is identical to one of the Philippines sequences (Philippines Sequence 1), which was present in the UP Diliman population of *Achatina fulica* and in both UP Diliman and Batasan Hills populations of *Laevicaulis alte*. A summary of the frequency and distribution of these nematode sequences is given in Table 7.3.

Table 7.3: Distribution and frequencies of the nematode sequences across global populations of *Achatina fulica*, West African populations of *Achatina* sp. and Philippine populations of *Laevicaulis alte*. Note that Philippines Sequence 1 and Tahiti Sequence 1 are 100% identical.

Locality	Host	Nematode Sequences	Number (Frequency relative to the total nematodes found in the whole snail population)
Nepal	<i>Achatina fulica</i>	None	None
Myanmar	<i>Achatina fulica</i>	None	None
Thailand	<i>Achatina fulica</i>	Thailand Sequence 1	1 (100.0%)
Ogasawara, Japan	<i>Achatina fulica</i>	Ogasawara Sequence 1	2 (2.3%)
		Ogasawara Sequence 2	86 (97.7%)
UP Diliman, Philippines	<i>Achatina fulica</i>	Philippines Sequence 1 (=Tahiti Sequence 1)	1 (16.7%)
		Philippines Sequence 2	5 (83.3%)
Batasan, Philippines	<i>Achatina fulica</i>	Philippines Sequence 5	6 (33.3%)
		Philippines Sequence 6	1 (5.6%)
		Philippines Sequence 7	11 (61.1%)
Subic Forest Reserve, Philippines	<i>Achatina fulica</i>	None	None
Singapore	<i>Achatina fulica</i>	Singapore Sequence 1	6 (9.7%)
		Singapore Sequence 2	1 (1.6%)
		Singapore Sequence 3	1 (1.6%)
		Singapore Sequence 4	54 (87.1%)
Tahiti	<i>Achatina fulica</i>	Tahiti Sequence 1 (=Philippines Sequence 1)	91 (98.9%)
		Tahiti Sequence 2	1 (1.1%)
Moorea	<i>Achatina fulica</i>	None	None
Ivory Coast	<i>Achatina</i> sp.	Ivory Coast Sequence 1	2 (8.7%)
		Ivory Coast Sequence 2	11 (47.8%)
		Ivory Coast Sequence 3	1 (4.3%)
		Ivory Coast Sequence 4	1 (4.3%)
		Ivory Coast Sequence 5	5 (21.7%)
		Ivory Coast Sequence 6	1 (4.3%)
		Ivory Coast Sequence 7	2 (8.7%)
Nigeria	<i>Achatina</i> sp.	Nigeria Sequence 1	1 (1.8%)
		Nigeria Sequence 2	1 (1.8%)
		Nigeria Sequence 3	54 (96.4%)
UP Diliman, Philippines	<i>Laevicaulis alte</i>	Philippines Sequence 1 (=Tahiti Sequence 1)	20 (95.2%)
		Philippines Sequence 2	1 (4.8%)
Batasan, Philippines	<i>Laevicaulis alte</i>	Philippines Sequence 1 (=Tahiti Sequence 1)	8 (80%)
		Philippines Sequence 3	1 (10%)
		Philippines Sequence 4	1 (10%)

Nine snails from six localities (UP Diliman-Philippines, Batasan Hills-Philippines, Ogasawara, Singapore, Tahiti and Ivory Coast) were infected by more than one type of worm based on the SSU rRNA sequences. Table 7.4 summarises the multiple infections of these snails.

Table 7.4: Snails infected with more than one type of worm based on the sequence of the 5' end of the SSU rRNA gene.

Locality	Host #	Nematode Sequence	Number (Frequency*)
UP Diliman, Philippines	<i>Achatina fulica</i> D51	Philippines Sequence 1	1 (16.7%)
		Philippines Sequence 2	5 (83.3%)
Batasan Hills, Philippines	<i>Achatina fulica</i> B42	Philippines Sequence 5	1 (33.3%)
		Philippines Sequence 7	2 (66.67%)
	<i>Achatina fulica</i> B43	Philippines Sequence 5	1 (50%)
		Philippines Sequence 7	1 (50%)
Ogasawara, Japan	<i>Achatina fulica</i> OJ2	Ogasawara Sequence 1	1 (8.3%)
		Ogasawara Sequence 2	11 (91.7%)
	<i>Achatina fulica</i> OJ3	Ogasawara Sequence 1	1 (14.3%)
		Ogasawara Sequence 2	6 (85.7%)
Singapore	<i>Achatina fulica</i> Si63	Singapore Sequence 1	4 (80.0%)
		Singapore Sequence 2	1 (8.3%)
Tahiti	<i>Achatina fulica</i> T1	Tahiti Sequence 1	11 (91.7%)
		Tahiti Sequence 2	1 (1.1%)
Ivory Coast	<i>Achatina</i> sp. IVC19	Ivory Coast Sequence 2	11 (84.6%)
		Ivory Coast Sequence 3	1 (7.7%)
		Ivory Coast Sequence 4	1 (7.7%)
	<i>Achatina</i> sp. IVC22	Ivory Coast Sequence 6	2 (66.7%)
		Ivory Coast Sequence 7	1 (33.3%)

*Frequency is relative to the total nematodes found per snail.

The 25 nematode sequences were then subjected to a BLAST search in GenBank (GB Release No. 158, 15 February 2007) to identify an exact match or closest identity to named nematode species. Two of these sequences, namely Tahiti Sequence 1/ Philippines Sequence 1 and Tahiti Sequence 2, are angiostrongylids based on the BLAST results. However, only Tahiti Sequence 1/Philippines Sequence 1 has a 100% match with *Angiostrongylus cantonensis* (AY295804, Carreno & Nadler, 2003). Among the non-angiostrongylids, Philippines Sequence 5 is the only other nematode sequence to have a 100% BLAST identity to a named species in GenBank. This

sequence is identical to *Ancylostoma caninum* (AJ920347, Chilton *et al.*, 2006). As the other sequences did not match any named taxon in GenBank based on BLAST results, only the closest sequence identity to a named nematode species could be assigned; their exact species identification based on the BLAST results could therefore not be known as of GB Release No. 158. For instance, the second highest sequence similarity attained was 99%; this was the case for Philippines Sequence 2 (for *Oslerus osleri*), Singapore Sequence 4 (for *Oslerus osleri*), Tahiti Sequence 2 (for *Angiostomngylus cantonensis*), Ivory Coast Sequence 1 (for *Metastrongylus elongatus* and *M. salmi*) and Nigeria Sequence 2 (for *Ancylostoma caninum*). On the other hand, Singapore Sequence 1 and Philippines Sequence 3 have the lowest sequence identity (87% BLAST identity for both with *Pellioiditis marina*, a marine nematode, which, being terrestrial nematodes, Singapore Sequence 1 and Philippines Sequence 3 clearly are not). A summary of the BLAST results is shown in Table 7.5.

Table 7.5: GenBank BLAST results for the nematode sequences

Sequence	BLAST Result
Thailand Sequence 1	96% identity with <i>Heterakis</i> sp. 14690 (AF083003) [Ascaridida]
Ogasawara Sequence 1	93% identity with <i>Panagrolaimus subelongatus</i> (AY284681) [Panagrolaimidae]
Ogasawara Sequence 2	93% identity with <i>Phasmarhabditis hermaphrodita</i> (DQ639981) [Rhabditida]
Philippines Sequence 1 (=Tahiti Sequence 1)	100% identity with <i>Angiostrongylus cantonensis</i> (AY295804) [Strongyloidea]
Philippines Sequence 2	99% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea] (but different from Singapore Sequence 4)
Philippines Sequence 3	87% identity with <i>Pellioiditis marina</i> (AF038021) [Rhabditidae] (but different from Singapore Sequence 1)
Philippines Sequence 4	97% identity with <i>Panagrolaimus</i> c.f. <i>rigidus</i> (DQ285636) [Panagrolaimidae]
Philippines Sequence 5	100% identity with <i>Ancylostoma caninum</i> (AJ920347) [Strongyloidea]
Philippines Sequence 6	93% identity with <i>Pellioiditis marina</i> (AF038021) [Rhabditidae]
Philippines Sequence 7	97% identity with <i>Caenorhabditis vulgaris</i> (U13931) [Rhabditidae]
Singapore Sequence 1	87% identity with <i>Pellioiditis marina</i> (AF038021) [Rhabditidae]
Singapore Sequence 2	95% identity with <i>Heterakis</i> sp. 14690 (AF083003) [Ascaridida]
Singapore Sequence 3	98% identity with <i>Caenorhabditis briggsae</i> (U13929) [Rhabditidae]
Singapore Sequence 4	99% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea] (but different from Singapore Sequence 2)
Tahiti Sequence 1 (=Philippines Sequence 1)	100% identity with <i>Angiostrongylus cantonensis</i> (AY295804) [Strongyloidea]
Tahiti Sequence 2	97% identity with <i>Angiostrongylus cantonensis</i> (AY295804) [Strongyloidea]
Ivory Coast Sequence 1	99% identity with <i>Metastrongylus elongatus</i> (AJ920363) [Strongyloidea] 99% identity with <i>Metastrongylus salmi</i> (AY295801) [Strongyloidea]
Ivory Coast Sequence 2	98% identity with <i>Metastrongylus elongatus</i> (AJ920363) [Strongyloidea] 98% identity with <i>Metastrongylus salmi</i> (AY295801) [Strongyloidea]
Ivory Coast Sequence 3	98% identity with <i>Protostrongylus rufescens</i> (AJ920364) [Strongyloidea]
Ivory Coast Sequence 4	98% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea] (but different from Ivory Coast sequence 6)
Ivory Coast Sequence 5	97% identity with <i>Heterakis gallinarum</i> (DQ503462) [Ascaridida]
Ivory Coast Sequence 6	98% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea] (but different from Ivory Coast sequence 4)
Ivory Coast Sequence 7	97% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea]
Nigeria Sequence 1	97% identity with <i>Protostrongylus rufescens</i> (AJ920364) [Strongyloidea]
Nigeria Sequence 2	99% identity with <i>Ancylostoma caninum</i> (AJ920347) [Strongyloidea]
Nigeria Sequence 3	97% identity with <i>Metastrongylus elongatus</i> (AJ920363) [Strongyloidea] 97% identity with <i>Metastrongylus salmi</i> (AY295801) [Strongyloidea]

To complement the BLAST results, a neighbor-joining tree of the Nematoda based on 376 unambiguously aligned nucleotide sites of the 5' end of the SSU rRNA gene (Fig. 7.2A) was then generated to visualise the relationships of the 25 nematode sequences with all 361 nematode taxa having at least the first 480 nucleotides of the 5' end of the SSU rRNA in GenBank (GB Release No. 158, see Appendix 6.2) as well as the new sequences from the four *Angiostrongylus* species in Chapter 6. The GTR+ Γ model was used to construct the tree after it was determined to be the best model

(Appendix 7.1A, p. 617). According to this phylogeny, all 25 nematode sequences fall within the Rhabditida clade (Meldal *et al.*, 2006). A smaller NJ phylogeny of the same gene fragment (376 nucleotide sites) was constructed (Fig. 7.2B) for the Rhabditida clade using all 241 rhabditid SSU sequences available in GenBank (GB Release No. 158) as well as the other *Angiostrongylus* sequences and rooted on the plectid nematode *Plectus acuminatus* (AF037628, Blaxter *et al.*, 1998). The GTR+ Γ model was determined to be the best model and was subsequently used for this analysis (Appendix 7.1B, p. 617). This NJ phylogeny shows that 22 out of the 25 nematode sequences, including the two angiostrongylid sequences (Philippines Sequence 1/ Tahiti Sequence 1 and Tahiti Sequence 2), are distributed across three major clades in the tree: the Strongyloidea, the Rhabditidae and the Panagrolaimidae. The three remaining nematode sequences (Ivory Sequence 5, Singapore Sequence 2 and Thailand Sequence 1) group with the two *Heterakis* species found outside the three aforementioned clades. The groupings of the nematode sequences to the different clades correspond to the taxonomic grouping of the closest match(es) of each of the nematode sequences according to BLAST (see also Table 7.5).

The Strongyloidea clade includes the 2 angiostrongylid sequences: Tahiti Sequence 1/ Philippines Sequence 1 and Tahiti Sequence 2. Tahiti Sequence 1/ Philippines Sequence 1 has 100% BLAST identity to the sequence of *Angiostrongylus cantonensis* on Genbank (AY295804, Carreno & Nadler, 2003) and clusters with *A. cantonensis* in the tree, while the sequences from the other *Angiostrongylus* species sequenced in Chapter 6 fall outside this cluster. Tahiti Sequence 2 also clusters with *A. cantonensis* in the tree but has a BLAST identity of 97% with *A. cantonensis*.

There are 12 non-angiostrongylid sequences that are also included in the Strongyloidea; these are the 3 Nigeria sequences, 6 out of 7 Ivory Coast sequences, 2

Philippines sequences and Singapore Sequence 4. Nigeria Sequence 1 clusters with the deer muscieworm *Parelaphostrongylus odocoilei*, the hair lungworm *Muellerius capillaris*, and the sheep lungworm *Dictyocaulus filaria*, though it has a 97% BLAST identity with the goat and sheep lungworm *Protostrongylus rufescens* (AJ920364, Chilton *et al.*, 2006) (see Table 7.5). Nigeria Sequence 2 and Philippines Sequence 5 cluster with the dog hookworms *Ancylostoma caninum* and *Necator americanus*; both sequences are most similar to *Ancylostoma caninum* (AJ920347, Chilton *et al.*, 2006) with 99% and 100% BLAST identities, respectively. Nigeria Sequence 3 clusters with Ivory Coast Sequence 7, though they have different BLAST identities; Nigeria Sequence 3 has a 97% BLAST identity with both pig lungworms *Metastrongylus elongatus* (AJ920363, Chilton *et al.*, 2006) and *M. salmi* (AY295609, Carreno & Nadler, 2003) while Ivory Coast Sequence 7 has a 97% BLAST identity with the filaroid dog lungworm *Oslerus osleri* (AY295812, Carreno & Nadler, 2003). Ivory Coast Sequence 1 groups with *Metastrongylus elongatus* (AJ920363) and *M. salmi* (AY295609) with 99% BLAST identity for both taxa. Ivory Coast Sequence 2 clusters with the cat lungworm *Aelurostrongylus abstrusus* (Chilton *et al.*, 2006), though it has a 98% BLAST identity with both *Metastrongylus elongatus* (AJ920363) and *M. salmi* (AY295609) (see Table 7.5). Ivory Coast Sequence 3 clusters with *Protostrongylus rufescens* (AJ920364) with 98% BLAST identity. Ivory Coast Sequences 4 and 6 cluster together and both have a 98% BLAST identity with *Oslerus osleri* (AY295812) though the two sequence types are not identical with each other (see Table 7.5). Both Philippines Sequence 2 and Singapore Sequence 4 cluster with *Oslerus osleri* (AY295812) with 99% BLAST identity though they are not identical with each other (see Table 7.5).

The Rhabditidae clade includes six nematode sequences: Singapore Sequences 1 and 3, Philippines Sequences 3, 6 and 7 and Ogasawara Sequence 2. Singapore Sequence 1 and Philippines Sequence 3 cluster together, and in turn group with the free-living marine nematode *Pellioditis mediterranea*, though both have an 87% BLAST identity with *Pellioditis marina* (AF038021, Fitch, 2000). Philippines Sequence 6 forms a sister group taxon with a group of rhabditids that include the *Pellioditis* species and has a 93% BLAST identity with *Pellioditis marina* (AF038021). Singapore Sequence 3 clusters with *Caenorhabditis* spp., free-living soil nematodes, along with Philippines Sequence 7; Singapore Sequence 3 is 98% identical with *Caenorhabditis briggsae* (U13929, Fitch *et al.*, 1995) while Philippines Sequence 7 is 97% identical with *Caenorhabditis vulgaris* (U13931) based on BLAST results. Ogasawara Sequence 2 groups with the two *Phasmarhabditis* taxa and has a 93% BLAST identity with *Phasmarhabditis hermaphrodita*.

Two nematode sequences, Ogasawara Sequence 1 and Philippines Sequence 4, are included in the Panagrolaimidae clade; the former has a 93% BLAST identity with the soil nematode *Panagrolaimus subelongatus* (AY284681, Holterman *et al.*, 2006) while the latter has a 97% identity with *Panagrolaimus* c.f. *rigidus* (DQ285636, Houthoofd *et al.*, 2006).

The three remaining nematode sequences, Singapore Sequence 2, Thailand Sequence 1 and Ivory Coast Sequence 5, all cluster with the bird parasites in the genus *Heterakis*; Singapore Sequence 2 and Thailand Sequence 1 are 95% and 96% identical, respectively, with *Heterakis* sp. isolate 14690 (AF083003, Fitch, 2000) based on BLAST results whereas Ivory Coast Sequence 5 has a 97% BLAST identity with *Heterakis galinarum* (DQ503462, Smythe *et al.*, 2006).

7.4. Discussion

7.4.1. *Angiostrongylus cantonensis* infection in snail and slug populations

The results of this study demonstrate the presence of *Angiostrongylus cantonensis* from *Achatina fulica* populations in the Philippines and Tahiti among the areas within the route of dispersal of *Achatina fulica*; in the case of the Philippines, the worm was also detected in the black slug *Laevicaulis alte*. This was demonstrated by the 100% BLAST identity of Philippines Sequence 1/Tahiti 1 with *A. cantonensis* based on the 5' end of the SSU rRNA gene and its distinction from all other *Angiostrongylus* species sequenced to date. Historically, cases of rat and mollusc infections in the Philippines (Garcia, 1979) and rat, gastropod and human infections in Tahiti (Alicata, 1966; Wallace & Rosen, 1969; Bronstein *et al.*, 1977) have been recorded. Although Wallace and Rosen (1969) surveyed Tahiti for the presence of *Angiostrongylus cantonensis* in gastropod intermediate hosts between 1965 and 1966 and found 3rd stage juveniles in areas where the rats were known to be infected with the parasite, they did not confirm the identity of the juveniles by feeding them to laboratory rats. Their survey did not include *A. fulica* among the gastropods they examined as they did not encounter the snail, noting that it was only reported in Tahiti in 1967 (Raut & Barker, 2002). This study therefore provides the first ever reported case of *A. cantonensis* infection in *A. fulica* in Tahiti. The first recorded case of human infection in Tahiti by the nematode parasite occurred in 1957 and rat infection in 1961 (Alicata, 1966), several years before *A. fulica* was supposedly introduced. In the case of Tahiti, at least, Alicata (1966) hypothesised that *A. cantonensis* was not brought there by *A. fulica*. Nevertheless, the results show that the Tahitian *Achatina fulica* is used as an intermediate host of *A. cantonensis*. Alicata (1966) also found *A. fulica* in Hawaii to be

infected with the parasite, and he concluded that if the snails and the worms are found in the same area, then it is highly likely that the majority of the snails in the population will be infected.

The other *A. fulica* populations surveyed in the current study that are within the dispersal route of *A. fulica* proved negative for *A. cantonensis*. Among these areas, Thailand, Japan and Singapore had prior reported cases of *A. cantonensis*. In the case of Thailand, *A. cantonensis* was “found” in the edible land snail, *Hemiplecta distincta*, from 16 provinces where a total of 467 snails were surveyed (Panha, 1988). However, it was not definitively identified by feeding the nematodes to laboratory rats; instead, identification relied more on the morphological features of the juveniles. The sampling site in Thailand for this study (with only 20 snails surveyed but with no *A. cantonensis* infection) is also near the border with Cambodia where the parasite was detected in both humans and rats (Brumpt *et al.*, 1968). In Japan, *A. cantonensis* was reported from 27 cases of human infections, the majority of which are from the Okinawa Prefecture in the Pacific south of the Japanese main islands (Nakazawa *et al.*, 1992). In Ogasawara (with 12 snails surveyed in this study), there are no reported cases of *A. cantonensis* to date; Ogasawara is another group of islands in the Pacific southeast of the main Japanese islands. In Singapore (33 snails surveyed), there is one previous case of eosinophilic meningoencephalitis in a woman who was suspected of acquiring the parasite from eating contaminated raw vegetables, though it was more likely that these vegetables were imported from a nearby country where these worms are found (Lim *et al.*, 2004). For the remaining areas, Nepal, Myanmar and Moorea, there are no known reported cases to date that demonstrate the presence of *Angiostrongylus cantonensis*, though no systematic attempts to survey the snails and rats for worms in these areas have been done so far.

For West Africa, neither the *Achatina* sp. sample population from Ibadan, Nigeria (3 snails surveyed) nor that of the Ivory Coast (22 snails surveyed) yielded *A. cantonensis*. However, both countries had prior reported cases of *A. cantonensis* infections. A previous survey conducted on 22 rats in Ibadan in 1989 also failed to recover the parasite (Kliks & Palumbo, 1992), although it was detected in rats in Port Harcourt 450 km to the southeast (Udonsi, 1989; Kliks & Palumbo, 1992). In the Ivory Coast, the only recorded incidence was based on a single patient in 1979 that manifested the disease (Nozais *et al.*, 1980).

Angiostrongylus cantonensis was found in only two of the global populations of *A. fulica* sampled in this study. These results are therefore insufficient to permit any consideration of the role of *A. fulica* in the dispersal of *A. cantonensis*. The low success in finding the parasite in the snail intermediate host could have been caused by several factors. For one, the absence of *A. cantonensis* in *A. fulica* populations within the snail's route of dispersal, with the exception of the Philippines and Tahiti, may be due to the patchy distribution of the parasite. This was shown in a previous study by Bisseru (1971) in which he surveyed the presence of *A. cantonensis* in *A. fulica* populations from 27 sites in West Malaysia and found two sites with no parasite in any of the snails. One of these sites, Padang Besar, Perlis, with 100 snails sampled, is less than 65 km from Alor Star, Kedah, with 70 snails sampled and with an infection rate of 28.5%. Further to the patchy distribution of the parasite, the snail intermediate host itself could also have a patchy distribution and be uncommon in some of the sampling sites, thus reducing the probability of finding the parasites. For example, the Subic Bay Forest Reserve in the Philippines only yielded four individuals despite rigorous searching. In addition to the issue of patchy distribution, this study was also affected by sampling limitation; in particular, sampling was severely hampered by the availability

of live snail samples that could be brought out of the sampling sites. For instance, governmental restrictions in India and Sri Lanka prevent the export of live snails from these countries. Based on these factors, a comprehensive global survey of *A. fulica* populations was not possible.

Angiostrongylus cantonensis is not stringent as to its choice of intermediate and definitive hosts (Prociv *et al.*, 2000); therefore, the current distribution range of the parasite could be attributed to more than just one host. Indeed, the parasite is known to have been established in many localities where *A. fulica* is absent or has arrived after *A. cantonensis*. Already mentioned is Tahiti, to which the parasite could have been brought either by contaminated rats or by molluscs other than *A. fulica* that were probably imported by labourers from Indochina and Indonesia in the 1950s (Alicata, 1966). Other examples include Brisbane, Australia, where the parasite was found in the exotic rats *R. norvegicus* and *R. rattus* (Yong *et al.*, 1981), and New Orleans, USA where the worm was recovered from *R. rattus* (Campbell & Little, 1988). In both cases, local snails and slugs were also found to harbor *A. cantonensis* either in the wild (Yong *et al.*, 1981) or experimentally (Campbell & Little, 1988). Furthermore, other intermediate hosts with similar routes of dispersal to that of *A. fulica* could just as easily have aided the dispersal of *A. cantonensis*. One candidate is the black slug, *Laevicaulis alte*, which originated from Africa (Solem, 1964) and has spread across the globe in association with humans (Hoffman, 1925); the slug is also known to harbor *A. cantonensis* as demonstrated by Salazar and Cabrera (1969) and by the results from this study. It is therefore highly likely that the dispersal of the nematode is attributed to a complex combination of rats and mollusc intermediate hosts, and a systematic survey of all possible taxa that could harbour *A. cantonensis* as well as the population genetic analysis of global populations of the parasite and its hosts, juxtaposed with the

information regarding the first reported cases of infection in new areas, is perhaps the only way to trace the dispersal of *A. cantonensis*.

7.4.2. The presence of other worms in snail and slug populations

Nematodes other than *Angiostrongylus cantonensis* can likewise infect terrestrial snails and slugs such as *Achatina fulica* and *Laevicaulis alte*. The BLAST results and the phylogenetic analysis in this study clearly revealed 23 unique nematode species from the snails and slugs that fall within the Rhabditida clade and which have not yet been characterised molecularly using the 18S rRNA gene as they did not have a match with any named nematode taxa in GenBank. Some of these nematode species were also found to infect the same snail, as in the case of two *Achatina* sp. from the Ivory Coast, three *Achatina fulica* from the Philippines and one *A. fulica* each from Singapore, Ogasawara and Tahiti. Infection in gastropods by more than one species of nematodes is not uncommon, although localization of these parasites to specific organs does occur (Morand *et al.*, 2004). In another study, Viyada (2005) surveyed 200 adult *A. fulica* from two provinces in Thailand in which he found *Rhabditis* sp. in the alimentary tract. He suggested that this species could have been an accidental parasite of the snail as no noticeable damage to the host was observed.

Some species of nematodes pass through a dauer stage, a non-feeding alternative 3rd juvenile stage that is resistant to stress and can survive for several months without feeding. In free living soil nematodes such as *Caenorhabditis elegans*, the dauer stage is utilised for dispersal and forms associations with invertebrate hosts or even feeds on carcasses of animals. For instance, *C. elegans* was found in snails (*Helix*, *Oxychilus* and *Pomatias*), isopods (*Oniscus asellus*) and a glomeris myriapod, indicating a wide host specificity for a nematode originally thought to be free-living (Barriere & Felix,

2005). Some of the nematode infections in this study could have been attributed to associations with the dauer stage; for example, Singapore Sequence 3 has a 98% identity with the free-living soil nematode *Caenorhabditis briggsae* while Philippines Sequence 7 has a 97% identity with *C. vulgaris*.

Grewal *et al.* (2003) listed 108 nematodes species that are known to be parasitic on gastropods and divided them into those that use gastropods as intermediate hosts and those that use gastropods as definitive hosts. For the first group of gastropod-parasitizing nematodes, the majority belong to the order Strongylida while the rest are from the Ascaridia, Spirurida and Rhabditida. Based on the results of this study, 13 nematode sequences fall within the Strongyloidea: Tahiti Sequence 2, Nigeria Sequences 1, 2 and 3, Ivory Coast Sequences 1, 2, 3, 4, 6 and 7, Philippines Sequences 2 and 5 and Singapore Sequence 4. A further three sequences cluster with the *Heterakis* species from the Ascaridia: Singapore Sequence 2, Thailand Sequence 1 and Ivory Coast Sequence 5. Based on their phylogenetic positions, these nematodes were probably juveniles and most likely used *Achatina fulica* and *Achatina* sp. as intermediate hosts. For the second group of gastropod-parasitizing nematodes, the majority belong to the Rhabditida (Grewal *et al.*, 2003). Six other sequences in this study fall within the Rhabditidae: Singapore Sequences 1 and 3, Philippines Sequences 3, 6 and 7, and Ogasawara Sequence 2. These nematodes were probably adults that used the snails and slugs as definitive hosts. On the other hand, Ogasawara Sequence 1 and Philippines Sequence 4 did not fall in either group described by Grewal *et al.* (2003) as they cluster with the Panagrolaimidae, a group of bacteria-feeding soil nematodes. It is highly likely that the *A. fulica* samples from Ogasawara and the Philippines *L. alte* that was infected with the nematode bearing Philippines Sequence 4 became accidental hosts of these probable free-living soil nematodes.

There were instances in the study when a nematode sequence clustered with a particular taxon or group of taxa in the phylogenetic analysis and yet was most similar to another species based on the BLAST result. This was true for Nigeria Sequences 1 and 3 and Ivory Coast Sequence 2. It should be noted that the BLAST results were based on approximately 480 nucleotide sites whereas the phylogenetic analysis utilised only 376 sites as the rest could not be unambiguously aligned and that the clades generated were not well supported at the tips. For this reason, BLAST results take precedence as they identify the closest sequence match to named nematode taxa found in GenBank using all available nucleotide sites obtained while the phylogenetic analyses only determine which clades the unknown sequence types belong to based on a subset of the sequence information obtained.

7.4.3. Limitations and the next step

This study was heavily dependent on the availability of live samples of *Achatina fulica* and other gastropod intermediate hosts. Alicata's hypothesis was not sufficiently tested precisely because of the patchy and limited sampling of live snails and parasites, and the second objective (comparison of the population genetics of the snail and the parasite to check for similarity in their dispersal) was therefore not met as only the population genetics of global samples of *Achatina fulica* based on the 16S rRNA gene was available (see Chapter 5). Many areas were not covered because of governmental restrictions on the export of live snails, the difficulty of finding these snails by collectors and keeping them alive whilst in transit, the unwillingness of some couriers to ship live specimens, and the UK government's restrictions on the importation of such live snails from different countries. One practical solution will be to collect the samples and extract the worms in the field. Another is to develop a technique that will detect the

presence of *A. cantonensis* through PCR in ethanol-preserved specimens, though this technique will not be able to determine the parasite load (number of parasites present) per individual snail.

Owing to the fact that *Angiostrongylus cantonensis* has a patchy distribution as demonstrated by the survey from three sampling sites in the Philippines, several sampling sites per locality should also be covered. In addition, other gastropod intermediate hosts that could have had a role in the dispersal of *A. cantonensis* should be surveyed alongside *A. fulica*.

7.5. Summary

Within the route of dispersal of *Achatina fulica*, *Angiostrongylus cantonensis* was only found in *A. fulica* populations in the Philippines and Tahiti as well as the *Laevicaulis alte* populations in the Philippines. The results for Tahiti provide the first recorded case of *A. cantonensis* infection for *A. fulica* in that island. Due to the patchy distribution of *A. cantonensis* and the limited sampling of the snail intermediate host, the results from this study are insufficient to test Alicata's hypothesis that the Giant African Land Snail is primarily responsible for the spread of the parasite. Further sampling is therefore required. *Angiostrongylus cantonensis* has a wide range of suitable intermediate hosts, and gastropods such as the black slug *Laevicaulis alte*, which this study clearly demonstrated to harbor the parasite, should also be considered in future studies to determine their role in the dispersal of the parasite. Other worms found in *Achatina fulica*, *Achatina* sp. and *Laevicaulis alte* include 12 strongyloid species and 3 ascarid species that probably used the snails as intermediate hosts and 5 rhabditid species that probably used the snails as definitive hosts. One panagrolaimid

nematode species each from Ogasawara and the Philippines most likely used *A. fulica* and *L. alte*, respectively, as accidental hosts.

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CHAPTER 8 – Summary

The Giant African Snail *Achatina* (*Lissachatina*) *fulica* is a pest species that originated in East Africa but currently has a pantropical distribution brought about by human activities. It is also a known intermediate host of the rat lungworm, *Angiostrongylus cantonensis*, which can infect humans and cause eosinophilic meningoencephalitis.

Achatina fulica belongs to the Stylommatophora, a group of land snails and slugs that has two pairs of caudal tentacles, a contractile pneumostome and no operculum. Recent molecular phylogenetic study based on the rRNA gene cluster has shown a clear dichotomy of the Stylommatophora into the ‘non-achatinoid’ clade and the ‘achatinoid’ clade, which includes *A. fulica*. Within the ‘achatinoid clade’ is the Achatinoidea, which includes the Achatinidae, Coeliacidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophorellidae. The phylogeny of the Achatinoidea was investigated further using 24 taxa across 6 families based on 3 nuclear genes (rRNA cluster, actin and histone 3) and 2 mitochondrial genes (CO1 and 16S rRNA). The 3rd codon position of the CO1 gene and the 16S rRNA were later excluded as they were found to be saturated. All strongly-supported groups in the rRNA phylogeny were recovered in the combined gene phylogenies with longer internal branches and receiving similar or slightly better statistical support. The study demonstrated the monophyly of the Achatinidae but not the Ferussaciidae, Coeliacidae and the Subulinidae. These results suggest the need to re-evaluate of the Ferussaciidae, Coeliacidae and Subulinidae as valid groupings.

The first systematic attempt to reconstruct the phylogeny of the Achatinidae was also conducted using 26 species from 7 genera based on the same genes used to evaluate the relationships of the Achatinoidea. The nuclear actin gene and the 3rd codon position of the mitochondrial CO1 gene were subsequently excluded due to the presence of paralogous sequences in the former that affected the phylogeny and the severe saturation of the latter. Results showed four distinct groups that to a large extent follow the geographical distributions of the taxa used in the survey. These were the West African group that fell at the base of the tree, a mixed group consisting of members with distributions ranging from the western, central, eastern and southern regions, and two groups that formed sister clades and which probably arose from the mixed group: the Southern African group and the East African group, which included *A. fulica*. The East African group incorporated all members of the subgenus *Lissachatina* of *Achatina*, which also exhibit distinct features of the reproductive organ, as well as the subgenus *Euaethiopina* of *Achatina*. Evidence from both molecular and morphological data would suggest that *Lissachatina* should be elevated to genus status.

The extent of genetic variation among global populations of *A. fulica* was also investigated using a fragment of the 16S rRNA gene to determine how many genetic types emerged from East Africa. Of the 15 distinct haplotypes observed, 13 were found in East Africa and nearby Indian Ocean islands while the rest of the global populations shared only haplotype C that came out of East Africa. Two non-African haplotypes (E from the Philippines and F from New Caledonia and Barbados) were probably recent mutations derived from C. This would indicate that the global populations of *A. fulica* descended

from a small group of individuals that shared the same haplotype and that these populations could be experiencing genetic bottlenecks as a consequence of lack of variation.

The role of *A. fulica* in spreading the rat lungworm *Angiostrongylus cantonensis* was evaluated. It is believed that the rat lungworm, like *A. fulica*, also originated in East Africa, and the nearly parallel distribution of the snail intermediate host and the parasite is noted. A molecular marker was first developed using a small segment of the SSU rRNA gene to rapidly identify the worm from snail tissues. This marker was also found to discriminate between closely related species of *Angiostrongylus*. Using this molecular marker, global populations of *A. fulica* were surveyed to detect the presence of *A. cantonensis* within the route of dispersal of *A. fulica*. The parasite was only found in the Philippines and Tahiti. The results for Tahiti provide the first recorded case of *A. cantonensis* infection for *Achatina fulica* in that country. The limited sampling of *A. fulica* and the patchy distribution of *A. cantonensis* could explain the lack of *A. cantonensis* among the populations sampled, with the exception of the Philippines and Tahiti. It is therefore difficult to assess if the snail has a role in the spread of the parasite, and the possibility of other gastropods such as *Laevicaulis alte* contributing to the spread of *A. cantonensis* is not discounted.

APPENDIX

Appendix 2.1: Command lines for generating likelihood scores for the 12 models in PAUP*

2.1.1. JC69

1. paup> log file=filename.JC69models.paupout (this saves an output file)
2. paup> execute dataset.nex
3. paup> set criterion=distance (sets the optimality criterion to distance)
4. paup> dset distance=ml (specifies the distance measure to maximum likelihood for the estimation of the likelihood score for the model, which will be used for the LRT)
5. paup> lset nst=1 basefreq=equal rates=equal pinvar=0 (this specifies the assumptions of the JC69 model, which are equal frequency for all bases and equal rate of substitutions)
6. paup> nj (this generates a neighbor-joining tree based on the assumptions of the model)
7. paup> likelihoods /basefreq=equal rates=equal pinvar=0 (this estimates a likelihood score based on the model)

2.1.2. JC69+ Γ (continued from JC69)

8. paup> likelihoods /basefreq=equal rates=gamma shape=estimate ncat=16 pinvar=0 (this adds the among site gamma rate variation with 16 categories dividing the discrete approximation of the gamma distribution)

9. paup> lset nst=1 basefreq=equal rates=gamma shape=previous ncat=16
pinvar=0
10. paup> nj
11. repeat 8-10 until the parameters and likelihood score do not change; note the likelihood score.

2.1.3. F81

1. paup> log file=filename.F81models.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml
5. paup> lset nst=1 basefreq=estimate rates=equal pinvar=0 (this specifies the assumptions of the F81 model, which are unequal base frequencies and equal rate of substitutions)
6. paup> nj
7. paup> likelihoods /basefreq=estimate rates=equal pinvar=0
8. paup> lset nst=1 basefreq=previous rates=equal pinvar=0
9. nj
10. repeat 7-9 until parameters and likelihood score do not change; note the likelihood score.

2.1.4. F81+ Γ (continued from F81)

11. paup> likelihoods /basefreq=estimate rates=gamma shape=estimate ncat=16
pinvar=0
12. paup> lset nst=1 basefreq=previous rates=gamma shape=previous ncat=16
pinvar=0
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the
likelihood score.

2.1.5. K2P

1. paup> log file=filename.K2Pmodels.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml
5. paup> lset nst=2 basefreq=equal tratio=estimate rates=equal pinvar=0 (this
specifies the assumptions of the K2P model, which are equal frequency of bases
and different substitution rates for transtitions and transversions)
6. paup> nj
7. paup> likelihoods /basefreq=equal tratio=estimate rates=equal pinvar=0
8. paup> lset nst=2 basefreq=equal tratio=previous rates=equal pinvar=0
9. paup> nj
10. repeat 7-9 until parameters and likelihood score do not change; note the
likelihood score.

2.1.6. K2P+ Γ (continued from K2P)

11. paup> likelihoods /basefreq=equal tratio=estimate rates=gamma
 shape=estimate ncat=16 pinvar=0
12. paup> lset nst=2 basefreq=equal tratio=previous rates=gamma shape=previous
 ncat=16 pinvar=0
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the
 likelihood score.

2.1.7. HKY85

1. paup> log file=filename.HKY85models.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml
5. paup> lset nst=2 basefreq=estimate tratio=estimate rates=equal pinvar=0 (this
 specifies the assumptions of the HKY85 model, which are unequal base
 frequencies and different substitution rates for transitions and transversions)
6. paup> nj
7. paup> likelihoods /basefreq=estimate tratio=estimate rates=equal pinvar=0
8. paup> lset nst=2 basefreq=previous tratio=previous rates=equal pinvar=0
9. paup> nj

10. repeat 7-9 until parameters and likelihood score do not change; note the likelihood score.

2.1.8. HKY85+ Γ (continued from HKY85)

11. paup> likelihoods /basefreq=estimate tratio=estimate rates=gamma
 shape=estimate ncat=16 pinvar=0
12. paup> lset nst=2 basefreq=previous tratio=previous rates=gamma
 shape=previous ncat=16 pinvar=0
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the likelihood score.

2.1.9. TN93

1. paup> log file=filename.TN93models.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml
5. paup> lset nst=6 basefreq=estimate rmatrix=estimate rates=equal pinvar=0
 rclass=(a b a a e a) (this specifies the assumptions of the TN93 model, which
 are unequal base frequencies and different substitution rates for transitions in
 purines and pyrimidines and transversions)
6. paup> nj

7. paup> likelihoods /basefreq=estimate rmatrix=estimate rates=equal pinvar=0
rclass=(a b a a e a)
8. paup> lset nst=6 basefreq=previous rmatrix=previous rates=equal pinvar=0
rclass=(a b a a e a)
9. paup> nj
10. repeat 7-9 until parameters and likelihood score do not change; note the
likelihood score.

2.1.10. TN93+ Γ (continued from TN93)

11. paup> likelihoods /basefreq=estimate rmatrix=estimate rates=gamma
shape=estimate ncat=16 pinvar=0 rclass=(a b a a e a)
12. paup> lset nst=6 basefreq=previous rmatrix=previous rates=gamma
shape=previous ncat=16 pinvar=0 rclass=(a b a a e a)
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the
likelihood score.

2.1.11. GTR

1. paup> log file=filename.GTRmodels.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml

5. paup> lset nst=6 basefreq=estimate rmatrix=estimate rates=equal pinvar=0 (this specifies the assumptions of the GTR model, which are unequal base frequencies and different substitution rates)
6. paup> nj
7. paup> likelihoods /basefreq=estimate rmatrix=estimate rates=equal pinvar=0
8. paup> lset nst=6 basefreq=previous rmatrix=previous rates=equal pinvar=0
9. paup> nj
10. repeat 7-9 until parameters and likelihood score do not change; note the likelihood score.

2.1.12. GTR+ Γ (continued from GTR)

11. paup> likelihoods /basefreq=estimate rmatrix=estimate rates=gamma shape=estimate ncat=16 pinvar=0
12. paup> lset nst=6 basefreq=previous rmatrix=previous rates=gamma shape=previous ncat=16 pinvar=0
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the likelihood score.

Appendix 2.2: Command lines in PAUP* for estimating the uncorrected and corrected distances as well as the transition and transversion values

1. paup> log file=filename.distances.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup>dset distance=p
5. paup> showdist (this generates the uncorrected total pairwise distances)
6. paup> savedist /format=onecolumn file=filename.distances.p.1col (this saves the pairwise uncorrected total distances in one column)
7. paup> dset subst=ti (this specifies that the transition distances will be computed)
8. showdist (this generates the transition distances)
9. paup>savedist /format=onecolumn file=filename.distances.p.ti.1col
10. paup> dset subst=tv (this specifies that the transversion distances will be computed)
11. showdist (this generates the transversion distances)
12. paup>savedist /format=onecolumn file=filename.distances.p.tv.1col
13. paup> dset subst=all (this restores the setting to pertain to total distances)
14. paup> set criterion=distance
15. paup> dset distance=ml
16. set the predetermined optimal criterion using the lset command
17. paup> showdist (this generates the corrected total distances)
18. paup> savedist /format=onecolumn file=filename.distances.ml.model.1col

Appendix 2.3: Command lines for the different tree-construction methods

2.3.1. Maximum likelihood (ML) method

The ML method was done in PHYML Version 2.4.4 package (Guindon *et al.*, 2005). The parameters of the optimal model was specified when the dataset analyzed.

Enter the sequence file name > sequence.file.phy
er the sequence file name > sequence.file.phy

Settings:

D:	Data type (DNA/AA)	DNA
I:	Input sequences interleaved (or sequential)	interleaved
S:	Analyze multiple data sets	no
B:	Non parametric bootstrap analysis	yes
	Number of replicates >	1000
	Print bootstrap trees (and statistics) ? [Y/n]	y
M:	Model of nucleotide substitution	(depending on optimal model)
E:	Base frequency estimates (empirical/ML)	ML
V:	Proportion of invariable sites (fixed/estimated)	fixed (p-invar = 0.00)
R:	One category of substitution rate [yes/no]	no (if with gamma distribution)
C:	Number of substitution rate categories	16 (if with gamma distribution)

A:	Gamma distribution parameter (fixed/estimated)	estimated (if with gamma distribution)
	Optimize alpha ? [Y/n]	Y
U:	Input tree (BIONJ/user tree)	BIONJ
O:	Optimize tree topology	yes

2.3.2. Bayesian inference

Bayesian inference (BI) analysis was carried out in the MrBayes Version 3.1.2 package (Ronquist & Huelsenbeck, 2003). The program was set to run with four independent chains, each of which aimed to converge on a stable likelihood value from a number of generations. Below are the command lines for approximating the posterior probabilities.

1. MrBayes > log start file=filename.mbout (this saves an output file)
2. MrBayes > execute filename
3. MrBayes > lset nst=1/2/6 rates=equal/gamma Ngammacat=16 (only for gamma distributed rates) (this states the model in which the tree will be based)
4. MrBayes > showmodel (this is to show the model settings)
5. MrBayes > help mcmc (this is to show default settings of the Markov Chain Monte Carlo analysis)
6. MrBayes > mcmc Nruns=1 Ngen=2000000 Temp=0.075 or 0.1 or 0.125 (the number of generations to be run as defined by Ngen and the temperature to heat the chains as defined by Temp need to be optimized as described below;

initially, values of 2000000 for Ngen and 0.1 for Temp are used and later on changed if necessary)

7. MrBayes > mcmc (this is to run the analysis)

At the end of the run, the program asked if the analysis should be continued. The word 'yes' was typed when the values from the four columns did not converge to similar likelihood scores. The number of additional generations (e.g. 1000000) was indicated to resume the analysis. Otherwise, the word 'no' was typed. When the cold chain (value enclosed with bracket) did not switch randomly across the four chains, the analysis was repeated but the Temp setting was changed.

When the analysis was completed, the state exchange information was displayed. As a rule of thumb, the values found on the upper diagonal, which expressed the acceptance rates of the Metropolis-Hastings mcmc sampler, should range between 0.1 and 0.7. When the acceptance rates were too low, the analysis was repeated but using a lower Temp. When the rates were too high, the analysis was repeated with a higher Temp.

8. MrBayes > sump burnin=value (e.g. 19001 if 2000000 generations were made;
this summarizes the samples of model parameters)

Note: The value given indicated the number of samples to be discarded. A sample contained 100 generations; however, the first generation is always sampled and should be included in the samples to be discarded.

9. MrBayes > sumt burnin =value (e.g. 19001 if 2000000 generations were made;
this summarized the samples of trees and branch lengths)

2.3.3. Neighbor-joining (NJ) method

The NJ method was done in PAUP* by starting out with the same parameters set by the identified optimal model for DNA sequence evolution, which was then followed by the commands listed below.

1. paup> nj brlens=yes (to display the table of branch lengths)
2. paup> savetrees /format=phylip brlens file=filename.nj.tree.phy (to save the tree in phylip format)
3. paup> savetrees /format=nexus brlens file=filename.nj.tree.nex (to save the tree in nexus format)
4. paup>bootstrap nreps=1000 method=nj keepall=yes
treefile=filename.nj.boottree (to calculate and save the bootstrap values for 1000 replicates)
5. paup>quit

2.3.4. Maximum parsimony (MP) method

The MP method was also carried out in Paup*; this did not employ any models of DNA sequence evolution.

1. paup>set maxtrees=100 increase=auto
2. paup> set criterion=parsimony
3. paup> hsearch swap=tbr addseq=random nreps=10 start=stepwise (this is to make a heuristic or non-exhaustive search of the most parsimonious trees)
4. paup> showtrees all
5. paup> savetrees /fmt=phylip brlens=yes file=filename.mp.phy

6. paup> savetrees /fmt=nexus brlens=yes file=filename.mp.nex
7. paup> contree /majrule=yes file=filename.mp.constree (this generates a consensus tree of all the equally most parsimonious trees found)
8. paup> bootstrap nreps=1000 search=heuristic keepall=yes
file=filename.mpboot.treefile (to calculate and save the bootstrap values for 1000 replicates)

Appendix 2.4: Command lines for the partition homogeneity test in PAUP* to determine if individual genes could be combined as a single dataset.

1. paup> execute concatenated.file.nex
2. paup> charpartition gene1:1-3435, gene2:3436-4296, gene3:4297-4624. (this is to specify which segment of the concatenated sequence belongs to which gene)
3. paup> hompart partition=genes nreps=1000 seed=123 search=heuristic (this is to perform a homogeneity test with 1000 homogeneity replicates searched heuristically using a random seed number of 123)

Appendix 2.5: The command lines to run the Shimodaira-Hasegawa test in PAUP* to compare for significant difference between optimal and constrained trees

1. paup> execute concatenated.file.nex
2. set the parameters of the optimal model
3. paup> constraints taxonmonophyly (MONOPHYLY) = ((1,2)) (where numbers refer to the taxon numbers; this specifies which taxa were constrained to form a clade based on an *a priori* hypothesis that they are monophyletic)
4. paup> nj brlens=yes enforce=yes constraints=taxonmonophyly (this generated a constrained neighbor-joining tree)
5. paup> savetrees /format=nexus brlens file=taxonmonophyly.nex (this saved the constrained neighbor-joining tree)
6. execute trees.file (this file should include both the constrained tree from no. 5 as well as the optimal neighbor-joining tree)
7. paup> showtrees all (this displayed both trees)
8. paup> lsocres all/shtest=rell (this computed for the likelihood scores of the two trees based on the estimated likelihood method and compared for significant difference between the two likelihood scores at $P=0.05$ level of significance)

Appendix 3.1: Alignment of the LSU rRNA gene for the Achatinoidea and three streptaxid outgroup taxa. Positions included for the phylogenetic analyses were marked by “m” within “NUCEOTIDES INCLUDED”.

	10	20	30	40	50	60
<i>Achatina fulica</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Achatina achatina</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Achatina stuhlmanni</i>	-----AGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Cochlitoma ustulata</i>	-----	---TTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Allopeas clavulinum</i>	-----AGAAC	ACATTGAACA	TCGACACCTT	GAACGCAAAAT	GGCGGCCTCG	GG-GCCATCC
<i>Bocageia</i> sp.	ATTGCAGAAC	ACATTGAACA	TCGATACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Eutomopeas layardi</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCAAAAT	GGCGGCCTCG	GG-GCCATCC
<i>Leptinaria lamellata</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCAAAAT	GGCGGCCTCG	GG-GCCATCC
<i>Paropeas achatinaceum</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCAAAAT	GGCGGCCTCG	GG-GCCATCC
<i>Riebeckia</i> sp.	-----AAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Rumina decollata</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Subulina octona</i>	-----	-----	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Subulina striatella</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Subulina vitrea</i>	-----	-----ACA	TCGATACCTT	GAACGCATAT	GGCGGCCTCG	GG-TCCATCC
<i>Subulona</i> sp.	-----	-----	-----CCTT	GAACGCATAT	GGCGGCCTCG	GGTCCATCC
<i>Tortaxis erectus</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Xerocerastus</i> sp.	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Zootecus insularis</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Coelioxaxis blandii</i>	-----AAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Pyrgina umbilicata</i>	-----AAC	ACATTGAACA	TCGACATCTT	GAACGCAAAAT	GGCGGCCTCG	GG-GCCATCC
<i>Glessula ceylanica</i>	ATTGCAGAAC	ACATTGAACA	TCGATACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Ceciloides gokweanus</i>	-----	-----	---ACATCTT	GAACGCAAAAT	GGCGGCCTCG	GG-TCCATCC
<i>Ferussacia folliculus</i>	---GCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Thyrophorella thomensis</i>	ATTGCAGAAC	ACATTGAACA	TcGACATCTT	GAACGCAAAAT	GGCGGCCTCG	GG-GCCATCC
<i>Gibbulinella dewinteri</i>	-----	-----	-----NCTT	GNACGCANAT	GGCGGCCNCG	GG-TCNNTCC
<i>Gonaxis quadrilateralis</i>	-----AGAAC	ACATTGAACA	TCGACAACTT	GAACGCATAT	GGCGGCCTCG	GG-TCAATCC
<i>Gonospira</i> sp.	-----AGAAC	ACATTGAACA	TCGACAACTT	GAACGCATAT	GGCGGCCTCG	GG-TAAATCC
NUCLEOTIDES INCLUDED	-----	-----mmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mm-mmmmmmm

	70	80	90	100	110	120
<i>Achatina fulica</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG---TACAA	AGCCTTCGCT	TCGTATGAGG
<i>Achatina achatina</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG---TACAA	AGCCTTCGCT	TCGCATGTGG
<i>Achatina stuhlmanni</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG---TACAA	AGCCTTCGCT	TCGTATGCGG
<i>Cochlitoma ustulata</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG---TACAA	AGCCTTCGCT	TCGTATGTGG
<i>Allopeas clavulinum</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---AACCA	AACCATCGCT	TCTCTTCTTC
<i>Bocageia</i> sp.	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---TCAA	AGCAATCGCT	TCCTGTTTGG
<i>Eutomopeas layardi</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---ATCA	AACCATCGCT	CCTCTT-TTC
<i>Leptinaria lamellata</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---ACTA	AACCAATCGCT	TCAGTT-TTT
<i>Paropeas achatinaceum</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---ACCA	AACCATCGCT	TCTTTTTTTT
<i>Riebeckia</i> sp.	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---TCAA	AGCAATCGCT	TCGAGATGGC
<i>Rumina decollata</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG---TCAA	AGCCTTCGCT	TCGCTTCTGG
<i>Subulina octona</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG--TCGCAA	AGCAATCGCT	TCGCTTTTGT
<i>Subulina striatella</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG--TCTCAA	AGCAATCGCT	TCCGTTTATA
<i>Subulina vitrea</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---TCAA	AGCAATCGCT	TCTATTTTGG
<i>Subulona</i> sp.	CGGNGCCACG	CCCGTCTGAG	GGTCGGCGAG	AATCTTAATA	AGCAATCGCT	TCTTATCGGA
<i>Tortaxis erectus</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG--TATCAA	AGCAATCGCT	TCGCACTAAA
<i>Xerocerastus</i> sp.	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AR---TCAA	AGCAATCGCT	TCGCTTTTTT
<i>Zootecus insularis</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG---ACAA	AGCAATCGCT	TCGCTTTTTT
<i>Coelioxaxis blandii</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---TCAA	AGCAATCGCT	TCGAGCTGGG
<i>Pyrgina umbilicata</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---ATCA	AACCAATCGCT	TCTCCTCAGA
<i>Glessula ceylanica</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---GTCAA	AGCAATCGCT	TCGAATTTGA
<i>Ceciloides gokweanus</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AA---ACTA	CACCAATCGCT	TCGCTTTATG
<i>Ferussacia folliculus</i>	CGAGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AG-----AA	ACTCATCGCT	TTTCATTGTC
<i>Thyrophorella thomensis</i>	CGGGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AT---ATCA	AACCAATCGCT	TCTCTTCAG-
<i>Gibbulinella dewinteri</i>	CGAGGCCACG	CCCGTCTGAG	GGTCGGGAGT	CTTAAAAAAA	GCCGTTTCGCT	TTCATGTTGG
<i>Gonaxis quadrilateralis</i>	CGAGGCCACG	CCCGTCTGAG	GGTCGGCGAG	AC---TTATA	AGCAATCGCT	TTTTCCGTTG
<i>Gonospira</i> sp.	CGAGGCCACG	CCCGTCTGAG	GGTCGGCGAG	TC--TTAAAA	GCCGTTTCGCT	TTCTGCTGAC
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	-----	-----	-----

	130	140	150	160	170	180	
Achatina fulica	-CAGCAGGTC	TCGCTC---A	ATTATATCCG	AGATCCGTCC	AGCTCTTC--	---CTCTATC	
Achatina achatina	GCAGCAGGTC	TCG-----	ATTATATCCG	AGATCCGTCT	A-TACTTC--	NTGNTCTANT	
Achatina stuhlmanni	-CAGCAGGTC	TCGCTT--AA	ATTTCCTGCC	AGATCTTC--	-----	---CTCTATC	
Cochlitoma ustulata	-CAGCAGGTC	TCGCTC---A	ATTATATCCA	AGATCCGTCT	AGCTCTTC--	---CTCTATC	
Allopeas clavulinum	TTACCAGACC	AAAGGCCG-C	ACGCCGTCGG	GGTCTTTTCG	TCTTCCGAAG	CTCGAAGGGA	
Bocageia sp.	CTCGGGGTTT	TAACATAAAC	CGTTTCGCGC	CGTCCTTCTT	CTTTTTCCTG	AGCTCGTGCG	
Eutomopeas layardi	TATGTGACCA	AAGGCCACCA	CGCCGTCGTG	AGACAGGTCT	TTCGCTCATC	CGAAGCTCGA	
Leptinaria lamellata	ATGACAACTG	GCATTCCCGT	TTCTTCCGAG	CTCGAGGAAG	AGCGGGAGAG	CCGCCGAAGC	
Paropeas achatinaceum	CTTATCAGAC	CAAAGGCAGC	GGCCGTCGCG	TGT--ATCGT	TCTTCCGAAG	CTCGAAGGGA	
Riebeckia sp.	GGTAACAGGT	CGCCTC-AAA	CAAGCTGCTC	CGATCCGATC	GCCGCCGTTT	CTTCCCGGGC	
Rumina decollata	-CAGCTGGTC	CGGTTT---	-----	-----	-----	-----TA	
Subulina octona	-CTCGACAGC	GGGTCTCCAG	CCTCCAGAGA	AGGCTT----	-----	-----	
Subulina striatella	-GTGACGGCG	GGTCTCCAGC	TTCCGATCGT	CCCCACAACT	TCCTTCGCGC	TCGGCAGGGA	
Subulina vitrea	CACAGGGGTC	TCCCGTTGGG	GGGTTGGCTG	GCAAGGCAAG	GCAGGCATAT	TTTCTTTCTC	
Subulona sp.	GAA-----	NNNNNNNNNN	NNNNN-----	-----	-----	-----	
Tortaxis erectus	-CCGACGAGT	GGTCGTCTCT	CCCCAACTTG	GCTCTGAGTT	TGCGGGCGGG	CAGGCCGACC	
Xerocerastus sp.	GGTNAACGGG	TCCNCCTTGN	TCAGNNAATT	CCGTTT-CAT	TGNGCTCGCG	GGAAACAAAA	
Zootecus insularis	TTTTTYCTTT	AAAAAATAAA	ATGCACCGGG	TTTGCTTAT	TCCCACCATG	TCGTTCTCGC	
Coeliaxis blandii	TTGGGTCGTC	TCGAAGTCCG	ATCAGCTCAG	TCCGTTCTCT	CGCGCTCATC	GTGGGAGGAT	
Pyrgina umbilicata	GACCAAAGGC	ACTCGCCGTC	CGTCCGTCGG	CCTTCCTAAG	CTCGAAGGAA	GCCTTTGATG	
Glessula ceylanica	CATGGGTAC	ACAAATTCNA	ANANACTTCN	AANAATNCTT	TTTTNTCCCC	GCTTTTGAAA	
Cecilioides gokweanus	GCAACAGGTT	TCCCCG-TCT	TCTTCCGAGC	T-----	-----	-----	
Ferussacia folliculus	CCTAAGGATT	CGACCTTGCC	ACCTCTGTGT	TCGTGTTAGA	GGGAGAAGGT	GCAGAATTCT	
Thyrophorella thomensis	-ATCAAAGGC	ACTCTCGC-C	CGTCCGTCGG	CCTTCCTAAG	CTCGAAGGAA	AGGCTTGATG	
Gibbulinella dewinteri	ACTGGTAGGC	CTACGCGGGC	CGTCGCCAGC	AGGTTTGGCT	TTTGCGGGAG	GGCATATTTT	
Gonaxis quadrilateralis	GACTGCTGCT	CGGGCCACCG	TCTGTCTGGG	TGANGAGGGG	NATTGACGCN	TCAACTCGTT	
Gonospira sp.	TGGAAGGCAA	CCGTGCGTCT	GTCACAGGCG	CGGAGGAAAG	TCCCTCTCGT	GTTCGACGCT	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	190	200	210	220	230	240	
Achatina fulica	C-ATCCGCGG	-CTCGTGCGG	A--GGGGTTA	CAGAGAGGAA	GAAT-CGGGC	G-----	
Achatina achatina	G-NGGTCCCG	CTGCTCTGTG	T---GGGACA	CGGAAAAGAA	GAATTGACN	N-----	
Achatina stuhlmanni	T--TCCGCGG	-CTCGTGCGG	-----GGA	CAGAAAGAAA	GAAT-CTGTC	G-----	
Cochlitoma ustulata	T-ATCCGCGG	-CTCGTGCGG	T---GGAGGA	CAGAAAGGCG	-----	-----	
Allopeas clavulinum	GAGCGCAGAC	ATGGTGGCGA	TGGGCCGCG	CAGAAGCGCT	TTGGGCTCTC	GTAGGTCGAT	
Bocageia sp.	GGTTGAGA--	-----	-----	-----	-----	-----	
Eutomopeas layardi	AGGGAGGGCG	AAGACCGCGG	CGACTGCGAT	GGGCCGCGC	AGGAGCGCGT	TGGGCTCTCG	
Leptinaria lamellata	GCATTGGGCT	TTCGCGGGAC	GCGCAGGGCA	CGTGCTCGTC	CGTCGCGCTC	TCGTACCGCG	
Paropeas achatinaceum	GGGCGAAGAT	ACGATGACGT	CGGGCCGCGG	CAGAAGCGCG	TTGGGCTGTC	GTAGGTCGAT	
Riebeckia sp.	TCGTGGGAGG	ATTCAAGTGG	TTAGTTAGGA	GGATGCTTTC	GAGAGGTGAC	CGTCGAAGCG	
Rumina decollata	CCACGTCTGT	TTTCACTCCG	TGCTCTGTGC	AGGGAGAAGC	TGCGTCGTGG	ATGA-----	
Subulina octona	-----	-----	-----	-----	-----	-----	
Subulina striatella	GGGTTTGGGT	GACGTTT---	-----	-----	-----	-----	
Subulina vitrea	CTAATTTTTT	TCATTTTCTT	CCCGCCCGTG	CGCTCTGACG	GGCGCGGGTA	GAGAGTGAAA	
Subulona sp.	-----	-----	-----	-----	-----	-----	
Tortaxis erectus	GTCTGAAGCG	GCTGGGCATT	CCGAGGATGA	GTGCTCCTCC	ACATGTCATC	CCCG-----	
Xerocerastus sp.	A-----	-NNNNNNNNN	N-----	-----	-----	-----	
Zootecus insularis	TCCGCCGTTC	TTGGGATGAA	AAGCGGCTCG	TGGAGTGGCC	GAA-----	-----	
Coeliaxis blandii	ACAAAAGGCG	GGGCGCGACT	NAAATGNGAC	CGTACAATCG	CACTGGGCT-	----NNNNNN	
Pyrgina umbilicata	ATGACGACGG	GCTTGCTGCC	GCAGAAGCGC	TTTGAG-CTG	TCGTGGGTCG	TCAGTGACCC	
Glessula ceylanica	AAAAA-----	NNNNNNNNN-	-----	-----	-----	-----	
Cecilioides gokweanus	-----	-----	-----	-----	-----	-----	
Ferussacia folliculus	CGGAAGCGCA	CTGGGCTTTC	GAGAATCACA	CCATTCTCGC	TCGAAAATAA	AGAATGAAGT	
Thyrophorella thomensis	ACGACG-GGC	TGCTGCTGCC	GCAGAAGCGC	TTTGGGCCTT	TCGTGGGTGC	TCAGCGACCC	
Gibbulinella dewinteri	TTTTTTGCCC	ACTCGCGTTA	CCAGCGGCGT	CCGTGAAGAC	CGAAGAAAGC	GCTTCTGAGC	
Gonaxis quadrilateralis	ACGGTGTGGT	TGTCCCAGAG	AGGAGGAAAA	GCGCTCCGGG	CTTTCGCGGN	TGGGCAAAAA	
Gonospira sp.	CCGGTTGCCG	AGAAAGCGGT	ATGGGCCTTC	GCGGATGATG	ACGTGCATCC	CCGCGGCCCTC	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	250	260	270	280	290	300			
Achatina fulica	-----	-----	-----	-----	-----	-----			
Achatina achatina	-----	-----	-----	-----	-----	-----			
Achatina stuhlmanni	-----	-----	-----	-----	-----	-----			
Cochlitoma ustulata	-----	-----	-----	-----	-----	-----GC			
Allopeas clavulinum	TTCAACGACC	ACCGTGGCCT	CAAGTACATC	GTGCGCCTGC	CCCGTCCGTT	TT--AATCGG			
Bocageia sp.	-----	-----	-----	-----	-----	-----TTAA			
Eutomoepas layardi	TAGGTCGATT	TCTACGACCG	CCGTGGCCTC	AAGTACATCG	TGCGCCTGCC	CCTGTCCGTT			
Leptinaria lamellata	TTGCCTCAAG	TACGTAGTGC	GCCCGCCCTG	TCCGATCAAT	CGGTCCGTTG	TGCTCTCGAC			
Paropeas achatinaceum	TTCAACGACC	CCCGTAGCCT	CAAGTACATC	GTGCGCCTGC	CCTGTCCGTT	TTTTAATCGG			
Riebeckia sp.	CAATGGGCTT	TCGAGGATGC	TTGTCTGCCTA	TNATGGANGG	AAAAAAGGAT	GGCTCGCTCC			
Rumina decollata	-----	-----	-----	-----	-----	-----			
Subulina octona	-----	-----	-----	-----	-----	-----			
Subulina striatella	-----	-----	-----	-----	-----	-----			
Subulina vitrea	ATACAGGAAA	AGAAGAAGAG	CCGCGGCCGC	CGTCGCCGCA	CG-----CCA	TTTTACTTTT			
Subulona sp.	-----	-----NNNNN	NNNNNNNNNN	NNNNNNNNNN	---TTTTTTT	TTTTTGTTA-			
Tortaxis erectus	-----	-----	-----	-----	-----	-----			
Xerocerastus sp.	-----	-----	-----	-----	-----	-----			
Zootecus insularis	-----	-----	-----	-----	-----	-----			
Coeliaxis blandii	NNNN-----	-----	-----	-----	-----	-----			
Pyrquina umbilicata	TCGTGGCTTC	AAGTACAACG	TGCGCCTGCC	CCTGTCCGTT	TTAGTCGGTC	TCGTTAGCTC			
Glessula ceylanica	-----	-----	-----	-----	-----	-----			
Cecilioides gokweanus	-----	-----	-----	-----	-----	-----			
Ferussacia folliculus	GGTTTCCGTT	GCCTGAAGTA	CTGAAAAAAA	CTGCGTCCTC	TGTCCGTTCT	AGCTAGCTCA			
Thyrophorella thomensis	TCGTGGCCTC	AAGTACAACG	TGCGCCTGCC	CCTGTCCGTT	TTGGTCGGTC	TCGTTAGCTC			
Gibbulinella dewinteri	CTTCGCGGAT	CGCATGATCC	CCGTGGCTTC	AAGTGTAATC	GATGCGCGCG	CGCCCTTTGC			
Gonaxis quadrilateralis	TCANCCCCGT	GGCCACAAGT	GTAAGAAGATG	CGCGCGTGTC	TGCTGANCTG	CCGCCTGCCT			
Gonospora sp.	AAGTGCAAGC	GTTGAGGGAG	CGTCTTAGCT	GCTGCTTCTG	TCCCGCGACC	TAAACGTGGA			
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----			

	310	320	330	340	350	360			
Achatina fulica	-GAGAAGGAA	GAAGAAGAGA	CCG-----	TCGA-AGCGC	TGTGGGCCGT	--CGTGGGAG			
Achatina achatina	ATANNGAAGG	G-AAGAGAGG	CCG-----	TCGA-AGCGC	CTGGGCCCGT	--CGTGGGAG			
Achatina stuhlmanni	-----GAG	AAGAAAGAGA	CCG-----	TCGA-AGCGC	TGTGGGCCGT	--CGTGGGAT			
Cochlitoma ustulata	GAAGGAAGAA	GAAGAAGAGA	CCG-----	TCGA-AGCGC	TGTGGGCCGT	--CGTGGGAG			
Allopeas clavulinum	TCTCGATGGT	TCTC-----	--TAACCGTC	GGAGCGCTCG	CTCGCTCGCC	GTCCGTCTGG			
Bocageia sp.	TAGGGCCGTG	CTGATG----	--AAGA-AG	CGCACTGGGC	TCT--CGAGG	AGAGAAGAAC			
Eutomoepas layardi	TTAATCGGTG	TCGATGGCTC	TCTAGCCGTC	TGAGCGCTCG	CTCGCTCGCC	GTCCGTCTGG			
Leptinaria lamellata	CGGAGCGCTC	GCCGTCAGTC	TGGCAGGACT	CGGCTCGCAT	AAAAGCATGA	AATCGAGCCT			
Paropeas achatinaceum	TCTC-----	-----	---TACCGTC	GGAGCGCTCG	CTCGCTCGCC	GTCCGTCTGG			
Riebeckia sp.	TTTTTTTTTT	-----	-----N	NNNNNNNNNN	-----	-----			
Rumina decollata	-----	---TGAAGA	CCG-----	TCGA-AGCGC	ACTGGGCTTT	--CGAGGACG			
Subulina octona	-----	---GGTGA	CCG-----	TAGA-AGCGC	CCTGGGCTCT	--CGAGGAGG			
Subulina striatella	-----GAAGA	GGCTTGCGCA	CCG-----	TGGA-AGCGC	CCTGGGCTGT	--CGAGGAGG			
Subulina vitrea	AACGTGAGAG	GACCCG----	---GAGAAG	CGCACTGGGC	TTT--CGAGC	TAAAGAAACA			
Subulona sp.	-----	-----	-----	-----	-----	-----			
Tortaxis erectus	-----	-----TGGC	CTGAAGTACA	AAATTTATGC	GCCGTCCCGT	CCAACCGAGC			
Xerocerastus sp.	-----	-----	-----N	NNNNNNNNNN	-----	-----			
Zootecus insularis	-----	---GACG---	---TCGAAG	CGTTCTGGCC	TT---CGAGG	ACGAGGAGGA			
Coeliaxis blandii	-----	-----	-----	NNNNNNNNNN	-----	-----			
Pyrquina umbilicata	TCGATGATCG	TCTCCGAGCG	CTAGCTAGCT	CGCTCGCTCG	CCGTCCGTCT	GGCAGGACTC			
Glessula ceylanica	-----	-----	-----NN	NNNNNNNNNN	-----	-----			
Cecilioides gokweanus	-----C	GGTTGG----	---ATGA-AG	CGAGGTTAGC	CGC--CGAAG	CGCCTTGGGT			
Ferussacia folliculus	CGAGTACGCC	ATCTCGTCTA	CTTGCCGTTA	TGTCTGGCAG	GACTCGGCTC	GCTAATTCTGA			
Thyrophorella thomensis	TCGATGATCG	TCT--GAGCG	ATAGC-----	--TCGCTCG	CCGTCCGTCT	GGCAGGACTC			
Gibbulinella dewinteri	GCGACCCCTC	CCGCCATTTA	GCGGCCCGCC	GAACCGCGGC	AGGACTCGGC	TCGTGATGAC			
Gonaxis quadrilateralis	CTCGGGGAAT	TGTTTGNTT	CCCAGCGCNC	CNCGACGAAG	AAACCTTTCT	GCATCCNCCG			
Gonospora sp.	CAGGGCGGAG	AGCACACAAC	ATCTCCTTTT	TTTCTTGTC	TCCGCCGGAT	CGTGGCGGGA			
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----			

	370	380	390	400	410	420	
Achatina fulica	GAGGAAGAA-	---GGGTTT	T-----	---TTTCCAC	CC-TT-CCCC	CGTGGCCTCA	
Achatina achatina	AGGAGGAGAG	G-----	GATTTTGGAT	TTTCTTCTT	TC-TT-CCCC	CGTGGCCTCA	
Achatina stuhlmanni	GAGGAATAAC	T---GGGTTT	TTCCCAG---	--TTTTTTTC	TC-TT-CCCC	CGTGGCCTCA	
Cochlitoma ustulata	GAGGAAGAAG	T---GCTTTT	TTTT-----	-----TCTC	TC-TT-TCCC	CGGGGCCTCC	
Allopeas clavulinum	CAGGACTCGG	CTCG---CTT	TTTCGTGCAT	CGAGCCTACC	AGACCACGTC	CGAA---GAA	
Bocageia sp.	A-CA---CTT	TCCCCGTGGC	CTGAAGTACA	CGTTGCGTCG	TCGTTTGTC	TTTGAGCGGC	
Eutomopeas layardi	CAGG---ACT	CGGCTCGCTT	TTTCTTGCAT	CGAGCCTGCC	AGACCCCGT	CCGAAGAAGG	
Leptinaria lamellata	GTCG---ACC	GTCCGAAGAA	GCGGCCTGCG	CGCAAAATAT	CCGCTAGAGA	GAGAAGGACC	
Paropeas achatinaceum	CAGGACTCGG	CTCG---CTC	AATCTTGCAT	CGAGCCTGCC	AGACCACGTC	CCAA---GAA	
Riebeckia sp.	-----	-----N	NNNNNNNNN	-----	-----	-----NN	
Rumina decollata	AGGAGGAT-C	G---AGTTTT	CTCT-----	-----TTCCA	CC-TT-CCCT	CGTAGCCTCA	
Subulina octona	A-----	-----	-----	-----TGGCTC	TTTTCTCTCC	CGTGGCTTCA	
Subulina striatella	AGGAGACGTA	TG-----	---TATGCTC	TGTCGTCTGC	GTCCTCTCCC	CGTGGCTTCA	
Subulina vitrea	AAACCGAAGT	TAGCCCGAT-	--GCCTGAAG	TACACGTTGC	GTCGTACAGT	CTTTGAGCGG	
Subulona sp.	-----	-----	-----	-----	-----	-----	
Tortaxis erectus	GCCCACGC--	-CCAACACCA	CGCTCCGCTC	GCCGTCCGTC	TGGCAGGACT	CGGCTCGCTT	
Xerocerastus sp.	-----	-----N	NNNNNNNNN	-----	-----	-----NN	
Zootecus insularis	T-GC-----	-----	---TGCT	TTATTTTCTT	CCTTCTTCCC	CGTGGCCTCA	
Coelioxys blandii	-----	-----N	NNNNNNNNN	-----	-----	-----NN	
Pyrgina umbilicata	GGCT---CGC	TTTATATTAT	CGCCTCGAGC	CTGCAGACTC	CGTCCAAAGA	AGAAGGCGCG	
Glessula ceylanica	-----	-----NNN	NNNNNNNNN	-----	-----	-----NNNN	
Cecilioides gokweanus	CT-----TCG	CGGAGGACGC	GGATACTGTG	CTCGTTATGC	GTATCTGAAA	CCTCCCCGTG	
Ferussacia folliculus	GCCT---GTC	AAAGACAACT	TGTCCGTAAG	CGGCTCGTGT	CGTGACGATG	ATAGGTGCCG	
Thyrophorella thomensis	GGCT---CGC	ATTATAATAT	CGCCTCGAGC	CTGCAGACTC	GGTCC---GA	AGAAGGCGCG	
Gibbulinella dewinteri	GCTA---TCG	AGCCTGCCCG	TCGTCCGAG	CGTTGCGGTG	CCTAACGCGC	GAGGAGAGGT	
Gonaxis quadrilateralis	AACC---GTT	GCAGGACTCG	GCTCGTGTTT	GTGTGGGTGG	CGGCGGCGAC	GANGCTGTCC	
Gonospira sp.	CTCG---GCT	CGCGGTGACC	AGAACATCGA	GCCTGCTCGG	TCAGTCTTGT	ACTACGGCGG	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	430	440	450	460	470	480	
Achatina fulica	AGTACACGCA	TGCGCCGTCC	TCGTCAATTC	AGCGACTCTT	TCGCT---CG	CCCGGAGCGC	
Achatina achatina	AGTGACAC-GG	TGCGCCGTCC	-CGTCAGTTT	AGCGACTCTT	TCGCT---TG	CCCGGAGCGC	
Achatina stuhlmanni	AGTACACGCA	TGCGCCGTCC	TCGTCAATTC	AGCGACTCTT	TCGCT---CG	CCCGGAGCGC	
Cochlitoma ustulata	AGTACACCCA	TGCGCCGTCC	TCGGCATTTT	AGCAACTCCT	TCGCTGGTCC	CCCGGAGCGC	
Allopeas clavulinum	GGCGGCCTGC	TTGCATGTCC	TTCCGATCGC	GGTTG-----	-----GAGA	GTGGGACCGA	
Bocageia sp.	CGGTCTTCGC	TCATCTCATC	CGAAACGCTC	GTGATGTTCC	TTGCTCTGCC	GTCCTCTCGG	
Eutomopeas layardi	CGGCTTGCTT	GCTTGCGTTG	CGTTCTGATG	GCGGATGGAG	GGCCGAGATC	GACTTTGGCT	
Leptinaria lamellata	GATGGCGGCG	TCGCCAGGAG	CGCAGGGAAG	AGAGTAAGCT	TTGCTCTCTT	TCCTCTCTCC	
Paropeas achatinaceum	GGCGGCTTGC	TTGCACCGCG	TTCCGATGAG	TGGAT-----	--TATGAGAC	CGGTTTCCGC	
Riebeckia sp.	NNNNNNNN--	-----	-----	-----	-----	-----	
Rumina decollata	AGTACACGAA	TGCGCCGTCC	CCGTCATCTG	AGCA-----	---TCGCTCT	CTCGAAGCGC	
Subulina octona	AGTACTCGCG	ATGCGT---	-----	---CGCCCA	GTCATTTT	GTTTGCGGCA	
Subulina striatella	AGTGCTCGAT	GCGTCGT---	-----	-CCAGTCAAT	TTCTTTTGCG	GCGGCACGCT	
Subulina vitrea	TC-----	TCCTTCGCTC	TTCTACGAAG	CCATTCTTCG	TTGCTTGCC	-GTCCGTCT-	
Subulona sp.	-----	-----	-----	-----	-----	-----	
Tortaxis erectus	ACTCGAAACC	AAATC-----	-----	---GAGCCT	GTCGGACCAG	CCCGAATCGG	
Xerocerastus sp.	NNNNNNNN--	-----	---TTGGAAG	ACGAGGACCT	TTGAACCGCC	TTGGCATTGG	
Zootecus insularis	AGTACATGTA	TGCGTCGTCC	CGTCATCTGA	GCGGCTACAT	CGCTCTCTCG	GAGCGCTCAT	
Coelioxys blandii	NNNNNNNN--	-----	-----	-----	---NNNNNNN	NNN---AAAA	
Pyrgina umbilicata	TTGCTTGCTT	GCTGGCTGTG	CTCGTTACGA	TTGGATTGAG	GGTGAGATCG	ATTTGCGCGG	
Glessula ceylanica	NNNNNN--	-----	-----	-----	-----	-----	
Cecilioides gokweanus	GCCTGAAGTA	CGCCTATGCG	TCGTCTGCC	GGAGAGCAGC	TCACTATCGT	CCGCTCGCCG	
Ferussacia folliculus	TGCAGTGGGG	TCTGGCTTTG	AGGATGCCAG	GAAGATGTGA	GATGCCACCG	TTTGGCGGCT	
Thyrophorella thomensis	TTGCTTGCTT	TGCTCGATTA	CGGTCG---	-----GAG	GGTGAGATCG	ATTTGCGCGG	
Gibbulinella dewinteri	GGCCTTGCTT	GGAGGCTGCG	CGCGTCCGCA	GCTGCAGAGC	AGCAGCAGAA	TCGTACGATG	
Gonaxis quadrilateralis	CTTCCCATTC	NAGCNTGCTC	GGTCGCNNTG	CTGAAANAAG	GGTTNTTGCG	GACAAACGTG	
Gonospira sp.	TTCTTGAAAA	ACGAGAGCTT	GTTTTGTGCG	GTGAGGCGCC	TCCTCTGGAT	TCCTACAGAG	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	610	620	630	640	650	660	
Achatina fulica	GGG----	CG CGCAAGCGCA	TGGG----	TC TGCTGCGGCG	GCGCCA----	-----	
Achatina achatina	GGT---GGCG	GGCAAGCG-A	TCGG----	TC T---GCGGCG	GCGCCAGCA-	-----	
Achatina stuhlmanni	GGG---CGCC	CGCGAGCGGA	TTGGGGGGTC	TGCTGTGGCG	GCGCCA----	-----	
Cochlitoma ustulata	GGG---CGCGC	GAGCGCA---	TGGG----	TC TGTGCGGCG	GCGCCA----	-----	
Allopeas clavulinum	-----	-----	-----	-----	-----	-----	
Bocageia sp.	AGTTTTTGTA	TTTTTTCATA	-----	-----	-----	-----	
Eutomopeas layardi	-----	-----	-----	-----	-----	-----	
Leptinaria lamellata	-----	-----	-----	-----	-----	-----	
Paropeas achatinaceum	-----	-----	-----	-----	-----	-----	
Riebeckia sp.	TACTACATAA	ACCGTTTCGC	A-----	-----	-----	-----	
Rumina decollata	AGAGGGCGAT	TGCCGCGGCG	GCGCCAATTT	TTTTTTGTCC	GTTCTAAA--	-----	
Subulina octona	AGGCAGTCGA	GAGCCGCAAC	GACGACGGCG	CCCCGTGGT-	-----	-AATGTGTCA	
Subulina striatella	AAGCAGTCGA	TGGCTAGCGA	AAGCGACGGC	GCCCGTGGTC	TATTACCGT	ACGTCGTCGT	
Subulina vitrea	CCGCGACGTC	GCCTATTTGT	AAAAAAAATT	TGTTTATTTT	TCTTTAC---	-----	
Subulona sp.	-----	-----	-----	-----	-----	-----	
Tortaxis erectus	-----	-----	-----	-----	-----	-----	
Xerocerastus sp.	CTCGCCGTCC	GTCCGGCAGG	ACTCGGCTCG	CTGTGAAAAC	AAAGCACACT	ATTCGAGCCT	
Zootecus insularis	TTGCTGCGGC	GGCGCCAAC	TTTTGTTTTT	CTTTCTAAA-	-----	-----	
Coelioxys blandii	-----	-----	-----	-----	-----	-----	
Pyrgina umbilicata	-----	-----	-----	-----	-----	-----	
Glessula ceylanica	-----	-----	-----	-----	-----	-----	
Cecilioides gokweanus	CTTTTTTTTT	-----	-----	-----	-----	-----	
Ferussacia folliculus	-----	-----	-----	-----	-----	-----	
Thyrophorella thomensis	-----	-----	-----	-----	-----	-----	
Gibbulinella dewinteri	-----	-----	-----	-----	-----	-----	
Gonaxis quadrilateralis	-----	-----	-----	-----	-----	-----	
Gonospira sp.	-----	-----	-----	-----	-----	-----	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	670	680	690	700	710	720	
Achatina fulica	GTCTTTCT--	-----AAA	ATCTTTCTTT	A-----	-----	-----	
Achatina achatina	GTCTCTATAA	CGACGACGTT	TNNTTTAAAA	TTTTTA----	-----	-----	
Achatina stuhlmanni	GTCTTTCT--	-----AAA	ATTTTCTTTT	A-----	-----	-----	
Cochlitoma ustulata	GTCTTTCT--	-----	TTCTTTCTTT	A-----	-----	-----	
Allopeas clavulinum	-----	-----	-----	-----	-----	-----	
Bocageia sp.	-----	-----	-----	-----	-----	-----	
Eutomopeas layardi	-----	-----	-----	-----	-----	-----	
Leptinaria lamellata	-----	-----	-----	-----	-----	-----	
Paropeas achatinaceum	-----	-----	-----	-----	-----	-----	
Riebeckia sp.	-----	-----	-----	-----	-----	-----	
Rumina decollata	-----	-----	-----	-----	-----	-----	
Subulina octona	TACTATTATT	T-----	-----	-----	-----	-----	
Subulina striatella	ACGAATCATT	T-----	-----	-----	-----	-----	
Subulina vitrea	-----	-----	-----	-----	-----	-----	
Subulona sp.	-----	-----	-----	-----	-----	-----	
Tortaxis erectus	-----	-----	-----	-----	-----	-----	
Xerocerastus sp.	GCC-GACCCC	GTCCGAAGCG	GCTTGCGGTT	TTGAAGTTTG	AGGCGCGCCG	CGCCCCGCTC	
Zootecus insularis	-----	-----	-----	-----	-----	-----	
Coelioxys blandii	-----	-----	-----	-----	-----	-----	
Pyrgina umbilicata	-----	-----	-----	-----	-----	-----	
Glessula ceylanica	-----	-----	-----	-----	-----	-----	
Cecilioides gokweanus	-----	-----	-----	-----	-----	-----	
Ferussacia folliculus	-----	-----	-----	-----	-----	-----	
Thyrophorella thomensis	-----	-----	-----	-----	-----	-----	
Gibbulinella dewinteri	-----	-----	-----	-----	-----	-----	
Gonaxis quadrilateralis	-----	-----	-----	-----	-----	-----	
Gonospira sp.	-----	-----	-----	-----	-----	-----	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	730	740	750	760	770	780			
Achatina fulica	-----	-----	-----	-----	-----	-----	T-		
Achatina achatina	-----	-----	-----	-----	-----	-----	T-		
Achatina stuhlmanni	-----	-----	-----	-----	-----	-----	T-		
Cochlitoma ustulata	-----	-----	-----	-----	-----	-----	T-		
Allopeas clavulinum	-----	-----	-----	-----	-----	-----	T-		
Bocageia sp.	-----	-----	-----	-----	-----	-----	T-		
Eutomoepas layardi	-----	-----	-----	-----	-----	-----	T-		
Leptinaria lamellata	-----	-----	-----	-----	-----	-----	T-		
Paropeas achatinaceum	-----	-----	-----	-----	-----	-----	T-		
Riebeckia sp.	-----	-----	-----	-----	-----	-----	T-		
Rumina decollata	-----	-----	-----	-----	-----	-----	T-		
Subulina octona	-----	-----	-----	-----	-----	-----	C-		
Subulina striatella	-----	-----	-----	-----	-----	-----	C-		
Subulina vitrea	-----	-----	-----	-----	-----	-----	T-		
Subulona sp.	-----	-----	-----	-----	-----	-----	T-		
Tortaxis erectus	-----	-----	-----	-----	-----	-----	T-		
Xerocerastus sp.	GAGAGGTGGA	GGAGGGCGAT	TGCTGCCCGC	GAGGCGCCAG	TATTTANTAT	TTTCTAAAT-			
Zootecus insularis	-----	-----	-----	-----	-----	-----	T-		
Coeliaxis blandii	-----	-----	-----	-----	-----	-----	T-		
Pyrquina umbilicata	-----	-----	-----	-----	-----	-----	T-		
Glessula ceylanica	-----	-----	-----	-----	-----	-----	T-		
Cecilioides gokweanus	-----	-----	-----	-----	-----	-----	T-		
Ferussacia folliculus	-----	-----	-----	-----	-----	-----	T-		
Thyrophorella thomensis	-----	-----	-----	-----	-----	-----	T-		
Gibbulinella dewinteri	-----	-----	-----	-----	-----	-----	T-		
Gonaxis quadrilateralis	-----	-----	-----	-----	-----	-----	TN		
Gonospira sp.	-----	-----	-----	-----	-----	-----	T-		
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	m-		

	790	800	810	820	830	840			
Achatina fulica	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Achatina achatina	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Achatina stuhlmanni	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Cochlitoma ustulata	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Allopeas clavulinum	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Bocageia sp.	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Eutomoepas layardi	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Leptinaria lamellata	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Paropeas achatinaceum	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Riebeckia sp.	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Rumina decollata	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Subulina octona	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Subulina striatella	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Subulina vitrea	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Subulona sp.	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Tortaxis erectus	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Xerocerastus sp.	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Zootecus insularis	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Coeliaxis blandii	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Pyrquina umbilicata	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Glessula ceylanica	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Cecilioides gokweanus	CCGACCTCAG	TTCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Ferussacia folliculus	CCGACCTCAG	ATGGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Thyrophorella thomensis	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Gibbulinella dewinteri	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Gonaxis quadrilateralis	CCGACCTCAG	ATCGGAGGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
Gonospira sp.	CCGACCTCAG	ATCGGACGAG	ACTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA			
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm			

	850 860 870 880 890 900
Achatina fulica	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Achatina achatina	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Achatina stuhlmanni	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Cochlitoma ustulata	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Allopeas clavulinum	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Bocageia sp.	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Eutomoepas layardi	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Leptinaria lamellata	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Paropeas achatinaceum	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Riebeckia sp.	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Rumina decollata	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Subulina octona	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Subulina striatella	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Subulina vitrea	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Subulona sp.	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Tortaxis erectus	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Xerocerastus sp.	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Zootecus insularis	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Coeliaxis blandii	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Pyrgina umbilicata	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Glessula ceylanica	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGT GGAAGAGCC CAGCACCAGAA
Cecilioides gokweanus	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Ferussacia folliculus	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Thyrophorella thomensis	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAATAGCC CAGCACCAGAA
Gibbulinella dewinteri	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAAGAGCC CAGCACCAGAA
Gonaxis quadrilateralis	AGAAACTAAC AAGGATTTCC CCAATAACGG CGAGTGAAGC GGGATAGCC CAGCACTGAA
Gonospira sp.	AGAAACTAAC AAGGATTTCC CCAGTAACGG CGAGTGAAGC GGAACGCC CAGCACCAGAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	910 920 930 940 950 960
Achatina fulica	TCCCTCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Achatina achatina	TCCCTCAGTG TCACGCTGAC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Achatina stuhlmanni	TCCCTCAGTG TCACGCTGAC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Cochlitoma ustulata	TCCCTCAGTG TCACGCTGAC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Allopeas clavulinum	TCCCCCAGTG TCACGCTGGC GGAAGATGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Bocageia sp.	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Eutomoepas layardi	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Leptinaria lamellata	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Paropeas achatinaceum	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Riebeckia sp.	TCCCCCGTG TCACGCCGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Rumina decollata	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Subulina octona	TCCCTCAGTG TCACGCTGAC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Subulina striatella	TCCCTCAGTG TCACGCTGAC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Subulina vitrea	TCCCTCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Subulona sp.	TCCCCCAGTG TCATGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Tortaxis erectus	TCCCCCGTG TCACGCCGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCTTCAGAG
Xerocerastus sp.	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Zootecus insularis	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Coeliaxis blandii	TCCCCCGTG TCATGCCGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Pyrgina umbilicata	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Glessula ceylanica	TCCCCCAGTG TCATGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Cecilioides gokweanus	TCCCTCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Ferussacia folliculus	TCCCTCAGTG TCACGCTGAC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Thyrophorella thomensis	TCCCCCAGTG TCACGCTGGC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Gibbulinella dewinteri	TCCCTCAGTG TCACGCTGAC GGAAGATGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
Gonaxis quadrilateralis	TCCCTCAGTG TCACGCTGAC GGAAGCTTTG GTGTGTGGGA CCCCACCAGT CGCATCAGAG
Gonospira sp.	TCCCTCAGTG TCACGCTGAC GGAAGCTGTG GTGTGTGGGA CGCCACCAGT CGCATCAGAG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	970	980	990	1000	1010	1020
Achatina fulica	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Achatina achatina	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Achatina stuhlmanni	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Cochlitoma ustulata	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Allopeas clavulinum	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Bocageia sp.	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Eutomoepas layardi	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Leptinaria lamellata	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Paropeas achatinaceum	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Riebeckia sp.	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Rumina decollata	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Subulina octona	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Subulina striatella	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Subulina vitrea	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Subulona sp.	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Tortaxis erectus	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Xerocerastus sp.	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Zootecus insularis	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Coeliaxis blandii	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Pyrquina umbilicata	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Glessula ceylanica	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Ceciloides gokweanus	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Ferussacia folliculus	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Thyrophorella thomensis	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Gibbulinella dewinteri	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Gonaxis quadrilateralis	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
Gonospora sp.	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCG
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	1030	1040	1050	1060	1070	1080
Achatina fulica	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Achatina achatina	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Achatina stuhlmanni	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Cochlitoma ustulata	CCACTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Allopeas clavulinum	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Bocageia sp.	CCCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Eutomoepas layardi	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Leptinaria lamellata	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Paropeas achatinaceum	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Riebeckia sp.	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Rumina decollata	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Subulina octona	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Subulina striatella	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Subulina vitrea	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Subulona sp.	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Tortaxis erectus	CCACTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGT
Xerocerastus sp.	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Zootecus insularis	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Coeliaxis blandii	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Pyrquina umbilicata	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Glessula ceylanica	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Ceciloides gokweanus	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Ferussacia folliculus	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Thyrophorella thomensis	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Gibbulinella dewinteri	CTCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Gonaxis quadrilateralis	CTCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
Gonospora sp.	CTCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	1090	1100	1110	1120	1130	1140		
Achatina fulica	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Achatina achatina	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Achatina stuhlmanni	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Cochlitoma ustulata	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Allopeas clavulinum	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Bocageia sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Eutomopeas layardi	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Leptinaria lamellata	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Paropeas achatinaceum	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Riebeckia sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Rumina decollata	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-TGCACGAG	TCCGATAGCG	GACAAGTACC		
Subulina octona	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Subulina striatella	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Subulina vitrea	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Subulona sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Tortaxis erectus	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	G-TGCACGAG	TCCGATAGCG	GACAAGTACC		
Xerocerastus sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Zootecus insularis	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Coeliaxis blandii	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Pyrgina umbilicata	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Glessula ceylanica	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	G-TGCACGAG	TCCGATAGCG	GACAAGTACC		
Cecilioides gokweanus	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Ferussacia folliculus	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	G-TGCACGAG	TCCGATAGCG	GACAAGTACC		
Thyrophorella thomensis	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Gibbulinella dewinteri	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	TTTGCACGAG	TCCGATAGCG	GACAAGTACC		
Gonaxis quadrilateralis	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
Gonospira sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	m-mmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	1150	1160	1170	1180	1190	1200		
Achatina fulica	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Achatina achatina	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Achatina stuhlmanni	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Cochlitoma ustulata	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Allopeas clavulinum	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Bocageia sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Eutomopeas layardi	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Leptinaria lamellata	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Paropeas achatinaceum	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Riebeckia sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Rumina decollata	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Subulina octona	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Subulina striatella	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Subulina vitrea	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Subulona sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Tortaxis erectus	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Xerocerastus sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Zootecus insularis	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Coeliaxis blandii	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Pyrgina umbilicata	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Glessula ceylanica	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Cecilioides gokweanus	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Ferussacia folliculus	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Thyrophorella thomensis	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCTCAG		
Gibbulinella dewinteri	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Gonaxis quadrilateralis	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
Gonospira sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	1210	1220	1230	1240	1250	1260
Achatina fulica	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCTCGG
Achatina achatina	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCTCGG
Achatina stuhlmanni	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Cochlitoma ustulata	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCTCGG
Allopeas clavulinum	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Bocageia sp.	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Eutomepeas layardi	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Leptinaria lamellata	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Paropeas achatinaceum	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Riebeckia sp.	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Rumina decollata	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Subulina octona	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Subulina striatella	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Subulina vitrea	AGGTAAACGG	GTGGATCCGC	AAATTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Subulona sp.	AGGTAAACGG	GTGGATCCGC	AAATTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Tortaxis erectus	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCT	CGCGGAATTC	AGCGCGGCGC	GCGGCCTGTG
Xerocerastus sp.	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Zootecus insularis	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Coeliaxis blandii	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Pyrquina umbilicata	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Glessula ceylanica	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCT	CGCGGAATTC	AGCGCGGCGC	GCGATCTGTG
Ceciloides gokweanus	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCT	CGCGGAATTC	AGCGCGGCGC	GCGGTTT-GG
Ferussacia folliculus	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Thyrophorella thomensis	AGGTAAACGG	GTGGATCCGC	AAAGTCGGCC	CGCGGAATTC	AGCGCGGCGC	GCGGCCT-GG
Gibbulinella dewinteri	AGGTAAACGG	GTGGATCCGC	AATGTCGGCC	CGCGGAATTC	AGCGCGGCGT	GCGGCCT-GG
Gonaxis quadrilateralis	AGGTAAACGG	GTGGATCCGC	AATGTCGGCC	CGCGGAATTC	AGCGCGGCGT	GCGGCCT-GG
Gonospora sp.	AGGCAAACGG	GTGGATCCGC	AATGTCGGCC	CGCGGAATTC	AGCGCGGCGT	GCGGCTC-GG
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	1270	1280	1290	1300	1310	1320
Achatina fulica	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Achatina achatina	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Achatina stuhlmanni	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Cochlitoma ustulata	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Allopeas clavulinum	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Bocageia sp.	GGCGACTGCG	GCCGGGGATC	CCTGGGACCC	CGCTGCGGTG	TCGATCCGGG	TTCCGCCGCG
Eutomepeas layardi	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Leptinaria lamellata	GGCGATTGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Paropeas achatinaceum	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Riebeckia sp.	GACTATCGCA	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Rumina decollata	GGCTATTGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Subulina octona	GGCGATCGCT	GCCGGGGATC	CCTGGGACCC	CGCTGTGGTG	TCGATCCGGG	CTCCGCCGCG
Subulina striatella	GGCGATCGCT	GCCGGGGATC	CCTGGGACCC	CGCTGTGGTG	TCGATCCGGG	CTCCGCCGCG
Subulina vitrea	GGCGACTGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Subulona sp.	GGCGACTGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Tortaxis erectus	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Xerocerastus sp.	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Zootecus insularis	GGCGATCGCG	GCCGGGGATC	CCTGGGACCT	CGCCGTGGTG	TCGAACCGGG	CTCCGCCGCG
Coeliaxis blandii	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGTGC	TCGATCCGGG	CTCCGCCGCG
Pyrquina umbilicata	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Glessula ceylanica	GGCGATTGCG	GCCGGGGATC	CCTGGGACCC	CGCCGTGATG	TCGAACCGGG	TTCCGCCGCG
Ceciloides gokweanus	GGCGATTGCG	GCCGGGGATC	CCTGGGACCC	CGCCGTGGTG	TCGAACCGGG	CTCCGCCGCG
Ferussacia folliculus	GACAATCGTG	GCCGGGGATC	CCTGGGACCC	CGCTTCGGTG	TCGAGCCGGG	CTCCGCCGCG
Thyrophorella thomensis	GGCGATCGCG	GCCGGGGATC	CCTGGGACCC	CGCCGCGGTG	TCGATCCGGG	CTCCGCCGCG
Gibbulinella dewinteri	GGCCGTGCTG	GCAAGGGATC	CCTGGGACCC	TGCCGCGATG	TCGAGCCCGG	CCCGGCCGCG
Gonaxis quadrilateralis	GACCGTCTG	GCCAGGGATC	CCTGGGACCC	TGCCGCGATG	TCGAGCTGGG	CCCGGCCGCG
Gonospora sp.	GGACGTCTG	GCCAGGGATC	CCTGGGACCC	TGCCGCGATG	TCGAGCTGGG	CCTCGGCCG
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	1330	1340	1350	1360	1370	1380		
Achatina fulica	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGCTG	GA--GCCGTC	AGAGGCCGGG	
Achatina achatina	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGCTG	GA--GCCGTC	AGAGGCCGGG	
Achatina stuhlmanni	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGCTG	GA--GCCGTC	AGAGGCCGGG	
Cochlitoma ustulata	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGCTG	GA--GCCGTC	AGAGGCCGGG	
Allopeas clavulinum	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGCG	G---GTCGTC	AGAGGCCGGG	
Bocageia sp.	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGACG	G---GCCGTC	AGAGGCCGGG	
Eutomopeas layardi	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGCG	----GTCGTC	AGAGGCCGGG	
Leptinaria lamellata	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GTTTCTGCG	----GCCGTC	AGAGGCCGGG	
Paropeas achatinaceum	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGG	----GTCGTC	AGAGGCCGGG	
Riebeckia sp.	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGG-T	----TCCGTC	AGAGGCCGGG	
Rumina decollata	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGCGG	GA--GCCGTC	AGAGGTCGGA	
Subulina octona	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTTCGCTG	GT--GCCGTC	AGAGGCCGGG	
Subulina striatella	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTTCGCTG	GT--GTCGTC	AGAGGCCGGG	
Subulina vitrea	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGGCT	GG--GCCGTC	AGATGCCGGG	
Subulona sp.	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGGCT	GG--GCCGTC	AGATGCCGGG	
Tortaxis erectus	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGCGG	GA--GCCGTC	AGCAGCCGTG	
Xerocerastus sp.	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGCGG	GA--GCCGTC	AGAGGCCGGG	
Zootecus insularis	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGCGG	GA--GCCGTC	AGAGGCCGGA	
Coeliaxis blandii	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCCGG-C	----TCTGTC	AGAGGCCGGG	
Pyrgina umbilicata	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGT	----GTCGTC	AGAGGCCGGG	
Glessula ceylanica	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTTCGCTG	GT--GCCGTC	AGAGGCCGGG	
Cecilioides gokweanus	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGCG	----GCCGCC	AGAGGCCGGG	
Ferussacia folliculus	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGCGA	GA--ACGCTC	AGAAGTCCCG	
Thyrophorella thomensis	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCTGT	----GTCGTC	AGAGGCCAGG	
Gibbulinella dewinteri	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTTCGGA	----GCCGTC	AAAAGCCCGG	
Gonaxis quadrilateralis	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTTCGATT	GAAGGCCGTC	AAAAGCCCGG	
Gonospira sp.	TGCAC	TTTCC	GCGGGCAGAG	TGCCACAACC	GGTTTCGGG	----GCCGTC	AAAAGCCCGG	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1390	1400	1410	1420	1430	1440		
Achatina fulica	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Achatina achatina	C-GAGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Achatina stuhlmanni	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Cochlitoma ustulata	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Allopeas clavulinum	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Bocageia sp.	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Eutomopeas layardi	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Leptinaria lamellata	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Paropeas achatinaceum	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Riebeckia sp.	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Rumina decollata	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Subulina octona	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Subulina striatella	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Subulina vitrea	T-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Subulona sp.	AAGGGTTGTC	GGTGGGGGCG	CTCGCCGTCC	CTACCAGCCC	---AACTCGG	CGGACGGCCT		
Tortaxis erectus	T-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---GCAACGG	CGGACGGCCT		
Xerocerastus sp.	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Zootecus insularis	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Coeliaxis blandii	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCC	GCCGCCCGG	CGGACGGCCT		
Pyrgina umbilicata	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Glessula ceylanica	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---GTTACGG	CGGACGGCCT		
Cecilioides gokweanus	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Ferussacia folliculus	T-GGGTTGTC	GGTGGGGGCG	TTGCG-GTGC	CTACCAGCCC	---GCTGAGG	CGGACGGCCT		
Thyrophorella thomensis	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Gibbulinella dewinteri	C-GGGTTGTC	GGTGGGGGCG	TTGCG-GTCC	CTACCAGCCT	---GCCCCGG	CGGACGGCCT		
Gonaxis quadrilateralis	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT		
Gonospira sp.	C-GGGTTGTC	GGTGGGGGCG	CTCGC-GTCC	CTACCAGCCT	---TCCCGGG	CGGACGGCCT		
NUCLEOTIDES INCLUDED	m-mmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	1450	1460	1470	1480	1490	1500
Achatina fulica	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTACCCG	GCCCCCTCTGC	GCGAGTCCGA
Achatina achatina	-CGGGACCGA	GGGACCGCCG	-CGCGCTTCG	AGGCTACCCG	ACCCCTCCGC	GCGAGTCCGA
Achatina stuhlmanni	-CGGGACCGA	GGAACCTGCCG	-CGCGCTTCG	AGGCTACCCG	ACCCCTCCGC	GCGAGTCCGA
Cochlitoma ustulata	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTACCCG	ACCCCTCCGC	GCGAGTCCGA
Allopeas clavulinum	-CGGGACCGA	GGAACCGCCG	-CGCACTTCG	AGGCCGCCT-	GCCCCCTCCGC	GCGAGTTCGA
Bocageia sp.	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTACCT-	GCCCCCTCCGC	GCGAGTTCGA
Eutomepeas layardi	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCCGCCT-	GCCCCCTCCGC	GCGAGTTCGA
Leptinaria lamellata	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCCGCCT-	GCCCCCTCCGC	GCGAGTTCGA
Paropeas achatinaceum	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCCGCCT-	GCCCCCTCCGC	GCGAGTTCGA
Riebeckia sp.	-TGGGATCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTACCT-	GCTCCTCCGC	GCGAGTTCGA
Rumina decollata	-CGGGACCGA	GGGACCGCCG	-CACGCTTCG	AGGCTACCT-	GCTCCTCCGC	GCGAGTCCGA
Subulina octona	-CGGGACCGA	GGAACCTGCCG	-CGCGCTTCG	AGGCTACCC-	GCTCCTCCGC	GCGAGTTCGA
Subulina striatella	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTACCC-	GCTCCTCCGC	GCGAGTTCGA
Subulina vitrea	-CGGGACCGA	GGAACCGCCG	-CGTGTTCG	AGGCTACTC-	GCTCCTCCGC	GTGAGTTCGA
Subulona sp.	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTGCTC-	GCTCCTCCGC	GCGAGTTCGA
Tortaxis erectus	-CGGGACCGA	GGAACCTGCCG	-CGCGCTTCG	AGGCTACCT-	GCCCCCTCCGC	ACGAGTTCGA
Xerocerastus sp.	-CGGGACCGA	GGGACCGCCG	-CACGCTTCG	AGGCTACCT-	GCTCCTCAGC	GCGAGTCCGA
Zootecus insularis	-CGGGACCGA	GGGACCGCCG	-CGCGCTTCG	AGGCTACCT-	GCTCCTCCGC	GCGAGTTCGA
Coeliaxis blandii	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTACCT-	GCTCCTCCGC	GCGAGTCCGA
Pyrquina umbilicata	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTACCT-	GCCCCCTCCGC	GCGAGTTCGA
Glessula ceylanica	-CGGGACCGA	GGAACCTGCCG	-CGCGCTTCG	AGGCTACCT-	GCTCCTCCGC	ACGAGTTCGA
Cecilioides gokweanus	-CGGGACCGA	GGAACAGCCG	-CGCGCTTCG	AGGCCACAT-	GCCCCCTCCGT	GCGAGTCCGA
Ferussacia folliculus	-TGGGACCGA	GGAACAGCCG	TGCGCTTCG	AGGCTACCT-	GCTCCTCCGT	GTGAGTTCAA
Thyrophorella thomensis	-CGGGACCGA	GGAACCGCCG	-CGCGCTTCG	AGGCTGCTC-	GCCCCCTCCGC	GCGAGTTCGA
Gibbulinella dewinteri	GCCAGACCGA	GGAACCGCCG	-CTCGCTTCG	AGGCTACCC-	GCTCCTCAGG	GCGAGTTCGA
Gonaxis quadrilateralis	-CCGGACCGA	GGAACCGCCG	-CTCGCTTCG	AGGCTACCC-	GCTCCTCCGG	GTGAGTTCGA
Gonospora sp.	-CCGGACCGA	GGAACCGCCG	-CTCGCTTCG	AGGCTGTCC-	GCTCCTCAGG	GTGAGTTCGA
NUCLEOTIDES INCLUDED	-mmmmmmmm	mmmmmmmm	-mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	1510	1520	1530	1540	1550	1560
Achatina fulica	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CGCGACCGGG	CC-GGGCTAG
Achatina achatina	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	TGCGACCGGG	CC-GGGCTAG
Achatina stuhlmanni	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CAAGACCGGG	CC-GGGCTAG
Cochlitoma ustulata	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CGCGACCGGG	CC-GGGCTAG
Allopeas clavulinum	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	TGCGACCGGG	ATCGGGCCGG
Bocageia sp.	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CGTGATCGGG	TT-GGGCTAG
Eutomepeas layardi	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	TGCGACCGGG	ATCGGGCCGG
Leptinaria lamellata	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	-CTGACAGGG	ATCGGGCCGG
Paropeas achatinaceum	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	TGCGACCGGG	ATCGGGCCGG
Riebeckia sp.	CTGGGAGAGA	CTGGGCAACC	GTGTCTKCCG	ACCGCTCGCT	CGCGATCGGG	TTGGGGCTAG
Rumina decollata	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CGCGACCGGG	CC-GGGCTAG
Subulina octona	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	TGCGACCGGG	CC-GGGATAG
Subulina striatella	CTGGGAGAGA	CCGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	TGTGATCGGG	CC-GGGATAG
Subulina vitrea	CCGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACTGCTCGCG	CGCGACCGGG	TT-GGGTCAG
Subulona sp.	CCGGGAGAGA	CTGGGCAACC	GTGTCTTCCT	ACCGCTCGCG	CGCGATTGGG	TT-GGGCTAG
Tortaxis erectus	CTGGAAGAGA	CTGGGTAACC	GTGGCTTCGG	AATGCTCGAG	CGCGATCGGG	CC-GGGCTAG
Xerocerastus sp.	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CGCGACCGAG	CC-GGGCTAG
Zootecus insularis	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CGCGACCGGG	CC-GGGCTAG
Coeliaxis blandii	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCT	CGCGATCGGG	TTGGGGTTAG
Pyrquina umbilicata	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	TGCGACCGGG	ATCGGGCCGG
Glessula ceylanica	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	AATGCTCGAG	CGTGACTGGG	TC-GGGCTAA
Cecilioides gokweanus	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CGCGACCGGG	ATCGTGTCCG
Ferussacia folliculus	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCA	CGCGACCGGG	TC-GGGCTAG
Thyrophorella thomensis	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCGCG	CGCGACCGGG	ATCGGGCCGG
Gibbulinella dewinteri	CTGGAAGAGA	CTGGGCAACC	GTGACTTCGG	ACCGCTCATC	CTCGATCGGG	CC-GGGCTAG
Gonaxis quadrilateralis	CAGGAAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCATC	CTCGACCGGG	CC-GGGCTAG
Gonospora sp.	CTGGGAGAGA	CTGGGCAACC	GTGTCTTCGG	ACCGCTCATC	CTCGATCGGG	CC-GGGCCAG
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mm-mmmmm

	1570	1580	1590	1600	1610	1620
Achatina fulica	CCGGGCGT-C	GCG----ACT	GTAA---CAG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Achatina achatina	CCGGGCGC--	G-----ACT	GAAG---CAG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Achatina stuhlmanni	CCGGGCGC-C	GCG----ACT	GTAA---CAG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Cochlitoma ustulata	CCGGGCGT-C	GCG----ACT	GTAA---CAG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Allopeas clavulinum	CCGGGTGC-C	GCTTGT----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Bocageia sp.	CCGGGTGC--	GTTAGT----	-----GAG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Eutomopeas layardi	CCGGGCGC-C	GCTTGT----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Leptinaria lamellata	CCGGGCGC--	GCACGC----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Paropeas achatinaceum	CCGGGCGC-C	GCTTGT----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Riebeckia sp.	CCGGGCGC-C	GCCAGT----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Rumina decollata	CCGGGCGT--	GCACA-----	-----CGG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Subulina octona	CCGGGCGCGC	GCACGCA---	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Subulina striatella	CCGGGCGCGC	GCATCGAG--	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Subulina vitrea	CCGGGCAC--	GCCCGT----	-----CAG	GGTTCGTGGC	GAGTCCGTCG	GCATTCCACC
Subulona sp.	CCGGGCGC--	GCCCGTC----	-----CAG	GGTTCGTGGC	GAGTCCGTCG	GCATTCCACC
Tortaxis erectus	CCGGGCGC--	GCAACGCTT-	-----CAG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Xerocerastus sp.	CCGGGCGT--	GCACA-----	-----CTG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Zootecus insularis	CCGGGCGT--	GCACA-----	-----CGG	GGTTGGTGGC	GAGTCCGTCG	GCATTCCACC
Coelioxys blandii	CCGGGCGC-C	GCCAGT----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Pyrgina umbilicata	CCGGGCGC-C	GCTCGT----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Glessula ceylanica	CCGGGCGT--	GCAATTT---	-----CAG	GGTGTGTGGC	GAGTCCGTCG	GCATTCCACC
Cecilioides gokweanus	CCGGGTGC--	GTTCTG----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Ferussacia folliculus	CCGGGCGC--	GCGTCAGT--	-----CAG	GGTTGGTGGC	AAGTCTGTCT	GCATTCCACC
Thyrophorella thomensis	CCGGGCGC-C	GCTCGT----	-----CAG	GGTTTGTGGC	GAGTCCGTCG	GCATTCCACC
Gibbulinella dewinteri	CCGGGCGA--	GCCGATTGG-	-----CAG	GGTTGGTGGC	GAATCTGTCT	GCATTCCACC
Gonaxis quadrilateralis	CCGGGCGA--	GCCTGTCTGG-	-----CAG	GGTTGGTGGC	GAATCTGTCT	GCATTCCACC
Gonospora sp.	CCGGGCGA--	GCCGATTGG-	-----CAG	GGTTGGTGGC	GAATCTGTCT	GCATTCCACC
NUCLEOTIDES INCLUDED	mmmmmmmm--	mm-----	-----mm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	1630	1640	1650	1660	1670	1680
Achatina fulica	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Achatina achatina	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Achatina stuhlmanni	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Cochlitoma ustulata	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Allopeas clavulinum	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Bocageia sp.	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Eutomopeas layardi	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Leptinaria lamellata	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Paropeas achatinaceum	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Riebeckia sp.	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Rumina decollata	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Subulina octona	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Subulina striatella	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Subulina vitrea	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Subulona sp.	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Tortaxis erectus	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Xerocerastus sp.	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Zootecus insularis	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Coelioxys blandii	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Pyrgina umbilicata	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Glessula ceylanica	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Cecilioides gokweanus	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Ferussacia folliculus	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Thyrophorella thomensis	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Gibbulinella dewinteri	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Gonaxis quadrilateralis	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
Gonospora sp.	CGACCCGTCT	TGAAACACGG	-ACCAAGGAG	TCTAACATGC	GCGCGAGTCA	TTGGGCGGTA
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	-mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	1690	1700	1710	1720	1730	1740
Achatina fulica	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGCGC	GACCCAGGTG
Achatina achatina	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGCGC	GACCCAGGTG
Achatina stuhlmanni	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGCGC	GACCCAGGTG
Cochlitoma ustulata	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGCGC	GACCCAGGTG
Allopeas clavulinum	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTTT---	-----CGGC	GACCCAGGTG
Bocageia sp.	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGGC	GACCCAGGTG
Eutomepeas layardi	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTTT---	-----CGGC	GACCCAGGTG
Leptinaria lamellata	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGGC	GACCCAGGTG
Paropeas achatinaceum	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTTT---	-----CGGC	GACCCAGGTG
Riebeckia sp.	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCTT--	-----AGCGC	GACCCAGGTG
Rumina decollata	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CNCGC	GACCCAGGTG
Subulina octona	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GCCT---	-----CGCGC	GGCCCAGGTG
Subulina striatella	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GCCT---	-----TGCGC	GGCCCAGGTG
Subulina vitrea	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTTT---	-----CGGC	GGCCCAGGTG
Subulona sp.	CGAAACTCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTTT---	-----CGGC	GACCCAGGTG
Tortaxis erectus	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGCGC	GACCCAGGTG
Xerocerastus sp.	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCTC--	-----GCGC	GGCCCAGGTG
Zootecus insularis	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGCGC	GACCCAGGTG
Coeliaxis blandii	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCGTTC	GTTTCGCGC	GACCCAGGTG
Pyrquina umbilicata	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTTT---	-----CGGC	GACCCAGGTG
Glessula ceylanica	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTTT---	-----CGCGC	GACCTAGGTG
Ceciloides gokweanus	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTCT---	-----CGCGC	GACCCAGGTG
Ferussacia folliculus	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GCC----	-----TGCGC	GACCCAGGTG
Thyrophorella thomensis	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-GTTT---	-----CGGC	GACCCAGGTG
Gibbulinella dewinteri	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TCCTCTC--	-----GCGG	GACCCAGGTG
Gonaxis quadrilateralis	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TCCTCTC--	-----GCGGA	GACCCAGGTG
Gonospora sp.	CGAAACCCAA	AGGCGCAGTG	AAAGCGAGGG	TC-TCCTC--	-----GCGGA	GACCCAGGTG
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mm-----	-----nn	nnnnnnnnnn

	1750	1760	1770	1780	1790	1800
Achatina fulica	GGATCCT-CC	GCTCCT----	-----TC	G---CGGGAG	CGGT-GGCGC	
Achatina achatina	GGATCCT-CC	GCTCCT----	-----TC	G---CGGGAG	CGGT-GGCGC	
Achatina stuhlmanni	GGATCCT-CC	GCTCCT----	-----TC	G---CGGGAG	CGGT-GGCGC	
Cochlitoma ustulata	GGATCCT-CC	GCTCCT----	-----TC	G---CGGGAG	CGGT-GGCGC	
Allopeas clavulinum	GGATCCT-CC	CTACCC--G	CGTTATCGCG	CGGGT-----	-----GCGT	GGGT-GGCGC
Bocageia sp.	GGATCCA-TC	CTCCGCT--	-----	-----CGCGC	GGGT-GGCGC	
Eutomepeas layardi	GGATCCT-CC	CTACCC--G	CGTTATCGCG	CGGGT-----	-----GCGT	GGGT-GGCGC
Leptinaria lamellata	GGATCCT-CC	CTACCC--G	CGTTTCCGCG	CGGGT-----	-----TCTTG	GGGT-GGCGC
Paropeas achatinaceum	GGATCCT-CC	CTACCC--G	CGTTATCGCG	CGGGT-----	-----GCGT	GGGT-GGCGC
Riebeckia sp.	GGATCCT-CC	TCCTCCTCCT	CCTTCCCCTC	--GCGGGGGT	G---GTGGTG	GGGT-GGCGC
Rumina decollata	GGATCCT-CC	ACCCCTCTCG	-----	-----GGGGG	CGGT-GGCGC	
Subulina octona	GGATCCT-CC	ACTTC-----	-----	-----TGG	TGGT-GGCGC	
Subulina striatella	GGATCCT-CC	ACTTC-----	-----	-----TGG	TGGT-GGCGC	
Subulina vitrea	GGATCCATCC	CTCCGCC---	----GCGAGC	GCAAGCCCGC	G---CGTCGG	GGGT-GGCGC
Subulona sp.	GGATCCATCC	CTCCTGTCTG	CGCGAGCGCA	AGCCCGCGTC	GGCGGTCTGG	GGGT-GGCGC
Tortaxis erectus	GGATCCT-CC	GCTCTCTC--	-----	-----GGGAG	CGGT-GGCGC	
Xerocerastus sp.	GGATCCT-CC	ACCCCTCT--	-----	-----CGGGGG	CGGT-GGCGC	
Zootecus insularis	GGATCCT-CC	ACTCCTC--	-----	-----TCGGGGG	CGGT-GGCGC	
Coeliaxis blandii	GGATCCT-CC	T-----	-----	-----GAT	AGGA-GGCGC	
Pyrquina umbilicata	GGATCCT-CC	CTACCC--G	CGTTCTCGCG	CGGGT-----	-----GCGT	GGGT-GGCGC
Glessula ceylanica	GGATCCT-CC	GCTCTC----	-----	-----TCGGGAG	CGGT-GGCGC	
Ceciloides gokweanus	GGATCCT-CC	CTGCCG--	CCCCTTTACG	CGGGT-----	-----TCGTCGT	GGGT-GGCGC
Ferussacia folliculus	GGATCCT-CT	CCCGTCC---	-----	-----GAGG	GGGA-GGCGC	
Thyrophorella thomensis	GGATCCT-CC	CTACCC--G	CGTTCTCGCG	CGGGT-----	-----GCGT	GGGT-GGCGC
Gibbulinella dewinteri	GGATCCTTCC	TCTCCCACGC	CTTCT-GGCG	CG-----	-----GGGGG	GAGTTGGCGC
Gonaxis quadrilateralis	GGATCCT-TC	CCTCCCCGTC	ACATTTGGCG	C-----	-----GGG	GGGCTGGCGC
Gonospora sp.	GGATCCT-GT	CCTTCGCGCC	ACATTTGGCG	CG-----	-----GGTG	AGGCTGGCGC
NUCLEOTIDES INCLUDED	mmmmmm--	-----	-----	-----	-----mm	mmmm-mmmmm

	1810	1820	1830	1840	1850	1860	
Achatina fulica	ACCACCGGCC	CGTCCCGTCC	GCGTCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Achatina achatina	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Achatina stuhlmanni	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Cochlitoma ustulata	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Allopeas clavulinum	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Bocageia sp.	ACCACCGGCC	CGTCCCATCC	GCGCCGCGCG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Eutomopeas layardi	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Leptinaria lamellata	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Paropeas achatinaceum	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Riebeckia sp.	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Rumina decollata	ACCACCGGCC	CGTCCCGTCC	GCGYCYCYCY	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Subulina octona	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Subulina striatella	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Subulina vitrea	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Subulona sp.	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Tortaxis erectus	ACCACCGGCC	CGTCCCGTCC	GCGCTGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Xerocerastus sp.	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Zootecus insularis	ACCACCGGCC	CGTCCCGTCC	GCGCTGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Coeliaxis blandii	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Pyrgina umbilicata	ACCACCGGCC	CGTCCCGTCC	GCGCCGCGCG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Glessula ceylanica	ACCACCGGCC	CGTCCCGTCC	GCGTCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Cecilioides gokweanus	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Ferussacia folliculus	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Thyrophorella thomensis	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Gibbulinella dewinteri	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Gonaxis quadrilateralis	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
Gonospira sp.	ACCACCGGCC	CGTCCCGTCC	GCGTCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1870	1880	1890	1900	1910	1920	
Achatina fulica	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Achatina achatina	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Achatina stuhlmanni	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Cochlitoma ustulata	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Allopeas clavulinum	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Bocageia sp.	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Eutomopeas layardi	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Leptinaria lamellata	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Paropeas achatinaceum	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Riebeckia sp.	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Rumina decollata	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Subulina octona	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Subulina striatella	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Subulina vitrea	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Subulona sp.	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Tortaxis erectus	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Xerocerastus sp.	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Zootecus insularis	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Coeliaxis blandii	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Pyrgina umbilicata	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Glessula ceylanica	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Cecilioides gokweanus	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Ferussacia folliculus	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Thyrophorella thomensis	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Gibbulinella dewinteri	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Gonaxis quadrilateralis	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
Gonospira sp.	CCCAGAAAGAT	GGTGAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGGTGGAGGT	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1930	1940	1950	1960	1970	1980
Achatina fulica	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Achatina achatina	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Achatina stuhlmanni	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Cochlitoma ustulata	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Allopeas clavulinum	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Bocageia sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Eutomepeas layardi	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Leptinaria lamellata	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Paropeas achatinaceum	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Riebeckia sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Rumina decollata	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Subulina octona	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Subulina striatella	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Subulina vitrea	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Subulona sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Tortaxis erectus	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Xerocerastus sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Zootecus insularis	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Coeliaxis blandii	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Pyrquina umbilicata	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Glessula ceylanica	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Ceciloides gokweanus	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Ferussacia folliculus	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Thyrophorella thomensis	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Gibbulinella dewinteri	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Gonaxis quadrilateralis	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
Gonospora sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTTGG	GTATAGGGGC	GAAAGACTAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1990	2000	2010	2020	2030	2040
Achatina fulica	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Achatina achatina	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Achatina stuhlmanni	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Cochlitoma ustulata	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Allopeas clavulinum	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Bocageia sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Eutomepeas layardi	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Leptinaria lamellata	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Paropeas achatinaceum	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Riebeckia sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Rumina decollata	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Subulina octona	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Subulina striatella	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Subulina vitrea	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Subulona sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Tortaxis erectus	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Xerocerastus sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Zootecus insularis	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Coeliaxis blandii	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Pyrquina umbilicata	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Glessula ceylanica	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Ceciloides gokweanus	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Ferussacia folliculus	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Thyrophorella thomensis	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Gibbulinella dewinteri	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Gonaxis quadrilateralis	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Gonospora sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	2170	2180	2190	2200	2210	2220			
Achatina fulica	-NTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Achatina achatina	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Achatina stuhlmanni	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Cochlitoma ustulata	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Allopeas clavulinum	GTTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Bocageia sp.	--TCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Eutomepeas layardi	GTTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Leptinaria lamellata	GTTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Paropeas achatinaceum	GTTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Riebeckia sp.	-GTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Rumina decollata	-NTCGAATGC	NTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Subulina octona	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Subulina striatella	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Subulina vitrea	TGTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Subulona sp.	GTTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Tortaxis erectus	GTACGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Xerocerastus sp.	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Zootecus insularis	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Coeliaxis blandii	--TCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Pyrquina umbilicata	GTTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Glessula ceylanica	GTCCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Ceciloides gokweanus	TGTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Ferussacia folliculus	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Thyrophorella thomensis	GTTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Gibbulinella dewinteri	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Gonaxis quadrilateralis	-TTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
Gonospora sp.	-CTCGAATGC	GTGTGCCAAG	TGGGCCACTT	TTGGTAAGCA	GAAGTGGCGC	TGTGGGATGA			
NUCLEOTIDES INCLUDED	---mmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm			

	2230	2240	2250	2260	2270	2280			
Achatina fulica	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Achatina achatina	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Achatina stuhlmanni	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Cochlitoma ustulata	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Allopeas clavulinum	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Bocageia sp.	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Eutomepeas layardi	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Leptinaria lamellata	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Paropeas achatinaceum	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Riebeckia sp.	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Rumina decollata	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Subulina octona	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Subulina striatella	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Subulina vitrea	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Subulona sp.	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Tortaxis erectus	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Xerocerastus sp.	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Zootecus insularis	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Coeliaxis blandii	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Pyrquina umbilicata	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Glessula ceylanica	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Ceciloides gokweanus	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Ferussacia folliculus	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Thyrophorella thomensis	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Gibbulinella dewinteri	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Gonaxis quadrilateralis	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
Gonospora sp.	ACCAAACGCC	CGGTTAAGGT	GCCAAACGCT	GACGCTCATC	AGACACCATA	AAAGGTGTTG			
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm			

	2290 2300 2310 2320 2330 2340
Achatina fulica	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Achatina achatina	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Achatina stuhlmanni	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Cochlitoma ustulata	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Allopeas clavulinum	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Bocageia sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Eutomopeas layardi	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Leptinaria lamellata	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Paropeas achatinaceum	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Riebeckia sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Rumina decollata	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Subulina octona	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Subulina striatella	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Subulina vitrea	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Subulona sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Tortaxis erectus	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Xerocerastus sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Zootecus insularis	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Coeliaxis blandii	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Pyrgina umbilicata	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Glessula ceylanica	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Cecilioides gokweanus	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Ferussacia folliculus	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Thyrophorella thomensis	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Gibbulinella dewinteri	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Gonaxis quadrilateralis	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
Gonospira sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
NUCLEOTIDES INCLUDED	mmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	2350 2360 2370 2380 2390 2400
Achatina fulica	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Achatina achatina	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Achatina stuhlmanni	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Cochlitoma ustulata	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Allopeas clavulinum	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Bocageia sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Eutomopeas layardi	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Leptinaria lamellata	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Paropeas achatinaceum	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Riebeckia sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Rumina decollata	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Subulina octona	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Subulina striatella	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Subulina vitrea	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Subulona sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Tortaxis erectus	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Xerocerastus sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Zootecus insularis	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Coeliaxis blandii	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Pyrgina umbilicata	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Glessula ceylanica	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Cecilioides gokweanus	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Ferussacia folliculus	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Thyrophorella thomensis	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Gibbulinella dewinteri	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Gonaxis quadrilateralis	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
Gonospira sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
NUCLEOTIDES INCLUDED	mmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	2530	2540	2550	2560	2570	2580	
Achatina fulica	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Achatina achatina	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Achatina stuhlmanni	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Cochlitoma ustulata	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Allopeas clavulinum	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Bocageia sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Eutomopeas layardi	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Leptinaria lamellata	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Paropeas achatinaceum	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Riebeckia sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Rumina decollata	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Subulina octona	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Subulina striatella	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Subulina vitrea	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Subulona sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Tortaxis erectus	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Xerocerastus sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Zootecus insularis	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Coeliaxis blandii	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Pyrgina umbilicata	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Glessula ceylanica	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Cecilioides gokweanus	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Ferussacia folliculus	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Thyrophorella thomensis	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Gibbulinella dewinteri	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Gonaxis quadrilateralis	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
Gonospira sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2590	2600	2610	2620	2630	2640	
Achatina fulica	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Achatina achatina	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Achatina stuhlmanni	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Cochlitoma ustulata	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Allopeas clavulinum	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Bocageia sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Eutomopeas layardi	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Leptinaria lamellata	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Paropeas achatinaceum	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Riebeckia sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Rumina decollata	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Subulina octona	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Subulina striatella	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Subulina vitrea	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Subulona sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Tortaxis erectus	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Xerocerastus sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Zootecus insularis	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Coeliaxis blandii	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Pyrgina umbilicata	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Glessula ceylanica	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Cecilioides gokweanus	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Ferussacia folliculus	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Thyrophorella thomensis	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Gibbulinella dewinteri	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Gonaxis quadrilateralis	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
Gonospira sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2650 2660 2670 2680 2690 2700
Achatina fulica	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT CTTTCT----- ----AGAAGA
Achatina achatina	CCGTTCTGAC CCCGGGGC-A TTCTTTGTCA GCAGT----- ----CAA
Achatina stuhlmanni	CCGTTCTGAC CCCGGGGC-A TTCTTTATAA ---TTCGTTA TT----- ----AGA
Cochlitoma ustulata	CCGTTCTGAC CCCGGGGC-A TTCTTTCTGTG ----- ----ATTAAA
Allopeas clavulinum	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT CTTTCTTCT- ----CGAA GCCAGAAGTA
Bocageia sp.	CCGTTCTGAC CCCGGGGC-A TTCTTTACTA ACA----- ----AAA AGGAAGAAAA
Eutomepeas layardi	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT CTTCTC----- --GAAGCAGA
Leptinaria lamellata	CCGTTCTGAC CCCGGGGC-A TTCTTTCTG- ----- --AATGAGTA
Paropeas achatinaceum	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTA CTTCTTCTTC T--CGCGAAC GAAGAGCAGA
Riebeckia sp.	CCGTTCTGAC CCCGGGGCAA TTCTTTCTTT CTTTCT----- ----CGAAC GAACGATCGA
Rumina decollata	CCGTTCTGAC CCCGGGGC-A ATCTTTCTCG AGCTA----- --AGA AGAAAAAGAA
Subulina octona	CCGTTCTGAC CCCGGGGC-A ATCTTTTCTT C----- ----AAGA
Subulina striatella	CCGTTCTGAC CCCGGGGC-A ATCTTCATTT TT----- ----GA
Subulina vitrea	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT AATCTTTT-- ----GCAGA
Subulona sp.	CCGTTCTGAC CCCGGGGC-G TTTTTTTGTA ATAATCTT-- ----A
Tortaxis erectus	CCGTTCTGAC CCCGGGGC-A TTCTTTCTC- ----- --AGCGAATA
Xerocerastus sp.	CCGTTCTGAC CCCGGGGC-A CTCTTT----- --CTTGACG ATTCAATTGA
Zootecus insularis	CCGTTCTGAC CCCGGGGC-A ATCTTTCTTT CTTTCTC----- --G AGAAGAAGAA
Coeliaxis blandii	CCGTTCTGAC CCCGGGGCAA TTCTTT----- ----CGTTGA
Pyrquina umbilicata	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT CACT----- ----CGA AGAAGCAAGA
Glessula ceylanica	CCGTTCTGAC CCCGGGGC-A TTCTTTCTAG- ----- ----AAAA
Ceciloides gokweanus	CCGTTCTGAC CCCGGGGCAA TTCTTTCTTT CTGTTT----- ----GA
Ferussacia folliculus	CCGTTCTGAC CCCGGGGC-A CTCTTT----- ----ACGT CGGTAAACGA
Thyrophorella thomensis	CCGTTCTGAC CCCGGGGC-A TTCTTTCTT- ----- ----ACTCGAAGCA
Gibbulinella dewinteri	CCGTTCTGAC CCCGGGGC-G CTCTTTACG- ----- --C AACGTTTCGA
Gonaxis quadrilateralis	CCGTTCTGAC CCCGGGGC-G CTCTTTTCTT- ----- --GTTGATCGA
Gonospora sp.	CCGTTCTGAC CCCGGGGC-G ATCTTTTACA- ----- --CAAC AACACGCGAG TTGTCGTCGA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmm-- ----- ----m

	2710 2720 2730 2740 2750 2760
Achatina fulica	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Achatina achatina	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Achatina stuhlmanni	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Cochlitoma ustulata	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Allopeas clavulinum	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Bocageia sp.	GTTTGGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Eutomepeas layardi	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Leptinaria lamellata	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Paropeas achatinaceum	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Riebeckia sp.	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Rumina decollata	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Subulina octona	GTTTGGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Subulina striatella	GTTTGGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Subulina vitrea	G-TTTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Subulona sp.	ACACTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Tortaxis erectus	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Xerocerastus sp.	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Zootecus insularis	G-TAAGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Coeliaxis blandii	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Pyrquina umbilicata	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Glessula ceylanica	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Ceciloides gokweanus	A-TCAGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Ferussacia folliculus	GCTCGGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Thyrophorella thomensis	GNGTCGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Gibbulinella dewinteri	G-TACGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Gonaxis quadrilateralis	G-CACGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
Gonospora sp.	G-CACGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
NUCLEOTIDES INCLUDED	m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2770	2780	2790	2800	2810	2820	
Achatina fulica	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Achatina achatina	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Achatina stuhlmanni	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Cochlitoma ustulata	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Allopeas clavulinum	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Bocageia sp.	ATTGGTCCTC	TGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGAGAGCC	
Eutomopeas layardi	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Leptinaria lamellata	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Paropeas achatinaceum	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Riebeckia sp.	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Rumina decollata	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Subulina octona	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Subulina striatella	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Subulina vitrea	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Subulona sp.	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Tortaxis erectus	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGAAGCC	
Xerocerastus sp.	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Zootecus insularis	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Coelioxaxis blandii	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Pyrgina umbilicata	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Glessula ceylanica	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGAAGCC	
Ceciloides gokweanus	ATTGGTCCTC	TGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Ferussacia folliculus	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Thyrophorella thomensis	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Gibbulinella dewinteri	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCT	
Gonaxis quadrilateralis	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Gonospira sp.	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2830	2840	2850	2860	2870	2880	
Achatina fulica	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Achatina achatina	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Achatina stuhlmanni	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Cochlitoma ustulata	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Allopeas clavulinum	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Bocageia sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Eutomopeas layardi	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Leptinaria lamellata	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Paropeas achatinaceum	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Riebeckia sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Rumina decollata	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Subulina octona	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Subulina striatella	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Subulina vitrea	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Subulona sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Tortaxis erectus	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Xerocerastus sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Zootecus insularis	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Coelioxaxis blandii	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Pyrgina umbilicata	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Glessula ceylanica	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Ceciloides gokweanus	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Ferussacia folliculus	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Thyrophorella thomensis	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Gibbulinella dewinteri	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Gonaxis quadrilateralis	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Gonospira sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3010 3020 3030 3040 3050 3060
Achatina fulica	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Achatina achatina	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Achatina stuhlmanni	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Cochlitoma ustulata	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Allopeas clavulinum	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Bocageia sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Eutomopeas layardi	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Leptinaria lamellata	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Paropeas achatinaceum	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Riebeckia sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Rumina decollata	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Subulina octona	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Subulina striatella	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Subulina vitrea	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Subulona sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Tortaxis erectus	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Xerocerastus sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Zootecus insularis	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Coeliaxis blandii	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Pyrgina umbilicata	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Glessula ceylanica	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Cecilioides gokweanus	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Ferussacia folliculus	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Thyrophorella thomensis	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Gibbulinella dewinteri	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Gonaxis quadrilateralis	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
Gonospira sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTAGGTAAG GGAAGTCGGC
NUCLEOTIDES INCLUDED	mmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	3070 3080 3090 3100 3110 3120
Achatina fulica	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Achatina achatina	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Achatina stuhlmanni	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Cochlitoma ustulata	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Allopeas clavulinum	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Bocageia sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Eutomopeas layardi	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Leptinaria lamellata	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Paropeas achatinaceum	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Riebeckia sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Rumina decollata	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Subulina octona	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Subulina striatella	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Subulina vitrea	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Subulona sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Tortaxis erectus	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Xerocerastus sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Zootecus insularis	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Coeliaxis blandii	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Pyrgina umbilicata	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Glessula ceylanica	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Cecilioides gokweanus	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Ferussacia folliculus	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Thyrophorella thomensis	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Gibbulinella dewinteri	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Gonaxis quadrilateralis	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
Gonospira sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCCG
NUCLEOTIDES INCLUDED	mmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	3130	3140	3150	3160	3170	3180		
Achatina fulica	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Achatina achatina	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Achatina stuhlmanni	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Cochlitoma ustulata	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Allopeas clavulinum	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Bocageia sp.	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
Eutomoepas layardi	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Leptinaria lamellata	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Paropeas achatinaceum	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Riebeckia sp.	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Rumina decollata	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
Subulina octona	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Subulina striatella	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Subulina vitrea	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Subulona sp.	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Tortaxis erectus	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
Xerocerastus sp.	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
Zootecus insularis	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
Coeliaxis blandii	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Pyrquina umbilicata	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Glessula ceylanica	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
Ceciloides gokweanus	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Ferussacia folliculus	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TCTCTGGCGG		
Thyrophorella thomensis	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGCCGGCGG		
Gibbulinella dewinteri	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
Gonaxis quadrilateralis	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
Gonospora sp.	GGTACGAAGC	GGGACTGGGA	TGGGCCCGGG	CTGGGCGAGG	CCGCCGCCGC	TAGTCGGCGG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	3190	3200	3210	3220	3230	3240		
Achatina fulica	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Achatina achatina	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Achatina stuhlmanni	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Cochlitoma ustulata	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Allopeas clavulinum	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Bocageia sp.	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Eutomoepas layardi	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Leptinaria lamellata	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Paropeas achatinaceum	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Riebeckia sp.	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Rumina decollata	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Subulina octona	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Subulina striatella	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Subulina vitrea	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Subulona sp.	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Tortaxis erectus	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Xerocerastus sp.	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Zootecus insularis	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Coeliaxis blandii	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Pyrquina umbilicata	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Glessula ceylanica	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Ceciloides gokweanus	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Ferussacia folliculus	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Thyrophorella thomensis	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Gibbulinella dewinteri	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Gonaxis quadrilateralis	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
Gonospora sp.	GCCGGCCGAG	CTCGGAACGC	GGCTGCAACC	TTCCCGTGGA	CCGCCCCAGC	TATGCGGCGG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	3250 3260 3270 3280 3290 3300
Achatina fulica	CGCCT----- -CCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Achatina achatina	CGCCT----- -CCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Achatina stuhlmanni	CGCCT----- -CCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Cochlitoma ustulata	CGCCT----- -CCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Allopeas clavulinum	CGCCT----- TCCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Bocageia sp.	CGCCT----- -CCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Eutomopeas layardi	CGCCT----- TCCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Leptinaria lamellata	CGCCT----- TCCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Paropeas achatinaceum	CGCCT----- TCCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Riebeckia sp.	CGCCT----- -CCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Rumina decollata	CGCCT----- -CCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Subulina octona	CGCCT----- -CCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Subulina striatella	CGCCT----- -CCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Subulina vitrea	CGCCT----- TCCCCGGCGT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Subulona sp.	CGCCT----- TCACGCGCT CGTTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Tortaxis erectus	CGCCT----- TCCCCAGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Xerocerastus sp.	CGCCT----- -CCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Zootecus insularis	CGCCT----- -CCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Coelioxys blandii	CGCCT----- -CCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Pyrgina umbilicata	CGCCT----- TCCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Glessula ceylanica	CACCG----- TCCCCGGTGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Cecilioides gokweanus	CGCCT----- TCCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Ferussacia folliculus	CGCCTCCTCC CGCGGGGTGW CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Thyrophorella thomensis	CGCCT----- TCCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Gibbulinella dewinteri	CGCCT----- -CCCCGGCGT TGTCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Gonaxis quadrilateralis	CGCCT----- -CCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
Gonospora sp.	CGCCT----- -CCCCGGCGT CGTCCGCGTC GGCTGGCATT CAACAGCCAA CTCAGAAGCTG
NUCLEOTIDES INCLUDED	mmmmmm----- --mmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	3310 3320 3330 3340 3350 3360
Achatina fulica	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Achatina achatina	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Achatina stuhlmanni	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Cochlitoma ustulata	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Allopeas clavulinum	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Bocageia sp.	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Eutomopeas layardi	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Leptinaria lamellata	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Paropeas achatinaceum	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Riebeckia sp.	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Rumina decollata	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Subulina octona	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Subulina striatella	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Subulina vitrea	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Subulona sp.	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Tortaxis erectus	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Xerocerastus sp.	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Zootecus insularis	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Coelioxys blandii	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Pyrgina umbilicata	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Glessula ceylanica	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Cecilioides gokweanus	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Ferussacia folliculus	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Thyrophorella thomensis	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Gibbulinella dewinteri	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Gonaxis quadrilateralis	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
Gonospora sp.	GTACGGACCA GGGGAATCCG ACTGTCTAAT TAAACAAAAG CATTGCGACG GCCGTCACCC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	3370	3380	3390	3400	3410	3420
Achatina fulica	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Achatina achatina	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Achatina stuhlmanni	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Cochlitoma ustulata	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Allopeas clavulinum	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Bocageia sp.	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Eutomepeas layardi	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Leptinaria lamellata	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Paropeas achatinaceum	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Riebeckia sp.	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Rumina decollata	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Subulina octona	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Subulina striatella	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Subulina vitrea	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Subulona sp.	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Tortaxis erectus	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Xerocerastus sp.	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Zootecus insularis	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Coeliaxis blandii	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Pyrquina umbilicata	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Glessula ceylanica	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Ceciloides gokweanus	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Ferussacia folliculus	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Thyrophorella thomensis	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Gibbulinella dewinteri	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Gonaxis quadrilateralis	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
Gonospora sp.	GGTGTGACG	CAATGTGATT	TCTGCCCAGT	GCTCTGAATG	TCAAAGTGAA	GAAATTCAAC
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	3430	3440	3450	3460	3470	3480
Achatina fulica	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Achatina achatina	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Achatina stuhlmanni	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Cochlitoma ustulata	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Allopeas clavulinum	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Bocageia sp.	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Eutomepeas layardi	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Leptinaria lamellata	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Paropeas achatinaceum	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Riebeckia sp.	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Rumina decollata	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Subulina octona	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Subulina striatella	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Subulina vitrea	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Subulona sp.	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Tortaxis erectus	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Xerocerastus sp.	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Zootecus insularis	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Coeliaxis blandii	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Pyrquina umbilicata	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Glessula ceylanica	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Ceciloides gokweanus	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Ferussacia folliculus	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Thyrophorella thomensis	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Gibbulinella dewinteri	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Gonaxis quadrilateralis	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
Gonospora sp.	CAAGCGCGGG	TAAACGGCGG	GAGTAACTAT	GACTCTCTTA	AGGTAGCCAA	ATGCCTCGTC
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	3490 3500 3510 3520 3530 3540
Achatina fulica	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Achatina achatina	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Achatina stuhlmanni	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Cochlitoma ustulata	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Allopeas clavulinum	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Bocageia sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Eutomopeas layardi	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Leptinaria lamellata	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Paropeas achatinaceum	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Riebeckia sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Rumina decollata	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Subulina octona	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Subulina striatella	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Subulina vitrea	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Subulona sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Tortaxis erectus	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Xerocerastus sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Zootecus insularis	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Coeliaxis blandii	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Pyrgina umbilicata	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Glessula ceylanica	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Cecilioides gokweanus	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Ferussacia folliculus	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Thyrophorella thomensis	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Gibbulinella dewinteri	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Gonaxis quadrilateralis	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
Gonospira sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3550 3560 3570 3580 3590 3600
Achatina fulica	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Achatina achatina	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Achatina stuhlmanni	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Cochlitoma ustulata	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Allopeas clavulinum	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Bocageia sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Eutomopeas layardi	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Leptinaria lamellata	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Paropeas achatinaceum	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Riebeckia sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Rumina decollata	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Subulina octona	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Subulina striatella	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Subulina vitrea	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Subulona sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Tortaxis erectus	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Xerocerastus sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Zootecus insularis	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Coeliaxis blandii	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Pyrgina umbilicata	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Glessula ceylanica	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Cecilioides gokweanus	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Ferussacia folliculus	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Thyrophorella thomensis	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Gibbulinella dewinteri	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Gonaxis quadrilateralis	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
Gonospira sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3610	3620	3630	3640	3650	3660		
Achatina fulica	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Achatina achatina	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Achatina stuhlmanni	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Cochlitoma ustulata	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Allopeas clavulinum	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Bocageia sp.	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Eutomopeas layardi	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Leptinaria lamellata	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Paropeas achatinaceum	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Riebeckia sp.	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Rumina decollata	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Subulina octona	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Subulina striatella	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Subulina vitrea	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Subulona sp.	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Tortaxis erectus	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Xerocerastus sp.	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Zootecus insularis	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Coeliaxis blandii	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Pyrquina umbilicata	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Glessula ceylanica	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Ceciloides gokweanus	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Ferussacia folliculus	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Thyrophorella thomensis	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Gibbulinella dewinteri	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Gonaxis quadrilateralis	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
Gonospora sp.	TGAGCTTGAC	TCTAGTCCGA	CTTTGTGAAG	ACACATGAAG	GGTGTAGCAT	AGGTGGGAGC		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	3670	3680	3690	3700	3710	3720		
Achatina fulica	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Achatina achatina	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Achatina stuhlmanni	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Cochlitoma ustulata	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Allopeas clavulinum	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Bocageia sp.	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Eutomopeas layardi	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Leptinaria lamellata	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Paropeas achatinaceum	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Riebeckia sp.	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Rumina decollata	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Subulina octona	GCGAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Subulina striatella	GCGAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Subulina vitrea	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Subulona sp.	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Tortaxis erectus	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Xerocerastus sp.	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Zootecus insularis	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Coeliaxis blandii	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Pyrquina umbilicata	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Glessula ceylanica	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Ceciloides gokweanus	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Ferussacia folliculus	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Thyrophorella thomensis	GCAAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Gibbulinella dewinteri	GCGAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Gonaxis quadrilateralis	GCGAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
Gonospora sp.	GCGAGCGCAA	TTGAAATACC	ACTACTTTTA	TCGTTTCCTT	ACTTATTCAG	TCAAGCGGAG		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	3730	3740	3750	3760	3770	3780	
Achatina fulica	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Achatina achatina	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Achatina stuhlmanni	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Cochlitoma ustulata	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	TCGGCGCTCG	TCGTCGGCCG	
Allopeas clavulinum	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Bocageia sp.	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Eutomopeas layardi	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Leptinaria lamellata	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Paropeas achatinaceum	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Riebeckia sp.	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Rumina decollata	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAAT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Subulina octona	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Subulina striatella	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Subulina vitrea	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Subulona sp.	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	TCGGCGCTCG	TCGTCGGCCG	
Tortaxis erectus	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Xerocerastus sp.	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCA-	CCGGCGCTCG	TCGTCGGCCG	
Zootecus insularis	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Coeliaxis blandii	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Pyrgina umbilicata	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Glessula ceylanica	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCTCG	TCGTCGGCCG	
Cecilioides gokweanus	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	TCGGCGCTCG	TCGTCGGCCG	
Ferussacia folliculus	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAC	TCGGCGCTCG	TCGTCGGCCG	
Thyrophorella thomensis	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	CCGGCGCCCG	TCGTCGGCCG	
Gibbulinella dewinteri	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	TCGGCGCTCG	TCGTCGGCCG	
Gonaxis quadrilateralis	AGCGGGGCGA	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	TCGGCGCTCG	TCGTCGGCCG	
Gonospira sp.	AGCGGGGCGC	AAGCCCTCG	CTTCTGGAGT	TAAGCGGCAA	TCGGCGCTCG	TCGTCGGCCG	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	3790	3800	3810	3820	3830	3840	
Achatina fulica	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Achatina achatina	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Achatina stuhlmanni	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Cochlitoma ustulata	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Allopeas clavulinum	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Bocageia sp.	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Eutomopeas layardi	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Leptinaria lamellata	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Paropeas achatinaceum	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Riebeckia sp.	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Rumina decollata	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Subulina octona	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Subulina striatella	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Subulina vitrea	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Subulona sp.	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Tortaxis erectus	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Xerocerastus sp.	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Zootecus insularis	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Coeliaxis blandii	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Pyrgina umbilicata	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Glessula ceylanica	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Cecilioides gokweanus	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Ferussacia folliculus	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Thyrophorella thomensis	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Gibbulinella dewinteri	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Gonaxis quadrilateralis	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
Gonospira sp.	CGATCCGCTC	TGAAGACAGT	GTCAGGCGGG	GAGTTTGACT	GGGGCGGTAC	ATCTGTCAAA	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	3850	3860	3870	3880	3890	3900			
Achatina fulica	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Achatina achatina	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Achatina stuhlmanni	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Cochlitoma ustulata	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Allopeas clavulinum	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Bocageia sp.	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Eutomopeas layardi	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Leptinaria lamellata	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Paropeas achatinaceum	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Riebeckia sp.	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Rumina decollata	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Subulina octona	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Subulina striatella	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Subulina vitrea	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Subulona sp.	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Tortaxis erectus	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Xerocerastus sp.	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Zootecus insularis	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Coeliaxis blandii	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAAAGCAAAA			
Pyrquina umbilicata	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Glessula ceylanica	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Ceciloides gokweanus	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Ferussacia folliculus	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Thyrophorella thomensis	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Gibbulinella dewinteri	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Gonaxis quadrilateralis	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
Gonospora sp.	AGGTAACGCA	GGTGTCTCTAA	GGCGAGCTCA	GCGAGGACGG	AAACCTCGCG	TAGAGCAAAA			
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm			

	3910	3920	3930	3940	3950	3960			
Achatina fulica	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Achatina achatina	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Achatina stuhlmanni	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Cochlitoma ustulata	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Allopeas clavulinum	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Bocageia sp.	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Eutomopeas layardi	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Leptinaria lamellata	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Paropeas achatinaceum	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Riebeckia sp.	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Rumina decollata	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Subulina octona	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Subulina striatella	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Subulina vitrea	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Subulona sp.	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Tortaxis erectus	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Xerocerastus sp.	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Zootecus insularis	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Coeliaxis blandii	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Pyrquina umbilicata	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Glessula ceylanica	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Ceciloides gokweanus	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Ferussacia folliculus	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Thyrophorella thomensis	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Gibbulinella dewinteri	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Gonaxis quadrilateralis	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
Gonospora sp.	GGGCAAAAGC	TCGCTTGATT	TTGATTTTCA	GTACGAATAC	AGACCGTGAA	AGCGTGGCCT			
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm			

	3970 3980 3990 4000 4010 4020
Achatina fulica	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Achatina achatina	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Achatina stuhlmanni	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Cochlitoma ustulata	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Allopeas clavulinum	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Bocageia sp.	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Eutomopeas layardi	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Leptinaria lamellata	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Paropeas achatinaceum	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Riebeckia sp.	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Rumina decollata	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Subulina octona	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Subulina striatella	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Subulina vitrea	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Subulona sp.	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Tortaxis erectus	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Xerocerastus sp.	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Zootecus insularis	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Coeliaxis blandii	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Pyrgina umbilicata	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Glessula ceylanica	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Cecilioides gokweanus	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Ferussacia folliculus	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Thyrophorella thomensis	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Gibbulinella dewinteri	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Gonaxis quadrilateralis	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
Gonospira sp.	ATCGATCCTT TTGACTTTAA GAGTTTAAAG CAAGAGGTGT CAGAAAAAGTT ACCACAGGGA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	4030 4040 4050 4060 4070 4080
Achatina fulica	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Achatina achatina	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Achatina stuhlmanni	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Cochlitoma ustulata	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Allopeas clavulinum	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Bocageia sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Eutomopeas layardi	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Leptinaria lamellata	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Paropeas achatinaceum	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Riebeckia sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Rumina decollata	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Subulina octona	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Subulina striatella	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Subulina vitrea	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Subulona sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Tortaxis erectus	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Xerocerastus sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Zootecus insularis	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Coeliaxis blandii	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Pyrgina umbilicata	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Glessula ceylanica	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Cecilioides gokweanus	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Ferussacia folliculus	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Thyrophorella thomensis	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Gibbulinella dewinteri	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Gonaxis quadrilateralis	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
Gonospira sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTATCC TTCGATGTCC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	4090	4100	4110	4120	4130	4140		
Achatina fulica	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Achatina achatina	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Achatina stuhlmanni	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Cochlitoma ustulata	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Allopeas clavulinum	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Bocageia sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Eutomopeas layardi	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Leptinaria lamellata	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Paropeas achatinaceum	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Riebeckia sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Rumina decollata	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Subulina octona	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Subulina striatella	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Subulina vitrea	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Subulona sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Tortaxis erectus	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Xerocerastus sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Zootecus insularis	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Coeliaxis blandii	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Pyrgina umbilicata	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Glessula ceylanica	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Ceciloides gokweanus	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Ferussacia folliculus	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Thyrophorella thomensis	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Gibbulinella dewinteri	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Gonaxis quadrilateralis	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
Gonospora sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTTCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	4150	4160	4170	4180	4190	4200		
Achatina fulica	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Achatina achatina	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Achatina stuhlmanni	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Cochlitoma ustulata	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Allopeas clavulinum	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Bocageia sp.	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Eutomopeas layardi	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Leptinaria lamellata	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Paropeas achatinaceum	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Riebeckia sp.	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Rumina decollata	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA		
Subulina octona	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Subulina striatella	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Subulina vitrea	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Subulona sp.	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Tortaxis erectus	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Xerocerastus sp.	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA		
Zootecus insularis	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA		
Coeliaxis blandii	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Pyrgina umbilicata	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Glessula ceylanica	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Ceciloides gokweanus	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Ferussacia folliculus	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA		
Thyrophorella thomensis	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Gibbulinella dewinteri	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA		
Gonaxis quadrilateralis	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
Gonospora sp.	GGAACGTGAG	CTGGGTTTAG	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	4210	4220	4230	4240	4250	4260	
Achatina fulica	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Achatina achatina	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Achatina stuhlmanni	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Cochlitoma ustulata	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Allopeas clavulinum	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Bocageia sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Eutomopeas layardi	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Leptinaria lamellata	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Paropeas achatinaceum	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Riebeckia sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Rumina decollata	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Subulina octona	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Subulina striatella	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Subulina vitrea	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Subulona sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Tortaxis erectus	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Xerocerastus sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Zootecus insularis	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Coeliaxis blandii	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Pyrgina umbilicata	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Glessula ceylanica	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Cecilioides gokweanus	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Ferussacia folliculus	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Thyrophorella thomensis	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Gibbulinella dewinteri	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Gonaxis quadrilateralis	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
Gonospira sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTTC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	4270	4280	4290	4300	4310	
Achatina fulica	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Achatina achatina	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Achatina stuhlmanni	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Cochlitoma ustulata	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Allopeas clavulinum	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Bocageia sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Eutomopeas layardi	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Leptinaria lamellata	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Paropeas achatinaceum	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Riebeckia sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Rumina decollata	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Subulina octona	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Subulina striatella	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Subulina vitrea	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Subulona sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Tortaxis erectus	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Xerocerastus sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Zootecus insularis	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Coeliaxis blandii	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Pyrgina umbilicata	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Glessula ceylanica	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Cecilioides gokweanus	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Ferussacia folliculus	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Thyrophorella thomensis	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Gibbulinella dewinteri	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Gonaxis quadrilateralis	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
Gonospira sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmm

Appendix 3.2: Alignment of the actin gene for the Achatinoidea and three streptaxid outgroup taxa.

	
	10 20 30 40 50 60	
<i>Achatina achatina</i>	TCCAAGAGAG	GTATYCTCAC TCTCAAGTAT CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Achatina fulica</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Achatina stuhlmanni</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Cochlitoma ustulata</i>	TCCAAGAGAG	GCATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Allopeas clavulinum</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGAATTGT CACCAACTGG
<i>Bocageia</i> sp.	TCCAAGAGAG	GTATCCTCAC ACTCAAGTAY CCCATTGARC ATGGTATTGT CACCAACTGG
<i>Eutomopeas layardi</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGAATTGT CACCAACTGG
<i>Leptinaria lamellata</i>	TCCAAGAGAG	GTATCCTAAC TCTCAAGTAT CCCATCGAGC ATGGCATTGT CACCAACTGG
<i>Paropeas achatinaceum</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGAATTGT CACCAACTGG
<i>Riebeckia</i> sp.	TCYAAGAGAA	GKATCCTCAC CYTCAAGTAC CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Rumina decollata</i>	TCYAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGTATTGT CACYAACTGG
<i>Subulina octona</i>	TCTAAGAGAG	GTATACTTAC TCTCAAGTAT CCCATTGAGC ATGGTATTGT CACAAACTGG
<i>Subulina striatella</i>	TCTAAGAGAG	GTATCCTCAC TCTCAAGTAC CCCATTGAGC AYGGTATTGT CACAAACTGG
<i>Subulina vitrea</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAY CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Subulona</i> sp.	TCCAAGAGAG	GTATCCTCAC TCTAAGTAC CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Tortaxis erectus</i>	TCTAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGARC ATGGTATTGT CACCAACTGG
<i>Xerocerastus</i> sp.	TCCAAGAGAG	GTATCTCAC CCTCAAGTAT CCCATTGAGC ATGGTATTGT CACTAACTGG
<i>Zootecus insularis</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Coeliaxis blandii</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Pyrquina umbilicata</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAT CCCATTGAGC ATGGWATTGT CACCAACTGG
<i>Glessula ceylanica</i>	TCCAAGAGAG	GTATCCTCAC TCTYAAGTAT CCCATTGAGC ATGGTRTTGT CACCAACTGG
<i>Ceciloides gokweanus</i>	TCCAAGAGAG	GTATCCTCAC ACTCAAGTAT CCCATTGAGC ACGGYATTGT CACCAACTGG
<i>Ferussacia folliculus</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAC CCCATTGAGC ATGGTATTGT CACCAACTGG
<i>Thyrophorella thomensis</i>	TCYAAGAGAG	GTATCCTTAC TCTCAARTAT CCCATYGAGC ATGGTATTGT CACCAACTGG
<i>Gibbulinella dewinteri</i>	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAC CCTATTGAAC ATGGCATTGT CACCAACTGG
<i>Gonaxis quadrilateralis</i>	TCCAAGAGAG	GCATCCTCAC TCTCAAGTAC CCCATTGAGC ACGGCATTGT CACCAACTGG
<i>Gonospora</i> sp.	TCCAAGAGAG	GTATCCTCAC TCTCAAGTAC CCCATTGAGC ATGGCATTGT CACCAACTGG

	
	70 80 90 100 110 120	
<i>Achatina achatina</i>	GATGATATGG	AGAAGATCTG GCATCACACY TTCTACAATG AGTTGAGAGT TGCWCCAGAA
<i>Achatina fulica</i>	GATGATATGG	AGAAGATCTG GCATCACACC TTCTACAATG AGTTSAGAGT TGCACCAGAA
<i>Achatina stuhlmanni</i>	GATGATATGG	AGAAAATATG GCATCACACC TTCTACAATG AGCTGAGAGT TGCACCAGAA
<i>Cochlitoma ustulata</i>	GATGATATGG	AGAAGATCTG GCACCACACC TTCTACAATG AGCTGAGAGT TGCACCAGAA
<i>Allopeas clavulinum</i>	GACGATATGG	AGAAGATCTG GCATCACACT TTCTACAACG AGCTCAGAGT TGCTCCTGAA
<i>Bocageia</i> sp.	GAYGATATGG	AGAAGATCTG GCATCACACT TTCTACAAYG AGCTKAGAGT TGWCCTGAA
<i>Eutomopeas layardi</i>	GACGATATGG	AGAAGATCTG GCATCACACT TTCTACAACG AGCTCAGAGT TGCTCCTGAA
<i>Leptinaria lamellata</i>	GACGATATGG	AGAAGATCTG GCATCACACT TTCTACAACG AGCTCAGAGT TGCTCCTGAA
<i>Paropeas achatinaceum</i>	GATGATATGG	AAAAGATCTG GCATCACACT TTCTACAACG AGCTCAGAGT TGCTCCTGAA
<i>Riebeckia</i> sp.	GAYGATATGG	AGAAGATCTG GCATCACACY TTCTACAACG AGCTGAGAGT TGCWCCTGAA
<i>Rumina decollata</i>	GAYGATATGG	AGAAGATCTG GCATCACACY TTCTACAATG AGCTGAGAGT TGCWCCTGAA
<i>Subulina octona</i>	GATGATATGG	AAAAGATCTG GCATCACACT TTCTACAATG AGCTGAGAGT TGCTCCTGAA
<i>Subulina striatella</i>	GATGATATGG	AGAAGATCTG GCATCACACT TTCTACAATG ARCTGAGAGT TGCTCCTGAA
<i>Subulina vitrea</i>	GATGATATGG	AGAAGATCTG GCATCACACY TTCTACAATG ARCTGAGAGT TGCCCCAGAA
<i>Subulona</i> sp.	GACGATATGG	AGAAGATCTG GCACCATACT TTCTACAACG AGCTTAGAGT TGCTCCAGAA
<i>Tortaxis erectus</i>	GATGACATGG	AGAAAATTGT GCATCACACT TTCTACAATG AGCTCAGAGT TGCACCTGAA
<i>Xerocerastus</i> sp.	GATGATATGG	AGAAGATCTG GCATCATACC TTCTACAATG AGCTGAGAGT TGCCCCTGAG
<i>Zootecus insularis</i>	GATGATATGG	AGAAGATCTG GCATCACACY TTCTACAATG AGCTGAGAGT TGCCCCTGAA
<i>Coeliaxis blandii</i>	GATGATATGG	AGAAGATCTG GCATCACACC TTCTACAATG AGCTGAGAGT TGCCCCTGAA
<i>Pyrquina umbilicata</i>	GAYGATATGG	AGAAGATCTG GCATYAYACY TTCTACAACG AGCTSAGAGT TGCYCCTGAA
<i>Glessula ceylanica</i>	GATGACATGG	AGAAGATCTG GCACCATACT TTCTACAATG AGCTCAGAGT TGCCCCTGAA
<i>Ceciloides gokweanus</i>	GATGATATGG	AGAAGATCTG GCATCACACC TTCTACAACG AGCTGAGAGT TGCCCCAGAA
<i>Ferussacia folliculus</i>	GATGATATGG	AGAAGATCTG GCATCACACC TTCTACAATG AGCTCAGAGT TGCCCCAGAG
<i>Thyrophorella thomensis</i>	GAYGATATGG	AGAAGATCTG GCATCAYACY TTCTACAACG AGCTSAGAGT TGCCCCTGAA
<i>Gibbulinella dewinteri</i>	GATGATATGG	AGAAGATCTG GCATCACACC TTCTACAACG AGCTGAGAGT TGCCCCTGAG
<i>Gonaxis quadrilateralis</i>	GATGACATGG	AGAAGATCTG GCATCACACC TTCTACAACG AGCTGAGAGT TGCCCCTGAG
<i>Gonospora</i> sp.	GATGATATGG	AGAAGATCTG GCATCACACC TTCTACAACG AGTTGAGAGT TGCCCCTGAG

	130 140 150 160 170 180
Achatina achatina	GAGCATCCAG TTCTRCKTAC AGAGGCTCCA CTCAATCCCA AGGCCAACAG AGAGAAGATG
Achatina fulica	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Achatina stuhlmanni	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Cochlitoma ustulata	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Allopeas clavulinum	GAACATCCAG TTTTACTTAC AGAAGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Bocageia sp.	GARCAYCCAG TWYTKYTKAC AGAGGCTCCA CTCAAYCCMA AGGCCAACAG AGARAAGATG
Eutomopeas layardi	GAGCATCCAG TTTTACTCAC AGAAGCTCCA CTCAACCCCA AGGCCAATAG AGAGAAGATG
Leptinaria lamellata	GAGCATCCAG TTTTACTCAC AGAGGCTCCA CTGAATCCCA AGGCCAACAG AGAGAAGATG
Paropeas achatinaceum	GAGCATCCAG TTTTACTTAC AGAGGCTCCA CTCAACCCCA AGGCCAATCG AGAAAAGATG
Riebeckia sp.	GARCAYCCAG TAYTTCTGAC AGAGGCTCCM CTCAATCCCA AGGCCAACAG AGAGAAGATG
Rumina decollata	GAGCACCAG TYTSTCTKAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Subulina octona	GAGCATCCAG TTTTGCTGAC AGAAGCACC A CTCAACCCCA AGGCCAATAG AGAGAAGATG
Subulina striatella	GAGCATCCAG TTTTGCTGAC AGAAGCACC A CTCAACCCCA AGGCCAACAG AGAGAAGATG
Subulina vitrea	GAGCAYCCAG TTCTCCTTAC AGAGGCTCCA CTCAATCCCA AGGCCAACAG AGAGAAGATG
Subulona sp.	GAGCATCCAG TTCTCCTGAC AGAGGCCCCA CTTAACCCCA AGGCCAACAG AGAAAAGATG
Tortaxis erectus	GAACATCCAG TTCTCCTTAC AGAGGCYCCA CTCAATCCCA AAGCCAACAG AGAAAAGATG
Xerocerastus sp.	GAGCACCAG TCCTCCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Zootecus insularis	GAGCACCAG TCCTCCTGAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Coeliaxis blandii	GAGCACCAG TACTTCTGAC AGAGGCTCCA CTCAATCCCA AGGCCAACAG AGAGAAGATG
Pyrgina umbilicata	GARCAYCCAG TWYTMCTYAC AGAGGCTCCW CTTAACCCCA AAGCCAAYAG AGARAAGATG
Glessula ceylanica	GAACAYCCAG TTYTCTTYAC AGAGGCYCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Ceciloides gokweanus	GAGCACCAG TACTCCTTAC AGAGGCCCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Ferussacia folliculus	GAGCACCAG TCCTCCTCAC TGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
Thyrophorella thomensis	GAGCAYCCAG TWCTCCTCAC AGAGGCTCCA CTTAACCCCA AAGCTAAYAG AGAAAAGATG
Gibbulinella dewinteri	GAACACCAG TCCTACTCAC GGAGGCTCCA CTCAACCCCA AAGCYAACAG AGAGAAGATG
Gonaxis quadrilateralis	GAACACCAG TYCTACTCAC GGAGGCTCCW CTCAATCCCA AAGCCAACAG AGAGAAGATG
Gonospira sp.	GAACACCAG TCCTACTCAC GGARGCTCCW CTCAATCCCA AAGCCAAYAG AGAGAAGATG

	190 200 210 220 230 240
Achatina achatina	ACCCAGATCA TGTTTGAGAC YTTCAAYWSY CCAGCYATGT ATGTYGCIAT TCAAGCNGTG
Achatina fulica	ACCCAGATCA TGTTTGAAAC YTTCAAYTCT CCAGCCATGT ATGTCGCCAT YCAAGCYGTG
Achatina stuhlmanni	ACCCAGATCA TGTTTGAAAC YTTCAACTCT CCAGCCATGT ATGTTGCCAT TCAAGCAGTC
Cochlitoma ustulata	ACCCAGATCA TGTTTGAGAC CTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCCGTG
Allopeas clavulinum	ACACAAATTA TGTTTGAAAC CTTCAATACT CCAGCCATGT ATGTCGCCAT CCAAGCCGTT
Bocageia sp.	ACCCAGATCA TGTTTGAGAC YTTCAAYACY CCAGCYATGT ACGTSKCIAT CCARGCTGTS
Eutomopeas layardi	ACACAGATTA TGTTTGAAAC CTTCAATACT CCAGCCATGT ATGTCGCCAT CCAAGCCGTT
Leptinaria lamellata	ACCCAGATTA TGTTTGAGAC CTTCAAACTT CCAGCAATGT ACGTCGCCAT TCAAGCTGTG
Paropeas achatinaceum	ACACAGATTA TGTTTGAAAC CTTCAATACC CCAGCCATGT ATGTCGCCAT CCAAGCCGTT
Riebeckia sp.	ACCCAGATCA TGTTTGAGAC YTTCAACWCY CCAGCCATGT AYGTGCIAT YCARGCWGTY
Rumina decollata	ACACAGATCA TGTTTGAGAC CTTCAAYACC CCAGCNATGT AYGTGCIAT YCARGCCGTG
Subulina octona	ACACAGATCA TGTTTGAGAC ATTCAACACC CCAGCTATGT ATGTTGCTAT CCAGGCTGTA
Subulina striatella	ACACAGATCA TGTTTGAGAC ATTCAAYTCY CCAGCCATGT ATGTTGCTAT YCARGCTGTS
Subulina vitrea	ACCCAGATCA TGTTTGAGAC CTTCAACACA CCAGCCATGT ATGTTGCIAT CCAGGCCGTA
Subulona sp.	ACCCAGATCA TGTTTGAGAC CTTCAACTCT CCAGCCATGT ACGTTGCAAT TCAAGCCGTG
Tortaxis erectus	ACCCAGATAA TGTTTGAGAC CTTCAAYWCT CCAGCGATGT ATGTTGCMAT CCAGGCAGTT
Xerocerastus sp.	ACACAGATCA TGTTTGAAAC CTTCAACTCT CCAGCCATGT ATGTAGCAAT TCAAGCAGTG
Zootecus insularis	ACACAGATCA TGTTTGAAAC CTTCAATACC CCAGCAATGT ATGTMGCCAT CCAGGCNGTG
Coeliaxis blandii	ACCCAGATCA TGTTTGAGAC CTTCAACACC CCAGCCATGT ACGTCGCCAT CCAGGCCGTA
Pyrgina umbilicata	ACCCAGATTA TGTTTGAGAC CTTCAAYWCY CCAGCCATGT ACGTNGCCAT CCAAGCTGTY
Glessula ceylanica	ACCCAGATMA TGTTTGAGAC CTTCAACTCT CCAGCMATGT WTGTGCMCAT TCAAGCCGTA
Ceciloides gokweanus	ACCCAGATCA TGTTTGAGAC CTTCAAACTT CCAGCCATGT ACGTTGCCAT CCAGGCCGTA
Ferussacia folliculus	ACACAGATCA TGTTTGAAAC CTTCAACTCC CCAGCAATGT ACGTCGCCAT TCAAGCCGTA
Thyrophorella thomensis	ACCCAGATTA TGTTTGAGAC YTTCAAYWCT CCAGCYATGT AYGTGCTAT CCAAGCTGTC
Gibbulinella dewinteri	ACTCAGATCA TGTTTGAGAC ATTCAAACTT CCAGCCATGT ATGTCGCCAT CCAGGCCGTA
Gonaxis quadrilateralis	ACCCAGATCA TGTTTGAAAC ATTCAACTCW CCAGCCATGT ATGTGCIAT CCAGGCAGTY
Gonospira sp.	ACCCAGATCA TGTTTGAGAC ATTYAACWCT CCAGCCATGT ATGTYGCSAT YCAGGCNGTT

	250 260 270 280 290 300
Achatina achatina	CTTTCYTTRT ATGCTTCAGG TCGTACAAC TGGATTGTGC TGGATTCTGG WGATGGTGTG
Achatina fulica	CTTTCCTTTRT ATGCWTCAGG TCGTACAAC TGGATTGTGC TGGATTCTGG TGATGGTGTG
Achatina stuhlmanni	CTTTCCTTTAT ATGCATCCGG TCGTACAAC TGGATTGTGC TGGATTCTGG TGATGGTGTG
Cochlitoma ustulata	CTTTCCTTTAT ATGCATCCAG TCGTACAAC TGGATTGTGC TGGATTCTGG TGATGGTGTG
Allopeas clavulinum	CTCTCCCTGT ATGCCTCAGG TCGTACAAC TGGATTGTGC TGGATTCTGG AGATGGTGTG
Bocageia sp.	CTGTCCCTGT AYGCCTCAGG TCGKACNAC GGTATTGTGC TGGATTCTGG WGATGGWGTG
Eutomoepas layardi	CTTTCCTCTGT ATGCCTCTGG TCGTACTAC TGGATTGTGC TGGATTCTGG AGATGGTGTG
Leptinaria lamellata	CTGTCCCTGT ATGCCTCAGG TCGTACAAC GGTATTGTGC TAGATTCTGG AGATGGAGTG
Paropeas achatinaceum	CTCTCCCTGT ATGCCTCTGG TCGTACAAC GGCATTGTGC TCGATTCTGG AGATGGTGTG
Riebeckia sp.	CTGTCCCTGT AYGCCTCAGG TCGTACAAC GGGATTGTGC TAGATTCTGG WGATGGTGTG
Rumina decollata	CTKTCTCTGT ATGCCTCTGG TCGTACAAC GGTATTGTGC TAGATTCTGG WGATGGTGTG
Subulina octona	CTGTCTCTGT ATGCCTCTGG TCGTACGAC TGGATTCTGG AGATGGTGTG
Subulina striatella	CTATCTCTGT ATGCCTCTGG TCGTACGAC TGGATTCTGG AGATGGTGTG
Subulina vitrea	CTTTCCTTGT ATGCCTCTGG TCGTACAAC TGGATTGTGC TAGATTCTGG AGATGGTGTG
Subulona sp.	CTGTCCCTGT ATGCCTCAGG TAGGACAACT GGTATTGTGC TAGATTCTGG AGATGGTGTG
Tortaxis erectus	CTCTCACTGT ATGCCTCAGG TCGTACAAC GGTATTGTGT TAGATTCTGG TGATGGTGTG
Xerocerastus sp.	CTTTCCTCTGT ATGCCTCTGG TCGTACAAC GGTATTGTGC TAGATTCTGG TGATGGTGTG
Zootecus insularis	CTATCCCTGT ATGCCTCTGG TCGTACAAC GGTATTGTGC TAGATTCTGG TGATGGTGTG
Coeliaxis blandii	CTTTCCTCTGT ATGCCTCAGG TCGTACAAC GGTATTGTGC TAGATTCTGG TGATGGTGTG
Pyrquina umbilicata	CTGTCCCTGT ATGCCTCAGG TCGTACAAC GGAATTGTGC TAGATTCTGG AGATGGWGTG
Glessula ceylanica	CTATCGCTNT ATGCCTCAGG TCGTACAAC GGTATTGTGC TAGATTCTGG TGATGGTGTG
Cecilioides gokweanus	CTGTCCCTGT ATGCCTCAGG TCGTACAAC GGTATTGTGC TAGATTCTGG TGATGGTGTG
Ferussacia folliculus	CTTTCCTCTGT ATGCCTCAGG TCGTACCAC GGTATTGTGC TGGATTCTGG TGATGGTGTG
Thyrophorella thomensis	CTCTCCCTGT ATGCCTCAGG TCGTACAAC GGAATTGTGC TAGATTCTGG MGAATGGWGTG
Gibbulinella dewinteri	CTTTCCTCTGT ATGCCTCAGG TCGTACCAC GGTATTGTGC TCGATTCTGG TGATGGTGTG
Gonaxis quadrilateralis	CTGTCTTGT ATGCCTCAGG TCGTACTAC GGTATTGTGC TGGATTCTGG TGATGGTGTG
Gonospora sp.	CTKTCTTGT ATGCCTCAGG TCGWACCAC GGTATTGTGC TGGATTCTGG KGATGGTGTG

	310 320 330 340 350 360
Achatina achatina	ACYCAYACTG TNCCAAATMTA TGAAGGTTAT GCTCTTCCYC AYGCCATCAT GAGACTGGAY
Achatina fulica	ACCCACACTG TCCCAATTTA TGAAGGTTAT GCTCTACCTC ATGCCATCAT GAGACTGGAC
Achatina stuhlmanni	ACCCACACTG TCCCAATTTA TGAAGGTTAT GCTCTTCCCC ATGCCATCAT GAGACTGGAC
Cochlitoma ustulata	ACTCACACTG TCCCAATTTA TGAAGGTTAT GCTCTTCCCC ATGCCATCAT GAGACTGGAC
Allopeas clavulinum	ACTCACACTG TCCCTATCTA TGAAGGCTAT GCTCTACCAC ACGCCATCAT GAGATTAGAC
Bocageia sp.	WCYCAACWG TSCCYATMTA TGAAGGTTAT GCCCTTCCCTC ATGCCATCAT GAGRYTKGAC
Eutomoepas layardi	ACTCATACTG TCCCATCTA TGAAGGCTAT GCTCTCCAC ACGCCATCAT GAGATTAGAC
Leptinaria lamellata	ACTCACACTG TCCCTATATA TGAAGGTTAT GCCCTCCCTC ATGCTATATAT GAGACTGGAC
Paropeas achatinaceum	ACTCATACTG TCCCTATCTA TGAAGGCTAT GCTCTCCAC ATGCCATCAT GAGATTAGAC
Riebeckia sp.	ACACACACTG TCCCATCTA TGAAGGTTAT GCMCTTCCCC AYGCCATCAT GAGACTGGAT
Rumina decollata	ACCCAYACTG TCCCATCTA TGAAGGTTAT GCCCTTCCCTC ATGCCATCAT GAGAYTGGAC
Subulina octona	ACCCACACTG TTCCCATCTA TGAAGGTTAT GCTCTTCCCTC ATGCCATCAT GAGACTGGAC
Subulina striatella	ACCCACACTG TWCCMATCTA TGAAGGCTAT GCTCTTCCCTC ACGCCATCAT GAGACTGGAC
Subulina vitrea	ACTCACACTG TYCCCATCTA TGAAGGTTAT GCTCTTCCCC ATGCCATCAT GAGACTGGAT
Subulona sp.	ACCCATACTG TCCCATCTA CGAAGGTTAT GCTCTCCAC ATGCCATCAT GAGACTGGAC
Tortaxis erectus	ACCCACACTG TTCCCATCTA TGAAGGATAT GCCCTTCCCTC ATGCCATCAT GAGACTGGAC
Xerocerastus sp.	ACCCATACTG TCCCATATA TGAAGGTTAT GCCCTTCCCC ACGCCATCAT GAGACTGGAC
Zootecus insularis	ACCCACACTG TCCCYATCTA TGAAGGTTAT GCCCTTCCCTC ATGCCATCAT GAGACTGGAY
Coeliaxis blandii	ACWCACACTG TCCCATCTA TGAAGGTTAT GCCCTTCCCTC ATGCCATCAT GAGACTGGAT
Pyrquina umbilicata	ACTCAYACWG TCCCYATCTA YGAAGGTTAT GCTCTTCCCTC ACGCCATCAT GAGACTGGAT
Glessula ceylanica	ACCCACACTG TCCCTATYTW TGAAGGTTAT GCCCTTCCCTC ATGCCATCAT GAGATTGGAC
Cecilioides gokweanus	ACTCACACTG TTCCCATCTA TGAAGGTTAT GCTCTTCCCTC ATGCCATCAT GAGACTGGAC
Ferussacia folliculus	ACCCACACTG TCCCATCTA TGAAGGTTAT GCTTTGCCCC ACGCCATCAT GAGACTGGAT
Thyrophorella thomensis	ACTCAYACWG TCCCYATMTA YGAAGGCTAT GCTCTTCCCTC ACGCCATCAT GAGAYTGGAT
Gibbulinella dewinteri	ACCCACACTG TCCCATCTA TGAAGGTTAT GCYCTTCCCC ACGCCATCAT GAGACTGGAT
Gonaxis quadrilateralis	ACCCACACTG TCCCYATCTA TGAAGGTTAT GCTCTYCCCC AYGCCATCAT GAGACTGGAY
Gonospora sp.	ACCCAYACAG TCCCYATCTA TGAAGGTTAT GCCCTTCCCTC AYGCCATCAT GAGATTGGAT

	
	370 380 390 400 410 420	
Achatina achatina	TTGGCTGGTC GTGACCTCAC AGATTATCTC ATGAARATCC TCACAGAGAG AGGTTACAGC	
Achatina fulica	TTGGCTGGTC GTGACCTCAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC	
Achatina stuhlmanni	TTGGCTGGTC GTGACCTCAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC	
Cochlitoma ustulata	TTGGCTGGTC GTGACCTCAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC	
Allopeas clavulinum	TTGGCTGGCC GTGACCTCAC CGACTACCTC ATGAAGATTC TGACTGAGCG AGGATACAGC	
Bocageia sp.	YTGGCTGGYC GTGACCTTAC AGAYTACCTS ATGAAGATYC TSACWGAGAG AGGCTACAGC	
Eutomopeas layardi	TTGGCTGGCC GTGACCTCAC TGACTACCTC ATGAAGATCC TCACTGAGCG AGGATACAGC	
Leptinaria lamellata	TTGGCTGGTC GTGATCTCAC AGATTATTTG ATGAAGATTC TAACAGAGAG AGGCTACAGC	
Paropeas achatinaceum	TTGGCTGGCC GTGACCTCAC TGACTACCTC ATGAAGATTC TCACTGAGCG AGGCTACAGC	
Riebeckia sp.	CTGGCYGGCC GTGACCTCAC AGACTACCTC ATGAAGATCC TCACAGAGAG GGGCTACAGC	
Rumina decollata	TTGGCTGGTC GTGACCTYAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC	
Subulina octona	TTGGCTGGTC GCGACCTGAC AGATTATCTA ATGAAGATTC TCACGGAGAG AGGTTACAGC	
Subulina striatella	TTGGCTGGTC GAGACCTGAC AGATTATCTG ATGAAGATTC TCACAGAGAG AGGTTACAGC	
Subulina vitrea	CTGGCTGGCC GTGACCTCAC AGACTAYCTC ATGAAGATCC TGACAGAGAG AGGCYACAGC	
Subulona sp.	TTGGCTGGTC GAGACCTCAC AGACTACCTC ATGAAGATCC TCACTGAGAG AGGTTACAGC	
Tortaxis erectus	TTGGCTGGCC GTGACCTCAC AGATTACCTC ATGAARATCC TCACAGAGAG AGGTTACAGC	
Xerocerastus sp.	TTGGCTGGTC GTGACCTCAC AGATTACCTC ATGAAAATCC TCACAGAGAG AGGCTACAGC	
Zootecus insularis	TTGGCTGGTC GTGACCTCAC AGATTAYCTC ATGAAGATCC TCACAGAGAG AGGCTAYAGY	
Coeliaxis blandii	CTGGCTGGCC GTGACCTCAC AGACTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC	
Pyrgina umbilicata	CTGGCTGGCC GTGACCTCAC AGAYTAYCTC ATGAAGATCC TMCACAGAGM AGGMTACAGC	
Glessula ceylanica	CTGGCTGGAC GAGACCTCAC AGATTATCTC ATGAAGATTC TTMACAGAGAG AGGCTACCCC	
Ceciloides gokweanus	TTGGCTGGCC GTGACCTTAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC	
Ferussacia folliculus	CTGGCTGGTC GTGATCTCAC AGATTACCTC ATGAAGATCC TYACAGAGAG AGGCTACAGC	
Thyrophorella thomensis	CTRGCTGGCC GTGAYCTCAC AGATTAYCTC ATGAAGATCC TCACAGAGAG AGGATACAGC	
Gibbulinella dewinteri	CTGGCTGGCC GTGACCTTAC AGATTAYCTG ATGAAGATCC TCACAGAGAG AGGCTACAGT	
Gonaxis quadrilateralis	TTGGCYGGGC GTGACCTTAC AGATTACCTG ATGAAGATCC TCACAGAGAG AGGCTACAGC	
Gonospira sp.	TTGGCTGGAC GTGACCTKAC AGATTAYYTG ATGAAGATCC TCACAGAGAG AGGCTAYAGC	

	
	430 440 450 460 470 480	
Achatina achatina	TTCACCACCA CAGCTGAGAG AGAAATTGTT CGAGAYATCA ARGAGAAGCT GTGCTATGTG	
Achatina fulica	TTCACCACCA CAGCTGAGAG AGAAATTGTT CGAGACATCA AAGAGAAGCT TTGCTATGTG	
Achatina stuhlmanni	TTCACCACCA CAGCTGAGAG AGAAATTGTT CGAGACATCA AAGAGAAAAT TTGCTATGTG	
Cochlitoma ustulata	TTCACCACCA CAGCTGAGAG AGAAATTGTT CGAGACATCA AAGAGAAGCT TTGCTATGTG	
Allopeas clavulinum	TTCACACAA CAGCCGAGAG AGAAATTGTA CGTGACATCA AGGAGAAAAT TTGCTACGTT	
Bocageia sp.	TTCACYACMA CWCWGAGAG AGAAATTGTT CGAGAYATCA AGGAAAAACT TTSCTATGTY	
Eutomopeas layardi	TTCACACAA CAGCCGAGAG AGAAATTGTA CGCGACATCA AGGAGAAAAT TTGCTACGTT	
Leptinaria lamellata	TTCACAACAA CGGCTGAGAG AGAAATTGTA CGTGACATCA AGGAGAAAAT TTGCTATGTT	
Paropeas achatinaceum	TTCACAACAA CAGCCGAGAG AGAAATTGTA CGTGACATCA AAGAGAAAAT TTGCTACGTT	
Riebeckia sp.	TTCACCACCA CTGCTGAGAG AGAAATTGTT CGAGACATCA AAGARAAGCT TTSYATYGTG	
Rumina decollata	TTCACCACCA CTGCTGAGAG AGAAATTGTT CGAGACATCA AGGAGAAGCT GTSCTATGTT	
Subulina octona	TTCACAACCA CAGCTGAGAG AGAAATTGTT CGAGACATCA AGGAGAAAAT TTGCTACGTC	
Subulina striatella	TTCACCACCA CAGCYGAGAG AGAAATTGTY CGAGACATCA AAGAGAAAAT TTGCTACGTC	
Subulina vitrea	TTCACCACCA CWCCTGAGAG AGAAATTGTT CGAGACATCA AGGAGAAAAT CTGYTATGTT	
Subulona sp.	TTCACCACAA CCGCCGAGAG AGAAATTGTT CGAGACATCA AGGAGAAAAT CTGCTACGTC	
Tortaxis erectus	TTCACYACMA CAGCYGAGAG AGAAATTGTT CGAGAYATCA AGGAGAAGCT KTSCTAYGTK	
Xerocerastus sp.	TTCACCACCA CCGCTGAGAG AGAAATTGTT CGAGACATCA AGGAGAAGCT TTGCTATGTT	
Zootecus insularis	TTCACCACCA CCGCTGAGAG AGAAATTGTT CGAGACATCA AGGARAAGCT TTSCTATGTT	
Coeliaxis blandii	TTCACCACCA CTGCTGAGAG AGAAATYGTT MGAGACATCA ARGAGAAGCT TTSCTATGTT	
Pyrgina umbilicata	TTYACTACCA CAGCYGAGAG AGAAATTGTT MGWGACATCA ARGAGAARCT KTYTAYGTK	
Glessula ceylanica	TTCACYACMA CAGCCGAGAG GRAAATYGTK CGAGACATGA AAGAGAAGCT TGCGTATGTT	
Ceciloides gokweanus	TTCACCACCA CAGCCGAGAG GGAATTTGTC CGAGACATCA AGGAGAAGCT TTCATACGTT	
Ferussacia folliculus	TTCACCACCA CTGCTGAGAG AGAAATTGTS AGGGAYATCA AGGAGAAGCT CTGTTATGTT	
Thyrophorella thomensis	TTYACTACCA CAGCYGAGAG AGAAATTGTT MGRGAYATCA ARGAGAARCT NTSYATACGTK	
Gibbulinella dewinteri	TTCACCACCA CTGCTGAGAG AGAGATTGTA CGAGACATCA AGGAGAAGCT CTGCTACGTT	
Gonaxis quadrilateralis	TTCACCACCA CTGCTGAGAG AGAAATTGTA MGAGACATCA AGGAGAARCT CTGCTACGTT	
Gonospira sp.	TTCACCACCA CTGCGWAGAG AGAGATYGTA CGAGACATMA ARGARAAGCT MTGYTATGTT	

	490 500 510 520 530 540
Achatina achatina	GCYCTTGACT TTGAGCAAGA GATGTCRACA GCTTCCACAT CATCTTCATT AGAGAAGAGY
Achatina fulica	GCCCTTGACT TTGAGCAAGA RATGGCAACA GCTTCCACAT CATCTTCATT AGAGAAGAGT
Achatina stuhlmanni	GCCCTTGACT TTGAGCAAGA GATGGCAACA GCTGCCACAT CATCTTYATT AGAGAAGAGT
Cochlitoma ustulata	GCCCTTGACT TTGAGCAAGA GATGGCAACA GCTTCCACAT CATCTTCATT AGAGAAGAGT
Allopeas clavulinum	GCCCTAGATT TTGAGCAGGA GATGGGAACA GCTGCTACAT CGTCATCTTT GGAGAAGAGC
Bocageia sp.	GCTCTTGACT TTGAACAGGA AATGSACACA GCTGCCWCAT CATCTTCYCT RGAGAAGAGC
Eutomoepas layardi	GCTCTTGACT TTGAGCAGGA GATGGGAACA GCTGCTACAT CATCATCGTT GGAAGAGC
Leptinaria lamellata	GCTCTTGATT TTGAGCAGGA GATGGGTACA GCTGCAACGT CATCATCATT AGAGAAGAGC
Paropeas achatinaceum	GCCCTAGATT TCGAACAGGA GATGGGAACA GCTGCCACAT CATCATCTTT GGAGAAGAGC
Riebeckia sp.	GCTCTYGACT TTGAGCAAGA AAWGGSTACG GCTGCCACAT CWTCTTCAYT AGAGAAGAGT
Rumina decollata	GCTCTTGACT TTGAACAAGA AATGGCAACA GCTGCCACTT CATCTTCWCT GGAGAAAAGT
Subulina octona	GCCCTTGACT TTGAACAAGA AATGGGTACA GCTGCCACAT CGTCATCTTT AGAGAAGAGC
Subulina striatella	GCCCTTGACT TCGAACAGA AATGGGTACA GCTGCCACAT CCTCATCTTT GGAGAAGAGT
Subulina vitrea	GCYCTTGACT TCGARCAGA AATGTCTACA GCTGCCACRT CWTCTTCATT RGAGAAGAGT
Subulona sp.	GCACTTGACT TTGAGCAGGA GATGCAGACA GCTGCTACAT CATCATCTTT GGAGAAAAGC
Tortaxis erectus	GCTCTAGACT TTGAGCAGGA GATGGCAACA GCTGCCACTT CATCTWCTCT AGAGAAAAGT
Xerocerastus sp.	GCTCTTGACT TTGAACAAGA AATGGCAACA GCTGCCACTT CATCTTCATT AGAGAAGAGT
Zootecus insularis	GCTCTTGAYT TTGAACAAGA AATGGCAACA GCTGCCACAT CATCTTCATT AGAGAAGAGT
Coelioxys blandii	GCTCTTGACT TTGAGCAAGA AATGGCTACA GCTGCYACAT CATCTTCATT AGAGAAGAGT
Pyrquina umbilicata	GCTCTRGACT TTGARCAAGA RATGGCRACA GCWGCYWCTT CATCYWYCYT NGAGAAGAGC
Glessula ceylanica	GCTCTTGACT TTGAMCARGA GATGCAGWTA TCATCCAGYK CTTCAWCAAT TGAARAAAGT
Cecilioides gokweanus	GCWCTAGACT TTGAGCAAGA AATGGCAACA GCTGCAACTT CATCTTCATT RGAGAAGAGC
Ferussacia folliculus	GCTCTTGATT TTGAACAGGA GATGGCTACA GCTGCAACTT CCTCTCCCTT GGAGAAGAGC
Thyrophorella thomensis	GCTCTRGACT TTGAGCAAGA AATGGCAACA GCTGCYACTT CATCYTCYCT GGAGAAGAGY
Gibbulinella dewinteri	GCTCTTGACT TTGAGCAGGA GATGGCAACA GCTGCTACAT CATCATCATT TGAGAAGAGC
Gonaxis quadrilateralis	GCTCTTGACT TTGAGCARGA GATGGCMACA GCTGCTACAT CATCMTCCCT TGAGAAGAGY
Gonospora sp.	GCTCTTGACT TTGAACARGA GATGGSWACW GCWGCWWCAT CATCWTCCCT TGAGAAGAGC

	550 560 570 580 590 600
Achatina achatina	TATGAATTGC CTGATGGACA RGTCATYACT ATTGGTAAYG AGCGYTTTCAG RTGYCCAGAA
Achatina fulica	TATGAATTTC CTGATGGACA GGTTCATCACC ATTGGTAAYG AGCGTTTCAG ATGTCCAGAA
Achatina stuhlmanni	TATGAATTGC CTGATGGACA GGTTCATCACC ATTGGTAAYG AGCGATTTCAG GTGCCAGAA
Cochlitoma ustulata	TATGAATTAC CTGATGGACA GGTTCATAACT ATTGGTAAYG AGCGCTTCAG GTGCCAGAA
Allopeas clavulinum	TACGAACCTC CAGATGGACA GGTTCATCACC ATTGGCAACG AGCGCTTCAG ATGCCCTGAG
Bocageia sp.	TAYGAACCTC CTGATGGACA GGTTCATCACC ATTGGCAATG ARCGWTTTCAG ATGTCCWGAR
Eutomoepas layardi	TACGAGCTTC CAGATGGACA GGTTCATCACA ATCGGCAATG AGCGCTTCAG ATGCCCTGAG
Leptinaria lamellata	TATGAGCTGC CTGATGGACA AGTCATTACC ATTGGAAATG AGCGCTTCAG ATGCCCTGAG
Paropeas achatinaceum	TACGAACCTC CAGATGGACA AGTTATCACA ATTGGCAATG AGCGCTTCAG ATGCCCTGAG
Riebeckia sp.	TAYGAATTGC CTGATGGACA AGTCATCACC ATTGGYAAAY ARCGKTTTCAG GWGYCCAGAA
Rumina decollata	TATGAATTGC CTGAYGGACA GGTTCATCACC ATCGGCAAYG AGCGTTTCAG ATGTCCAGAA
Subulina octona	TACGAACCTC CAGATGGACA AGTGATTACC ATTGGCAATG AACGTTTCAG ATGCCCTGAG
Subulina striatella	TACGAACCTC CAGATGGACA AGTTATTACC ATTGGCAATG AACGTTTCAG ATGCCCTGAG
Subulina vitrea	TACGAGYTCG CTGATGGACA GGTTCATCACC ATTGGCAAYG AGCGTTTCAG ATGYCCWGAA
Subulona sp.	TACGAACCTC CAGATGGACA AGTCATTACT ATTGGTAACG AGCGATTTCAG ATGTCCCGAA
Tortaxis erectus	TATGAACTRC CTGATGGACA GGTTCATCACC ATTGGCAAYG AAAGATTTCAG RTGNCCAGAA
Xerocerastus sp.	TACGAACCTC CTGATGGACA GGTTCATCACC ATTGGCAATG AGCGCTTCAG GTGCCAGAA
Zootecus insularis	TATGAATTGC CTGAYGGACA GGTTCATCACC ATTGGCAATG AGCGTTTCAG RTGTCCAGAA
Coelioxys blandii	TATGAATTGC CTGATGGACA AGTCATCACC ATCGGTAAYG AGCGTTTCAG ATGCCCWGAA
Pyrquina umbilicata	TACGARYTKC CWGATGGACA GGTTCATCACC ATTGGNAAYG AGCGYTTTCAG RTGTCCWGAR
Glessula ceylanica	TACGARCTKC CGGATGGACA SRTMATYACM ATWGGAAACG AGCGATTTCAG RTGTCCCGAA
Cecilioides gokweanus	TACGAACCTC CCGATGGTCA AGTCATCACC ATTGGCAACG AGCGTTTCAG ATGTCCAGAA
Ferussacia folliculus	TACGAACCTC CTGACGGACA GGTTCATCACC ATTGGCAACG AGCGATTTCAG RGCTCCGAA
Thyrophorella thomensis	TACGARYTNC CWGATGGACA GGTTCATCACC ATTGGWAAAY AGCGTTTCAG ATGTCCAGAA
Gibbulinella dewinteri	TAYGAACCTC CAGACGGACA GGTTCATCACC ATTGGCAATG ARCGATTTCAG GTGCCAGAA
Gonaxis quadrilateralis	TATGAACTRC CWGAYGGACA GGTTCATYACY ATTGGMAACG ARCGWTTTCAG RTGCCAGAA
Gonospora sp.	TATGAAYTAC CTGAYGGACA GGTTCATYACC ATTGGCAAYG ARMGWTTTCAG GWSWCTGAA

	610	620	630	640	650	660	
Achatina achatina	GCCATGTTCC	AGCCATCTTT	CCTTGGTATG	GARWCTGCAG	GTATTCATGA	AACCACCTAC	
Achatina fulica	GCCATGTTCC	AGCCWTCTTT	CCTTGGTATG	GARWCCGCAG	GTATTCATGA	RACCACYTAC	
Achatina stuhlmanni	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCACGA	GACCACCTAC	
Cochlitoma ustulata	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
Allopeas clavulinum	GCAATGTTCC	AGCCATCCTT	CCTTGGTATG	GAGTCTGCTG	GTACCCATGA	AACAACGTAC	
Bocageia sp.	GCAATGTTYC	AGCCATCTTT	CCTKGGGAATG	GAGTCTGCWG	GTATTCATGA	GACYAYCTAC	
Eutomopeas layardi	GCAATGTTCC	AGCCATCCTT	CCTTGGTATG	GAGTCTGCTG	GTACCCATGA	AACAACGTAC	
Leptinaria lamellata	GCAATGTTCC	AGCCATCTTT	CCTTGGCATG	GAATCTGCTG	GCACACATGA	AACAACATAC	
Paropeas achatinaceum	GCAATGTTCC	AGCCATCATT	CCTTGGTATG	GAGTCTGCAG	GTACCCATGA	AACAACATAC	
Riebeckia sp.	GCAATGTTCC	AGCCATCCTT	YCTTGGYATG	GAGTCTGCTG	GTATTCAYGA	RACCACYTAC	
Rumina decollata	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATTCATGA	GACCACCTAC	
Subulina octona	GCAATGTTTC	AGCCATCTTT	TCTGGGCATG	GAATCTGCAG	GTACTCATGA	AACAACGTAC	
Subulina striatella	GCAATGTTYC	AGCCATCTTT	TCTGGGCATG	GAATCTGCAG	GTACTCACGA	AACAACATAC	
Subulina vitrea	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GARTCWGCTG	GTAYTCACGA	GACCACYTAC	
Subulona sp.	GCTGAGTTCC	AACCATCTTT	CCTTGGTATG	GAGTCAGCTG	GCATTACAG	AACCTACCTAT	
Tortaxis erectus	GCWATGTTNC	ARCCATCTTT	TCTTGGTATG	GAATCTGCTG	GWATCCATGA	GACCACATAC	
Xerocerastus sp.	GCAGAATTCC	AACCATCCTT	CCTTGGTATG	GAGTCTGCTG	GTATTCATGA	GACCACCTAC	
Zootecus insularis	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATYCATGA	GACCACCTAC	
Coeliaxis blandii	GCAATGTTCC	AGCCATCCTT	CCTTGGWATG	GAATCTGCTG	GTATTCACGA	GACCACCTAC	
Pyrgina umbilicata	GCAATRTTCC	ARCCATCWTT	CCTTGGYATG	GARTCTGCYG	GTATTCATGA	AACCACCTAC	
Glessula ceylanica	GCTTTGTTCC	AGCCATCTTT	TCTAGGTMTA	GAGATCRSTG	GTATTCACGA	AACCACCTAC	
Ceciloides gokweanus	GCAATGTTCC	AGCCATCCTT	CCTTGGTATG	GAGTCTGCTG	GTATTCACGA	GACCACATAC	
Ferussacia folliculus	GCAGAATTTT	AGCCATCTTT	CCTTGGCATG	GAATCTGCCG	GTATTCACGA	AACAACCTAC	
Thyrophorella thomensis	GCAATRTTCC	ARCCATCNTT	CCTTGGTATG	GAGTCWGCTG	GTATTCATGA	AACCACCTAC	
Gibbulinella dewinteri	GCAATGTTCC	AACCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATTCACGA	GACCACGTAC	
Gonaxis quadrilateralis	GCAATGTTYC	AGCCATCWTT	CYTGGTATG	GARTCTGCYG	GYRTCCAYGA	GACMACATAC	
Gonospira sp.	GCAATGTTYC	AGCCATCYTT	CYTSGGYATG	GAGTCTGCTG	GTRTYCAYGA	GACCACATAC	

	670	680	690	700	710	720	
Achatina achatina	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTA	
Achatina fulica	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
Achatina stuhlmanni	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTACTT	
Cochlitoma ustulata	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
Allopeas clavulinum	AATTCCATCA	TGAAGTGCGA	TGTTGACATC	AGAAAAGACC	TCTATGCCAA	CACTGTCTCTG	
Bocageia sp.	AAATCYATCA	TGAAGTGTGA	TGYGAYATC	CGTAAAGAYY	TGTAYGCCAA	YAYTGTACTG	
Eutomopeas layardi	AATTCCATCA	TGAAATGCGA	TGTTGATATC	AGAAAAGACC	TCTATGCCAA	CACTGTCTCTG	
Leptinaria lamellata	AATTCAATTA	TGAAGTGTGA	TGTTGATATC	AGAAAAGACC	TCTATGCCAA	CACTGTACTA	
Paropeas achatinaceum	AATTCCATTA	TGAAGTGCGA	TGTTGATATC	AGAAAAGACC	TTTATGCTAA	CACTGTATTG	
Riebeckia sp.	AAATCCWTYA	TGAARTGTGA	YGTYGACATY	CGTAAAGACT	TGTATGCCAA	CWCCGTCTTG	
Rumina decollata	AATTCCATCA	TGAARTGTGA	TGYGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTCTTG	
Subulina octona	AATTCAATCA	TGAAGTGTGA	CGTTGATATC	CGAAAAGACC	TTTATGCTAA	CACTGTACTG	
Subulina striatella	AATTCAATCA	TGAAATGTGA	CGTCGATATC	CGAAAAGACC	TATATGCTAA	CACTGTACTG	
Subulina vitrea	AACTCCATCA	TGAAGTGTGA	TGYGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTAYTG	
Subulona sp.	AACTCCATCA	TGAAGTGTGA	TGTTGACATC	AGAAAAGACC	TTTACGCCAA	TACTGTGTTG	
Tortaxis erectus	AACTCCATCA	TGAARTGTGA	TGTTGACATC	CGTAAAGACT	TGTATGCAAA	YACAGTCTCTG	
Xerocerastus sp.	AACTCCATCA	TGAAGTGTGA	TGTTGACATT	CGTAAAGACT	TGTATGCCAA	CACTGTGTTG	
Zootecus insularis	AACTCCATCA	TGAARTGCGA	TGTTGACATY	CGTAAAGACY	TGTATGCCAA	CACTGTATTG	
Coeliaxis blandii	AACTCCATCA	TGAAGTGTGA	CGTAGACATY	CGTAAAGACT	TGTACKCCAA	CAYCGTATTG	
Pyrgina umbilicata	AACTCCATCA	TGAAATGYGA	YGTYGACATY	CGTAAAGAYT	TGTATGCCAA	CACTGTATTG	
Glessula ceylanica	AACTCCATMA	TSAAATGTGA	CGTCGACATC	CGTAAAGACC	TGTAYGCMMA	CACWGTCTYTR	
Ceciloides gokweanus	AACTCCATCA	TGAAGTGCGA	CGTTGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
Ferussacia folliculus	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTACGCCAA	CACTGTCTCTG	
Thyrophorella thomensis	AAATCYATCA	TGAAATGCGA	CGTYGAYATY	CGWAAAAGACY	TGTATGCCAA	CACTGTATTG	
Gibbulinella dewinteri	AACTCCATCA	TGAAGTGCGA	CGTTGACATC	CGTAAAGACT	TGTACGCCAA	CACCGTGCTG	
Gonaxis quadrilateralis	AACTCCATCA	TGAARTGCGA	YGTTGATATC	CGTAAAGAYT	TGTACKCCAA	CACYGNTTGT	
Gonospira sp.	AACTCCATCA	TGAARTGTGA	TGTRGAYATC	CGMAAAGACT	TGTATTCCAA	CACTGTCTCTG	

	730	740	750	760	770	780	
Achatina achatina	TCTGGTGGAW	CTACCATGTA	TCCTGGCATT	GCTGACCGCA	TGCAGAAAGGA	AATCACNAAY	
Achatina fulica	TCTGGTGGAW	CTACCATGTA	TCCAGGCATT	GCTGACCGCA	TGCAGAAAGGA	AATYGYCAAC	
Achatina stuhlmanni	TCTGGTGGCT	CCACAATGTT	TCCAGGCATC	GCTGACCGCA	TGCAGAAAGGA	AATCACCAGC	
Cochlitoma ustulata	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAAGGA	AATCACCAGC	
Allopeas clavulinum	TCTGGTGGTT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAAGA	GATAACCGCT	
Bocageia sp.	TCTGGMGGTT	CYACAATGTT	YCCTGGMATY	GCTGACCGCA	TGCAGAAARGA	AATSACCTCW	
Eutomopeas layardi	TCTGGTGGTT	CCACAATGTT	CCCTGGCATC	GCTGACCGCA	TGCAGAAAGA	GATAACCGCG	
Leptinaria lamellata	TCTGGTGGTT	CCACTATGTT	CCCTGGCATT	GCTGATCGTA	TGCAGAAAGA	AATCACTGCT	
Paropeas achatinaceum	TCTGGTGGGT	CCACAATGTT	CCCAGGCATA	GCTGACCGCA	TGCAGAAAGA	GATAACTGCT	
Riebeckia sp.	TCYGGWGGST	CCACCATGTT	CCCTGGSATC	GCTGACCGCA	TGCAGAAAGGA	AATCACYKCT	
Rumina decollata	TCTGGTGGNT	CAACAATGTT	CCCAGGCATC	GCTGACCGCA	TGCAGAAAGA	AATCACCAMT	
Subulina octona	TCTGGCGGTT	CTACAATGTT	CCCTGGCATT	GCCGATCGCA	TGCAAAAGGA	AATCACGGCT	
Subulina striatella	TCTGGCGGTT	CWACAATGTT	CCCTGGCATT	GCCGACCGCA	TGCAGAAAGGA	AATCACAGCT	
Subulina vitrea	TCTGGTGGTT	CYACAATGTT	YCCTGGCATY	GCCGACCGCA	TGCARAAGGA	AATYRTCAAT	
Subulona sp.	TCTGGAGGGT	CCACGATGTT	CCCAGGCATC	GCTGACCGAA	TGCAGAAAGGA	AATTACTTCT	
Tortaxis erectus	TCTGGWGGTT	CTACCATGTW	CCCTGGARTT	GCTGAYCGCA	TGCAGAAAGGA	AATCACAGMT	
Xerocerastus sp.	TCTGGTGGCT	CAACAATGTT	CCCAGGCATC	GCTGACCGCA	TGCAGAAAGA	AATCACCAGC	
Zootecus insularis	TCTGGTGGTT	CAACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAAGA	AATCACCNAAT	
Coeliaxis blandii	TCTGGTGGTW	CCWCAATGTT	CCCYGGTATY	GCTGACCGCA	TGCAGARGGA	AATCGTGAAC	
Pyrquina umbilicata	TCTGGWGGTT	CCACAATGTT	CCCTGGAATC	GCTGACCGCA	TGCAGAAAGA	AATCACTKCY	
Glessula ceylanica	TCTGGAGGAT	CCACAATGTW	TCCTGGAATT	GCTGACCGCA	TGCAGAAAGGA	AATCACMGCS	
Ceciloides gokweanus	TCTGGTGGCT	CCACCATGTT	CCCCGGCATC	GCTGACCGCA	TGCAGAAAGA	AATTACCTGC	
Ferussacia folliculus	TCTGGAGGTT	CAACAATGTT	CCCTGGCATC	GCTGACCGCA	TGCAGAAAGA	GATCACGCTT	
Thyrophorella thomensis	TCTGGWGGTT	CCACAATGTW	CCCTGGAATC	GCTGACCGCA	TGCAGAAAGA	AATCACTKCT	
Gibbulinella dewinteri	TCTGGTGGGT	CTACCCATGTT	CCCAGGCATC	GCCGACCGTA	TGCAGAAAGGA	AATTACTTCC	
Gonaxis quadrilateralis	TCTGGTGGTT	CCACCATGTA	YCCCGGCATT	GCTGACCGCA	TGCAGAAAGGA	AATYACYTCC	
Gonospora sp.	TCTGGAGGTT	CTACMATGTT	CCCWGSATC	GCWGAYCGCA	TGCAAAARGA	AATYACKKSY	

	790	800	810	820	830	840	
Achatina achatina	CTGGCTCCAS	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CAGAACGTAA	ATACTCTGTC	
Achatina fulica	CTGGCTCCTT	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Achatina stuhlmanni	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC	
Cochlitoma ustulata	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Allopeas clavulinum	TTGGCTCCAC	CAACAATGAA	GATCAAGATC	ATTGCCCCAC	CAGAAAGGAA	GTACTCAGTC	
Bocageia sp.	CTTGCTCCAY	CCACAATGAA	GATCAAGATC	ATTGCTCCTC	CYAGMGGGAA	ATACTCAGTC	
Eutomopeas layardi	TTGGCTCCAC	CAACGATGAA	GATCAAGATC	ATCGCTCCAC	CAGAGAGGAA	GTACTCGGTC	
Leptinaria lamellata	CTCGCACCAC	CAACAATGAA	AATCAAAATC	ATCGCTCCAC	CTGAGAGAAA	GTACTCCGTA	
Paropeas achatinaceum	TTGGCTCCAC	CAACAATGAA	AATCAAGATC	ATTGCCCCAC	CAGAGAGAAA	GTACTCAGTC	
Riebeckia sp.	CTKGCTCCAM	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAAMGCAA	ATACTCWGTC	
Rumina decollata	CTGGCTCCAC	CCACAATGAA	GATCAAAATC	ATTGCTCCTC	CTGAGAGAAA	ATACTCTGTC	
Subulina octona	CTAGCTCCAC	CCACAATGAA	AATCAAGATA	ATTGCGCCAC	CTGAAAGGAA	ATATTTCAGTC	
Subulina striatella	CTAGCTCCAC	CCACAATGAA	AATMAAAATA	ATTGCACCAC	CTGAACGGAA	ATATTTCGTT	
Subulina vitrea	CTTGCTCCAC	CMACAATGAA	GATCAAAATC	ATCGCACCCC	CAGAACGYAA	ATACTCTGTG	
Subulona sp.	CTTGCTCCAG	CAACAATGAA	GATCAAGATA	ATAGCTCCCC	CTGAGAGGAA	GTACTCTGTT	
Tortaxis erectus	TTRGCTCCAC	CCACAATGAA	GATCAAGATC	ATAGCTCCTC	CTGAACGCAA	GTACTCTGTC	
Xerocerastus sp.	CTGGCTCCAG	CCACAATGAA	GATCAAGATC	ATTGCTCCTC	CTGAGAGGAA	ATACTCTGTC	
Zootecus insularis	CTGGCYCCAC	CCACGATGAA	RATYAAAATC	ATTGCTCCTC	CTGAGAGAAA	ATACTCTGTC	
Coeliaxis blandii	CTTGCTCCAC	CCACAATGAA	GATCAAAATC	ATTGCTCCCC	CTGAACGTAA	ATACTCAGTC	
Pyrquina umbilicata	CTTGCTCCAA	CAACRATGAA	GATCAAGATC	ATYGCTCCAC	CWGARGRAA	GTACTCAGTC	
Glessula ceylanica	CTGGCWCCAA	SCACCATGAA	AATCAAGATC	ATTGCTCCCC	CAGARCGGAA	GTACTCTGTC	
Ceciloides gokweanus	TTGGCTCCAC	CCACAATGAA	GATCAAGATC	ATCGCTCCCC	CAGAGAGGAA	ATACTCAGTC	
Ferussacia folliculus	TTAGCCCCTG	CCACCATGAA	AATCAAGATC	ATCGCTCCAC	CAGAACGTAA	ATACTCCGTG	
Thyrophorella thomensis	CTTGCTCCAG	CAACAATGAA	GATCAAGATC	WTYGCTCCWC	CTGAARRRAA	GTACTCAGTC	
Gibbulinella dewinteri	CTCGCTCCAC	CCACAATGAA	GATCAAGATC	ATTGCTCCCC	CCGAGCGTAA	ATACTCTGTA	
Gonaxis quadrilateralis	CTKGCTCCAM	STACAATGAA	GATCAAGATY	ATTGCTCCYC	CWGAGCGNAA	ATACTCTGTA	
Gonospora sp.	YTKGCTCCAC	CCACTATGAA	RATCAAGATC	ATTGCTCCCC	CWGAGCGWAA	GTAYTCTGTC	

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      ....|....| ....|....| .
            850      860
Achatina achatina      TGGATTGGAG GTTCCATTCT G
Achatina fulica        TGGATTGGAG GATCCATCCT G
Achatina stuhlmanni    TGGATTGGAG GATCCATCCT G
Cochlitoma ustulata    TGGATTGGAG GATCCATCCT G
Allopeas clavulinum    TGGATTGGTG GCTCCATCTT G
Bocageia sp.           TGGATTGGAG GCTCCATCTT G
Eutomopeas layardi     TGGATCGGTG GCTCCATCTT G
Leptinaria lamellata   TGGATTGGTG GTTCAATCTT A
Paropeas achatinaceum  TGGATCGGTG GCTCCATCTT G
Riebeckia sp.          TGGATTGGAG GCTCCATCCT G
Rumina decollata       TGGATTGGAG GTTCCATCCT G
Subulina octona        TGGATTGGAG GCTCCATTTT G
Subulina striatella    TGGATTGGAG GCTCTATTTT G
Subulina vitrea        TGGATTGGAG GTTCCATCCT G
Subulona sp.           TGGATTGGTG GCTCCATCTT G
Tortaxis erectus       TGGATTGGAG GTTCCATCCT G
Xerocerastus sp.       TGGATTGGAG GTTCCATCCT G
Zootecus insularis     TGGATTGGAG GTTCCATCCT G
Coeliaxis blandii      TGGATTGGAG GTTCCATCCT G
Pyrgina umbilicata     TGGATCGGAG GTCCCATYTT G
Glessula ceylanica     TGGATTGGAG GTTCCATCCT G
Cecilioides gokweanus  TGGATCGGAG GCTCAATCCT G
Ferussacia folliculus  TGGATTGGTG GCTCCATCCT G
Thyrophorella thomensis TGGATCGGAG GTTCCATCTT G
Gibbulinella dewinteri TGGATCGGAG GCTCCATCTT G
Gonaxis quadrilateralis TGGATCGGAG GCTCCATCTT G
Gonospira sp.          TGGATCGGAG GTTCCATTTT G

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Appendix 3.3: Alignment of the histone 3 gene for the Achatinoidea and three streptaxid outgroup taxa. Note that sequencing was unsuccessful for the achatinoids *Ceciloides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilateralis* and *Gonospora* sp.

	10 20 30 40 50 60
<i>Achatina achatina</i>	TCGTAAATCC ACCGGAGGCA AGGCTCCCCG CAAGCARCTG GCCACCAAGG CKGCCAGRAA
<i>Achatina fulica</i>	TCGTAAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CCGCTAGAAA
<i>Achatina stuhlmanni</i>	TCGCAAATCC ACCGGAGGTA AGGCTCCTCG CAAACAGCTT GCCACCAAGG CCGCTAGAAA
<i>Cochlitoma ustulata</i>	TCGCAAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CCGCTAGGAA
<i>Allopeas clavulinum</i>	TCGTAAATCY ACCGGAGGCA ARGCTCCCCG CAARCARCTG GCCACCAARG CNGCCAGGAA
<i>Bocageia</i> sp.	TCGCAAGTCC ACCGGTGGTA AAGCACCCCG CAAACAGCTT GCCACAAAGG CCGCTAGAAA
<i>Eutomepeas layardi</i>	TCGCAAATCT ACCGGAGGCA AAGCACCCCG CAAGCAGCTT GCCACCAAGG CCGCTAGGAA
<i>Leptinaria lamellata</i>	CCGAAAATCT ACTGGTGGTA AAGCACCCCG CAAGCAACTT GCCACCAAGG CTGCTAGAAA
<i>Rumina decollata</i>	TCGCAAATCC ACAGGAGGAA AAGCTCCCCG CAAACAGCTT GCCACTAAGG CCGCTAGAAA
<i>Subulina octona</i>	CCGTAAATCA ACTGGAGGAA AGGCACCACG CAAACAATG GCCACTAAGG CAGCCAGAAA
<i>Subulina striatella</i>	CCGTAAATCA ACTGGAGGCA AGGCCCCGCG CAAACAGCTG GCCACTAAGG CCGCTAGAAA
<i>Subulona</i> sp.	GCGAAAGTCT ACCGGTGGCA AAGCTCCCCG CAAGCAGCTA GCCACCAAGG CCGCTAGAAA
<i>Tortaxis erectus</i>	CCGCAAATCT ACCGGAGGAA AGGCCCTCG TAAACAGCTT GCCACCAAGG CTGCTAGAAA
<i>Xerocerastus</i> sp.	TCGYAAATCY ACCGGAGGYA ARGCTCCCCG CAARCARCTK GCCACCAARG CRGCIYAGRAA
<i>Zootecus insularis</i>	CCGCAAGTCC ACCGGAGGCA AGGCTCCACG TAAACAGCTT GCCACCAAGG CCGCTAGAAA
<i>Coelioxaxis blandii</i>	TCGYAAATYY ACCGGAGGCA AGGCTCCYCG CAARCARCTK GCCACCAARG CCGCIYAGRAA
<i>Pyrquina umbilicata</i>	TCGTAAATCT ACCGGAGGCA AAGCACCCCG AAAGCAGCTT GCCACCAARG CCGCTAGGAA
<i>Glessula ceylanica</i>	CAGGAAATCA ACCGGAGGCA AGGCTCCGCG TAAACAGCTT GCCACCAAGG CTGCCAGAAA
<i>Ferussacia folliculus</i>	TCGTAAGTCT ACTGGTGGTA AGGCGCCACG CAAACAGTTG GCCACCAAGG CAGCCAGGAA
<i>Thyrophorella thomensis</i>	TCGTAAATCT ACCGGAGGCA AAGCACCCCG AAAGCAGCTT GCCACAAAGG CCGCTAGGAA
<i>Gibbulinella dewinteri</i>	ACGTAAATCC ACCGGAGGCA AGGCTCCACG AAAGCAGCTG GCCACCAAGG CAGCAAGGAA

	70 80 90 100 110 120
<i>Achatina achatina</i>	RTCGGCCCCR GCCACMGGRG GTGTSAAAGAA RCCCCATCGW TACAGGCCCG GCACNGTSGC
<i>Achatina fulica</i>	GTGCGCCCCG GCCACAGGAG GTGTCAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCGC
<i>Achatina stuhlmanni</i>	GTGCGCCCCA GCCACAGGAG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACAGTCGC
<i>Cochlitoma ustulata</i>	GTGCGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCGC
<i>Allopeas clavulinum</i>	GTGCGGCCCR GCCACCGGWG GTGTSAAAGAA ACCCCATCGT TACAGGCCCG GCACCGTGGC
<i>Bocageia</i> sp.	ATCGGCCCCA GCCACAGGAG GTGTGAAGAA ACCTCACCGA TACAGGCCCG GTACAGTGGC
<i>Eutomepeas layardi</i>	GTGCGGCCCA GCTACCGGAG GTGTCAAGAA ACCTCACAGA TACAGGCCCG GCACAGTCGC
<i>Leptinaria lamellata</i>	GTGCGGCCCG GCTACTGGGG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACTGTTCGC
<i>Subulina octona</i>	GTCTGCCCCA GCTACAGGAG GTGTCAAGAA RCCGCATCGT TACAGGCCCG GCACTGTTCGC
<i>Subulina striatella</i>	GTCTGCCCCA GCTACAGGAG GTGTCAAGAA GCCCCATCGT TACAGGCCCG GCACTGTTCGC
<i>Subulona</i> sp.	GTGCGGCCCA GCGACTGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCAG GCACCGTGGC
<i>Tortaxis erectus</i>	GTGCGGTCCG GCCACAGGAG GTGTGAAGAA GCCCCACAGG TACAGGCCCG GTACAGTCGC
<i>Xerocerastus</i> sp.	RTCKGCCCCR GCCACCGGWG GTGTSAAAGAA ACCCCAYCGW TACAGGCCCG GCACCGTSGC
<i>Zootecus insularis</i>	GTGCGGCCCG GCCACAGGAG GTGTGAAGAA ACCCCACCGA TACAGACCTG GCACCGTTCGC
<i>Coelioxaxis blandii</i>	GTCKGCCCCR GCCACCGGWG GTGTSAAAGAA RCCYCAYMGA TACMGGCCSG GCACNGTSGC
<i>Pyrquina umbilicata</i>	ATCCGCCCCA GCCACCGGAG GTGTCAAGAA ACCTCACAGA TACAGGCCCG GCACTGTTCGC
<i>Glessula ceylanica</i>	GTGCGGCCCG GCCACAGGAG GCGTGAAGAA ACCTCACAGR TACAGGCCCG GGACAGTCGC
<i>Ferussacia folliculus</i>	GTCCGCCCCA GCCACTGGTG GTGTGAAGAA ACCTCACCGT TACAGGCCCG GTACAGTTGC
<i>Thyrophorella thomensis</i>	ATCCGCCCCA GCCACCGGAG GTGTCAAGAA ACCTCACAGA TACAGGCCCG GCACTGTTCGC
<i>Gibbulinella dewinteri</i>	ATCTGCCCCA GCAACCGGTG GTGTGAAGAA GCCCCATCGC TACAGGCCAG GCACTGTTCGC

	
	130 140 150 160 170 180	
Achatina achatina	TCTKCGAGAR ATCCGTCGTT ACCAGAAGAG CACKGAGCTC CTCATCCGCA AGCTGCCCTT	
Achatina fulica	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACTGAGCTT CTCATCCGCA AACTGCCCTT	
Achatina stuhlmanni	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCATT	
Cochlitoma ustulata	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATTCGCA AACTGCCCTT	
Allopeas clavulinum	TCTGCGAGAR ATCCGTCGTT ACCAGAAGAG CACTGAGCTS CTCATCCGCA AGCTGCCCTT	
Bocageia sp.	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTC CTCATCCGTA AACTGCCCTT	
Eutomopeas layardi	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTC CTCATCCGCA AGCTGCCCTT	
Leptinaria lamellata	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAACCT CTCATCCGCA AGCTGCCCTT	
Rumina decollata	GCTTCGAGAG ATCCGACGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA AGCTGCCCTT	
Subulina octona	ACTTCGAGAG ATCCGCCGTT ATCAGAAGAG CACGGAGCTT CTCATCCGCA AGCTGCCCTT	
Subulina striatella	ACTTCGAGAG ATCCGCCGTT ACCAGAAGAG CACGGAGCTC CTCATCCGCA AGCTGCCCTT	
Subulona sp.	ACTCCGAGAG ATCCGCCGTT ACCAGAAGAG CACCGAGCTT CTCATCCGCA AACTGCCCTT	
Tortaxis erectus	ACTCAGAGAG ATCCGCCGTT ACCAGAAGAG CACAGAGCTG CTCATCCGCA AATTGCCCTT	
Xerocerastus sp.	TCTGCGAGAG ATCCGTCGTT ACCAGAAGAG CACTGAGCTK CTCATCCGCA AGCTGCCCTT	
Zootecus insularis	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA AACTGCCCTT	
Coeliaxis blandii	TCTKCGAGAG ATCCGTCGTT ACCAGAAGAG CACKGAGCTN CTCATCCGCA ARCTGCCCTT	
Pyrgina umbilicata	ACTCCGAGAG ATCCGTCGTT ACCARAAGAG CACGGAGCTM CTCATCCGCA AGCTGCCCTT	
Glessula ceylanica	ACTTCGAGAG ATCCGCCGTT ACCAGAAGAG CACGGAGCTG CTCATCCGCA AACTGCCCTT	
Ferussacia folliculus	ACTCCGTGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTG CTCATCCGCA AACTGCCCTT	
Thyrophorella thomensis	ACTCCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AGCTGCCCTT	
Gibbulinella dewinteri	TCTTCGTGAG ATTCGTCGTT ATCAGAAAAG CACGGAACCT CTCATCCGCA AACTGCCCTT	

	
	190 200 210 220 230 240	
Achatina achatina	CCAGCGCCTG GTCAGAGAAA TCGCCCAGGA CTTCAAGACY GACCTGCGCT TCCAGAGCTC	
Achatina fulica	CCAGCGGCTG GTCAGAGAGA TCGCCCAGGA CTTTAAGACC GATCTGCGTT TCCAGAGCTC	
Achatina stuhlmanni	CCAGCGGCTG GTCAGAGAGA TCGCACAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC	
Cochlitoma ustulata	CCAGCGGCTG GTCAGAGAGA TCGCCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC	
Allopeas clavulinum	CCAGCGCCTG GTSMGAGAAA TYGCYCAGGA CTTCAAGACY GACCTGCGYT TCCAGAGCTC	
Bocageia sp.	CCAGCGCCTG GTCAGAGAAA TCGCCCAGGA TTTCAAACC GACCTGCGTT TCCAGAGCTC	
Eutomopeas layardi	CCAGCGTCTG GTCAGAGAGA TCGCCCAGGA CTTTAAGACA GATCTGCGTT TCCAGAGCTC	
Leptinaria lamellata	CCAGCGCCTG GTCAGAGAAA TCGCTCAGGA CTTTAAACA GATCTGCGTT TCCAAAGTTC	
Rumina decollata	CCAGCGCCTT GTCAGAGAGA TCGCGCAGGA TTTCAAGACT GACCTGCGTT TTCAGAGCTC	
Subulina octona	CCAGCGCCTG GTCAGAGAAA TCGCCCAGGA TTTCAAGACG GACCTGCGCT TCCAGAGCTC	
Subulina striatella	CCAGCGACTG GTCAGAGAAA TCGCCCAGGA TTTCAAGACC GACCTGCGCT TCCAGAGCTC	
Subulona sp.	CCAGCGACTG GTGAGAGAAA TCGCCCAGGA CTTCAAGACC GATCTTCGAT TCCAGAGCTC	
Tortaxis erectus	CCAGCGCCTT GTCAGGAGGA TCGCACAGGA CTTTAAGACT GACCTGCGTT TCCAGAGCTC	
Xerocerastus sp.	CCAGCGCCTG GTSMGAGARA TYGCYCAGGA YTTCAAGACY GAYCTGCGYT TCCAGAGCTC	
Zootecus insularis	CCAGCGCCTG GTCAGAGAGA TCGCGCAGGA TTTCAAGACT GATCTGCGCT TCCAGAGCTC	
Coeliaxis blandii	CCARCGCCTG GTSMGAGAAA TCGCYCAGGA YTTCAAGACY GAYCTGCGYT TCCAGAGCTC	
Pyrgina umbilicata	CCAGCGCCTG GTCAGAGAAA TCGCTCAGGA CTTCAAGACC GATCTGCGCT TCCAGAGCTC	
Glessula ceylanica	CCAGCGCCTG GTCAGAGAGA TTGCCAGGA CTTCAAGACC GACCTGCGTT TCCAGAGCTC	
Ferussacia folliculus	CCAGCGACTG GTCAGAGAAA TCGCACAGGA CTTCAAGACT GACCTGCGCT TCCAAAGCTC	
Thyrophorella thomensis	CCAGCGCCTG GTCAGAGAAA TCGCTCAGGA CTTCAAGACC GATCTGCGCT TCCAAAGCTC	
Gibbulinella dewinteri	CCAACGCTTG GTAAGAGAAA TTGCACAGGA CTTCAAGACA GATCTGCGCT TCCAGAGCTC	

	
	250 260 270 280 290 300	
Achatina achatina	GGCYGTCATG GCCCTGCAAG AGGCTAGCGA RGCCTACCTG GTGGGTCTGT TYGAGGACAC	
Achatina fulica	GGCTGTCATG GCACTGCAAG AGGCTAGCGA GGCCTACCTG GTGGGTCTGT TTAGAGGACAC	
Achatina stuhlmanni	GGCTGTCATG GCACTGCAAG AGGCTAGCGA GGCCTACCTG GTGGGTCTGT TTAGAGGACAC	
Cochlitoma ustulata	GGCTGTCATG GCACTGCAAG AGGCTAGCGA GGCCTACCTG GTGGGTCTGT TTAGAGGACAC	
Allopeas clavulinum	SGCTGTCATG GCYCTGCARG AGGCCAGCGA RGCCTACCTG GTSGGYCTST TTAGAGGACAC	
Bocageia sp.	GGCTGTCATG GCTCTGCAAG AGGCTAGCGA GGCCTACCTG GTCGGTCTGT TTAGAGGACAC	
Eutomopeas layardi	GGCTGTCATG GCACTGCAGG AAGCCAGCGA GGCCTACCTC GTAGGTCTGT TTGAAGACAC	
Leptinaria lamellata	AGCTGTCATG GCTCTGCAGG AGGCTAGCGA GGCCTACCTT GTGGGTCTGT TCGAAGACAC	
Rumina decollata	RGCYGTCATG GCACTGCAGG AGGCTAGCGA GGCMTACYTR GTGGGCTGT TTAGAGGACAC	
Subulina octona	GGCTGTCATG GCGCTGCAGG AGGCCAGCGA GGCCTACCTC GTAGGTCTGT TTAGAGGACAC	
Subulina striatella	GGCTGTCATG GCGCTGCAGG AGGCCAGCGA GGCCTACCTG GTAGGTCTGT TTAGAGGACAC	
Subulona sp.	TGCTGTCATG GCACTGCAAG AGGCTAGCGA GGCTTACCTG GTCGGCTGT TCGAGGACAC	
Tortaxis erectus	GGCCGTCATG GCTCTGCAGG AAGCAAGCGA GGCCTATCTG GTGGGTCTGT TCGAGGACAC	
Xerocerastus sp.	SGCTGTCATG GYCTGCARG AGGCTAGCGA RGCCTACCTG GTSGGYCTST TTAGAGGACAC	
Zootecus insularis	GGCTGTCATG GCACTGCAGG AGGCTAGCGA GGCCTACCTG GTGGGTCTGT TTAGAGGACAC	
Coeliaxis blandii	SGCTGTCATG GCYCTGCAAG AGGCTAGCGA RGCCTACCTG GTSGGTCTST TTAGAGGACAC	
Pyrgina umbilicata	GGCTGTCATG GCACTGCAGG AAGCCAGCGA GGCCTACCTC GTAGGTCTGT TTGAAGACAC	
Glessula ceylanica	AGCTGTCATG GCACTGCAGG AAGCCAGCGA AGCCTAYTTG GTGGGTTTGT TCGAGGACAC	
Ferussacia folliculus	SGCCGTCATG GCCCTGCAAG AGGCAAGYGA GGCCTACCTT GTGGGTCTGT TTGAAGACAC	
Thyrophorella thomensis	GGCTGTCATG GCACTGCAGG AAGCCAGCGA GGCCTACCTC GTAGGTCTGT TTGAAGACAC	
Gibbulinella dewinteri	RGCCGTCATG GCATTGCAGG AGGCCAGCGA GGCTTACCTC GTGGGTCTCT TTAGAGGACAC	

	310 320
Achatina achatina	YAACYTGTGC GCYATCCACG CCAARCGT
Achatina fulica	CAACCTGTGC GCCATCCACG CCAAACGT
Achatina stuhlmanni	CAACCTGTGC GCCATYCACG CCAAACGT
Cochlitoma ustulata	CAACCTGTGC GCCATCCACG CCAAACGT
Allopeas clavulinum	CAACYTGTGC GCCATCCAYG CCAAGCGT
Bocageia sp.	CAACCTGTGC GCCATCCATG CCAAGCGG
Eutomopeas layardi	CAATCTGTGC GCAATTCACG CCAAGCGT
Leptinaria lamellata	CAACCTGTGC GCCATCCATG CCAAACGT
Rumina decollata	CAACCTGTGC GCCATCCACG CCAAACGT
Subulina octona	CAACCTGTGC GCCATCCACG CCAAGCGT
Subulina striatella	CAACCTGTGC GCCATCCACG CCAAACGT
Subulona sp.	CAACCTGTGC GCCATCCATG CCAAGCGC
Tortaxis erectus	AAACCTGTGC GCCATCCACG CCAAGCGA
Xerocerastus sp.	CAACTTGTGC GCCATCCAYG CCAARCGT
Zootecus insularis	CAACCTGTGC GCCATCCACG CCAAACGT
Coelioxaxis blandii	CAACYTGTGC GCYATCCAYG CYAAGCGY
Pyrgina umbilicata	CAACCTGTGC GCCATCCATG CCAAGCGT
Glessula ceylanica	CAACCTGTGC GCCATCCACG CCAAACGT
Ferussacia folliculus	CAACCTGTGT GCCATCCACG CCAAGCGT
Thyrophorella thomensis	TAACCTGTGC GCCATCCATG CCAAGCGT
Gibbulinella dewinteri	CAACCTGTGC GCTATCCACG CCAAGCGT

Appendix 3.4: Alignment of the CO1 gene for the Achatinoidea and three streptaxid outgroup taxa. Note that sequencing was unsuccessful for *Ceciloides gokweanus* as well as the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp.

	10	20	30	40	50	60
Achatina achatina	AGGTACAGGC	CTATCATTGT	TAATTCGATT	AGAATTAGGG	ACAGCTGGGG	TTTTAACCGA	
Achatina fulica	TGGCACAGGC	TTGTCACCTCT	TAATTCGGTT	AGAGCTTGGA	ACAGTGGGAA	CCTTAACCTGA	
Achatina stuhlmanni	TGGTACCGGC	TTGTCCTTTAC	TTATCCGGCT	AGAACTAGGG	AGTATTGGAA	CACTGACTGA	
Cochlitoma ustulata	GGGCACAGGT	CTATCTTTAT	TGATCCGACT	AGAGCTAGGA	ACGACCTCTA	CTTTAACTGA	
Coeliaxis blandii	TGGGACTGGG	TTATCTTTAC	TTATTCGTTT	AGAATTAGGT	ACTGTGGAG	TCTTAACCTGA	
Pyrgina umbilicata	CGGGACTGGT	CTATCTTTAT	TAATCCGTTT	AGAACTCGGA	ATAGGGGGCG	TGTTAATTGA	
Glessula ceylanica	CGGTACAGGA	TTATCATTAT	TAATTCGGTT	AGAATTAGGA	ACAACCTGGT	TATTGACGGA	
Ferussacia folliculus	TGGAACGGC	TTATCTTTGC	TTATTCGTTT	AGAATTAGGT	ACATCGGGCG	TCTTAACAGA	
Thyrophorella thomensis	TGGAACAGGT	TTGTCATTAC	TGATCCGTCT	GGAGTTAGGA	ACAGCCGGTG	TACTAGTTGA	
Allopeas clavulinum	TGGGACAGGA	TTATCTCTTC	TCATTCGTTT	TGAGTTAGGT	ACTAGTGGGG	TGTTAACCGA	
Bocageia sp.	AGGTACCGGC	TTATCCTTAT	TAATTCGTCT	GGAATTAGGA	ACCTCTGGTG	TTTTGACTGA	
Eutomopeas layardi	GGGGACAGGT	TTATCCTTAC	TAATCCGTTT	AGAATTAGGA	ACTAGCGGAG	TCTTAACCTGA	
Leptinaria lamellata	AGGTACAGGT	TTATCGTTAT	TAATTCGTTT	AGAATTAGGG	ACAATGGAAG	TGTTAACAGA	
Paropeas achatinaceum	TGGGACTGGT	TTATCTTTAC	TAATTCGTCT	TGAGTTAGGA	ACTAGGGGAG	TGTTAACCGA	
Riebeckia sp.	TGGGACTGGT	TTATCTTTAT	TAATTCGATT	AGAGTTAGGT	ACTTCAGGTG	TATTAACCTGA	
Rumina decollata	TGGTACTGGT	TTATCTCTTC	TTATTCGTTT	AGAACTAGGT	ACAGCAGGAG	TGTTAACAGA	
Subulina octona	TGGGACTGGT	TTATCATTAT	TAATTCGTTT	AGAGCTTGGG	ACGGCTGGAG	TGTTAACTGA	
Subulina striatella	TGGGACGGGT	TTGTCATTAT	TAATTCGACT	TGAGTTAGGT	ACTGCTGGTG	TTTTAACCGA	
Subulina vitrea	TGGTACAGGG	CTTTCCTTTT	TAATTCGTAT	TGAGCTTGGA	ACTGCTGGGG	TCTTAACTGA	
Subulona sp.	AGGGACAGGA	TTATCTTTAT	TAATTAGTTT	AGAATTAGGG	ACTGTAAGGG	TATTAGTAGA	
Tortaxis erectus	TGGGACAGGG	TTGTCCTTAT	TGATTCGTCT	TGAACCTAGGT	ACTTCTGGTG	TATTAACAGA	
Xerocerastus sp.	TGGTACTGGA	CTATCTTTAT	TAATTCGGCT	TGAGTTAGGA	ACAGCTGGTG	TTTTAACTGA	
Zootecus insularis	TGGGACAGGG	TTGTCGTTAC	TAATTCGGCT	TGAATTAGGT	ACCTCAGGTG	TGTTAACAGA	
Gibbulinella dewinteri	TGGGACCGGT	TTGTCCTTAT	TAATTCGGCT	AGAATTAGGT	ACAGCTGGGG	TTCTTATTGA	
	70	80	90	100	110	120
Achatina achatina	TGATCATTTT	TTTAATGTAG	TGGTGACAGC	CCATGCTTTT	GTTATAATTT	TTTTTATAGT	
Achatina fulica	TGATCACTTT	TTCAACGTGG	TTGTAACCTGC	GCATGCTTTT	GTCATAATTT	TTTTTATGGT	
Achatina stuhlmanni	CGATCATTTT	TTTAATGTTG	TAGTTACTGC	TCATGCTTTT	GTGATAATTT	TTTTTATAGT	
Cochlitoma ustulata	TGATCATTTT	TTTAATGTAA	TCGTGACAGC	GCACGCTTTT	GTAATAATTT	TTTTCATAGT	
Coeliaxis blandii	TGATCATTTT	TTTAATGTGG	TTGTAACCTGC	TCATGCTTTT	GTAATAATTT	TTTTTATAGT	
Pyrgina umbilicata	TGACCACTTT	TTCAATGTTG	TTGTAACAGC	TCATGCGTTT	GTAATAATTT	TTTTTATAGT	
Glessula ceylanica	CGATCATTTT	TTTAATGTAA	TTGTTACCGC	TCATGCTTTT	GTTATAATTT	TTTTTATAGT	
Ferussacia folliculus	TGATCACTTT	TTTAATGTAA	TTGTTACTGC	ACACGCTTTT	GTTATGATTT	TTTTTATGGT	
Thyrophorella thomensis	TGATCACTTT	TTTAATGTGG	TCGTACAGC	CCACGCTTTT	GTTATAATCT	TTTTTATAGT	
Allopeas clavulinum	TGACCACTTT	TTTAATGTTG	TAGTTACAGC	TCATGCTTTC	GTAATAATTT	TTTTTATAGT	
Bocageia sp.	CGATCACTTT	TTTAACGTTG	TAGTAACAGC	TCATGCGTTT	GTTATAATTT	TCTTTATAGT	
Eutomopeas layardi	TGATCACTTT	TTTAATGTTG	TAGTAACAGC	TCACGCTTTC	GTGATGATTT	TTTTTATAGT	
Leptinaria lamellata	TGATCATTTT	TTTAACGTAG	TGGTTACTGC	TCACGCTTTT	GTAATAATTT	TTTTTATAGT	
Paropeas achatinaceum	TGATCATTTT	TTTAATGTTG	TAGTAACCTGC	TCATGCATT	GTAATAATTT	TTTTTATAGT	
Riebeckia sp.	TGATCACTTT	TTTAATGTAA	TTGTAACCGC	ACATGCTTTT	GTTATAATTT	TTTTTATAGT	
Rumina decollata	TGACCACTTT	TTTAATGTGG	TAGTTACTGC	CCATGCATT	GTGATAATTT	TTTTTATAGT	
Subulina octona	TGATCATTTT	TTTAATGTAA	TTGTAACCTGC	TCATGCATT	GTTATGATTT	TTTTTATGGT	
Subulina striatella	TGACCACTTT	TTTAATGTAA	TTGTGACAGC	TCATGCTTTT	GTTATAATTT	TTTTTATAGT	
Subulina vitrea	CGATCATTTT	TTTAATGTAA	TTGTGACTGC	TCATGCTTTT	GTTATAATTT	TTTTTATAGT	
Subulona sp.	CGATCATTTT	TTTAACGTAA	TTGTTACTGC	TCATGCTTTT	GTAATAATTT	TCTTTATAGT	
Tortaxis erectus	TGATCATTTT	TTTAATGTTG	TAGTTACAGC	CCATGCTTTT	GTTATGATTT	TTTTTATAGT	
Xerocerastus sp.	TGATCACTTT	TATAATGTTG	TTGTTACGGC	TCATGCTTTT	GTAATAATTT	TTTTTATAGT	
Zootecus insularis	CGACCATTTT	TTTAATGTAA	TTGTAACAGC	TCATGCATT	GTAATAATTT	TTTTTATAGT	
Gibbulinella dewinteri	TGATCATTTT	TTTAAGTTAA	TTGTTACWGC	TCACGCTTTT	GTTATAATTT	TTTTTATAGT	

	130	140	150	160	170	180	
<i>Achatina achatina</i>	CATACCAATT	ATGATTGGGG	GTTTGGGTAA	TTGAATAGTC	CCACTATTAA	TTGGGGCTCC	
<i>Achatina fulica</i>	TATACCAATT	ATAATTGGCG	GATTTGGAAA	CTGGATGGTC	CCAATACTTA	TTGGTGCTCC	
<i>Achatina stuhlmanni</i>	GATACCAATT	ATAATTGGTG	GGTTTGGAAA	CTGGATGGTC	CCATTACTGA	TTGGTGCTCC	
<i>Cochlitoma ustulata</i>	AATACCCATT	ATAATTGGGG	GTTTGGGTAA	TTGAATAACT	CCCTTTTAA	TCGGGGCTCC	
<i>Coelioxaxis blandii</i>	TATACCTATT	ATAATTGGGG	GATTTGGAAA	TTGAATGGTT	CCATTATTAA	TTGGAGCTCC	
<i>Pyrgina umbilicata</i>	TATACCAATT	ATGATTGGTG	GGTTTGGGAA	TTGAATAGTG	CCGCTTTTAA	TTGGTGCTCC	
<i>Glessula ceylanica</i>	TATGCCTATT	ATAATTGGAG	GATTTGGTAA	TTGATAGTT	CCGTTATTAA	TTGGTGCTCC	
<i>Ferussacia folliculus</i>	TATACCTATT	ATAATTGGAG	GGTTTGGAAA	TTGAATGGTG	CCCTTTTGA	TTGGTGCTCC	
<i>Thyrophorella thomensis</i>	TATACCAATT	ATAATTGGTG	GGTTTGGTAA	TTGAATAGTA	CCACTTTTAA	TTGGTGCTCC	
<i>Allopeas clavulinum</i>	TATGCCTATT	ATAATTGGTG	GCTTTGGGAA	CTGAATGGTC	CCACTATTAA	TTGGGGCTCC	
<i>Bocageia sp.</i>	AATACCTATC	ATAATTGGTG	GGTTTGGGAA	CTGAATAGTT	CCACTATTAA	TTGGAGCTCC	
<i>Eutomepeas layardi</i>	TATACCAATC	ATGATTGGGG	GGTTTGGAAA	CTGAATAGTC	CCATTACTGA	TTGGGGCCCC	
<i>Leptinaria lamellata</i>	TATACCTATT	ATAATTGGGG	GATTTGGAAA	TTGAATAGTG	CCTTTACTAA	TTGGTGACCC	
<i>Paropeas achatinaceum</i>	TATACCAATT	ATAATTGGAG	GTTTGGGAA	TTGATAGTC	CCCTTATTAA	TTGGTGCTCC	
<i>Riebeckia sp.</i>	TATACCAATT	ATAATTGGAG	GATTTGGAAA	TTGAATAGTC	CCTTTACTAA	TTGGGGCCCC	
<i>Rumina decollata</i>	AATGCCTATT	ATAATCGGTG	GTTTGGGAAA	TTGAATAGTG	CCCTATTAA	TTGGGGCACC	
<i>Subulina octona</i>	AATACCTATT	ATAATTGGAG	GATTTGGGAA	TTGAATAGTT	CCTTTGCTTA	TTGGGGCTCC	
<i>Subulina striatella</i>	CATACCTATT	ATGATTGGAG	GGTTTGGGAA	TTGATAGTT	CCTTTACTGA	TTGGTGCTCC	
<i>Subulina vitrea</i>	TATACCTATT	ATAATTGGGG	GGTTTGGGAA	TTGATAGTA	CCACTACTAA	TTGGTGCGCC	
<i>Subulona sp.</i>	AATACCTATT	ATGATTGGGG	GATTTGGAAA	TTGAATAGTG	CCTTTACTTA	TTGGGGCTCC	
<i>Tortaxis erectus</i>	AATGCCTATT	ATAATTGGTG	GTTTGGGTAA	TTGAATAGTT	CCACTTTTAA	TTGGTGCTCC	
<i>Xerocerastus sp.</i>	TATACCAATT	ATAATTGGTG	GATTTGGTAA	TTGAATAGTT	CCATTATTAA	TTGGTGCTCC	
<i>Zootecus insularis</i>	TATGCCCATC	ATAATTGGCG	GCTTCGGAAA	TTGAATGGTC	CCACTATTAA	TTGGTGCTCC	
<i>Gibbulinella dewinteri</i>	TATACCACTA	ATAATTGGAG	GTTTGGGAAA	TTGAATAGTA	CCTCTTCTTA	TTGGGGCTCC	

	190	200	210	220	230	240	
<i>Achatina achatina</i>	AGATATAAGA	TTCCCTCGGA	TGAATAATAT	AAGGTTCTGA	TTATTACCAC	CTTCATTTAC	
<i>Achatina fulica</i>	TGATATAAGA	TTTCCACGAA	TAAATAATAT	AAGGTTTGA	CTTTTACCAC	CTTCATTTAC	
<i>Achatina stuhlmanni</i>	TGATATAAGG	TTTCCGCGGA	TAAATAACAT	AAGATTTTGA	CTACTTCCCC	CATCATTTTC	
<i>Cochlitoma ustulata</i>	TGATATAAGC	TTTCTCGTA	TAAATAACAT	GAGCTTTTGA	TTGTTACCCC	CTTCCTTTAC	
<i>Coelioxaxis blandii</i>	TGATATAAGA	TTTCCCCGAA	TAAATAATAT	AAGATTTTGG	CTTTTACCTC	CATCTTTTAT	
<i>Pyrgina umbilicata</i>	TGATATAAGT	TTCCCACGAA	TAAATAATAT	AAGATTTTGA	CTTTTACCTC	CATCTTTTAT	
<i>Glessula ceylanica</i>	AGATATAAGA	TTCCCCCGAA	TAAATAATAT	AAGATTTTGG	TTATTACCAC	CATCTTTTGT	
<i>Ferussacia folliculus</i>	TGATATAAGG	TTTCTCGTA	TAAATAATAT	GAGGTTTGA	CTTCTCCCGC	CTTCCTTTAT	
<i>Thyrophorella thomensis</i>	TGATATGAGT	TTTCCCCGTA	TAAACAACAT	AAGCTTCTGG	CTCCTTCCAC	CATCATTTAT	
<i>Allopeas clavulinum</i>	AGATATAAGG	TTTCTCGAA	TAAATAATAT	AAGTTTGTGG	CTTCTTCCAC	CATCTTTTAT	
<i>Bocageia sp.</i>	GGACATAAGT	TTTCTCGTA	TAAATAATAT	AAGATTTTGA	CTGCTTCTCT	CGTCATTTAT	
<i>Eutomepeas layardi</i>	GGATATGAGA	TTTCCCCGGA	TAAATAATAT	AAGTTTCTGG	TTACTTCCAC	CATCTTTCGT	
<i>Leptinaria lamellata</i>	AGATATAAGA	TTTCTCGAA	TAAATAATAT	AAGATTTTGG	TTATTGCCGC	CTTCCTTTAT	
<i>Paropeas achatinaceum</i>	TGATATAAGA	TTCCCTCGTA	TAAATAATAT	AAGTTTGTGA	TTACTTCCGC	CATCTTTTGT	
<i>Riebeckia sp.</i>	AGATATAAGA	TTTCTCGAA	TAAATAATAT	AAGATTTTGA	CTTTTACCTC	CATCATTTAT	
<i>Rumina decollata</i>	AGATATAAGG	TTTCTCGAA	TAAATAATAT	GAGATTTTGA	CTACTTCTCT	CTTCCTTTAT	
<i>Subulina octona</i>	TGATATAAGA	TTTCCGCGTA	TGAATAATAT	AAGGTTTGG	TTGTTACCTC	CTTCATTTTT	
<i>Subulina striatella</i>	TGACATAAGT	TTTCCCCGTA	TAAATAATAT	AAGATTTTGG	TTGCTACCAC	CTTCCTTTAT	
<i>Subulina vitrea</i>	TGACATAAGA	TTTCCACGGA	TAAATAATAT	AAGATTTTGG	CTTCTACCAC	CCTCATTTAT	
<i>Subulona sp.</i>	AGATATAAGT	TTTCCACGAA	TAAATAATAT	AAGGTTTGA	TTATTGCCTC	CTTCGTTTAT	
<i>Tortaxis erectus</i>	TGATATAAGG	TTTCCACGAA	TAAATAATAT	AAGATTTTGA	TTATTACCTC	CATCTTTTAT	
<i>Xerocerastus sp.</i>	CGATATAAGA	TTTCTCGGA	TAAATAACAT	AAGATTTTGA	TTGTTACCAC	CATCATTTAT	
<i>Zootecus insularis</i>	GGATATAAGT	TTTCTCGAA	TAAATAATAT	AAGATTTTGG	CTTCTTCCGC	CTTCCTTTAT	
<i>Gibbulinella dewinteri</i>	AGATATAAGT	TTTCCACGCA	TAAATAATAT	AAGATTTTGA	TTATTACCCC	CTTCCTTTGT	

	
	250 260 270 280 290 300	
Achatina achatina	CTTACTTATT CTTTCTAGTA TAGTTGAGGG AGGAGCTGGT ACTGGGTGAA CAGTTTATCC	
Achatina fulica	TTTGTTAATC TGTTC AAGTA TAGTGAAGG AGGGGCTGGA ACTGGGTGGA CTGTGTACCC	
Achatina stuhlmanni	TTTGCTAATT CTCTCAAGTA TAGTAGAAGG TGGGGCTGGG ACAGGATGGA CAGTTTACCC	
Cochlitoma ustulata	ACTTCTTATC TTGTCAAGAA TAGTGAAGG GGGTGCAGGT ACAGGTTGAA CTGTGTATCC	
Coeliaxis blandii	TTTTTTAATT GTCTCGAGAA TGGTAGAAGG GGGAGCCGGG ACTGGGTGAA CTGTTTACCC	
Pyrgina umbilicata	TTTCCTTATT ATGTCTAGTA TAATAGAGGG TGGAGCTGGT ACAGGTTGGA CAGTTTATCC	
Glessula ceylanica	TCTTTTAATT GTATCTAGTA TAATTGAAGG TGGTGCTGGT ACAGGGTGAA CTGTTTATCC	
Ferussacia folliculus	TCTTTTAATT ACATCTAGTT TGGTAGAAGG TGGTGCAGGG ACAGGATGGA CAGTTTACCC	
Thyrophorella thomensis	TTTCTTGATC ACGTCAAGGT TAATAGAAGG CGGAGCTGGC ACAGGTTGGA CTGTATACCC	
Allopeas clavulinum	TTTTTTAATT ATGTCAAGAA TAATAGAAGG AGGAGCAGGA ACAGGTTGAA CCGTTTATCC	
Bocageia sp.	TTTTTTAATT ATGTCTAGAA TAGTTGAAGG TGGGGCTGGA ACTGGTTGAA CTGTGTACCC	
Eutomopeas layardi	TTTTTTAATT ATGTCTAGTA TAATAGAGGG GGGGGCTGGT ACAGGCTGAA CTGTTTACCC	
Leptinaria lamellata	ATTTTTAATT ATTTCTAGTT TAGTGAAGG GGGTGCTGGA ACAGGTTGAA CAGTCTACCC	
Paropeas achatinaceum	TTTTCTAATT GTTTC AAGGA TAATAGAGGG GGGAGCCGGA ACTGGTTGAA CTGTCTACCC	
Riebeckia sp.	TCTACTTATT GTATCAAGTA TAGTAGAAGG AGGTGTAGGA ACAGGGTGAA CTGTTTACCC	
Rumina decollata	TTTATTAATT ATATCTAGGA TAGTAGAAGG TGGGGCAGGA ACTGGTTGAA CAGTGTATCC	
Subulina octona	ATTATTAATT TTATCTAGAA TAGTGGAAGG TGGTGCTGGT ACTGGTTGAA CAGTTTATCC	
Subulina striatella	TTTTTTAATT TTATCAAGCA TGGTARAAGG AGGTGCGGGG ACAGGTTGAA CCGTTTATCC	
Subulina vitrea	TTTTTTAATT GTCTCTAGGA TAGTGAAGG AGGGGCTGGA ACAGGGTGAA CAGTTTACCC	
Subulona sp.	TTTTTTAATT ATTTCTAGGT TGATTGAAGG TGGAGCCGGA ACTGGTTGAA CTGTTTACCC	
Tortaxis erectus	TTTATTAATT TTATCTAGAA TAGTTGAAGG TGGGGCTGGG ACTGGTTGGA CTGTTTACCC	
Xerocerastus sp.	TTTTATTAATT ATATCGAGGA TAGTTGAAGG GGGTGCTGGT ACTGGTTGAA CTGTTTACCC	
Zootecus insularis	TTTACTAATT GTTTC AAGTT TAGTTGAGGG GGGTGCAGGG ACAGGCTGAA CCGTATATCC	
Gibbulinella dewinteri	TTTTTTAATT ATTTCTAGAA TAGTAGAAGG TGGAGCTGGA ACAGGTTGAA CCGTATACCC	

	
	310 320 330 340 350 360	
Achatina achatina	ACCTTTAAGA TCTACTTTAG GTCATAGAGG TGCCTCGGTA GATTTAGCGA TTTTTCATT	
Achatina fulica	GCCCTTAAGT TCTTGCTTAG GACACAGAGG GGCCTCAGTT GATTTAGCTA TTTTTCCTTT	
Achatina stuhlmanni	TCCTCTAAGT TCATTAATTG GCCATAGTGG AGCATCAGTG GACTTGGCAA TTTTTCCTCT	
Cochlitoma ustulata	ACCTTTAAGG TCTATCATAG GTCATAGGGG AGCATCAGTG GATTTGGCTA TCTTCTCTCT	
Coeliaxis blandii	GCCCTTAAGT GGTATTATTG GGCACAGCGG TGCCTCAGTT GATCTGGCTA TTTTTCCTTT	
Pyrgina umbilicata	ACCCCTTAGC TCAATAATTG GCCACAGAGG TCCATCTGTT GATTTGGCAA TTTTCTCCCT	
Glessula ceylanica	ACCATTAAGA AGTATTGTTG GACATAGCGG TCGCTCTGTT GACTTAGCTA TTTTTCCTCT	
Ferussacia folliculus	CCCTTTAAGC AGATTGGTAG GCCATAGAGG TGCTTCTGTA GATTTGGCGA TTTTCTCTCT	
Thyrophorella thomensis	CCCCCTTAGA TCAATGATCG GTCATAGTGG TCCGTCTGTT GATTTAGCAA TCTTCTCTCT	
Allopeas clavulinum	ACCATTAAGA TCAATATTAG GGCACGGAGG ACCTTCTGTA GACTTAGCTA TTTTTCCTCT	
Bocageia sp.	TCCTTTAAGA AGTCTGATAG GTCATAGAGG TGCCTCCGTG GATTTGGCTA TTTTTCCTCT	
Eutomopeas layardi	ACCTTTAAGA TCAATATTAG GGCACGGTGG GCCCTCAGTA GATCTAGCTA TTTTTCCTCT	
Leptinaria lamellata	ACCTTTAAGT GGGATTTTAG GTCATAGGGG CGCTTCTGTC GACTTAGCTA TTTTTCCTCT	
Paropeas achatinaceum	ACCATTAAGG TCAATACTAG GTCATAGAGG CCCATCAGTA GACTTAGCTA TTTTTCCTCT	
Riebeckia sp.	TCCTTTAAGA GGTATTTTAG GACATAGGGG AGCATCTGTT GATTTAGCTA TCTTTCCTCT	
Rumina decollata	ACCCCTAAGC AGAATCATAG GACATAGTGG GGCATCTGTT GACTTAGCTA TTTTCTCTCT	
Subulina octona	ACCTTTAAGC AGAATTATAG GTCATGCTGG GGCCTCAGTT GATCTTGCTA TTTTTCCTTT	
Subulina striatella	CCCTCTTAGA GGAATTTTAG GACATGCTGG TGCCTCAGTG GATCTAGCTA TTTTTCCTTT	
Subulina vitrea	CCCCCTAAGA GGGATCTTAG GCCATAGTGG CGCTTCTGTT GATTTAGCCA TTTTTCCTTT	
Subulona sp.	TCCATTAAGT GGGATTAGAG GTCATGGTGG AGCTTCTGTT GACTTAGCAA TTTTTCCTTT	
Tortaxis erectus	TCCTTTAAGT GGAATTATGG GCCATAGTGG TGCTTCTAGTT GACTTAGCTA TTTTTCCTTT	
Xerocerastus sp.	CCCATTAAGT AGTATTATTG GACACAGTGG TGCTTCTGTA GATTTAGCTA TCTTCTCTCT	
Zootecus insularis	ACCTCTAAGA AGAAGCTTGG GACATAGTGG AGCTTCTGTC GATCTGGCTA TTTTCTCACT	
Gibbulinella dewinteri	ACCATTAAGT AGCCTATTAG GACATGGAGG AGCTTCTGTA GATTTAGTTA TTTTTCCTTT	

	370 380 390 400 410 420
Achatina achatina	ACATTTGGCT GGTATATCAT CTATCTTAGG GGCTATTAAC TTTATTACTA CAATTTTAA
Achatina fulica	ACATTTGGCT GGTGTATCCT CAATTTTAGG GGCAATCAAT TTTATTACGA CCGTGTATAA
Achatina stuhlmanni	ACACTTAGCC GGAATATCCT CTATTCTCGG AGCTATCAAC TTTATTACTA CAATTTTAA
Cochlitoma ustulata	ACACTTAGCT GGTATATCTT CTATTCTCGG GGCTATTAAC TTTATCACA CTATCTTTAA
Coelioxaxis blandii	ACATTTAGCT GGTATGTCTT CAATTTTAGG TGCTATTAAT TTTATTACAA CTATTTTAA
Pyrgina umbilicata	CCATTTGGCA GGAATTTCTT CAATCTTAGG GGCAATTAAC TTTATTACCA CTATTTTAA
Glessula ceylanica	TCACCTAGCA GGAATATCAT CAATTTTAGG TGCTATCAAT TTTATTACAA CTATTTTAA
Ferussacia folliculus	ACATCTTGCT GGAATATCTT CAATTTTAGG GGCAATTAAT TTTATTACTA CTATCTTTAA
Thyrophorella thomensis	TCACCTAGCA GGGATATCCT CTATTTTGGG TGCAATTAAC TTTATTACAA CAATCTTTAA
Allopeas clavulinum	TCATTTAGCT GGGATGTCTT CTATCTTG GGTATCAAT TTTATTACTA CGATTTTAA
Bocageia sp.	TCATCTTGCG GGTATATCTT CAATCTTAGG TGCCATCAAT TTTATTACTA CTATTTTAA
Eutomepeas layardi	TCACCTAGCG GGGATATCTT CAATTTTAGG GGCTATCAAT TTTATTACCA CAATTTTAA
Leptinaria lamellata	CCATTTAGCT GGAATATCTT CAATTTTAGG TGCAATTAAT TTTATTACTA CAATTTTAA
Paropeas achatinaceum	CCATTTGGCT GGGATATCCT CAATTTTAGG GGCTATTAAC TTTATTACAA CAATCTTTAA
Riebeckia sp.	TCATTTAGCA GGAATATCTT CAATCTTAGG GGCTATTAAT TTTATTACAA CTATTTTAA
Rumina decollata	TCATTTAGCA GGGATATCTT CTATTTTAGG TGCAATTAAT TTTATTACTA CTATTTTAA
Subulina octona	GCATTTAGCA GGAATGTCTT CTATTTTAGG TGCAATTAAT TTTATTACTA CTGTATTTAA
Subulina striatella	ACATTTGGCT GGTATGTCTT CAATTTTAGG TGCAATTAAT TTTATTACGA CTGTTTTAA
Subulina vitrea	ACATTTAGCA GGAATATCTT CTATTTTAGG GGCCATTAAT TTTATTACTA CAATTTTAA
Subulona sp.	ACATCTAGCT GGTATATCTT CCATTTTAGG TGCAATTAAC TTTATTACAA CTATTTTAA
Tortaxis erectus	ACATTTAGCT GGACTTTCTT CTATTTTAGG GGCTATTAAC TTTATTACAA CTATTTTAA
Xerocerastus sp.	CCACTTAGCC GGGATGTCTT CTATCTTG GGTATTAAT TTTATTACTA CCATTTTAA
Zootecus insularis	TCACCTGGCC GGTATATCTT CAATCTTG AGCTATTAAT TTTATTACTA CTATCTTTAA
Gibbulinella dewinteri	ACATCTAGCA GGTATATCTT CTATTTTAGG GGCTATTAAT TTTATTACTA CTATCTTTAA

	430 440 450 460 470 480
Achatina achatina	TATACGAGCA TCAGGGATAT CTATAGAACG GGTTAGACTG TTTGTATGAT CTATTTTAGT
Achatina fulica	TATACGAGCC TCTGGGCTAA CTATAGAACG GGTAAAGCTTA TTCGTGTGGT CAATTTTAGT
Achatina stuhlmanni	CATACGAGTA TCTGGACTAT CTATGGAACG GGTGAGGTTA TTTGTATGAT CAATCTTGT
Cochlitoma ustulata	TATACGAGCT CAAGGCTTAT CTATGGAGCG TATAAGATTA TTTGTATGAT CTATTTTAGT
Coelioxaxis blandii	TATACGTGCT TCCGGATTAA CAATAGAACG TGTAAGTCTT TTTGTATGAT CTATTTTAA
Pyrgina umbilicata	TATACGTTCG TCAGGAGTTA AATATGAACG GTTAAGCTTG TTTGTTTGT CAATTTTAGT
Glessula ceylanica	CATACGAGCT CCTGGTCTTA CCATAGAACG AGTGAGGTTG TTTGTATGAT CTATTTTAGT
Ferussacia folliculus	TATACGATCC CCAGGTATAA CTATGGAACG GGTTAATTTA TTTGTGTGGT CTATTTTGT
Thyrophorella thomensis	CATACGTTCA TCTGGAATAA GTTATGAGCG TCTAAGATTA TTTGTGTGAT CAATTTTAGT
Allopeas clavulinum	TATACGATCG TCAGGAATAA AGTACGAACG AATAAGGTTA TTTGTTTGT CTATTTTGT
Bocageia sp.	TATACGATCA CCTGGTATAA CAATAGAACG AACAAGATTA TTTGTATGAT CTATCTTAGT
Eutomepeas layardi	TATACGTTCA TCTGGGTTGA AATATGAGCG AATAAGATTA TTTGTTTGT CTATCTTAGT
Leptinaria lamellata	TATACGATCA CCTGGATTAA AATAGAACG AGTAAGATTA TTTGTTTGT CAATTTTAGT
Paropeas achatinaceum	TATACGCTCA TCAGGGATGA AGTATGAACG TATAAGTCTA TTTGTGTGGT CTATTTTAA
Riebeckia sp.	CATACGTGCC TCAGGATTAA CATGAGAACG TACTAGTTTA TTTGTTTGT CAATTTTAGT
Rumina decollata	TATACGTGCC CCAGGTCTTT CAATAGAACG GGTAAGCCTA TTTGTGTGAT CTATTTTAGT
Subulina octona	TATACGTCTT TCTGGTTTAT CAATAGAACG AGTTAGATTG TTTGTTTGT CAATTTTAA
Subulina striatella	TATACGATCT TCCGGTTTAA GTTTTGAGCG GGTGAGTTTA TTTGTATGAT CTATTTTAGT
Subulina vitrea	TATACGAGCC CCTGGTATAA CCCCAGAACG AATTAGGCTA TTTGTTTGT CAATTTTAGT
Subulona sp.	CATACGAGCA CCAGGCTTAA CTCCAGAACG AACTAGACTA TTTGTTTGT CTATTTTAGT
Tortaxis erectus	TATACGTCTT TCAGGAATAT CTATGGAACG GGTAAGTCTA TTTGTTTGT CTATTTTAGT
Xerocerastus sp.	TATACGATCC TCTGGAATAA GTATGGAACG TGTCAGGTTG TTTGTATGAT CAATCTTGT
Zootecus insularis	TATGCGCTCT CCCGGATTAA CTATGGAACG GGTGAGCTTA TTTGTATGAT CTATTTTGT
Gibbulinella dewinteri	TATACGATCG CCTGGAATAA CAATAGAACG GGTGCTTTTA TTTGTTTGT CTATTTTAGT

	490	500	510	520	530	540		
Achatina achatina	TACAGTATTT	TTACTGCTTT	TATCTCTCCC	AGTATTAGCG	GGTGCAATTA	CTATACTTCT		
Achatina fulica	TACAGTGTTT	CTACTATTGT	TATCATTGCC	AGTTCTAGCG	GGGGCAATTA	CTATACTATT		
Achatina stuhlmanni	TACTGTTTTC	CTTTTACTTT	TATCTCTACC	AGTCTTAGCT	GGGGCGATTA	CTATACTATT		
Cochlitoma ustulata	TACTGTTTTC	CTACTGCTCT	TATCTCTACC	AGTACTAGCA	GGGGCTATTA	CAATACTCCT		
Coeliaxis blandii	TACAGTATTT	TTATTACTCC	TATCTTTACC	AGTCTTGGA	GGGGCAATTA	CTATGTTATT		
Pyrgina umbilicata	TACTGTATTT	CTTCTCCTTC	TTTCCCTTCC	TGTATTAGCT	GGTGCAATTA	CCATATTATT		
Glessula ceylanica	TACAGTTTTT	TTACTTCTTC	TTTCATTACC	TGTGTTAGCT	GGAGCTATTA	CTATACTATT		
Ferussacia folliculus	TACAGTTTTC	CTCTTATTAT	TATCACTCCC	TGTACTTGCG	GGGGCTATTA	CTATACTATT		
Thyrophorella thomensis	CACTGTTTTT	CTTCTTCTCT	TATCTCTTCC	TGTTCTAGCT	GGGGCTATTA	CTATACTCTT		
Allopeas clavulinum	GACAGTTTTT	TTACTTCTTT	TATCTCTTCC	AGTTCTTGCC	GGTGTATCA	CTATACTTTT		
Bocageia sp.	CACAGTATTT	CTATTACTTT	TATCATTACC	TGTTTTAGCT	GGTGTATTA	CTATGCTTCT		
Eutomopeas layardi	AACAGTTTTT	TTGCTTTTAC	TATCTCTTCC	TGTTCTAGCT	GGCGCGATTA	CGATACTTTT		
Leptinaria lamellata	AACGTTTTTT	TTACTTCTCC	TATCGCTTCC	TGTTTTAGCT	GGGGCAATTA	CAATATTATT		
Paropeas achatinaceum	CACAGTTTTC	TTATTACTTC	TTTCGTTACC	TGTTCTTGCT	GGTGAATCA	CTATACTTCT		
Riebeckia sp.	AACAGTTTTT	TTACTTCTTT	TATCTCTTCC	TGTTTTAGCT	GGAGCAATTA	CTATACTTTT		
Rumina decollata	AACCGTTTTT	CTATTGCTTT	TATCCCTACC	AGTACTAGCA	GGAGCTATTA	CAATACTACT		
Subulina octona	TACTGTTTTT	TTATTATTAC	TATCATTGCC	TGTATTGGCT	GGTGCAATTA	CAATATTATT		
Subulina striatella	AACGGTGTTT	TTACTATTAT	TGCTTTTACC	TGTATTGGCT	GGGGCAATCA	CTATGTTATT		
Subulina vitrea	AACAGTATTT	TTATTACTTT	TATCTTTACC	GGTTTTAGCT	GGGGCTATCA	CAATGTTATT		
Subulona sp.	TACTGTGTTT	TTATTATTGT	TATCATTACC	AGTACTAGCT	GGCGCAATTA	CTATACTACT		
Tortaxis erectus	TACAGTTTTC	TTGTTACTTC	TTTCACTTCC	TGTTTTAGCA	GGTGTATTA	CTATGTTATT		
Xerocerastus sp.	TACTGTATTC	CTTTTATTAC	TTTCACTCCC	AGTATTAGCT	GGTGTATTA	CTATACTATT		
Zootecus insularis	TACTGTATTT	CTATTACTGC	TATCATTACC	CGTTTTAGCG	GGTGCAATTA	CTATGCTATT		
Gibbulinella dewinteri	TACTGTTTTC	CTCTTACTTT	TATCACTTCC	AGTTCTTGCT	GGAGCAATTA	CTATACTGTT		

	550	560	570	580	590	600		
Achatina achatina	GACCGATCGT	AACTTTAAATA	CATCTTTTTT	TGATCCAGCT	GGAGGTGGTG	ATCCAGTGCT		
Achatina fulica	AACGGATCGT	AACTTTAAACA	CATCTTTTTT	TGACCCAGCT	GGAGGGGGGG	ACCCAGTCTT		
Achatina stuhlmanni	AACGTATCGA	AATTTTAAATA	CATCTTTTTT	TGATCCGGCA	GGGGGTGGTG	ATCCAATCCT		
Cochlitoma ustulata	TACTGATCGT	AATTTTAAACA	CATCCTTTTT	CGACCCGGCA	GGGGGAGGAG	ATCCTATCTT		
Coeliaxis blandii	AACAGACCGG	AACTTTAAATA	CGTCATTTTT	TGACCCGGCC	GGGGGCGGAG	ATCCTGTTCT		
Pyrgina umbilicata	AACGTATCGT	AATTTTAAATA	CTTCTTTTTT	TGATCCAGCT	GGGGGTGGTG	ATCCGCTGCT		
Glessula ceylanica	GACAGATCGA	AATTTTAAATA	CATCTTTTTT	TGACCCAGCT	GGAGGTGGTG	ACCCAATTTT		
Ferussacia folliculus	AACGGATCGT	AATTTTAAATA	CGTCTTTTTT	TGATCCAGCA	GGAGGTGGAG	ACCCAATTTT		
Thyrophorella thomensis	AACGTATCGA	AATTTTAAATA	CTTCTTTCTT	TGACCCGCG	GGAGGGGGCG	ATCCGGTGTT		
Allopeas clavulinum	AACGTATCGG	AATTTTAAATA	CATCTTTTTT	TGATCCTGCT	GGTGGAGGGG	ACCCAGTCTT		
Bocageia sp.	TACAGATCGT	AATTTTAAATA	CATCCTTTTT	TGATCCGGCT	GGGGGAGGGG	ATCCTATTTT		
Eutomopeas layardi	AACGTATCGA	AACTTTAAACA	CATCTTTTTT	TGATCCGGCT	GGAGGGGGGG	ATCCTATCTT		
Leptinaria lamellata	AACAGATCGA	AATTTTAAATA	CATCATTCTT	TGATCCAGCA	GGTGGTGGAG	ATCCAATTTT		
Paropeas achatinaceum	TACAGACCGT	AATTTTAAATA	CATCTTTTTT	TGACCCGCT	GGTGGCGGTG	ACCCTGTGTT		
Riebeckia sp.	AACAGACCGA	AATTTTAAATA	CATCATTCTT	TGACCCAGCT	GGAGGAGGTG	ATCCTATTTT		
Rumina decollata	AACGTACCGT	AACTTTAAACA	CTTCATTCTT	TGATCCTGCA	GGGGGAGGTG	ATCCAATTCT		
Subulina octona	AACGTATCGT	AATTTTAAATA	CATCTTTTTT	TGATCCAGCT	GGTGGGGGGG	ACCCTATCTT		
Subulina striatella	AACGTATCGT	AATTTTAAATA	CATCTTTTTT	TGACCCAGCA	GGTGGAGGGG	ACCCTATCTT		
Subulina vitrea	AACCGATCGT	AATTTTAAACA	CATCTTTCTT	TGATCCGGCT	GGAGGGGGAG	ACCCTGTTTT		
Subulona sp.	GACGGACCGT	AATTTTAAATA	CATCATTCTT	TGATCCTGCT	GGGGGAGGAG	ATCCAGTCTT		
Tortaxis erectus	GACTGATCGA	AATTTTAAATA	CATCATTCTT	TGACCCAGCT	GGTGGTGGTG	ATCCAATTCT		
Xerocerastus sp.	AACGTATCGT	AATTTTAAATA	CTTCATTCTT	TGATCCAGCA	GGCGGAGGTG	ACCCTGTACT		
Zootecus insularis	GACTGATCGA	AATTTTAAACA	CGTCGTTTTT	TGATCCAGCT	GGGGGTGGTG	ATCCTATTTT		
Gibbulinella dewinteri	AACGTATCGG	AATTTTAAATA	CGTCTTTTTT	TGATCCTGCT	GGGGGTGGGG	ATCCTATTTT		

....|..

<i>Achatina achatina</i>	TTACCAG
<i>Achatina fulica</i>	GTACCAG
<i>Achatina stuhlmanni</i>	ATACCAG
<i>Cochlitoma ustulata</i>	GTATCAG
<i>Coelioxaxis blandii</i>	TTACCAA
<i>Pyrgina umbilicata</i>	ATATCAA
<i>Glessula ceylanica</i>	ATATCAG
<i>Ferussacia folliculus</i>	ATACCAA
<i>Thyrophorella thomensis</i>	ATATCAA
<i>Allopeas clavulinum</i>	ATATCAG
<i>Bocageia</i> sp.	ATATCAA
<i>Eutomopeas layardi</i>	ATACCAA
<i>Leptinaria lamellata</i>	ATATCAA
<i>Paropeas achatinaceum</i>	GTATCAA
<i>Riebeckia</i> sp.	GTATCAA
<i>Rumina decollata</i>	ATATCAG
<i>Subulina octona</i>	TTATCAG
<i>Subulina striatella</i>	ATATCAG
<i>Subulina vitrea</i>	ATACCAA
<i>Subulona</i> sp.	ATATCAA
<i>Tortaxis erectus</i>	TTATCAA
<i>Xerocerastus</i> sp.	TTATCAA
<i>Zootecus insularis</i>	GTACCAA
<i>Gibbulinella dewinteri</i>	ATATCAA
<i>Gonaxis quadrilateralis</i>	ATATCAA
<i>Gonospira</i> sp.	GTATCAA

Appendix 3.5: Alignment of the 16S rRNA gene for the Achatinoidea and three streptaxid outgroup taxa. Positions included for the phylogeny analyses were marked by “m” within “NUCEOTIDES INCLUDED”.

		10	20	30	40	50	60
<i>Achatina fulica</i>	TAGCTTAAAG	A-----	-----GTGC	GTAACACAGA	TTTTAAGTAA	CCCCTGCCCG	
<i>Achatina achatina</i>	TAGCTTTAGG	A-----	-----GTGC	---TTTAA	TCTTTAGTAC	CTCCTGCCCA	
<i>Achatina stuhlmanni</i>	TAGCTAATAG	G-----	-----CT	CTTTCCATAA	TATTATAGTG	ACCCTGCCCG	
<i>Cochlitoma ustulata</i>	TAGCCTGCTG	T-----	-----	---AGATTTA	TTACACGGTA	TCCCTGCCCG	
<i>Allopeas clavulinum</i>				---AATTAA	TATAAGGTTA	GACCTGCTCA	
<i>Bocageia</i> sp.					---AGATAT	ATTCTGCCCA	
<i>Leptinaria lamellata</i>	TTTCCTAAAG	A-----	-----	---TTTTTA	TTTTTAGGTC	AGCCTGCCCA	
<i>Paropeas clavulinum</i>	TAGCCTTAAG	G-----	-----	---TATTA	TTTAAGGTAA	AGCCTGCTCA	
<i>Riebeckia</i> sp.				---GATGGCTA	TACTTAGCAT	CCCCTGCCCA	
<i>Rumina decollata</i>					---TTTAGGTAA	CCCCTGCCCA	
<i>Subulina octona</i>	TAGCTTGAAG	A-----	-----	---AAATTGG	TTTTTAGGTTA	TTTCTGCCCA	
<i>Subulina striatella</i>				---TACTA	TTTTTAAGTGA	CTTCTGCCCG	
<i>Subulina vitrea</i>	TAGCTTAAAG	A-----	-----	-GTATTTTTA	TTTTAAGTAC	CTTCTGCCCA	
<i>Subulona</i> sp.	TKRCTTAAAG	G-----	-----	ATATDGKGRA	TGTTAAGCAT	SSCCTGCCCA	
<i>Tortaxis erectus</i>					CTTTATGTAA	CTTCTGCCCG	
<i>Xerocerastus</i> sp.					---TTAGGTGA	TTTCTGCCCG	
<i>Zootecus insularis</i>		-CATATTTTA	TTAATTGTGA	AGTTTATTTA	TTTCAAGCTT	CTCCTGCCCA	
<i>Coelioxys blandii</i>					TTATAGATTA	ATCCTGCCCA	
<i>Pyrgina umbilicata</i>						AGCCTGTCTA	
<i>Cecilioides gokwaeus</i>	TAACCTAAAG	A-----	-----	---AATTTA	TTTTAAGTAT	ACCCTGCCCG	
<i>Ferussacia folliculus</i>					-----A	CTCCTGCCCG	
<i>Thyrophorella thomensis</i>						AGCCTGTCTA	
<i>Gibbulinella dewinteri</i>	TAGCCTAGTG	A-----	-----	---GTTTTA	TATTAGGTTG	CTTCTGCCCA	
<i>Gonaxis quadrilateralis</i>					---AGGTGA	CTTCTGCCCA	
<i>Gonospira</i> sp.					---AGTGT	TGCTGTGCCA	
NUCLEOTIDES INCLUDED						mmmmmmmmmm	

	70	80	90	100	110	120
<i>Achatina fulica</i>	GTGAGTTATA	GA---AACTT	TAACGGCCGC	AGTACCTTGA	CTGTGCAAAG	GTAGCATAAT
<i>Achatina achatina</i>	GTGATAGGTT	AC---CTATT	TAACGGCCGC	AGTACCTTGA	CTGTGCAAAG	GTAGCATAAT
<i>Achatina stuhlmanni</i>	GTGAAGTTAA	----CCTTT	TAACGGCCGC	AGTACCTTGA	CTGTGCTAAG	GTAGCATAAT
<i>Cochlitoma ustulata</i>	GTGGGCGCTA	TT---ACCCT	AAACGGCCGC	AGTACCTTGA	CTGTGCAAAG	GTAGCATAAT
<i>Allopeas clavulinum</i>	ATGAAAA---	-----TGTTT	TAACGACCGC	AGTACCCTGA	CTGTGCTAAG	GTAGCATAAT
<i>Bocageia</i> sp.	ATGAACCTTA	-----AGTTT	TAATGGCCGC	AGTACCTTGA	CTGTGCAAAG	GTAGCATAAT
<i>Leptinaria lamellata</i>	ATGATA----	-----TATTT	TAATGGCCGC	AGTACCCTGA	CTGTGCTAAG	GTAGCATAAT
<i>Paropeas clavulinum</i>	ATGAAT----	-----AATTT	TAATAGCCGC	AGTACCCTGA	CTGTGCTAAG	GTAGCATAAT
<i>Riebeckia</i> sp.	GTGAGATATA	-----TTCTT	AAATGGCCGC	GGTACTTTGA	CCGTGCTAAG	GTAGCATAAT
<i>Rumina decollata</i>	GCGATATAT-	-----AAGTT	TAGCGGCCGC	AGTACTTTGA	CTGTGCTAAG	GTAGCATAAT
<i>Subulina octona</i>	GTGACGGTAA	A-----AAGTT	AAATGGCCGC	AGTACATTGA	CTGTGCTAAG	GTAGCATAAT
<i>Subulina striatella</i>	GTGAATAG--	-----AAATT	TAACGGCCGC	AGTACATTGA	CTGTGCTAAG	GTAGCATAAT
<i>Subulina vitrea</i>	ATGAAATT--	-----AATTT	TAATGGCCGC	AGTACTTTGA	CTGTGCAAAG	GTAGCATAAT
<i>Subulona</i> sp.	AWGAATAATT	T----ATTKT	TAATGGCCGC	AGTACCTTGA	CTGTRCTAAG	GTAGCATAAT
<i>Tortaxis erectus</i>	GTGATT----	-----TTTTT	TAACGGCCGC	AGTACCCTGA	CTGTGCTAAG	GTAGCATAAT
<i>Xerocera</i> sp.	GTGATA----	-----TATTT	TAACGGCCGC	AGTACCTTGA	CTGTGCTAAG	GTAGCGTAAT
<i>Zootecus insularis</i>	GTGATTTTT-	-----ATATT	TAACGGCCGC	AGTACCCTGA	CTGTGCTAAG	GTAGCGTAAT
<i>Coeliaxis blandii</i>	GTGAATACTT	GTGAAAATTT	TAACGGCCGC	GGTACCTTGA	CCGTGCTAAG	GTAGCATAAT
<i>Pyrgina umbilicata</i>	ATGAATAAAA	A-----ACTTT	TAATGACCGC	AGTACCCTGA	CTGTGCTAAG	GTAGCATAAT
<i>Cecilioides gokweanus</i>	GTGAGTTATT	T----TTCTT	TAACGGCCGC	AGTACCTTGA	CTGTGCAAAG	GTAGCATAAT
<i>Ferussacia folliculus</i>	GTGATGGT--	-----TTATT	TAACGGCCGC	AGTACCCTGA	CTGTGCAAAG	GTAGCGTAAT
<i>Thyrophorella thomensis</i>	ATGAA-----	-----TATT	TAATGACCGC	AGTACCCTGA	CTGTGCTAAG	GTAGCATAAT
<i>Gibbulinella dewinteri</i>	CTGATTAGTT	AACT-AACTT	TAACGGCCGC	AGTACTCTGA	CTGTGCTAAG	GTAGCATAAT
<i>Gonaxis quadrilateralis</i>	GTGAA-----	-----AATTT	TAACGGCCGC	AGTACCCTGA	CTGTGCTAAG	GTAGCATAAT
<i>Gonospira</i> sp.	GTGATGT-	-----TTATT	CAACGGCCGC	AGTACTTTGA	CTGTGCTAAG	GTAGCATAAT
NUCLEOTIDES INCLUDED	mmmmmm----	-----mmmm	mmmmmmmmmmmm	mmmmmmmmmmmm	mmmmmmmmmmmm	mmmmmmmmmmmm

	130	140	150	160	170	180	
Achatina fulica	AATTTGTCTT	CTAATTAAGG	TCTGGAATGA	AGGGGGACA-	-CAGGGGAGA	GCTGTCTCCA	
Achatina achatina	AATTTGTCCC	TTAATTAGGG	TCTGGTATGA	AAGGATTTT-	-CGGGAGTTT	ACTGTCTTAG	
Achatina stuhlmanni	AATTTGTCTT	TTAATTAAGG	TCTTGCATGA	ATGGGTGTT-	-CGAGGAGTC	GCTGTCTTTA	
Cochlitoma ustulata	CACCTGTCTT	TTAATTGAGG	ACTGGAATGA	AAGGGAAGAA	ACGGGTATGA	GCTGTCTCTA	
Allopeas clavulinum	AATTTGTCTT	TTAATTGGAG	TCTGGAATGA	ATGGATTAA-	-TGGGGGATA	ACTGTCTCTT	
Bocageia sp.	AATTTGTCTT	TTAATTAGGG	TCTAGAATGA	ATGAAAAATT	TTGAAAAACC	TCTGTCTCAT	
Leptinaria lamellata	AATTAGTCCT	TTAATTGAGG	TCTGGAATAA	ATGGTATAA-	-CGGATAATA	ACTGTCTCAT	
Paropeas clavulinum	AATTTGTCTT	TTAATTAAGG	TCTAGAATGA	ATGGTTTAT-	-TGGGGAAAA	GCTGTCTTAA	
Riebeckia sp.	AATTTGTCTT	TTAATTAAGG	TCTAGAATGA	ATGGGGTTA-	-CTGAAAAAGT	ATTGTCTCTG	
Rumina decollata	AACCTGTCTT	TTAATTGAGG	TCTTGAATGA	ACGGGTTTA-	-CGAAAAATT	TCTGTCTCCT	
Subulina octona	AATTTGTCTT	TTAATTGAGG	TCTAGAATGA	ATGAAATTTA-	-TGGGGAATA	GCTGTCTCAA	
Subulina striatella	AAATTGTCTC	TTAATTGGGG	TCTAGAATTA	ACGGAGTTA-	-TGGGGGATA	ACTGTCTCTA	
Subulina vitrea	AATTTGTCTT	TTAATTGGGG	ACTAGAATGA	ATGAAAAAAA-	-CGGAGGAAA	GCTGTCTCTT	
Subulona sp.	CACCTGTCTT	TTAATTGAGG	ACTAGAATGA	ATGGGGGCA-	-CGGGAACCR	GCTGTCTGTA	
Tortaxis erectus	AATTTGTCTT	TTAATTAGGG	TCTAGAATGA	AAGAAAGATT-	-TAGGGATAA	ACTGTCTCTG	
Xerocerastus sp.	AATTTGGCTT	TTAATTGAGG	TCTTGAATGA	AAGGGATAA-	-CGTGGGGGA	ATTGTCTCTT	
Zootecus insularis	AATTTGTCTT	TTAATTGGGG	TCAGGTATGA	ACGGAGATCA	-CACATAGCC	GCTGTCTCTT	
Coelioxys blandii	AATTTGTCTT	TTAATTGAGG	TCGGGAATGA	ACGGATATT-	-CTGAAATAA	CCTGTCTTAA	
Pyrquina umbilicata	AATTTGTCTT	TTAATTGAGG	TCTAGAATGA	ATGGCACCT-	-TGGGGTGAA	GCTGTCTTAA	
Ceciloides gokweanus	AATTTGTCTT	TTAATTGGAG	TCTTGAATAA	ATGGGAATTT	-GGGGAGG-G	GCTGTCTCTT	
Ferussacia folliculus	AATTTGTCTT	TTAATTGGGG	TCGAGAATGA	AAGGAATTTT	-GGGAAGA-A	GGTGTCTCAT	
Thyrophorella thomensis	AACCTGTCTC	CTAATTAGGG	TCTAGAATAA	ACGGCATCTA	-GAAGAGA-A	TCTGTCTTGG	
Gibbulinella dewinteri	AAATTGATTT	TTAATTGGAG	TCTAGAATGA	AAGGGATTA-	-TGGGGGGAT	ACTGTCTCTA	
Gonaxis quadrilateralis	AAATTGATTT	TTAATTGAAG	TCTAGAATGA	ATGGAAATT-	-TGGAAAAAA	TCTGTCTTAA	
Gonospira sp.	AAATTGTCTT	ATAATTAGG	ACTAGAATCA	AAGAAATTTA-	-TGGAGTTTA	ACTGTCTTAA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	190	200	210	220	230	240	
Achatina fulica	ATAAGGTTAA	TT--TAACCT	TC--TTATCAG	GTGAAAAATTC	CTGAGCCT--	--CCGATGAA	
Achatina achatina	TGAGGTTAAT	AT--GAATTT	ACTCAATAAG	GTGAAAAATTC	CTTAAAAAT--	--TAGATAAA	
Achatina stuhlmanni	TTAGGGTTTA	ATT-AAATTT	AC--TCAAGAG	GTGAAAAATTC	CTCTAAATTT-	--AAGATAAA	
Cochlitoma ustulata	GTTAGCTCTT	TT--AAATTT	AA--TTGTTAG	GTGAAAAATAC	CTATACCT--	--TTAACAGA	
Allopeas clavulinum	TAAGATTAAT	TT--TAACCT	GT--TTAAAAG	GTGAAAAAAC	CTTTGAAAA-	--CAGAAAAA	
Bocageia sp.	TTAGAAGTTT	TG--TAAGTT	TC--TAATAAG	GTGAAAAATTC	CTTAGTGG--	--CCGAAGAA	
Leptinaria lamellata	TGATTAATTT	TAATTAACCT	AC--TAATAAG	GTGAAAAATTC	CTTAAAA--	--TTGAAAAA	
Paropeas clavulinum	TTCAATTAAC	TT--TAATTT	AC--TTAGAAG	GTGAAAAAAC	CTTCAAAT--	--TAGAAAAA	
Riebeckia sp.	CTTTATTACT	T---GAAATT	TC--TTATCAG	GTGAAAAATTC	CTGAATT---	--TTGAAAAA	
Rumina decollata	GTACAAATTT	TACTGAACCT	AC--TTAAGAG	GTGAAAAATTC	CTTTAGTA--	--ATGAAAAA	
Subulina octona	TAAGAATTTA	TAT-GAATTT	AT--TAATAAG	GTGAAAAATAC	CTTAAGT---	--TTGAAAAA	
Subulina striatella	TAGGGTTTAT	AT--GAAGTT	AA--TTATTAG	GTGCAAAAGC	CTAATTGT--	--TAGAAAAA	
Subulina vitrea	GTAAGTTAAC	TT--TAATTT	AC--TTAAAAG	GTGAAAAATAC	CTTTATTT--	--AACATGAA	
Subulona sp.	TTATATTATC	TA--TAATTT	GC--TAAGGAG	GTGAAAAATTC	CTCTAGC---	--TATAAGGT	
Tortaxis erectus	TAGAGGTTAA	TT--GAATTT	CC--TTAAAAG	GTGAAAAATTC	CTTTATAA--	--ATGAAGTT	
Xerocerastus sp.	TAAAAGTTTG	TA--GAATTT	GT--TTATTAG	GTGAAAAATAC	CTATATTT--	--AATATATT	
Zootecus insularis	AGAAGTTTTT	GT--GAAGTT	AC--TTATAAG	GTGAAAAATTC	CTTTGAGCT-	--TAGATAAA	
Coelioxys blandii	TTTAATTAAA	TT--TAAGTT	CC--TAATAAG	GTGCAAAATTC	CTTAATTTTG	TTATGAAAAAT	
Pyrquina umbilicata	TTGGGGATTA	TTT-TAACCT	ACTTTATAGG	GTGAAAAATTC	CCTTAAAT--	--ATTTATTT	
Ceciloides gokweanus	ATAAATTTTT	TT--TAACCT	AC--TAAAGAG	GTGAAAAATTC	CTCTGGTTA-	--TTTTTAAA	
Ferussacia folliculus	TTAAACTTTT	TT--TAATTT	TC--TTAGGGT	GTGAAAAATTC	ACCTATTAACT	T-TTTAAGAA	
Thyrophorella thomensis	GTAGGTTAAT	AC--TAACCT	AC--TTAAAAG	GTGAAAAATTC	CCTTGAAA--	--CTTTATTT	
Gibbulinella dewinteri	GTTTCGATTT	ATTTGAACCT	AT--ATTTTAG	GTGAAAAATAC	CTAAGAATT-	--TTAATAAT	
Gonaxis quadrilateralis	ATTAATTATT	TT--TAAATT	AC--TTATAAG	GTGAAAAATAC	CTTAAATT--	--TTAATTAA	
Gonospira sp.	TTTAATTGTT	TT--GAATTT	AC--TTAAAAG	GTGAAAAATTC	CTTTATTA--	--AAAAATAAT	
NUCLEOTIDES INCLUDED	-----	-----mmmmmm	mm-mmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	250	260	270	280	290	300		
Achatina fulica	AGACGAGAAG	ACCCTTAGAG	TTTTTATTAT	AACCTTGATA-	-----	-----	TA	
Achatina achatina	AGACGAGAAG	ACCCTTGGAG	TTTTTATTAT	ATAGATAGA-	-----	-----		
Achatina stuhlmanni	AGACGAGAAG	ACCCTTAGAG	TTTTTAATGAA	AAGTAAAC--	-----	-----		
Cochlitoma ustulata	AGACGAGAAG	ACCCTTAGAG	TTTTTATGAT	AAGCATTCT-	-----	-----		
Allopeas clavulinum	AGACGAGAAG	ACCCTTAGAG	TTTTTAATTA	ATTGAATTT-	-----	-----		
Bocageia sp.	AGACGAGAAG	ACCCTAAGAG	TTTTTATAAA	CTTCTACAT-	-----	-----		
Leptinaria lamellata	AGACGAGAAG	ACCCTAAGAA	TTTTTATGAA	AAAG-----	-----	-----		
Paropeas clavulinum	AGACGAGAAG	ACCCTTAGAG	TTTTAATAAA	ATGTTAATT-	-----	-----		
Riebeckia sp.	AGACGAGAAG	ACCCTTAGAA	TTTTTATAAT	ACCTTAC---	-----	-----		
Rumina decollata	AGACGAGAAG	ACCCTTAGAA	TTTTAATAAA	AACTGTCAT-	-----	-----		
Subulina octona	AGACGAGAAG	ACCCTTAGAG	TTTTTATAAG	TAGTAAAAA-	-----	-----	T	
Subulina striatella	AGACGAGAAG	ACCCTATGAA	TTTTAATTAA	AATATTTGA-	-----	-----		
Subulina vitrea	AGACGAGAAG	ACCCTTAGAA	TTTTTATAAA	CATAATTTCT-	-----	-----		
Subulona sp.	AGACGAGAAG	ACCCTAAGAA	TTTTACACCT	GACCAGCGC-	-----	-----		
Tortaxis erectus	AGACGAGAAG	ACCCTTAGAA	TTTTTATAAT	GTAACCTCTT-	-----	---	TATAATA	
Xerocerastus sp.	AGACGAGAAG	ACCCTTAGAA	TTTTAAATTAT	TG-----	-----	-----		
Zootecus insularis	AGACGAGAAG	ACCCTTAGAA	TTTTTATAAT	AGGTAAGAA-	-----	-----		
Coeliaxis blandii	AGACGAGAAG	ACCCTTAGAA	TTTTTATTTT	ATTTAAATA-	-----	-----		
Pyrgina umbilicata	AGACGAGAAG	ACCCTTAGAA	CTTTTAATTA	AAGAGG----	-----	-----		
Cecilioides gokweanus	AGACGAGAAG	ACCCTTAGAG	TTTTAATAAA	ATTTAATTTT	ATATAATTTA	AGTAAGAATT		
Ferussacia folliculus	AGACGAGAAG	ACCCTTAGAG	TTTTAATCGTG	TGAGAGTAT-	-----	-----		
Thyrophorella thomensis	AGACGAGAAG	ACCCTCAGAA	CTTTAACTAA	TGAAGG----	-----	-----		
Gibbulinella dewinteri	AGACGAGAAG	ACCCTTAGAA	TTTTTGAAAAT	AAATAAAA--	-----	-----		
Gonaxis quadrilateralis	AGACGAGAAG	ACCCTTAGAG	TTTTTATAAA	AGG-----	-----	-----		
Gonospira sp.	AGACGAGAAG	ACCCTATGAA	TTTTTATAAT	TGTTAGA---	-----	-----		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmm----	-----	-----	-----		

	310	320	330	340	350	360		
Achatina fulica	TTAAGATTCT	TGTTCTGTTT	TTGTTGGGGC	GACAGGGTTA	CA-----	-----	A	
Achatina achatina	-----TT	CTATTCTTTT	TCGCTGGGGC	AGCGAGATTA	CA-----	-----		
Achatina stuhlmanni	-----TTT	ACTTAAGTTT	TTGTTGGGGC	AACAAGATGG	CA--AAAATT	AGTAATACTT		
Cochlitoma ustulata	---ACATATG	CTGTCAGTTT	TCGTTGGGGC	AACGAGATTG	CA-----	-----	TG	
Allopeas clavulinum	--TTTTAATG	TTTATTTTTT	TTGTTGGGGC	GACATAGTAA	CA-----	-----		
Bocageia sp.	CAACTTTAGA	ATAGGCGTTT	TCGTTGGGGC	GACGAAAATTA	CA-----	---	ATAGTA	
Leptinaria lamellata	-----TTAA	ACTTAAGTTT	TCGTTGGGGC	GACGGGTTTT	CA-----	-----		
Paropeas clavulinum	-----ATT	TTCTTTTTTT	TTGTTGGGGC	GACAAAAGTAA	CA-----	-----		
Riebeckia sp.	-----TAAA	GGGAAAGTTT	TTGTTGGGGC	GACAAAAGTTA	CA-----	-----	AG	
Rumina decollata	--TGCGTGAT	TGTTTAGTTT	TCGTTGGGGC	GACGAGGTAG	CA-----	-----	AT	
Subulina octona	TATTAGATTT	ACATAAGTTT	TCGTTGGGGC	GACGGGATTA	CA-----	-----		
Subulina striatella	-GAAGCAAAG	TATTAGATTT	TCGTTGGGGC	GACGGAATTA	CA-----	---	TAAAGAGTT	
Subulina vitrea	---AACTTAT	TATTGTATTT	TCGTTGGGGC	AACGGGCTGA	CA-----	-----	CG	
Subulona sp.	-AACTCGTCT	GWATAGTTT	TCGTTGGGGC	GACGATTAA	CA-----	-----	ACTC	
Tortaxis erectus	TTATTGTAGG	TTAATTTTTT	TCGTTGGGGC	GACGAAAATTA	CA-----	---	AAAATTTAT	
Xerocerastus sp.	--TTTTAAAAG	TATAACATTT	TTGTTGGGGC	GACAAAAGTTA	CA-----	-----		
Zootecus insularis	-----CTTTT	ACTTTTGTTT	TCGTTGGGGC	GACGAGGTGA	CATGTTGCTT	TTAGGTGGAC		
Coeliaxis blandii	ATTAATTTTA	ATTAATGTTT	TCGTTGGGGC	GACGGGTTTA	CA-----	-----		
Pyrgina umbilicata	---TTACTCA	CTTCTCTTTT	TCATTGGGGC	GATGAATTAA	CA-----	-----		
Cecilioides gokweanus	ATTATATTTT	AAATTAGTTT	TCGTTGGGGC	GACGTAGTTA	CA-----	-----		
Ferussacia folliculus	---GTCTCTT	CTTGACGATT	TTGTTGGGGC	GACAAAGTTT	CA-----	-----		
Thyrophorella thomensis	--TTATATAT	CTTCTGTTTT	TTGTTGGGGC	GACAAGTTAA	CA-----	-----		
Gibbulinella dewinteri	--ATTATTTT	ATGAGTTTTT	TTGTTGGGGT	GACAAAATAA	CA-----	-----	GGTT	
Gonaxis quadrilateralis	---TAATTA	ACTTTTTTTT	TTGTTGGGGT	GACAAAAGTAR	AA-----	-----	A	
Gonospira sp.	---TTTTTC	TAATTATTTT	TTGTTGGGGC	AACAAGATAG	CA-----	-----		
NUCLEOTIDES INCLUDED	-----	-----mm	mmmmmmmm	mmmmmmmm	mm-----	-----		

	370	380	390	400	410	420
Achatina fulica	TAGATAACTT	ACCCTACCAT	ATTTTATATAT	GGCGATCATT	ATACTTT---	-----	
Achatina achatina	----TCTTTA	CTTTTAATTA	ATCTTTACTT	ATCTAAACAT	TTGCCAAGTA	TTTATCCTAA	
Achatina stuhlmanni	AAACCCATCT	AATTTTCTAA	AGTGAATCAG	TCAATTATTA	TAAGAA----	-----	
Cochlitoma ustulata	AAATAAACCA	ATCATACGTT	AATAGTAGAT	AAGTCAATCT	TAATAGGTA-	-----	
Allopeas clavulinum	AAATTAACCT	ACTATATTAT	TTTATTTTAC	ACAACGCTTA	AGCTATA---	-----	
Bocageia sp.	ACCATAACTA	ATTAAGATTA	ATCTTAAACA	AATCAAAAGA	ATTTTAACT	-----	
Leptinaria lamellata	--CATACTAT	AACTTTACCT	GATAATGATT	ATTTTACTAA	CCAAGACAA	TATTTATTA-	
Paropeas clavulinum	-----TATT	ACTTATAACT	TACTTTTTTA	TTAGTTTATC	TGACATCTTT	ATTGAG----	
Riebeckia sp.	TTATATAATA	ACTAACTATA	AATAATTATT	TTTGCCATAG	TTTTATTGTG	AT-----	
Rumina decollata	CAAATAACCT	ACCTATTAAT	ATTTAATACA	CCGAAATCTA	TTAAACA---	-----	
Subulina octona	-TATAATTTA	AACTTTTAA	TTATAGAATA	GGGATATTTC	AAATTTTGT	TAGAA----	
Subulina striatella	AAAGAATATA	ACTTTGTAA	ACTTTTTTAA	TTATTGAGTT	TGTATATACC	AGGTGATACC	
Subulina vitrea	GATTAAATTC	GGCCAAAAAG	ATTTATATAT	AAACCTATAT	TTTCAAGAG-	-----	
Subulona sp.	TACATAACTT	AGCTGATATG	AGTAAATTTA	YTATATARAT	CTTGTTRT	AATGA----	
Tortaxis erectus	GAACTTATTG	TTTTTAGTTT	TT-----	-ATGTGCCGA	ATCATATTAA	GAA-----	
Xerocerastus sp.	-----TTTA	ACTTACTATA	TTATTTAAGC	TACTAGCCGA	GTTAATTATG	AG-----	
Zootecus insularis	CCTGAAACTC	ACCTTTAATT	TTAA-AATTT	TATAAGCCGA	ATTTTATTAT	GTA-----	
Coelioxaxis blandii	-TTCTGCATA	AACTAATCTA	TAAATTTTTC	TTAATAAAC	GAGGATTTT	ATCTT-----	
Pyrquina umbilicata	-----TTTT	AGGTTTAACT	TAATTTTTTT	GTTTAGTACA	AGACACTTTC	TACAA-----	
Ceciloides gokweanus	--TGATGTGA	ACTAACTTAA	TGAATTATTG	TATTTTAAATA	AGTTTATTAT	AATTAATTT	
Ferussacia folliculus	TAAATAACAA	ACCTTTATGA	AGTTTTC	GACGATTATA	TTATGGT---	-----	
Thyrophorella thomensis	GCAATAACTT	AACTTTAAGT	TCATTTTACA	AGGTATCGTC	TTAGGA----	-----	
Gibbulinella dewinteri	AAATCAACTT	ATTTTGGTA	TTAGTTTAC	AAGAAAGTTT	TAATAACAGG	AA-----	
Gonaxis quadrilateralis	AAATTAACCT	ACATTATAAT	TCAAGCCGAG	TTATACTATG	AA-----	-----	
Gonospira sp.	TAAAAAATCT	ATCTTAAAAAT	GATTTACTAC	ATTTCAAAGT	TTTTTATAA-	-----	
NUCLEOTIDES INCLUDED	-----mm	mmmm-----	-----	-----	-----	-----	

	430	440	450	460	470	480
Achatina fulica	-----	-----TA	ATTAAGCTAC	CTAAGGGATA	ACAGCGTAAT	CTTTTTTTTT	
Achatina achatina	GAA-----	-----AA	ACTAAACTAC	CCAAGGGATA	ACAGCATAAT	TCCTT-AGGG	
Achatina stuhlmanni	-----	-----CA	GAAAACTAC	CTAAGGGATA	ACAGCGTAAT	CTAAT-ATTA	
Cochlitoma ustulata	-----	-----TA	ATTAAGCTAC	CTAAGGGATA	ACAGCGTAAT	CTTAT-TTAT	
Allopeas clavulinum	-----	-----AG	AAGAACTAC	CTTAGGGATA	ACAGCATAAT	TTTTT-AAAA	
Bocageia sp.	-----	-----TG	AATAACTAC	CTTAGGGATA	ACAGCATAAT	CTATT-TTTA	
Leptinaria lamellata	-----	-----AG	ATTAAATTAC	CTTAGGGATA	ACAGCATAAT	ATTTT-ATAA	
Paropeas clavulinum	-----	-----AA	ATTAACTAC	CTTAGGGATA	ACAGCATAAT	TTTA--ATCA	
Riebeckia sp.	-----	-----TA	AAAAAATTAC	CTAAGGGATA	ACAGCATAAT	CTTAA-ATAA	
Rumina decollata	-----	-----TA	ATTAAATTAC	CTAAGGGATA	ACAGCATAAT	ACTAA-GTAG	
Subulina octona	-----	-----TA	AGTAACTAC	CTTAGGGATA	ACAGCATAAT	ACTT--TTAG	
Subulina striatella	TTTTTA----	-----TA	AGAAAATTAC	CTTAGGGATA	ACAGCATAAT	ACTT--TTAG	
Subulina vitrea	-----	-----TA	TTTAAATTAC	CTAAGGGATA	ACAGCATAAT	TTTTT-AAAA	
Subulona sp.	-----	-----CA	ATTAAATTAC	CTTAGGGATA	ACAGCATAAT	CTTTW-ATAA	
Tortaxis erectus	-----	-----AA	TTTAAATTAC	CTAAGGGATA	ACAGCATAAT	ATTTT-TTTA	
Xerocerastus sp.	-----	-----TG	ATAAAATTAC	CTGAGGGATA	ACAGCATAAT	TTAT--TTTA	
Zootecus insularis	-----	-----AA	ATTAAATTAC	CTGAGGGATA	ACAGCATAAT	ATCCT-TTGA	
Coelioxaxis blandii	-----	-----TA	TATAAATTAC	CTAAGGGATA	ACAGCATAAT	CTTAG-ATAA	
Pyrquina umbilicata	-----	-----AG	GAGAAGTTAC	CTGAGGGATA	ACAGCATTAT	TTATT-TTTA	
Ceciloides gokweanus	AACGAGAAAT	TTTAATTTTA	ATTAAGCTAC	CTTAGGGATA	ACAGCATAAT	ATTT--TTAA	
Ferussacia folliculus	-----	-----TG	AGAAAACTAC	CTAAGGGATA	ACAGCATAAT	TCTAT-AAAG	
Thyrophorella thomensis	-----	-----AG	ATTAAGTTAC	CTGAGGGATA	ACAGCATTAT	TTATT-AGTA	
Gibbulinella dewinteri	-----	-----AG	ATTAAATTAC	CTTAGGGATA	ACAGCATAAT	TCTTT-TACG	
Gonaxis quadrilateralis	-----	-----AG	AAAAACTAC	CTTAGGGATA	ACAGCATAAT	TTTTT-TTAA	
Gonospira sp.	-----	-----AG	GAGAAATTAC	CTTAGGGATA	ACAGCGTAAT	TTCACTTTGA	
NUCLEOTIDES INCLUDED	-----	-----	---mmmmmm	mmmmmmmm	mmmmmmmm	mmmm-----mm	

	490	500	510	520	530	540	
Achatina fulica	GGTTTTCGAC	CTCGATGTTG	GAC-TAGGGG	CC---TACTG	GTTAGTTACC	ATTA-----T	
Achatina achatina	AGATTGTGAC	CTCGATGTTG	GAC-TAGGGC	TC---TATTG	GTTAACTACC	AAGA-----A	
Achatina stuhlmanni	GGATTGCGAC	CTCGATGTTG	GAC-TAGGGA	CC---TAATG	ATGAGATATC	AAGGTGACTA	
Cochlitoma ustulata	GGATTGCGAC	CTCGATGTTG	GAC-TAGGGA	CC---CAGTG	GTGAGAAACC	AGCT-----	
Allopeas clavulinum	AGATTGTGAC	CTCGATGTTG	GAC-TAGGGA	CT---TTATG	GTTAACCACC	TTAA-----	
Bocageia sp.	GGATTGTGAC	CTCGATGTTG	GAC-TAGGGT	CA---GAAAG	GTAAGCAATC	AAAA-----	
Leptinaria lamellata	AGATTGTGAC	CTCGATGTTG	GAC-TAGGGA	CC---TATAG	GTTAACCACC	AATA-----	
Paropeas clavulinum	AGATTGTGAC	CTCGATGTTG	GAC-TAGGGA	CT---TTATG	GTTAACTACC	TGAT-----	
Riebeckia sp.	GGTTTGTGAC	CTCGATGTTG	GAT-TAGGAA	CA---TAATG	ATTAACAATC	AGCT-----	
Rumina decollata	TGTTTGTGAC	CTCGATGTTG	GAC-TAGGGA	CT---AATTG	GTTAACCACC	AGGA-----	
Subulina octona	TGATTGTGAC	CTCGATGTTG	GAC-TAGGGA	CAAAATGAATG	GTTAGATGCC	TGTT-----	
Subulina striatella	TGTTTGTGAC	CTCGATGTTG	GAC-TAGGGA	CA--TTAGTG	GCGAGATGTC	AAAT-----	
Subulina vitrea	AGATTGTGAC	CTCGATGTTG	GAT-TAGGAA	CT---TAATG	GTTAGCCACC	AAGT-----	
Subulona sp.	GGATTRTGAC	CTCGAWGTTG	GAT-TAGGGA	TT--AGATAG	GTTAGCCATC	TTTT-----T	
Tortaxis erectus	TGATTGTGAC	CTCGATGTTG	GAC-TAGGGA	CT---TAATG	GTTAATAACT	AAAA-----	
Xerocerastus sp.	AGTTTGTGAC	CTCGATGTTG	GAT-TAGGGA	AA--ATAAAG	GTTAGTAATT	TTTA-----	
Zootecus insularis	TGATTGTGAC	CTCGATGTTG	GACTTAGGGC	CC---AAATG	GTTAGCCACC	AAGC-----	
Coeliaxis blandii	TGATTGTGAC	CTCGATGTTG	GAT-TAGGGC	CT---AAAAG	GTAAGTAACC	TAGT-----	
Pyrgina umbilicata	AGATTGTGAC	CTCGATGTTG	GAC-TAGGGC	CC---TGAGG	GTTAACCACC	CAAA-----	
Cecilioides gokweanus	TGATTGTGAC	CTCGATGTTG	GAC-TAGGGA	CT---TTCTG	ATTAACCATC	AAAC-----	
Ferussacia folliculus	AGATTATGAC	CTCGATGTTG	GAC-TAGGAA	AC---TTATG	GTTAGACACC	AAAA-----	
Thyrophorella thomensis	AGATTGTGAC	CTCGATGTTG	GAC-TAGGGC	CC--ATT-AG	GTTAACCACC	TTAA-----	
Gibbulinella dewinteri	AGTTTGTGAC	CTCGATGTTG	GAC-TAGGAA	CC---TAATA	ATAAGCAATT	AAAT-----	
Gonaxis quadrilateralis	AGATTGTGAC	CTCGATGTTG	GAC-TAGGAA	CT---TAATA	ATTAGTAATT	AAAA-----	
Gonospira sp.	AGATTGCGAC	CTCGATGTTG	GAC-TAGGAA	CC---TTATG	ACTAAAAGTC	TAAA-----	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	550	560	570				
Achatina fulica	GGG-GCGGTT	CTGTTCTGAAC	TTTCTCTACC	CTAC			
Achatina achatina	TGT-AAGGTT	CTGTTCTGAAC	TTT-TTATCC	CTAC			
Achatina stuhlmanni	TGG-ATTGTT	CTGTTCTGAAC	AC--TAGACC	CTAC			
Cochlitoma ustulata	AGG-CAAGTT	CTGTTCTGAAC	TTT-CCACCC	CTAC			
Allopeas clavulinum	TAGATTGCT	CTGTTCTGAGC	TT--TAACCC	CTAC			
Bocageia sp.	CTG-GATGTT	CTGTTCTGAAC	ATT-TTACCC	CTAC			
Leptinaria lamellata	AGG-ATAGTT	CTGTTCTGAAC	TC--TGTACC	CTAC			
Paropeas clavulinum	TAG-ATTGCT	CTGTTCTGAGC	AC--TTGTTC	CTAC			
Riebeckia sp.	TTG-TAAGTT	CTGTTCTGAAC	TTT-TACTTC	CTAC			
Rumina decollata	TAG-ATGATT	CTGTTCTGAAT	CC--TTCACC	CTAC			
Subulina octona	TTG-CTGGTT	CTGTTCTGAAC	TT--AAAACC	CTAC			
Subulina striatella	TTG-CAGGTT	CTGTTCTGAAC	AT--ATAACC	CTAC			
Subulina vitrea	AAG-TTAGTT	CTGTTCTGAAC	CC--TTCCTC	CTAC			
Subulona sp.	TAA-TGAAYT	CTGTTCTGAAY	TWA-ATCNCC	CTAC			
Tortaxis erectus	AAG-ATTGTT	CTGTTCTGAAC	ATA-ACAACC	CTAC			
Xerocerastus sp.	-TA-TAAGTT	CTGTTCTGAAC	ATT-TTTACC	CTAC			
Zootecus insularis	TGG-ATGGTT	CTGTTCTGAAC	CG--TTTACC	CTAC			
Coeliaxis blandii	TAG-ATAGTT	CTGTTCTGAAC	TT--TTTACC	CTAC			
Pyrgina umbilicata	TGG-ACTGCT	CTGTTCTGAGC	AT--TGTTC	CTAC			
Cecilioides gokweanus	TAGATTGTT	CTGTTCTGAAC	TTT-TTTACC	CTAC			
Ferussacia folliculus	TGT-TATGTT	CTGTTCTGAAC	ATT-AAACTC	CTAC			
Thyrophorella thomensis	CGG-ATAGCT	CTGTTCTGAGC	TT--CTTACC	CTAC			
Gibbulinella dewinteri	TGG-AAAGTT	CTGTTCTGAAC	AA--ATTATC	CTAC			
Gonaxis quadrilateralis	TTG-ATGGTT	CTGTTCTGAAC	TT--ATTTTC	CTAC			
Gonospira sp.	AGG-AGTGTT	CTGTTCTGAAC	AAT-TATTTC	CTAC			
NUCLEOTIDES INCLUDED	-mmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

Appendix 3.6: Log likelihood scores of the different models used for the Achatinoidea using (A) 3435 unambiguously aligned nucleotides from the rRNA gene cluster; (B) 861 unambiguously aligned nucleotides of actin; (B.1) 574 unambiguously aligned nucleotides of the 1st and 2nd codon positions of actin; (B.2) 287 unambiguously aligned nucleotides of the 3rd codon position of actin; (C) 328 unambiguously aligned nucleotides of histone 3; (C.1) 218 unambiguously aligned nucleotides of the 1st and 2nd codon positions of histone 3; (D.2) 110 unambiguously aligned nucleotides of the 3rd codon position of histone 3; (D) 607 unambiguously aligned nucleotides of CO1; (D.1) 404 unambiguously aligned nucleotides of the 1st and 2nd codon positions of CO1; (D.2) 203 unambiguously aligned nucleotides of the 3rd codon position of CO1; and (E) 294 unambiguously aligned nucleotides of the 16S rRNA gene. Scores in **bold** belong to optimal models as determined by Likelihood Ratio Test. Note that the K2P and F81 models are non-nested and therefore cannot be compared for significant difference using the LRT.

	(A)	(B)	(B.1)	(B.2)	(C)	(C.1)	(C.2)	(D)	(D.1)	(D.2)	(E)
Model	rRNA gene cluster	Actin-combined	Actin-1 st +2 nd codons	Actin 3 rd codons	Histone 3	Histone 3 1 st +2 nd codons	Histone 3 3 rd codons	CO1	CO1 1 st +2 nd codons	CO1 3 rd codons	16S rRNA
JC69	8722.54544	7034.70607	1582.96160	4551.89700	2263.05590	387.26745	1519.62452	9562.61908	2707.77230	5655.47905	3720.10115
JC69+Γ	8166.65498	6318.61908	1457.51470	4364.19564	2055.16076	387.26745	1503.87233	8204.89347	2288.23519	5546.19261	3285.58073
F81	8696.59840	6997.51585	1577.09006	4404.85550	2259.84534	384.18591	1516.51436	9502.53463	2670.20603	5476.35282	3669.15845
F81+Γ	8139.63345	6263.18854	1452.88144	4302.40699	2050.07377	384.18591	1499.41097	8005.17426	2231.16774	5356.26679	3214.09399
K2P	8604.34705	6815.80245	1569.84105	4225.33326	2188.48052	385.69744	1444.53135	9484.47349	2553.77652	5631.30210	3670.91274
K2P+Γ	8038.52028	6058.78021	1439.71055	4120.55651	1977.17697	385.69744	1425.02655	8027.24407	2125.16439	5511.17394	3223.49673
HKY85	8577.13470	6776.05418	1567.69219	4205.14693	2183.03436	382.58884	1438.38403	9391.53749	2525.16880	5283.03501	3610.74395
HKY85+Γ	8000.79127	6026.10365	1439.81885	4092.36162	1969.91019	382.58884	1420.79835	7546.29497	2090.98831	4997.88304	3103.10529
TrN93	8524.93663	6730.23510	1540.64290	4201.59908	2172.17869	376.07479	1444.42135	9390.78281	2515.71421	5278.46766	3609.31276
TrN93+Γ	7970.59211	6024.01007	1420.57243	4092.24782	1965.63536	376.07479	1420.52557	7522.62965	2083.36045	4997.26819	3102.99238
GTR	8515.61458	6707.83141	1539.76409	4145.18258	2167.71615	370.44371	1411.10672	9096.60454	2506.42682	5222.31450	3536.04532
GTR+Γ	7951.71731	5993.90425	1420.05757	4051.76435	1960.84472	370.44371	1408.67625	7507.55611	2069.51524	4993.80631	3089.76804

Appendix 3.7: Summary of ambiguous sites for the actin gene for the Achatinoidea and the three streptaxid outgroup taxa

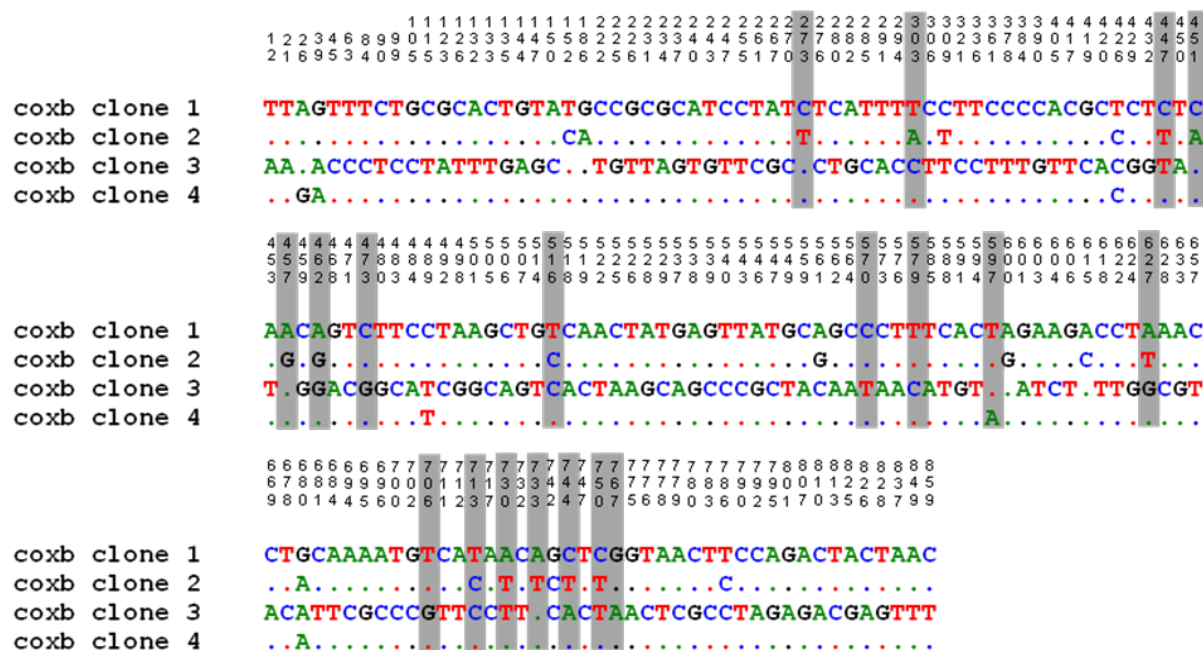
Species	Total ambiguous sites	% ambiguous sites (out of 873 total; 861 for subv)	Total ambiguous 3 rd codon positions	% ambiguous 3 rd codon positions relative to all ambiguous sites	Number of RY sites at ambiguous 3 rd codon positions	% RY sites relative to all ambiguous 3 rd codon positions	Number of ambiguous amino acids	% ambiguous amino acids (out of 291 total; 287 for subv)
ACHATINIDAE								
<i>Achatina achatina</i>	45	5.15	40	88.89	31	77.50	4	1.37
<i>Achatina fulica</i>	18	2.06	16	88.89	12	75.00	2	0.69
<i>Achatina stuhlmanni</i>	2	0.23	1	50.00	1	100.00	1	0.34
<i>Cochlitoma ustulata</i>	0	0	0	0	0	0	0	0
SUBULINIDAE								
<i>Allopeas clavulinum</i>	0	0	0	0	0	0	0	0
<i>Bocageia</i> sp.	101	11.57	85	84.16	51	60.00	14	4.81
<i>Eutomoepas layardi</i>	0	0	0	0	0	0	0	0
<i>Leptinaria lamellata</i>	0	0	0	0	0	0	0	0
<i>Paropeas achatinaceum</i>	1	0.11	0	0	0	0	1	0.34
<i>Riebeckia</i> sp.	76	8.71	59	77.63	44	74.58	15	5.15
<i>Rumina decollata</i>	29	3.32	24	82.76	16	66.67	3	1.03
<i>Subulina octona</i>	0	0	0	0	0	0	0	0
<i>Subulina striatella</i>	26	2.98	25	96.15	15	60.00	0	0
<i>Subulina vitrea</i>	37	4.30	32	86.49	26	81.25	4	1.39
<i>Subulona</i> sp.	0	0	0	0	0	0	0	0
<i>Tortaxis erectus</i>	32	3.67	26	81.25	20	76.92	7	2.41
<i>Xerocerastus</i> sp.	0	0	0	0	0	0	0	0
<i>Zootecus insularis</i>	20	2.29	18	90.00	16	88.89	1	0.34
COELIAXIDAE								
<i>Coeliaxis blandii</i>	21	2.41	13	61.90	9	69.23	7	2.41
<i>Pyrgina umbilicata</i>	88	10.08	78	88.64	55	70.51	7	2.41
THYROPHORELLIDAE								
<i>Thyrophorella thomensis</i>	62	7.10	51	82.26	37	72.55	7	2.41
GLESSULIDAE								
<i>Glessula ceylanica</i>	53	6.07	34	64.15	16	47.06	19	6.53
FERRUSSACIIDAE								
<i>Cecilioides gokweanus</i>	4	0.46	4	100.00	3	75.00	0	0
<i>Ferussacia folliculus</i>	4	0.46	4	100.00	4	100.00	0	0
STREPTAXIDAE (Outgroup)								
<i>Gibbulinella dewinteri</i>	6	0.69	6	100.00	4	66.67	0	0
<i>Gonaxis quadrilateralis</i>	55	6.30	49	89.09	34	69.39	4	1.37
<i>Gonospora</i> sp.	77	8.82	64	83.12	37	57.81	9	3.09

Appendix 3.8A: Summary of cloned actin sequences from three achatinoid species.

Species cloned	Total ambiguous sites	Total variable sites found in clones	Clone #	Sequence type*
<i>Coelioxys blandii</i>	29	163	coxb clone 1	a
			coxb clone 2	b
			coxb clone 3	c
			coxb clone 4	d
<i>Cochlitoma granulata</i>	9	72	coig clone 1	e
			coig clone 2	f
			coig clone 3	g
			coig clone 4	h
			coig clone 5	i
			coig clone 6	j
			coig clone 7	k
<i>Leptinaria lamellata</i>	0	4	letl clone 1	l
			letl clone 2	m
			letl clone 3	l
			letl clone 4	n
			letl clone 5	l
			letl clone 6	o

* NOTE: Clones with the same letter have identical actin sequences.

Appendix 3.8B: Variable sites across the actin sequence for the different clones of (1) *Coelioxys blandii*, (2) *Cochlitoma granulata* and (3) *Leptinaria lamellata*. The number shown directly above each site pertains to a variable site. Shaded positions refer to ambiguous sites found in the direct cycle sequences.



(1) *Coelioxys blandii* (coxb)

NOTE: A total of 163 variable positions were detected from the four clones, with 132 positions alone attributable to Clone 3; these positions were not picked up as ambiguities in the direct sequence.

coig clone 1 TGCCCCATCTTATCTCTTTATATGCTCCCCATCACAATTTCGAGAATCTCCTCCATCTACA
 coig clone 2 CA.....G.TGTCGCC.....C.....
 coig clone 3 ..TT.GCTACGCT.....G.TAG.T..TGTC.C.TATCATGC....C.....TG
 coig clone 4T.....
 coig clone 5 CA.....G.TGTC.CC.....T...C.....
 coig clone 6 ..T.TGCTAC.CTCTCCCGCTGTT..T.TGGTGT.C...CATG..CTTCATCATAC..
 coig clone 7 CA.....G..GTC.CC.....C.....

777777777788
 7777888889945
 678903490202

coig clone 1 CCAGCGATGCCA
 coig clone 2G.....
 coig clone 3 T.T.T.GACTG.
 coig clone 4G.....
 coig clone 5G.....
 coig clone 6 .TGCTTGGC..T
 coig clone 7G.....

(2) *Cochlitoma granulata* (coig)

NOTE: A total of 72 variable positions were found in the 7 clones, with 55 of these derived from Clones 3 and 6 and which were not picked up as ambiguities in the direct sequence.

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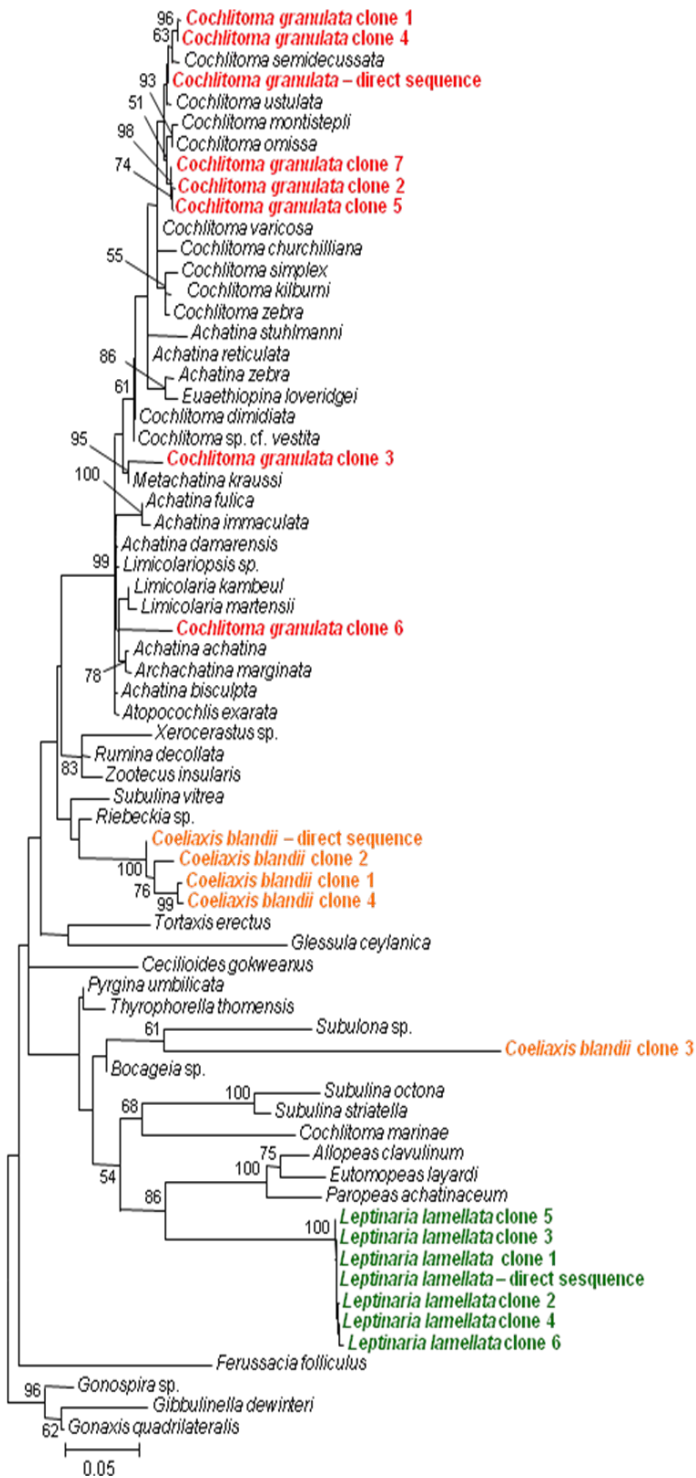
      2 3 7
      1 7 7 4
      9 4 6 0

let1 clone 1  AACT
let1 clone 2  ...C
let1 clone 3  ....
let1 clone 4  .G..
let1 clone 5  ....
let1 clone 6  G.T.

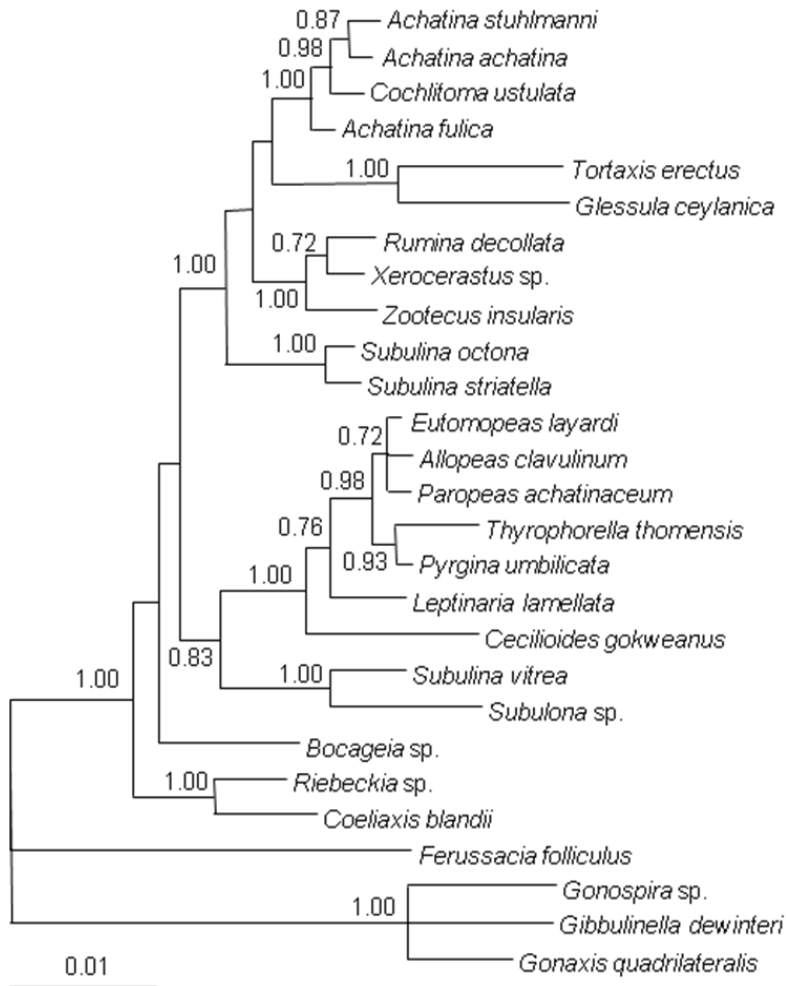
```

(3) *Leptinaria lamellata* (letl)

NOTE: *Leptinaria lamellata* did not exhibit any ambiguous positions in the direct sequence. Nevertheless, this taxon was cloned to determine if the direct sequence missed any of the ambiguous sites. Six clones were obtained, three of which (clones 1, 3 and 5) were identical with the direct sequence.



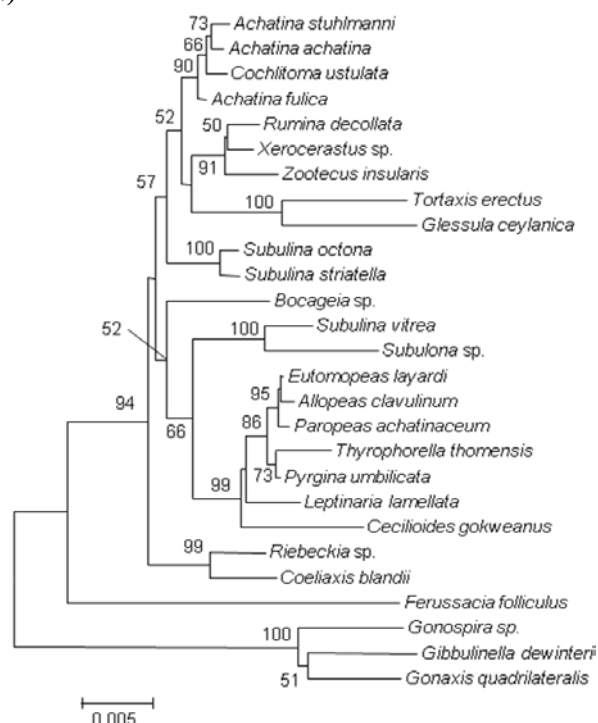
Appendix 3.9: Neighbor-joining phylogenetic tree of the Achatinoidea (including taxa from the Achatinidae) based on the actin and showing the clones of the three representative taxa (*Cochlitoma granulata*, *Coeliaxis blandii* and *Leptinaria lamellata*). The tree was constructed using 861 unambiguously aligned nucleotide sites and rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp. using the GTR+ Γ model of DNA sequence evolution. Bootstrap values lower than 50% are not shown. The scale bar represents 5 substitutional changes per 100 nucleotides.



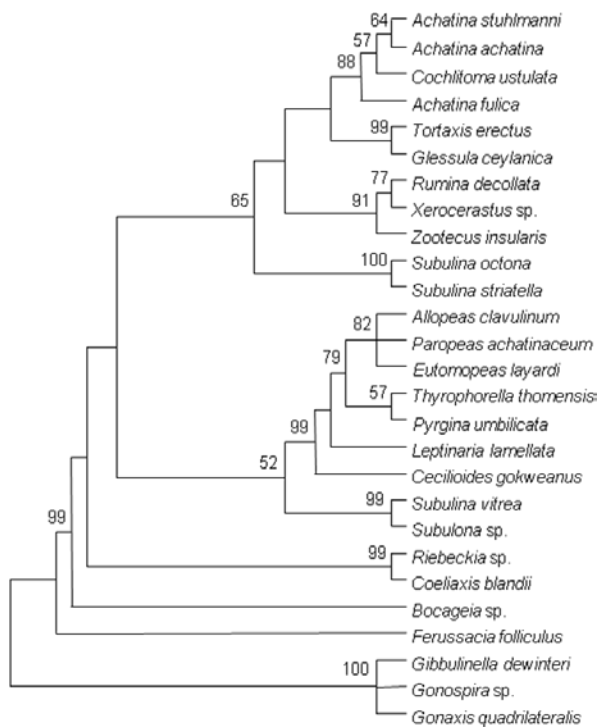
(A) BI (LSU rRNA)

Appendix 3.10: Phylogenetic trees of the Achatinoidea based on the large subunit rRNA gene using (A) Bayesian analysis, (B) neighbor-joining (p. 440) and (C) maximum parsimony based on two equally parsimonious trees (p. 440). The phylogenies were constructed from 3435 unambiguously aligned nucleotide sites and were rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 100 nucleotides and that of (B) represents 5 substitutional changes per 1000 nucleotides.

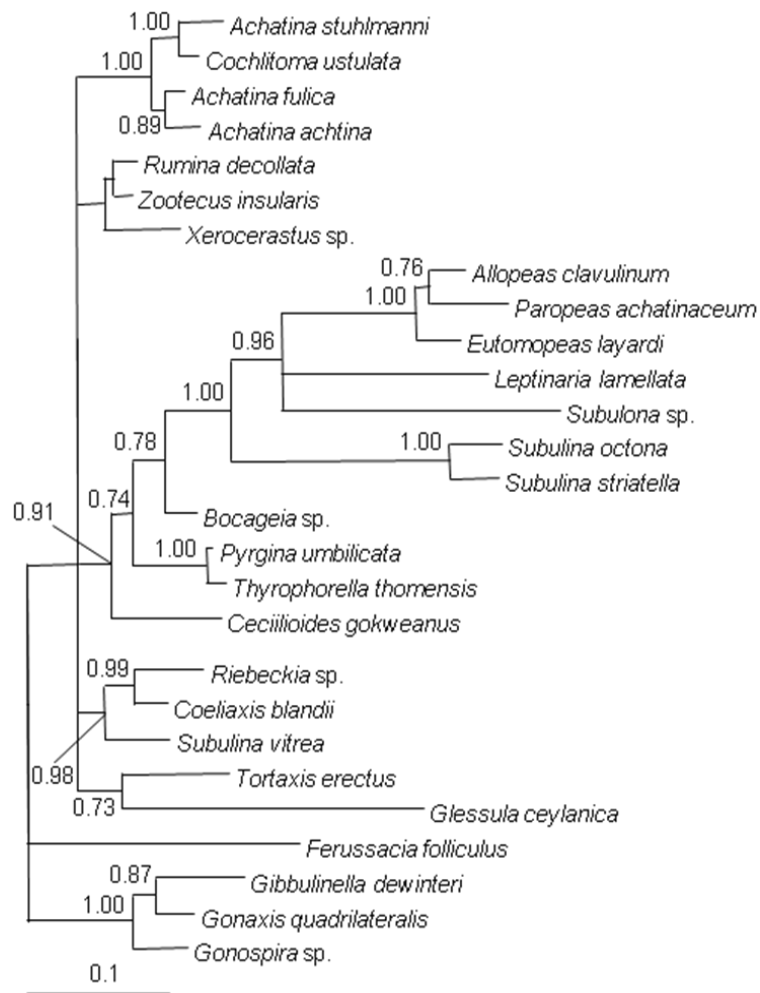
Appendix 3.10 (contd.)



(B) NJ (LSU rRNA)



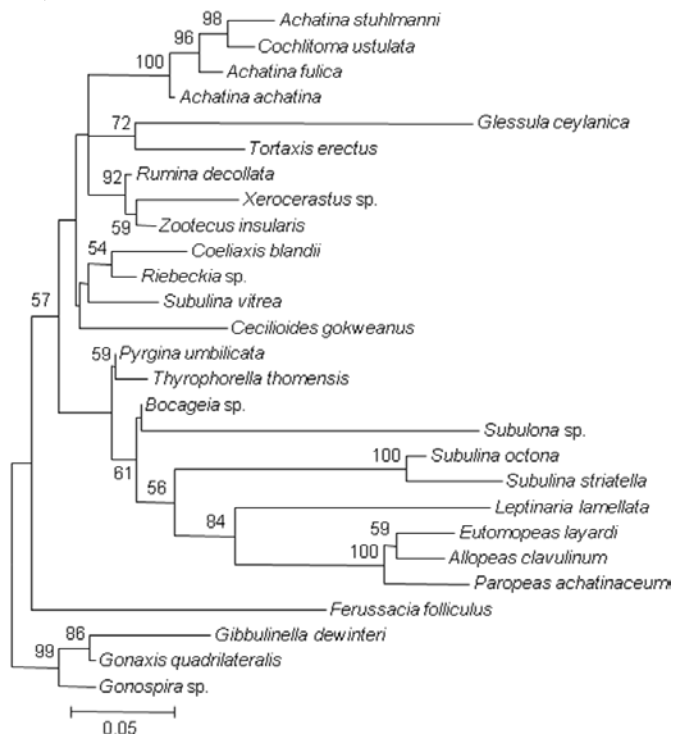
(C) MP (LSU rRNA)



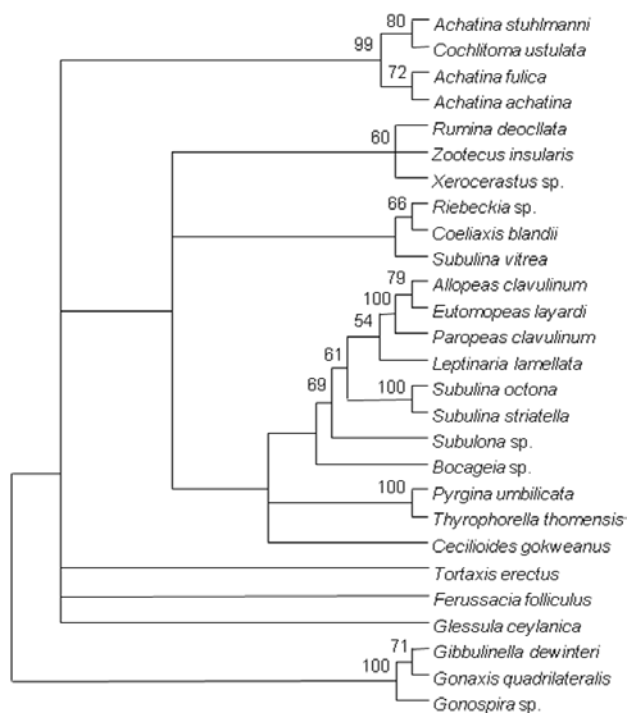
(A) BI (actin)

Appendix 3.11: Phylogenetic trees of the Achatinoidea based on the actin gene using (A) Bayesian analysis, (B) neighbor-joining (p. 442) and (C) maximum parsimony based on seven equally parsimonious trees (p. 442). The phylogenies were constructed from 861 unambiguously aligned nucleotide sites and were rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira sp.* Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 10 nucleotides and that of (B) represents 5 substitutional changes per 100 nucleotides.

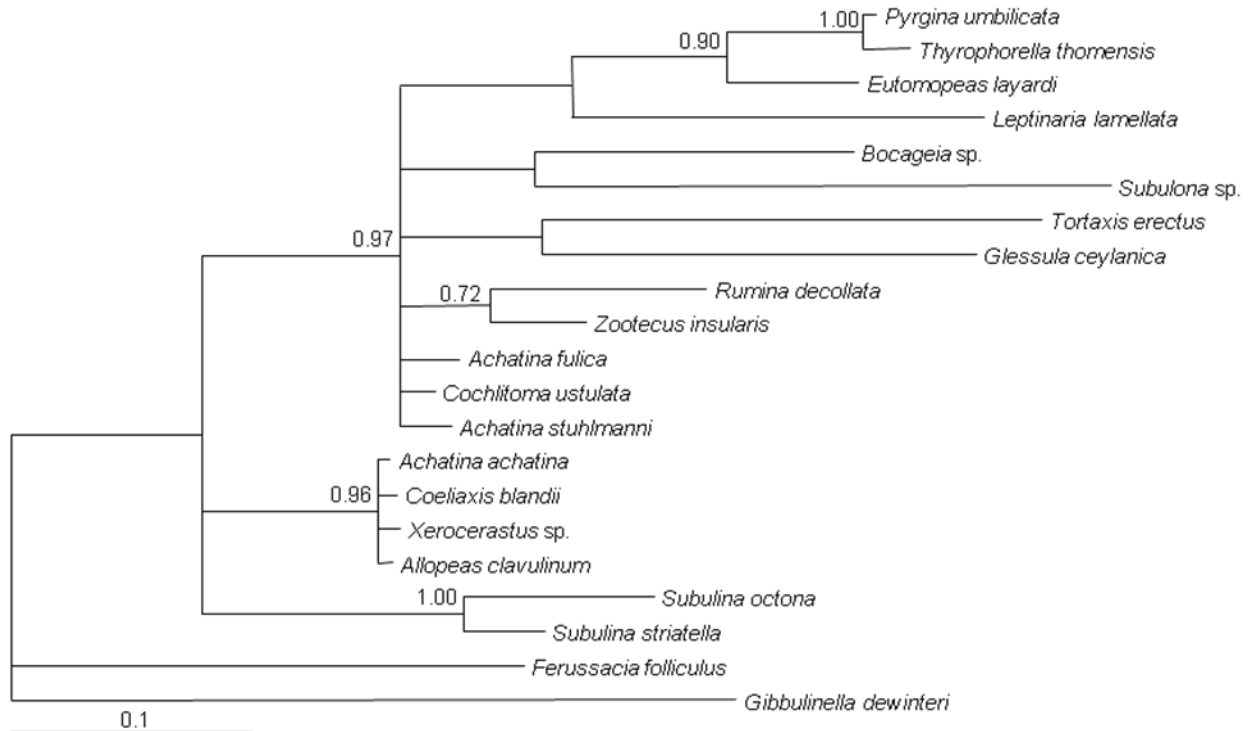
Appendix 3.11 (contd.)



(B) NJ (actin)



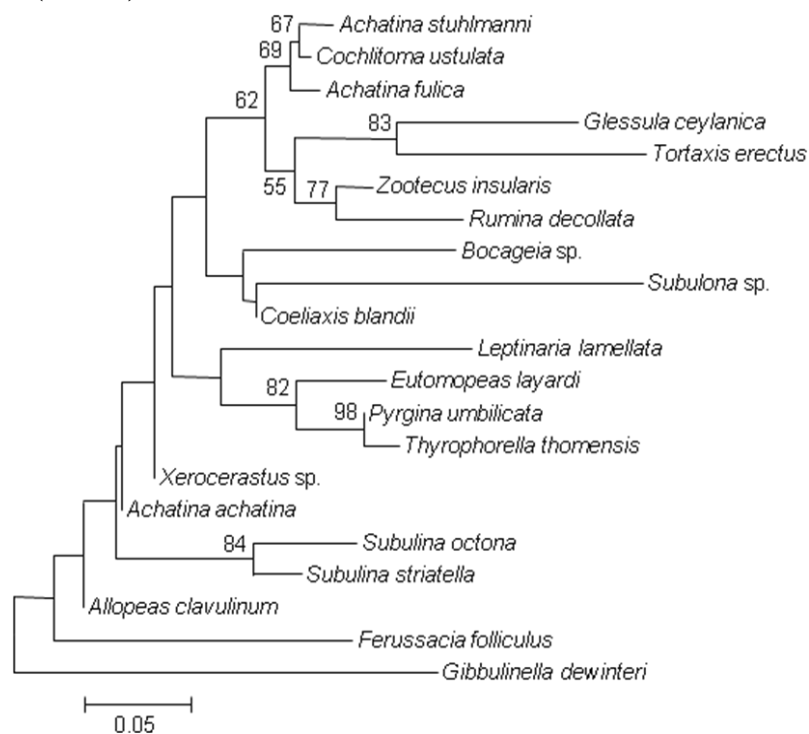
(C) MP (actin)



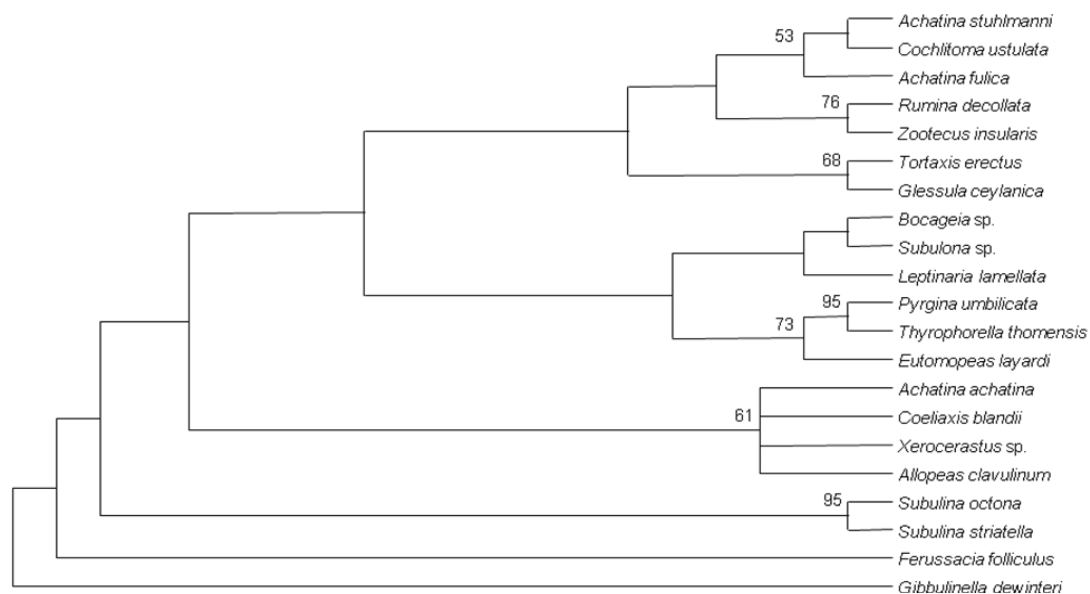
(A) BI (histone 3)

Appendix 3.12: Phylogenetic trees of the Achatinoidea based on the histone 3 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 444) and (C) maximum parsimony based on five equally parsimonious trees (p. 444). The phylogenies were constructed from 328 unambiguously aligned nucleotide sites and were rooted on the streptaxid *Gibbulinella dewinteri*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 10 nucleotides and that of (B) represents 5 substitutional changes per 100 nucleotides. Note that the achatinoids *Ceciloides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilateralis* and *Gonospora* sp. were not included in the phylogenies as they were not successfully sequenced.

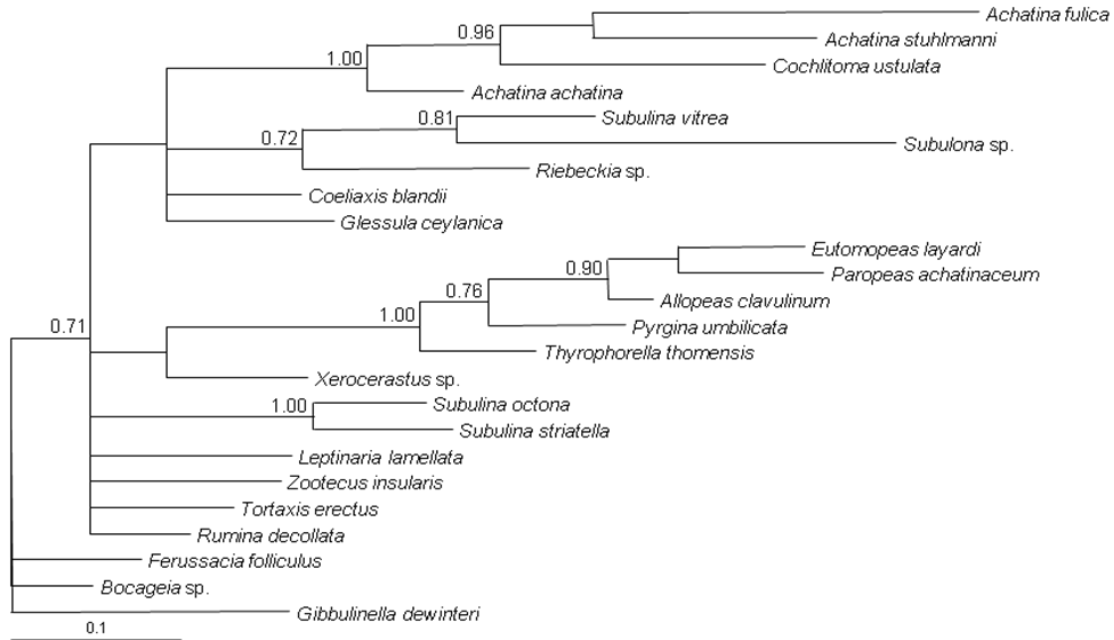
Appendix 3.12 (contd.)



(B) NJ (histone 3)



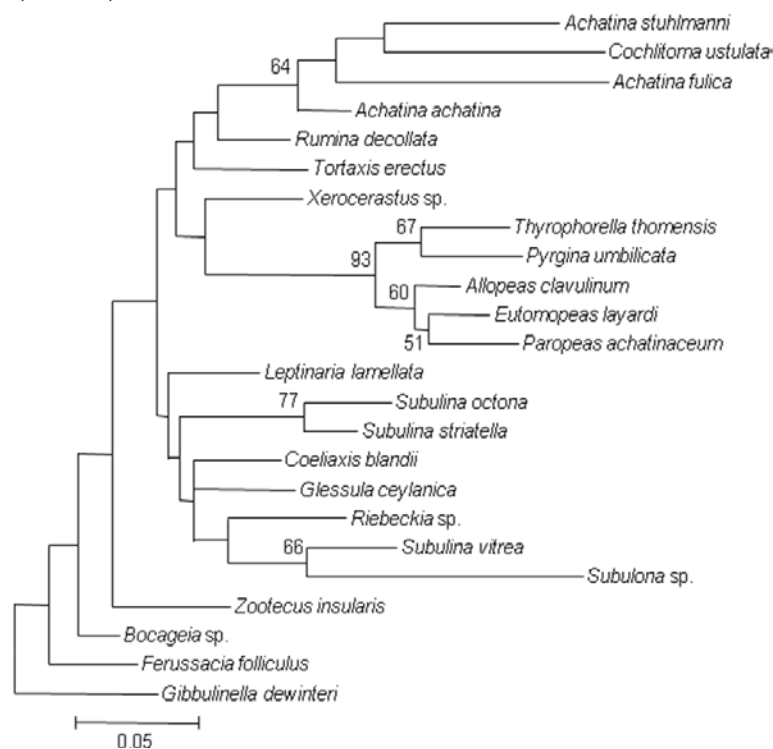
(C) MP (histone 3)



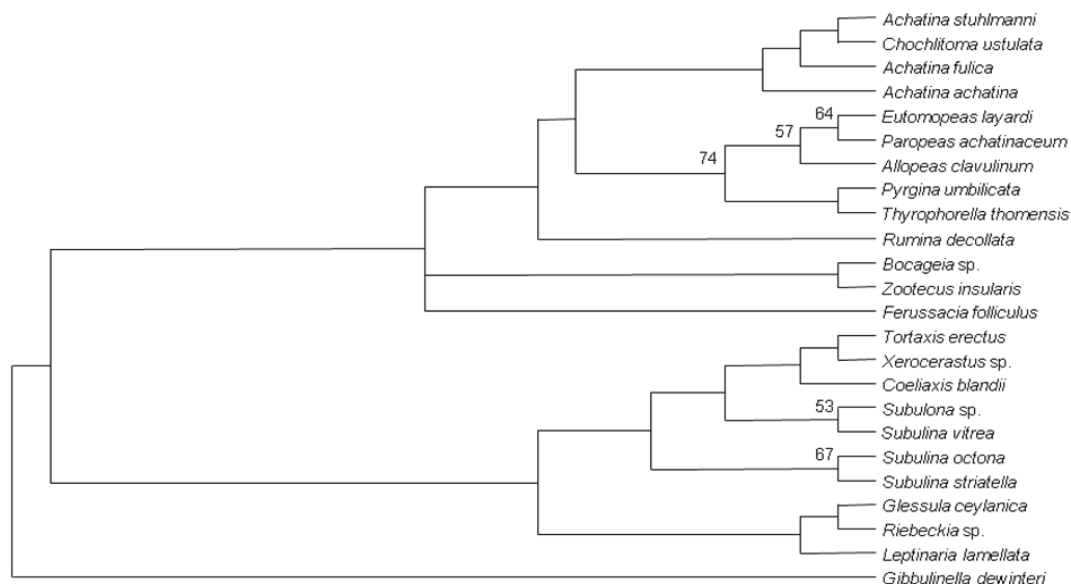
(A) BI (1st and 2nd codon positions of CO1)

Appendix 3.13: Phylogenetic trees of the Achatinoidea based on the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 446) and (C) maximum parsimony based on two equally parsimonious trees (p. 446). The phylogenies were constructed from 404 unambiguously aligned nucleotide sites and were rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. . The scale bar for (A) represents 1 substitutional change per 10 nucleotides and that of (B) represents 5 substitutional changes per 100 nucleotides. Note that *Ceceilioides gokweanus* and the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp. were excluded in the analyses as they were not successfully sequenced.

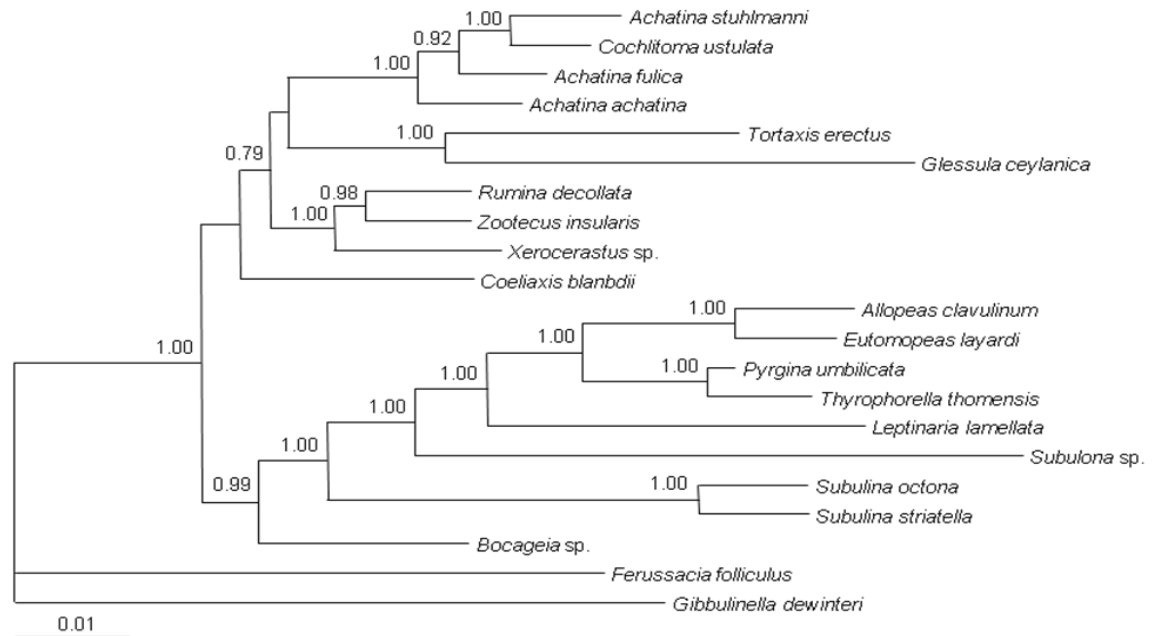
Appendix 3.13 (contd.)



(B) NJ (1st and 2nd codon positions of the CO1 gene)



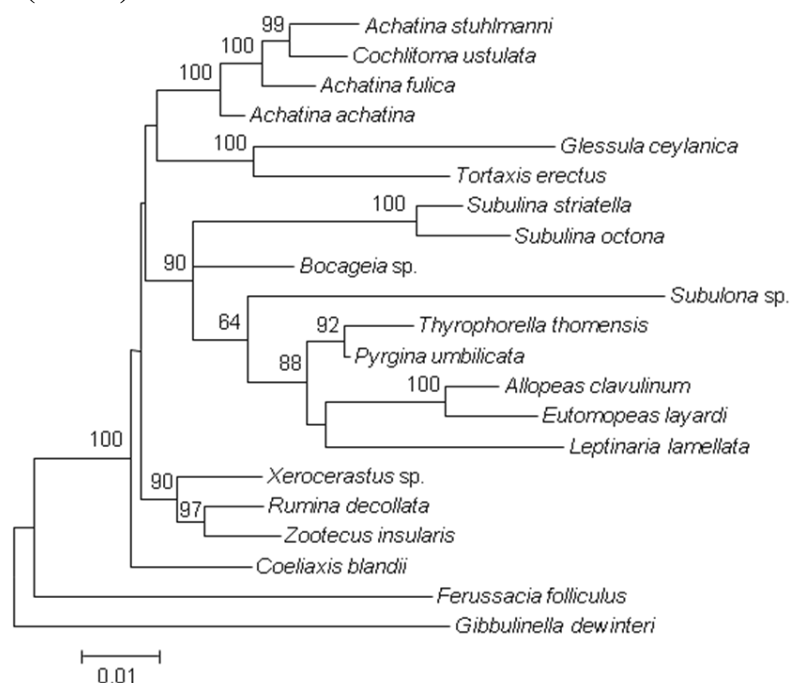
(C) MP (1st and 2nd codon positions of the CO1 gene)



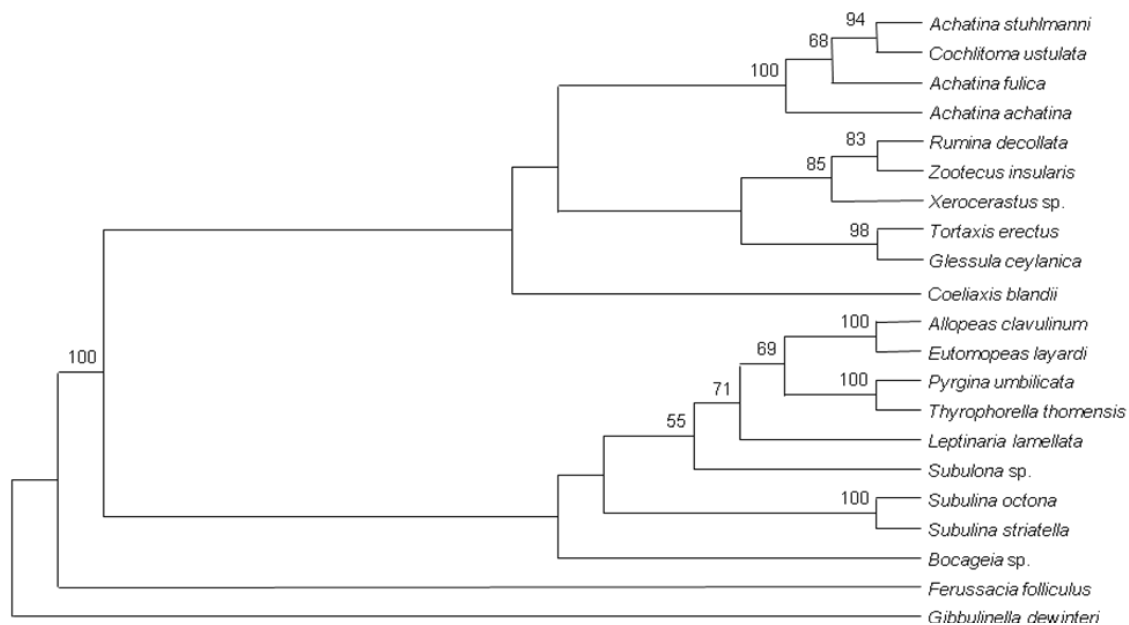
(A)BI (concatenated sequence – taxa with complete datasets only)

Appendix 3.14: Phylogenetic trees of the Achatinoidea (taxa with complete datasets only) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 448) and (C) maximum parsimony based on one most parsimonious tree (p. 448). The phylogenies were constructed from a concatenated sequence of 5028 nucleotides and were rooted on the streptaxid *Gibbulinella dewinteri*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bars for (A) and (B) represent 1 substitutional change per 100 nucleotides. Note that the achatinoid *Cecilioides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp. were not included in the phylogenies as they lack at least one of the genes used for the combined analyses.

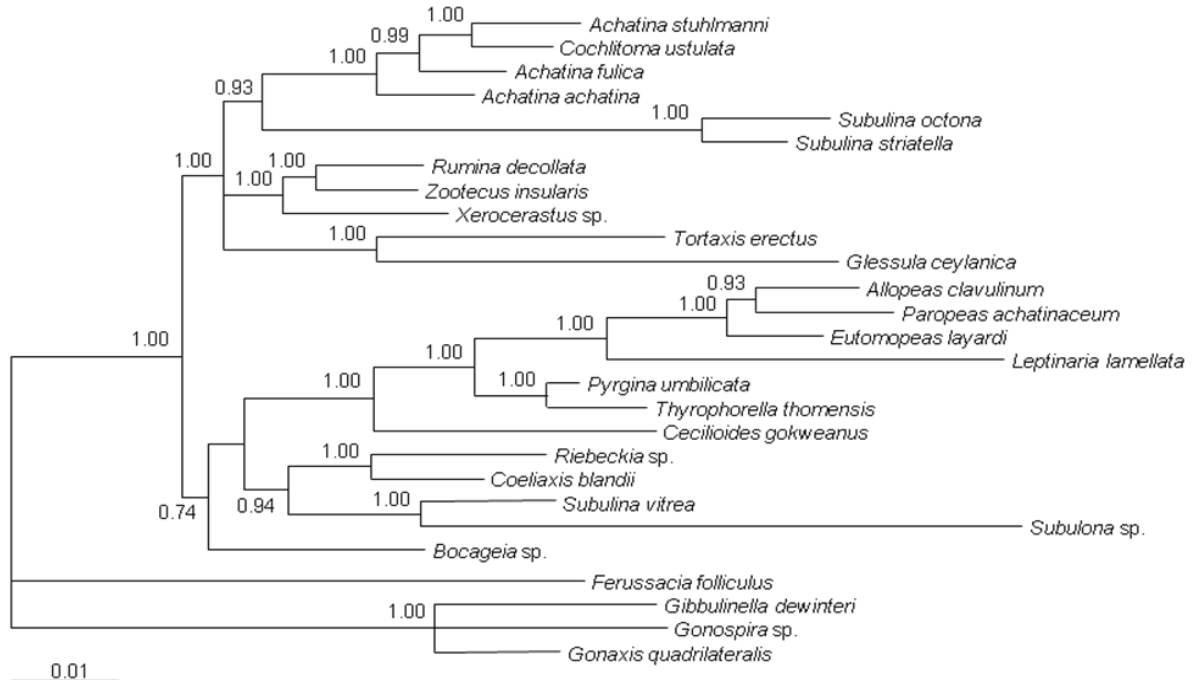
Appendix 3.14 (contd.)



(B) NJ (concatenated sequence – taxa with complete datasets only)



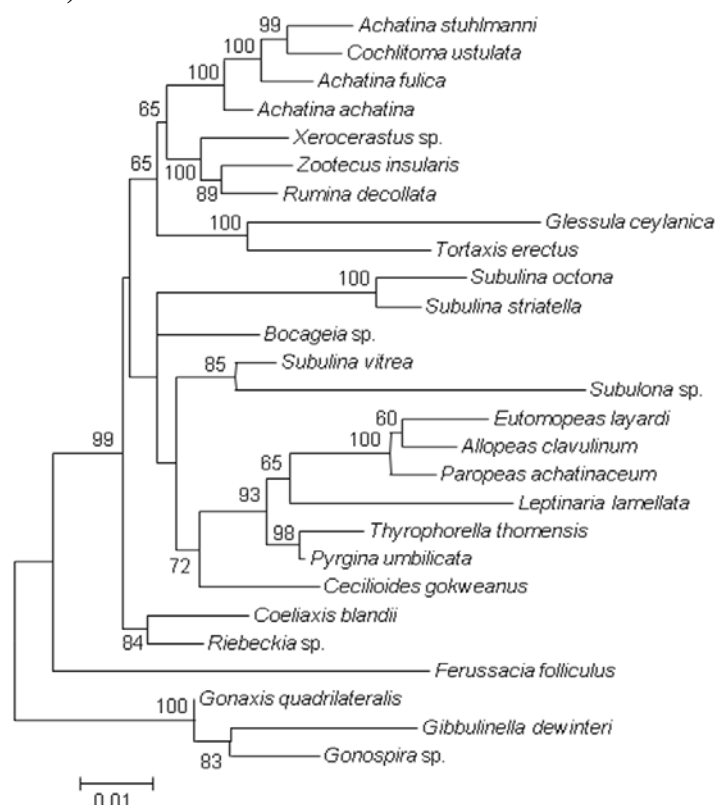
(C) MP (concatenated sequence – taxa with complete datasets only)



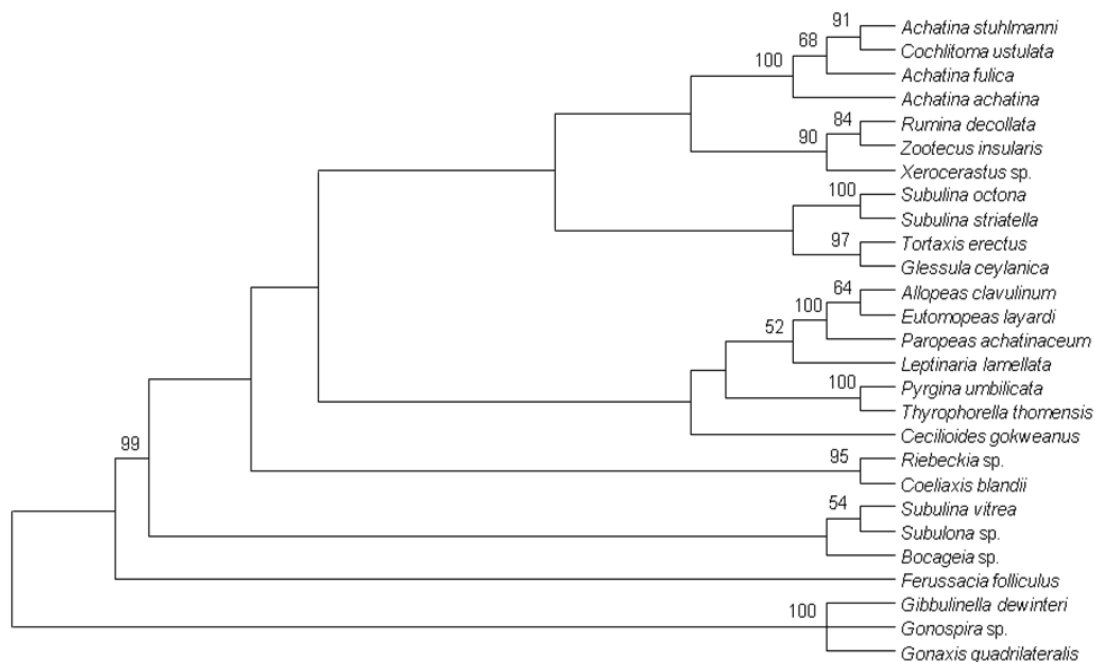
(A) BI (concatenated sequence – all taxa)

Appendix 3.15: Phylogenetic trees of the Achatinoidea (all taxa) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 450) and (C) maximum parsimony based on two equally parsimonious trees (p. 450). The phylogenies were constructed from a concatenated sequence of 5028 nucleotides and were rooted on the streptaxid *Gibbulinella dewinteri*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bars for (A) and (B) represent 1 substitutional change per 100 nucleotides.

Appendix 3.15 (contd.)



(B) NJ (concatenated sequence – all taxa)



(C) MP (concatenated sequence – all taxa)

	10	20	30	40	50	60
<i>Achatina fulica</i>	-----	-----A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Achatina immaculata</i>	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCACAT
<i>Achatina reticulata</i>	-----	-----	-----A-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Achatina zanzibarica</i>	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCATAT
<i>Euaethiopia loveridgei</i>	-----	-----A	TTGCA-GAAA	CCATTGAACA	TCGAC-CCTT	GAACGCACAT
<i>Achatina achatina</i>	-----AGAAT	TAATGTGA-A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Archachatina marginata</i>	-----	-----A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Achatina bisculpta</i>	-----	-----	-----A-GAAC	ACATTGAACA	TCGACACCTT	GAACGCATAT
<i>Achatina damarensis</i>	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCACAT
<i>Achatina stuhlmanni</i>	-----	-----	-----A-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Atopocochlis exarata</i>	-----	-----A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Limicolaria kamebul</i>	-----	-----A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Limicolaria martenssii</i>	-----	-----A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Limicolariopsis</i> sp.	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCATAT
<i>Limicolariopsis ruwenzori</i>	-----	-----	-----A-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma varicosa</i>	TGCGAGGAAA	TTATGAGA-A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma dimidiata</i>	-----	-----	-----GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma</i> sp. cf. <i>vestita</i>	-----	-----A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCATAT
<i>Cochlitoma marinae</i>	-----	-----	-----A-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma churchilliana</i>	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma granulata</i>	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma simplex</i>	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma kilburni</i>	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma montistempli</i>	-----	-----	-----	-----TTGAACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma omissa</i>	-----	-----	-----	-----TTGAACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma semidecussata</i>	-----	-----	-----	-----AACA	TCGACACCTT	GAACGCACAT
<i>Cochlitoma ustulata</i>	-----	-----	-----	-----AAC	ACATTGAACA	TCGACACCTT
<i>Cochlitoma zebra</i>	-----	-----	-----	-----TTGAACA	TCGACACCTT	GAACGCACAT
<i>Metachatina kraussi</i>	-----	-----	-----	-----TTGAACA	TCGACACCTT	GAACGCACAT
<i>Rumina decollata</i>	-----	-----A	TTGCA-GAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT
NUCLEOTIDES INCLUDED	-----	-----	-----	-----mmmm	mmmmmmmmmm	mmmmmmmmmm

451

	130	140	150	160	170	180
Achatina fulica	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCGC	TC---AATTT	ATTCCGAGAT	CCGTCCAGCT
Achatina immaculata	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCGC	TC---AATTT	ATTCCAGAT	CCGTCCAGCT
Achatina reticulata	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCAC	TC---AATTT	ATTCCGAGAT	CCGTCCAGCT
Achatina zanzibarica	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCGC	TC---AATTT	ATTCCGAGAT	CCGTCCAGCT
Euaethiopina loveridgei	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCGC	TC---AATTT	ATTCCGAGAT	CCGTCCAGCT
Achatina achatina	TTCGCTTCGC	ATGTGG-CAG	CAGGTCTTG-	-----ATTT	ATTCCGAGAT	CCGTCTAAAA
Archachatina marginata	TTCGCTTCGC	ATGTGGG-CAG	CAGGTCTCG-	-----ATTT	ATTCCGAGAT	CCGTCTA-TA
Achatina bisculpta	TTCGCTTCGT	ATGCGG-CAG	CAGGTCTCTC	TC--AAATTT	CCGTCCAGAT	CTTCCAC---
Achatina damarensis	TTCGCTTCGT	ATGCGG-CAG	CAGGTCTCTC	TC--AAATTT	CCGTCCAGAT	CTTCCCT----
Achatina stuhlmanni	TTCGCTTCGT	ATGCGG-CAG	CAGGTCTCTCG	TT--AAATTT	CCGTCCAGAT	CTTC-----
Atopocochlis exarata	TTCGCTTCGT	ATGCGG-CAG	CAGGTCTCTCG	TCC--AGCTT	CCTTCCAGCT	CTTAATTC--
Limicolaria kambeul	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC--AAATTT	CCGTCCAGCT	CTTCTTCTCTC
Limicolaria martenssii	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC--AAATTT	CCGTCCAGCT	CTTCTTCTCTC
Limicolariopsis sp.	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCAC	TC--AAATTT	CCGTCCAGCT	CTTCC-----
Limicolariopsis ruwenzori	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC--AAATTT	CCGTCCAGCT	CTTCTTCTCTC
Cochlitoma varicosa	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCGAGAT	CCGTCTAGCT
Cochlitoma dimidiata	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCGAGAT	CCGTCTAGCT
Cochlitoma sp. cf. vestita	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCAATAT	CCGTCTAGCT
Cochlitoma marinae	CTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AGTTT	ATTCCAAGAT	CCGTCCAGCT
Cochlitoma churchilliana	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TCCAAGATTT	ATTCCAAGAT	CCGTCCAGCT
Cochlitoma granulata	TTCGCTTCGT	ATGTGG-CAG	CAGGGCTCTCG	TCC---ATTT	ATTCCCAGAA	CCGGCTAGCT
Cochlitoma simplex	TTCGCTTCGT	ATGTGG-CAG	GAGGTCTCTCG	TC---AATTT	ATTCC-----	--GTCTAGCT
Cochlitoma kilburni	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCAGGAT	CCGTCTAGCT
Cochlitoma montistempli	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCAAGAT	CCGTCTAGCT
Cochlitoma omisa	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCAAGAT	CCGTCTAGCT
Cochlitoma semidecussata	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCAAGAT	CCGTCTAGCT
Cochlitoma ustulata	CTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCAAGAT	CCGTCTAGCT
Cochlitoma zebra	TTCGCTTCGT	ATGTGGG-CAG	CAGGTCTCTCG	TC---AATTT	ATTCCAAGAT	CCGTCTAGCT
Metachatina kraussi	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCTCG	TC--AATTTT	ATTCCAAGAT	CCGTCTAGCT
Rumina decollata	TTCGCTTCGC	TTCTGG-CAG	CTGGTCCGGT	TC-----	-----	-----
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmm-mmm	mmmmmmmmmm-	-----	-----	-----

	190	200	210	220	230	240
Achatina fulica	CTTC-----C	TCTATCC-AT	CCGCGG-CTC	GTGCGGA---	-----GGGG	TTACAGAGAG
Achatina immaculata	CTTCTCTTCC	TCTATCC-AT	CCGCGG-CTC	GTGCGGG---	-----GGGG	G-ACAGAAAG
Achatina reticulata	CTTC--AACC	TCTATCC-AT	CCGCGG-CTC	GTGCGGG---	-----GGG-	TTACAGAGAG
Achatina zanzibarica	CTTC-----C	TCTATCC-AC	CCGCGG-CTC	GTGCGGG---	-----GGGG	TTACAGAGAT
Euaethiopina loveridgei	CTTC-----C	TCTATCC-AT	CCGCGG-CTC	GCGCGGA---	-----GGGG	TTACAGAGAG
Achatina achatina	CTTC---TTG	CTCTGCCGTN	CCGCTG-CTC	GTGCTGG---	-----GG	ACACGGAAAA
Archachatina marginata	CTTCNTGNTC	TANTGNCGTC	CCGCTG-CTC	GTGCTGG---	-----G	ACACGGAAAA
Achatina bisculpta	-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAG
Achatina damarensis	-----C	TAAATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAG
Achatina stuhlmanni	-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAG
Atopocochlis exarata	-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAG
Limicolaria kambeul	G-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAG
Limicolaria martenssii	G-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACWGAAG
Limicolariopsis sp.	-----T	CGAATCT--T	CCGCGG-CTC	GTGCGG---	-----G	GGACAGAAAG
Limicolariopsis ruwenzori	G-----C	TCGATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAG
Cochlitoma varicosa	CTTC-----C	TCTATCTTAT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAG
Cochlitoma dimidiata	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAG
Cochlitoma sp. cf. vestita	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAG
Cochlitoma marinae	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGAAAGGAAG
Cochlitoma churchilliana	CTTC-----	-----TTCCT	CCGCGG-CTC	GTGCGGT---	-----GG	GGACACAAAG
Cochlitoma granulata	CTTC-----C	TCTATCT-AT	CCGCGGGCTC	GGGGGGG---	-----GGA	GGACAGAAAG
Cochlitoma simplex	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAG
Cochlitoma kilburni	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAG
Cochlitoma montistempli	CTTC-----C	TCYATCT-AT	CCGCGGGCTC	GTGCGGTGGA	GAAAGGAAGA	AGACAGAAAG
Cochlitoma omisa	CTTC-----C	TCTATCT-AT	CCGCGGGCTC	GTGCGGT---	-----GGA	GGACAGAAAG
Cochlitoma semidecussata	CTTC-----C	TCTATCT-AA	CCGCGGGCTC	GTGCGGT---	-----GGA	GGACAGAGAG
Cochlitoma ustulata	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAG
Cochlitoma zebra	CTTC-----C	-----T	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAG
Metachatina kraussi	CTTCCCT--C	TCTATCTTAT	CTGCGG-CTC	GTGCGGT---	---TT---	GGACAGAAAG
Rumina decollata	-TACCACGTC	GTTTTTCACT	CCGTG--CTC	GTGCGA---	-----	---GGAGAAAG
NUCLEOTIDES INCLUDED	-----	-----	-----mmmm	mmmm-----	-----	-----

	250	260	270	280	290	300			
Achatina fulica	GAAGAAT-CG	GG-CG-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Achatina immaculata	GAAGAAT-CG	GG-CG-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Achatina reticulata	GAAGAAT-CG	GG-CG-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Achatina zanzibarica	GAAGAAT-CG	GG-CG-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Euaethiopina loveridgei	GAAGAAT-CG	GG-CG-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Achatina achatina	GAAGAAT-TG	GGCG-----	-----GATA	AGGAAGGGGA	AGAGAGACCG	TCGAAGCGCT			
Archachatina marginata	GAAGAAT-TG	GA-CNN-----	-----ATA	NNGAAGGG-A	AGAGAGGCCG	TCGAAGCGCT			
Achatina bisculpta	AAAGAAT-CT	GG-CG-----	-----GAGAA	GCGGAGACCG	TCGAAGCGCT				
Achatina damarensis	AAAGAAT-CT	GG-CG-----	-----GAGAA	GCGGAGACCG	TCGAAGCGCT				
Achatina stuhlmanni	AAAGAAT-CT	GT-CG-----	-----GAGAA	AAAGAGACCG	TCGAAGCGCT				
Atopocochlis exarata	GAAGAAT-CT	GG-CG-----	-----GAGA	AGAGAGACCG	TCGAAGCGCT				
Limicolaria kambeul	GAAGAAT-CT	GG-CG-----	-----GA	GAAGAGACCG	TCGAAGCGCT				
Limicolaria martenssii	GAAGAAT-CT	GG-CG-----	-----GA	GAAGAGACCG	TCGAAGCGCT				
Limicolariopsis sp.	GAAGAAT-CT	GG-CG-----	-----GA	GAAGAGACCG	TCGAAGCGCT				
Limicolariopsis ruwenzori	GAAGAAT-CT	GG-CG-----	-----GA	GAAGAGGCCG	TCGAAGCGCT				
Cochlitoma varicosa	GCGGAGAA--	-----	-----GGAA	GAAGAGACCG	TCGAAGCGCC				
Cochlitoma dimidiata	GCGGAGAA--	-----GGA	AGAAGAAGAA	GAAGNGNCCG	TCGAAGCGCT				
Cochlitoma sp. cf. vestita	GCGGAGAA--	-----	-----GGAAGAA	GAAGAGACCG	TCGAAGCGCT				
Cochlitoma marinae	GCG-----	-----G	AGAAGGAAGA	AGAAGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Cochlitoma churchilliana	GAAGAA--TCG	GGCG-----	--GAGAAGGG	ACTAAGAAAG	GAAGAGACCG	TCGAAGCGCT			
Cochlitoma granulata	GCG-----	-----	-----GA	NAAAGAAGAA	GAAGANACCG	TCGAAGCGCT			
Cochlitoma simplex	GCG-----	-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Cochlitoma kilburni	GCG-----	-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Cochlitoma montistempli	GCG-----	-----	-----GAGAA	GGAAGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Cochlitoma omissa	GCG-----	-----	-----GAGAA	GGAAGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Cochlitoma semidecussata	GCG-----	-----G	AGAAGGAAGA	AGAAGAAGAA	GAAGAGACCG	CCGAAGCGCT			
Cochlitoma ustulata	GCG-----	-----	-----GCGAA	GGAAGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Cochlitoma zebra	GCG-----	-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT			
Metachatina kraussi	GAAGAA--TCG	GGCG--GAGA	AGGAAGAAGA	AGAAGAAGAA	GCAGAGACCG	TCGAAGCGCT			
Rumina decollata	CTGCGTCGTG	GATGA-----	-----	-----	-TGAAGACCG	TCGAAGCGCA			
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	-----	-----	-----

	310	320	330	340	350	360			
Achatina fulica	GTGGGCCGTC	GTGGGAGGAG	GAAGAA----	GGGTTTT---	-----T	TTCCACCTTT			
Achatina immaculata	GTGGGCCGTC	GTGGGAGGAG	GAGGAAGA-A	GGGTTTACC	CCCCTTTTTT	--ACACTCTT			
Achatina reticulata	GTGGGCCGTC	GTGGGCCGAG	GAGGAAGA-A	GGGTCTCACC	CCCCTTTTTT	CCCCTCTCTT			
Achatina zanzibarica	GTGGGCCGTC	GTGGGAGGAG	GAAGAA----	GGGTTTCACC	CCCCTTTTTT	CCC-ACCTTT			
Euaethiopina loveridgei	GTGGGCCGTC	GTGGGAGGAG	GAAGAA----	GGGTTTCACC	CCCCTTTTTT	TTCCACTCTT			
Achatina achatina	CTGGGCCGTC	GTGGGAGAGA	GG-AGGGA--	-----	--TTGTTTTT	TTTTTTTCTT			
Archachatina marginata	CTGGGCCGTC	GTGGGAGAGG	AGGAGAGG--	-----GATT	TTGGATTTTT	CTTCTTTCTT			
Achatina bisculpta	GTGGGCCGTC	GTGGGATGAG	GAAGAAGT--	GGGTTTTTCC	CAG---TTTT	TT-CCCTCTT			
Achatina damarensis	GTGGGCCGTC	GTGGGATGAG	GGTGAAGA--	-----ACC	CAG---TTTT	TTTTCTCTCT			
Achatina stuhlmanni	GTGGGCCGTC	GTGGGATGAG	GAATAACT--	GGGTTTTTCC	CAG---TTT	TTTTTCTCTT			
Atopocochlis exarata	GTGGGCCGTC	GCGGGATGAG	GAAGAAGT--	GGGTTTTCCC	CAG---TTT	TTTTTCTCTT			
Limicolaria kambeul	GTGGGCCGTC	GTGGGATGAG	GAAGAAGT--	GGGTT-----	-----T	TTCCCTCTTT			
Limicolaria martenssii	GTGGGCCGTC	GTGGGATGAG	GAAGAAGT--	GGGTT-----	-----	TTTCCCTCTT			
Limicolariopsis sp.	GTGGGCCGTC	GTGGGATGAG	GAATTACT--	GGGTTT-ACC	CAG-----TT	TTTTTCTATT			
Limicolariopsis ruwenzori	GTGGGCCGTC	GTGGGATGAG	GAAGAAGT--	GGCTT-----	-----TT	TTCCCTCTTT			
Cochlitoma varicosa	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	-GCTTTTTCT	TTT-----	--CCTCTCTT			
Cochlitoma dimidiata	GTGGGCCGTN	GTGGGAGGAG	GAGGAAAA--	AGTTTTTT--	-----	--CCTCTCTT			
Cochlitoma sp. cf. vestita	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	-GCTTTTTTTT	TT-----	--CCTCTCTT			
Cochlitoma marinae	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	-GCTTTTTTTT	T-----	--TCTCTCTT			
Cochlitoma churchilliana	GTGGGCCGTC	GTGGGAGGGA	GGAAGAAG--	TGCTTTTTTT	CCC-----	--CCTGTCTT			
Cochlitoma granulata	GTGGGCCGTN	GNGGGANNAC	GAAGAAGT--	-GNTNTTTTT	TCT-----	--NCTCTCTT			
Cochlitoma simplex	GTGGGCCGTC	GTGGGAGGAG	GAGGAAGAAG	TGCTCTTATG	TTTTT-----	CCCCCTCTTT			
Cochlitoma kilburni	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	GCTTTTTTTT	TCTTT-----	--CCTCTCTT			
Cochlitoma montistempli	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	GCTTTTTTTT	-----	CTGCTCTCTT			
Cochlitoma omissa	GTGGGCCGTC	GTGGGAGGAG	GAAGAATT--	GCTTTTTTTT	-----	CTCCTCTCTT			
Cochlitoma semidecussata	GTGGGCCGTC	GTGGGAGGAG	GAAGAAG--T	GCTTTTTTTT	T-----	--CCTCTCTT			
Cochlitoma ustulata	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	GCTTTTTTTT	-----	--TCTCTCTT			
Cochlitoma zebra	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	GCTTTTTTTT	CC-----	--CCTCTCTT			
Metachatina kraussi	GTGGGCCGTC	GTGGGAGGAG	GAAGAGTG--	CTTTTTTTTT	-----	--CCCCTCTT			
Rumina decollata	CTGGGCTTTC	G-AGGACGAG	GAGGATCG-A	GTTTTCTCT-	-----	-TTCCACCTT			
NUCLEOTIDES INCLUDED	-----	m-----	-----	-----	-----	-----	-----	-----	-----

	370 380 390 400 410 420
Achatina fulica	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Achatina immaculata	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Achatina reticulata	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Achatina zanzibarica	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Euaethiopina loveridgei	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Achatina achatina	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Archachatina marginata	CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Achatina bisculpta	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Achatina damarensis	-TCCCCGTGG CCTCAAGTAC ACTCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Achatina stuhlmanni	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Atopocochlis exarata	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTC---GCT
Limicolaria kambeul	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Limicolaria martenssii	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Limicolariopsis sp.	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Limicolariopsis ruwenzori	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma varicosa	-CCCCCGTGG CCTCAAGTAA ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma dimidiata	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGNGA CTCTTTCGCT
Cochlitoma sp. cf. vestita	-CCCCCGTGG TCTCAAGTAT ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCCTTTCGCT
Cochlitoma marinae	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGAGA CTCTTTCGCT
Cochlitoma churchilliana	TCCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma granulata	-CCCCCGTGN CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma simplex	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma kilburni	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma montistempli	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma omisa	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma semidecussata	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Cochlitoma ustulata	-TCCCCGGGG CCTCAAGTAC ACGCATGCGC CGTCCTCGGC ATTTTCAGCAA CTCCTTTCGCT
Cochlitoma zebra	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Metachatina kraussi	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTCGCT
Rumina decollata	-CCCTCGTAG CCTCAAGTAC ACGAATGCGC CGTCCCCGTC ATCTGAGCA- -----TC
NUCLEOTIDES INCLUDED	--mmmmmmmm mmmmmmmmm mm--mmmm mmmmm-mmm m---mmmm -----

	430 440 450 460 470 480
Achatina fulica	---CGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Achatina immaculata	---CGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Achatina reticulata	---CGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGTTTCGCC GTCCGTCCGG CAGGACTCGG
Achatina zanzibarica	---CGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Euaethiopina loveridgei	---CGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Achatina achatina	---TGCCCCG AGCGCC-CGT CTTCGTTT-G TTCGC-----C GTCCGTCCGG CAGGACTCGG
Archachatina marginata	---TGCCCCG AGCGCC-CGT CTTCGTTT-G TTCGC-----C GTCCGTCCGG CAGGACTCGG
Achatina bisculpta	---CGCCAGG AGCGCTCCAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Achatina damarensis	---CGCCCGG AGCGCTCCAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Achatina stuhlmanni	---CGCCCGG AGCGCTCCAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Atopocochlis exarata	---CGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Limicolaria kambeul	-----CNT GTTCG-----CTCGCC GTCCGTCCGG CAGGACTCGG
Limicolaria martenssii	-----CAT CTTCGTTT-G TTCGTTTCGCC GTCCGTCCGG CAGGACTCGG
Limicolariopsis sp.	---CGCCCGG AGCACT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Limicolariopsis ruwenzori	-----CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCAG CAGGACTCGG
Cochlitoma varicosa	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma dimidiata	GCTCNCNCCG ANNGNT-CAT NTNNNTTTG TTCGCTTNC GTCCNTCCGN CAGGACTCGG
Cochlitoma sp. cf. vestita	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma marinae	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma churchilliana	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma granulata	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma simplex	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma kilburni	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma montistempli	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma omisa	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma semidecussata	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Cochlitoma ustulata	GGTCGCCCGG AGCGT-CAT CTTCGTTT-G TTCGCTCGCC GKCCGTCCGG CAGGACTCGG
Cochlitoma zebra	GCTCGCCCGG AGCGCT-CGT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Metachatina kraussi	GCTCGCCCGG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
Rumina decollata	GCTCTCTCGA AGCGCT-CAT CTTCGTTT-G CTCGC-----C GTCCGTCTGG CAGGACTCGG
NUCLEOTIDES INCLUDED	-----mmmm mmmmm-----

	490 500 510 520 530 540
Achatina fulica	CTCGCTTCTT T----- AACAAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Achatina immaculata	CTCGCTTCTT T-----G AAAAAATCGA GCCTGCCAGA C---CGTCCA AAGCGGCTAG
Achatina reticulata	CTCGCTTCTT T----- AACAAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Achatina zanzibarica	CTCGCTTTT TTT----- AACAAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Euaethiopina loveridgei	CTCGCTTCTT T----- AACAAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Achatina achatina	CTCGCTT--- ----- --TTATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Archachatina marginata	GTCGCTT--- ----- --TTATCGA GCCTGCCAGA C---TGTCCG AAGCGGCTAG
Achatina bisculpta	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Achatina damarensis	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Achatina stuhlmanni	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Atopocochlis exarata	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Limicolaria kambeul	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Limicolaria martenssii	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Limicolariaopsis sp.	CTCGCTTCTT TTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Limicolariaopsis ruwenzori	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma varicosa	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma dimidiata	CTCGNTTNTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AATCGGCTAG
Cochlitoma sp. cf. vestita	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma marinae	CTCGCTTCTT TTTGTTT--- --AATCGA GCCTGCCAGA CAC-CGTCCG AAGCGGCTAG
Cochlitoma churchilliana	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma granulata	CTCGCTTCTT TTTTNTT--- --ANTCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma simplex	CTCGCTTCTT TTTTNTTNTT TTTTAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma kilburni	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma montistempli	CTCGCTTCAT TTTTATT--- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma omissa	CTCGCTTCAT TTTTATT--- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma semidecussata	CTCGCTTCTT TTT-ATT--- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma ustulata	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Cochlitoma zebra	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Metachatina kraussi	CTCGCTTCTT TTTAT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
Rumina decollata	CTCGCTGTG- ----- AATAAAACGA GCCTGCC- GA C---CGTCCG AAGTGGCATG
NUCLEOTIDES INCLUDED	mmmmmm----- -----m mmmmmmm-mm m---mmmmmm mmmmmmm--m

	550 560 570 580 590 600
Achatina fulica	GG-ATTG-CG AAGTGGGG-- --CGCGCAA GCGCATGGG-- --TCTGCTG CGGCGGCGCC
Achatina immaculata	GG-ATTG-CG AAGTGGGG-- --CGCGCAA GCGCATGGG-- --TCTGCTG CGGCGGCGCC
Achatina reticulata	GG-ATTG-CG AAGTGGGG-- --CGCGCAA GCGCATGGG-- --TCTGCTG CGGCGGCGCC
Achatina zanzibarica	GG-ATTG-CG AAGTGGGG-- --CGCGCAA GCGCATGGGG-- --TCTGCTG CGGCGGCGCC
Euaethiopina loveridgei	GG-ATTG-CG AAGTGGGG-- --CGCGCAA GCGCATGGG-- --TCTGCTG CGGCGGCGCC
Achatina achatina	CG-GTTG-CG AAGCGGGT-- --GGCGCGTAA GCG-ATCGG-- --TCT---G CGGCGGCGCC
Archachatina marginata	CG-GTTG-CG AAGTGGGT-- --GGCGGGCAA GCG-ATCGG-- --TCT---G CGGCGGCGCC
Achatina bisculpta	CG-ATTG-CG AAGTGGAG-- ----- GGG-- --TCTGCTG CGGCGGCGCC
Achatina damarensis	CG-ATTG-CG AAGTGGGG-C GCGTGAGCGC TGGCATGTTG GGGTCTGCTG CGGCGGCGCC
Achatina stuhlmanni	CG-ATTG-CG AAGTGGGG-- --CGCCCGCA GCGGATTGGG GGGTCTGCTG TGCGGCGCC
Atopocochlis exarata	CG-AATG-CG AAGTGGGG-- --CCCGGGAG CGGCATGGG-- --TCTGCTG CGGCGGCGCC
Limicolaria kambeul	CG-ATTGGCG AAGTG----- ----- --CATGGG-- --TCTGCTG AGGCGGCGCC
Limicolaria martenssii	CG-ATTGGCG AAGTG----- ----- --CATGGG-- --TCAGCTG AGGCGGCGCC
Limicolariaopsis sp.	TG-ATTG-CG AAGTGGGT-- --CGCGGGT GCGCATGGG-- --TCTGCTG CGGCGGCGCC
Limicolariaopsis ruwenzori	CG-ATTGGCG AAGTG----- ----- --CATGGG-- --TCTGCTG CGGCGGCGCC
Cochlitoma varicosa	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAGCATGGG-- --GTCTGCTG CGGCGGCGCC
Cochlitoma dimidiata	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAGCATGGGG GAGTCTGCTG CGGCGGCGCC
Cochlitoma sp. cf. vestita	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAGCATGGG-- --TCTGCTG CGGCGGCGCC
Cochlitoma marinae	CGGATTG-CG AAGTGGGG-- --CGCGCGA GCGCATGGG-- --TCTGTTG CGGCGGCGCC
Cochlitoma churchilliana	CG-AGTG-CG AAGTGGGG-- --GGCGCGCA GCGCATGGG-- --TCTGCTG CGGCGGCGCC
Cochlitoma granulata	CGGATTG-NG AAGNNGGG-- CGCGCGAGCG CAACATGGGG G--TCTGNTG CGGCGGCGCC
Cochlitoma simplex	CGGATTG-CG AAGTGGGGCG CACGCGAGCG CAGCATAGG-- --TCTGTTG CGGCGGCGCC
Cochlitoma kilburni	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAGCATGGG-- --TCTGCTG CGGCGGCGCC
Cochlitoma montistempli	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAACATGGGG GG-TCTGCTG CGGCGGCGCC
Cochlitoma omissa	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAACATGGGG G--TCTGCTG CGGCGGCGCC
Cochlitoma semidecussata	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAACATGGGG G--TCTGCTG CGGCGGCGCC
Cochlitoma ustulata	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CA---TGGG-- --TCTGTTG CGGCGGCGCC
Cochlitoma zebra	CGGATTG-CG AAGTGGGT-- CGCGCGAGCG CAGCATGGG-- --TCTGTTG CGGCGGCGCC
Metachatina kraussi	CG-ATTG-CG AAGTGGGG-- CGCGCGAGCG CA---TGGG-- --TCTGCTG CGGCGGCGCC
Rumina decollata	CG-GTTT-CG AAGTGTGA-- ----- GAGG GCGATTGCCG CGGCGGCGCC
NUCLEOTIDES INCLUDED	mm-----mm mmmmm----- -----m -----m mmmmmmmmm

	610	620	630	640	650	660		
Achatina fulica	A-----	----GTCTT	TCT-----	--AAAATCTT	TCTTTA----	-TCCGACCTC		
Achatina immaculata	A-----	----GTCTT	TCT-----	--AAAATCTT	TCTTTA----	-TCCGACCTC		
Achatina reticulata	A-----	----GTCTT	TCT-----	--AAAATCTT	TCTTTA----	-TCCGACCTC		
Achatina zanzibarica	A-----	----GTCTT	TCT-----	--AAAATCTT	TCTTTA----	-TCCGACCTC		
Euaethiopina loveridgei	A-----	----GTCTT	TCT-----	--AAAATCTT	TCTTTA----	-TCCGACCTC		
Achatina achatina	ACAACA----	----GTCTT	TATACGAACG	AACGTTCCGGT	TCTTTA----	-TCCGACCTC		
Archachatina marginata	AGCA-----	----GTCTC	TATAACGACG	ACGTTTNNTT	TAAAATTTT	ATCCGACCTC		
Achatina bisculpta	A-----	----GTCTT	TCT-----	--AAAATATT	TCTTTA----	-TCCGACCTC		
Achatina damarensis	A-----	----GTCTT	TCT-----	AAAAAATCTT	TCTTTA----	-TCCGACCTC		
Achatina stuhlmanni	A-----	----GTCTT	TCT-----	--AAAATTTT	TCTTTA----	-TCCGACCTC		
Atopocochlis exarata	A-----	----GTCTT	TCT-----	AAAAAATCTT	TCTTTA----	-TCCGACCTC		
Limicolaria kambeul	A-----	----GTCTT	TCT-----	--AAAATTTT	TCTTTA----	-TCCGACCTC		
Limicolaria martenssii	A-----	----GTCTT	TCT-----	--AAAATTTT	TCTTTA----	-TCCGACCTC		
Limicolariopsis sp.	A-----	----GTCTT	TCT-----	--AAAATCTT	TCTTTA----	-TCCGACCTC		
Limicolariopsis ruwenzori	A-----	----GTCTT	TCT-----	--AAAATTTT	TCTTTA----	-TCCGACCTC		
Cochlitoma varicosa	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma dimidiata	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma sp. cf. vestita	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma marinae	AA-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma churchilliana	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma granulata	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma simplex	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma kilburni	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma montistempli	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma omisa	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma semidecussata	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma ustulata	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Cochlitoma zebra	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Metachatina kraussi	A-----	----GTCTT	TCT-----	-----TTCAT	TCTTTA----	-TCCGACCTC		
Rumina decollata	A----ATTTT	TTTTTGTCGG	TTCT-----	--AAA-----	-----	-TCCGACCTC		
NUCLEOTIDES INCLUDED	m-----	-----	-----	-----	-----	-----	-----	-----

	670	680	690	700	710	720	
Achatina fulica	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Achatina immaculata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Achatina reticulata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Achatina zanzibarica	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Euaethiopina loveridgei	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Achatina achatina	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Archachatina marginata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Achatina bisculpta	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Achatina damarensis	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Achatina stuhlmanni	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Atopocochlis exarata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Limicolaria kambeul	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Limicolaria martenssii	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Limicolariopsis sp.	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Limicolariopsis ruwenzori	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma varicosa	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma dimidiata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma sp. cf. vestita	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma marinae	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma churchilliana	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma granulata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma simplex	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma kilburni	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma montistempli	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma omisa	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma semidecussata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma ustulata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Cochlitoma zebra	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Metachatina kraussi	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
Rumina decollata	AGATCGGACG	AGATTACCCG	CTGAATTTAA	GCATATAACT	AAGCGGAGGA	AAAGAAACTA	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	730	740	750	760	770	780	
Achatina fulica	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Achtina immaculata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Achatina reticulata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Achatina zanzibarica	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Euaethiopina loveridgei	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Achatina achatina	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Archachatina marginata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Achatina bisculpta	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Achatina damarensis	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Achatina stuhlmanni	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Atopocochlis exarata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Limicolaria kambeul	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Limicolaria martenssii	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Limicolariopsis sp.	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Limicolariopsis ruwenzori	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma varicosa	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma dimidiata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma sp. cf. vestita	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma marinae	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma churchilliana	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma granulata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma simplex	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma kilburni	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma montistempli	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma omissa	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma semidecussata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma ustulata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Cochlitoma zebra	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Metachatina kraussi	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
Rumina decollata	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAAGAG	CCCAGCACCG	AATCCCCCAG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	790	800	810	820	830	840	
Achatina fulica	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Achtina immaculata	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Achatina reticulata	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Achatina zanzibarica	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Euaethiopina loveridgei	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Achatina achatina	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Archachatina marginata	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Achatina bisculpta	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Achatina damarensis	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Achatina stuhlmanni	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Atopocochlis exarata	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Limicolaria kambeul	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Limicolaria martenssii	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Limicolariopsis sp.	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Limicolariopsis ruwenzori	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma varicosa	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma dimidiata	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma sp. cf. vestita	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma marinae	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma churchilliana	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma granulata	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma simplex	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma kilburni	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma montistempli	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma omissa	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma semidecussata	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma ustulata	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Cochlitoma zebra	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Metachatina kraussi	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
Rumina decollata	TGTCACGCTG	GCGGGAAGAG	TGGTGTGTGG	GACGCCACCA	GTGCGCATCAG	AGGGCGTCGA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	
	850 860 870 880 890 900	
Achatina fulica	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Achatina immaculata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Achatina reticulata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Achatina zanzibarica	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Euaethiopina loveridgei	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Achatina achatina	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Archachatina marginata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Achatina bisculpta	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Achatina damarensis	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Achatina stuhlmanni	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Atopocochlis exarata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Limicolaria kambeul	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Limicolaria martenssii	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Limicolariopsis sp.	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Limicolariopsis ruwenzori	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma varicosa	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma dimidiata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma sp. cf. vestita	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma marinae	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma churchilliana	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma granulata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma simplex	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma kilburni	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma montistempli	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma omissa	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma semidecussata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma ustulata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Cochlitoma zebra	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Metachatina kraussi	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
Rumina decollata	AGTCCTCCTG ATCGGGGCTT CACCCAGAGC GGGTGTAAGG CCTTTGCAGG CGCCTCTCTG	
NUCLEOTIDES INCLUDED	mmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm	

	
	910 920 930 940 950 960	
Achatina fulica	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Achatina immaculata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Achatina reticulata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Achatina zanzibarica	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Euaethiopina loveridgei	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Achatina achatina	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Archachatina marginata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Achatina bisculpta	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Achatina damarensis	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Achatina stuhlmanni	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Atopocochlis exarata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Limicolaria kambeul	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Limicolaria martenssii	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Limicolariopsis sp.	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Limicolariopsis ruwenzori	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma varicosa	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma dimidiata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma sp. cf. vestita	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma marinae	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma churchilliana	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma granulata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma simplex	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma kilburni	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma montistempli	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma omissa	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma semidecussata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma ustulata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Cochlitoma zebra	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Metachatina kraussi	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
Rumina decollata	TGCGGCCGCG AGCGTCTCAG GAGTCGGGTT GTTTGGGAAT GCAGCCCAAA GCGGGTGCGTA	
NUCLEOTIDES INCLUDED	mmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm	

	970	980	990	1000	1010	1020
Achatina fulica	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Achtina immaculata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Achatina reticulata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Achatina zanzibarica	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Euaethiopina loveridgei	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Achatina achatina	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Archachatina marginata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Achatina bisculpta	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Achatina damarensis	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Achatina stuhlmanni	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Atopocochlis exarata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Limicolaria kambeul	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Limicolaria martenssii	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Limicolariopsis sp.	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Limicolariopsis ruwenzori	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma varicosa	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma dimidiata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma sp. cf. vestita	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma marinae	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma churchilliana	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma granulata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma simplex	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma kilburni	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma montistempli	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma omissa	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma semidecussata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma ustulata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Cochlitoma zebra	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Metachatina kraussi	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
Rumina decollata	AACTCCATCT	AAGGCTAAAT	ACTGGCACGA	GTCCGATAGC	GGACAAGTAC	CGTGAGGGAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1030	1040	1050	1060	1070	1080
Achatina fulica	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Achtina immaculata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Achatina reticulata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Achatina zanzibarica	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Euaethiopina loveridgei	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Achatina achatina	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Archachatina marginata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Achatina bisculpta	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Achatina damarensis	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Achatina stuhlmanni	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Atopocochlis exarata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Limicolaria kambeul	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Limicolaria martenssii	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Limicolariopsis sp.	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Limicolariopsis ruwenzori	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma varicosa	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma dimidiata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma sp. cf. vestita	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma marinae	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma churchilliana	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma granulata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma simplex	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma kilburni	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma montistempli	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma omissa	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma semidecussata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma ustulata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Cochlitoma zebra	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Metachatina kraussi	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
Rumina decollata	AGTTGAAAAG	AAC TTTGAAG	AGAGAGTTCA	AGAGTACGTG	AAACCGCCCA	GAGGTAAACG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1090	1100	1110	1120	1130	1140		
Achatina fulica	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Achatina immaculata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Achatina reticulata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Achatina zanzibarica	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Euaethiopina loveridgei	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Achatina achatina	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Archachatina marginata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Achatina bisculpta	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCT-G	GGGCGATCGC		
Achatina damarensis	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCT-G	GGGCGATCGC		
Achatina stuhlmanni	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCT-G	GGGCGATCGC		
Atopocochlis exarata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Limicolaria kambeul	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCAGCCTCG	GGGCTATCGC		
Limicolaria martenssii	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCAGCCTCG	GGGCTATCGC		
Limicolariopsis sp.	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCTATCGC		
Limicolariopsis ruwenzori	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCAGCCTCG	GGGCTATCGC		
Cochlitoma varicosa	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma dimidiata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma sp. cf. vestita	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma marinae	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma churchilliana	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma granulata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma simplex	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma kilburni	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma montistempli	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma omissa	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma semidecussata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma ustulata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Cochlitoma zebra	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Metachatina kraussi	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC		
Rumina decollata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCT-G	GGGCTATTGC		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm-m	mmmmmmmm		

	1150	1160	1170	1180	1190	1200	
Achatina fulica	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Achatina immaculata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Achatina reticulata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Achatina zanzibarica	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Euaethiopina loveridgei	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Achatina achatina	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Archachatina marginata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Achatina bisculpta	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Achatina damarensis	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Achatina stuhlmanni	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Atopocochlis exarata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Limicolaria kambeul	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Limicolaria martenssii	GGCCGGGGAT	CCYTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Limicolariopsis sp.	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Limicolariopsis ruwenzori	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma varicosa	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma dimidiata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma sp. cf. vestita	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma marinae	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma churchilliana	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma granulata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma simplex	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma kilburni	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma montistempli	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma omissa	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma semidecussata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma ustulata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Cochlitoma zebra	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Metachatina kraussi	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
Rumina decollata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1210	1220	1230	1240	1250	1260	
Achatina fulica	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Achatina immaculata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Achatina reticulata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Achatina zanzibarica	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Euaethiopina loveridgei	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Achatina achatina	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GAGTTGTCCG	
Archachatina marginata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GAGTTGTCCG	
Achatina bisculpta	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Achatina damarensis	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Achatina stuhlmanni	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Atopocochlis exarata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Limicolaria kambeul	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Limicolaria martenssii	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Limicolariaopsis sp.	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Limicolariaopsis ruwenzori	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma varicosa	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma dimidiata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma sp. cf. vestita	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma marinae	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma churchilliana	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma granulata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma simplex	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma kilburni	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma montistempli	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma omissa	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma semidecussata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma ustulata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Cochlitoma zebra	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Metachatina kraussi	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG	
Rumina decollata	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCG	GGAGCCGTCA	GAGGTCCGGAC	GGGTTGTCCG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1270	1280	1290	1300	1310	1320
Achatina fulica	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Achatina immaculata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Achatina reticulata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Achatina zanzibarica	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Euaethiopina loveridgei	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Achatina achatina	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGGACCG
Archachatina marginata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGGACCG
Achatina bisculpta	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Achatina damarensis	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Achatina stuhlmanni	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Atopocochlis exarata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Limicolaria kambeul	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Limicolaria martenssii	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Limicolariaopsis sp.	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Limicolariaopsis ruwenzori	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma varicosa	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma dimidiata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma sp. cf. vestita	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma marinae	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma churchilliana	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma granulata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma simplex	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma kilburni	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma montistempli	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma omissa	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma semidecussata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma ustulata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Cochlitoma zebra	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Metachatina kraussi	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
Rumina decollata	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGGACCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1330	1340	1350	1360	1370	1380	
Achatina fulica	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina immaculata	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina reticulata	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina zanzibarica	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Euaethiopina loweridgei	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina achatina	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Archachatina marginata	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina bisculpta	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina damarensis	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina stuhlmanni	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Atopocochlis exarata	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Limicolaria kambeul	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Limicolaria martenssii	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Limicolariopsis sp.	CCGCGCGCTT	CGAGGCTACC	CGATCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Limicolariopsis ruwenzori	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma varicosa	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma dimidiata	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma sp. cf. vestita	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma marinae	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma churchilliana	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma granulata	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma simplex	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma kilburni	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma montistempli	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma omissa	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma semidecussata	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma ustulata	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma zebra	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Metachatina kraussi	CCGCGCGCTT	AGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Rumina decollata	CCGCACGCTT	CGAGGCTACC	T-GCTCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	m-mmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1390	1400	1410	1420	1430	1440	
Achatina fulica	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina immaculata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina reticulata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina zanzibarica	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Euaethiopina loweridgei	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina achatina	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Archachatina marginata	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina bisculpta	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina damarensis	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina stuhlmanni	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Atopocochlis exarata	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Limicolaria kambeul	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Limicolaria martenssii	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Limicolariopsis sp.	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Limicolariopsis ruwenzori	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma varicosa	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma dimidiata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma sp. cf. vestita	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma marinae	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma churchilliana	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma granulata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma simplex	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma kilburni	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma montistempli	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma omissa	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma semidecussata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma ustulata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma zebra	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Metachatina kraussi	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Rumina decollata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GC-AC---A-	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	m--mm----	

	1450	1460	1470	1480	1490	1500		
Achatina fulica	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Achatina immaculata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Achatina reticulata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Achatina zanzibarica	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Euaethiopina loveridgei	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Achatina achatina	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Archachatina marginata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Achatina bisculpta	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Achatina damarensis	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Achatina stuhlmanni	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Atopocochlis exarata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Limicolaria kambeul	CAGGGTTAGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Limicolaria martenssii	CAGGGTTAGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Limicolariaopsis sp.	CAGGGTTAGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Limicolariaopsis ruwenzori	CAGGGTTAGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma varicosa	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma dimidiata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma sp. cf. vestita	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma marinae	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma churchilliana	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma granulata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma simplex	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma kilburni	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma montistempli	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma omissa	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma semidecussata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma ustulata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Cochlitoma zebra	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Metachatina kraussi	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
Rumina decollata	CAGGGTTGGT	GGCGAGTCCG	TCGGCATTCC	ACCCGACCCG	TCTTGAACA	CGGACCAAGG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	1510	1520	1530	1540	1550	1560		
Achatina fulica	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Achatina immaculata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Achatina reticulata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Achatina zanzibarica	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Euaethiopina loveridgei	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Achatina achatina	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Archachatina marginata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Achatina bisculpta	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Achatina damarensis	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Achatina stuhlmanni	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Atopocochlis exarata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Limicolaria kambeul	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Limicolaria martenssii	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Limicolariaopsis sp.	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Limicolariaopsis ruwenzori	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma varicosa	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma dimidiata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma sp. cf. vestita	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma marinae	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma churchilliana	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma granulata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma simplex	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma kilburni	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma montistempli	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma omissa	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma semidecussata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma ustulata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Cochlitoma zebra	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Metachatina kraussi	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
Rumina decollata	AGTCTAACAT	GCGCGCGAGT	CATTGGGCGG	TACGAAACCC	AAAGGCGCAG	TGAAAGCGAG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	1570	1580	1590	1600	1610	1620		
Achatina fulica	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Achatina immaculata	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Achatina reticulata	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Achatina zanzibarica	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGTTCTTC	GCGGGAGCGG	TGGCGCACCA		
Euaethiopina loweridgei	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Achatina achatina	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Archachatina marginata	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Achatina bisculpta	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Achatina damarensis	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Achatina stuhlmanni	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Atopocochlis exarata	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Limicolaria kambeul	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Limicolaria martenssii	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Limicolariopsis sp.	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Limicolariopsis ruwenzori	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma varicosa	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGTTCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma dimidiata	GGTCGCTTCG	CGCGACCCAG	GTGGGATCCC	CCGCTCCTTC	GCGGGAGCGG	AGGCGCACCA		
Cochlitoma sp. cf. vestita	GGTCGCTTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma marinae	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma churchilliana	GGTCGCTTCG	CGCGACCCAG	GTGGGATCCC	CCACTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma granulata	GGTCGCTTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma simplex	GGTCGCTTCG	CGCGACCCAG	GTGGGATCCC	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma kilburni	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma montistempli	GGTCGCTTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma omisssa	GGTCGCTTCG	CGCGATCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma semidecussata	GGTCGCTTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	ACGGGAGCGG	TGGCGCACCA		
Cochlitoma ustulata	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Cochlitoma zebra	GGTCGCTTCG	CGCGATCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Metachatina kraussi	GGTCGCTTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA		
Rumina decollata	GGTCGTCTCN	CGCGACCCAG	GTGGGATCCT	CCACCCCTTC	G-GGGGCGG	TGGCGCACCA		
NUCLEOTIDES INCLUDED	mmmmmm--	-mmmmmmmm	mmmmmmmm	-----	--mmmmmm	mmmmmmmm		

	1630	1640	1650	1660	1670	1680		
Achatina fulica	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Achatina immaculata	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Achatina reticulata	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Achatina zanzibarica	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Euaethiopina loweridgei	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Achatina achatina	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Archachatina marginata	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Achatina bisculpta	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Achatina damarensis	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Achatina stuhlmanni	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Atopocochlis exarata	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Limicolaria kambeul	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Limicolaria martenssii	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Limicolariopsis sp.	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Limicolariopsis ruwenzori	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma varicosa	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma dimidiata	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma sp. cf. vestita	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma marinae	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma churchilliana	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma granulata	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma simplex	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma kilburni	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma montistempli	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma omisssa	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma semidecussata	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma ustulata	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Cochlitoma zebra	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Metachatina kraussi	CCGGCCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
Rumina decollata	CCGGCCCCGTC	CCGTCYCGCY	CGYCYGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	1690	1700	1710	1720	1730	1740	
Achatina fulica	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Achtina immaculata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Achatina reticulata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Achatina zanzibarica	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Euaethiopina loveridgei	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Achatina achatina	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Archachatina marginata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Achatina bisculpta	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Achatina damarensis	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Achatina stuhlmanni	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Atopocochlis exarata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Limicolaria kambeul	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Limicolaria martenssii	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Limicolariopsis sp.	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Limicolariopsis ruwenzori	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma varicosa	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma dimidiata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma sp. cf. vestita	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma marinae	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma churchilliana	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma granulata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma simplex	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma kilburni	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma montistempli	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma omissa	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma semidecussata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma ustulata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Cochlitoma zebra	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Metachatina kraussi	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
Rumina decollata	AAAGATGGTG	AACATATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1750	1760	1770	1780	1790	1800
Achatina fulica	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Achtina immaculata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Achatina reticulata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Achatina zanzibarica	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Euaethiopina loveridgei	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Achatina achatina	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Archachatina marginata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Achatina bisculpta	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Achatina damarensis	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Achatina stuhlmanni	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Atopocochlis exarata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Limicolaria kambeul	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Limicolaria martenssii	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Limicolariopsis sp.	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Limicolariopsis ruwenzori	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma varicosa	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma dimidiata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma sp. cf. vestita	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma marinae	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma churchilliana	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma granulata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma simplex	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma kilburni	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma montistempli	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma omissa	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma semidecussata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma ustulata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Cochlitoma zebra	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Metachatina kraussi	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
Rumina decollata	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1810	1820	1830	1840	1850	1860	
Achatina fulica	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Achatina immaculata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Achatina reticulata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Achatina zanzibarica	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Euaethiopina loveridgei	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Achatina achatina	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Archachatina marginata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Achatina bisculpta	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Achatina damarensis	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Achatina stuhlmanni	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Atopocochlis exarata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Limicolaria kambeul	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Limicolaria martenssii	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Limicolariopsis sp.	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Limicolariopsis ruwenzori	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma varicosa	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma dimidiata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma sp. cf. vestita	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma marinae	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma churchilliana	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma granulata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma simplex	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma kilburni	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma montistempli	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma omissa	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma semidecussata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma ustulata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Cochlitoma zebra	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Metachatina kraussi	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
Rumina decollata	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1870	1880	1890	1900	1910	1920	
Achatina fulica	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Achatina immaculata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Achatina reticulata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Achatina zanzibarica	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Euaethiopina loveridgei	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Achatina achatina	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Archachatina marginata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Achatina bisculpta	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Achatina damarensis	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Achatina stuhlmanni	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Atopocochlis exarata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Limicolaria kambeul	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Limicolaria martenssii	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Limicolariopsis sp.	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTTT	
Limicolariopsis ruwenzori	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma varicosa	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma dimidiata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma sp. cf. vestita	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma marinae	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma churchilliana	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma granulata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma simplex	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma kilburni	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma montistempli	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma omissa	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma semidecussata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma ustulata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Cochlitoma zebra	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Metachatina kraussi	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
Rumina decollata	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1930	1940	1950	1960	1970	1980	
Achatina fulica	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTNTCGAATG
Achtina immaculata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Achatina reticulata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Achatina zanzibarica	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Euaethiopina loveridgei	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Achatina achatina	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Archachatina marginata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Achatina bisculpta	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Achatina damarensis	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Achatina stuhlmanni	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Atopocochlis exarata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Limicolaria kambeul	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Limicolaria martenssii	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Limicolariopsis sp.	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AACCGGGCGC	GTTTGAATG
Limicolariopsis ruwenzori	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma varicosa	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma dimidiata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma sp. cf. vestita	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma marinae	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma churchilliana	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma granulata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma simplex	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma kilburni	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma montistempli	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma omissa	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma semidecussata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma ustulata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Cochlitoma zebra	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Metachatina kraussi	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGATTGG	AGCCGGGGCGC	GTTTCGAATG
Rumina decollata	CAAAC	TTTAA	ATGGGTAAGA	AGTCCGGCTC	GCTCGAYTGG	AGCCGGGGCGC	GTNTCGAATG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1990	2000	2010	2020	2030	2040
Achatina fulica	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Achtina immaculata	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Achatina reticulata	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Achatina zanzibarica	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Euaethiopina loveridgei	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Achatina achatina	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Archachatina marginata	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Achatina bisculpta	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Achatina damarensis	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Achatina stuhlmanni	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Atopocochlis exarata	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Limicolaria kambeul	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Limicolaria martenssii	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Limicolariopsis sp.	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Limicolariopsis ruwenzori	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma varicosa	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma dimidiata	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma sp. cf. vestita	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma marinae	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma churchilliana	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma granulata	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma simplex	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma kilburni	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma montistempli	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma omissa	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma semidecussata	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma ustulata	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Cochlitoma zebra	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Metachatina kraussi	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
Rumina decollata	CNTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	2050	2060	2070	2080	2090	2100	
Achatina fulica	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achtina immaculata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina reticulata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina zanzibarica	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Euaethiopina loveridgei	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina achatina	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Archachatina marginata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina bisculpta	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina damarensis	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina stuhlmanni	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Atopocochlis exarata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Limicolaria kambeul	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Limicolaria martenssii	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Limicolariopsis sp.	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Limicolariopsis ruwenzori	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma varicosa	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma dimidiata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma sp. cf. vestita	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma marinae	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma churchilliana	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma granulata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma simplex	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma kilburni	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma montistempli	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma omissa	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma semidecussata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma ustulata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma zebra	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Metachatina kraussi	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Rumina decollata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	2110	2120	2130	2140	2150	2160	
Achatina fulica	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achtina immaculata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina reticulata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina zanzibarica	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Euaethiopina loveridgei	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina achatina	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Archachatina marginata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina bisculpta	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina damarensis	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina stuhlmanni	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Atopocochlis exarata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Limicolaria kambeul	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Limicolaria martenssii	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Limicolariopsis sp.	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Limicolariopsis ruwenzori	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma varicosa	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma dimidiata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma sp. cf. vestita	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma marinae	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma churchilliana	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma granulata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma simplex	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma kilburni	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma montistempli	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma omissa	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma semidecussata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma ustulata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma zebra	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Metachatina kraussi	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Rumina decollata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	2170	2180	2190	2200	2210	2220	2230	2240
Achatina fulica	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Achatina immaculata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Achatina reticulata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Achatina zanzibarica	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Euaethiopina loveridgei	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Achatina achatina	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Archachatina marginata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Achatina bisculpta	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Achatina damarensis	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Achatina stuhlmanni	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Atopocochlis exarata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Limicolaria kambeul	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Limicolaria martenssii	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Limicolariopsis sp.	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Limicolariopsis ruwenzori	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma varicosa	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma dimidiata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma sp. cf. vestita	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma marinae	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma churchilliana	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma granulata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma simplex	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma kilburni	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma montistempli	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma omissa	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma semidecussata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma ustulata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Cochlitoma zebra	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Metachatina kraussi	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
Rumina decollata	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	2230	2240	2250	2260	2270	2280		
Achatina fulica	CTCGGCAATG	GGCCCTTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Achatina immaculata	CTCGGCAATG	GGCCCTTTCCA	TGGGG-CCGG	NAAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Achatina reticulata	CTCGGCAATG	GGCCCTTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Achatina zanzibarica	CTCGGCAATG	GGCCCTTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Euaethiopina loveridgei	CTCGGCAATG	GGCCCTTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Achatina achatina	CTCGGCAATA	GGCC-----	TTCCG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Archachatina marginata	CTCGGCAATA	GGCC---TTT	TTCCG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Achatina bisculpta	CTCGGCAATG	GGCTCTTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Achatina damarensis	CTCGGCAATG	GGCCCTTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Achatina stuhlmanni	CTCGGCAATG	GGCCCTTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Atopocochlis exarata	CTCGGCAATG	GGCCCTTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Limicolaria kambeul	CTCGGCAATG	GGCCCTTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Limicolaria martenssii	CTCGGCAATG	GGCCCTTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Limicolariopsis sp.	CTCGGCAATG	GGCCCTTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Limicolariopsis ruwenzori	CTCGGCAATG	GGCCCTTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma varicosa	CTCGGCAATG	GGCCCTTTTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma dimidiata	CTCGGCAATG	GGCCCTTTTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma sp. cf. vestita	CTCGGCAATG	GGCCCTTTTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma marinae	CTCGGCAATG	GGCCC-TTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma churchilliana	CTCGGCAATG	GGCCC-TTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma granulata	CTCGGCAATG	GGCCCTTTTCC	ACGGGGCCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma simplex	CTCGGCAATG	GGCCCTTTTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma kilburni	CTCGGCAATG	GGCCCTTTTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma montistempli	CTCGGCAATG	GGCCCTTTTCC	ACGGGGCCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma omissa	CTCGGCAATG	GGCCCTTTTCC	ACGGGGCCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma semidecussata	CTCGGCAATG	GGCCCTTTTCC	ACGGGGCCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma ustulata	CTCGGCAATG	GGCCC-TTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Cochlitoma zebra	CTCGGCAATG	GGCCCTTTTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Metachatina kraussi	CTCGGCAATG	GGCCC-TTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG		
Rumina decollata	CTCGGCAGTG	GCGATAGCCG	CG-----	-AATCGAGGC	CGAGACGAGT	AGGAGGGCCG		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	m-----	-----mmmm	-----mmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	2290	2300	2310	2320	2330	2340	
Achatina fulica	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Achatina immaculata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Achatina reticulata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Achatina zanzibarica	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Euaethiopina loveridgei	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Achatina achatina	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Archachatina marginata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Achatina bisculpta	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Achatina damarensis	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Achatina stuhlmanni	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Atopocochlis exarata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Limicolaria kambeul	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Limicolaria martenssii	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Limicolariopsis sp.	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Limicolariopsis ruwenzori	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma varicosa	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma dimidiata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma sp. cf. vestita	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma marinae	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma churchilliana	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma granulata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma simplex	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma kilburni	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma montistempli	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma omisssa	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma semidecussata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma ustulata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Cochlitoma zebra	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Metachatina kraussi	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
Rumina decollata	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	2350	2360	2370	2380	2390	2400
Achatina fulica	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Achatina immaculata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Achatina reticulata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Achatina zanzibarica	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Euaethiopina loveridgei	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Achatina achatina	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Archachatina marginata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Achatina bisculpta	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Achatina damarensis	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Achatina stuhlmanni	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Atopocochlis exarata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Limicolaria kambeul	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Limicolaria martenssii	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Limicolariopsis sp.	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Limicolariopsis ruwenzori	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma varicosa	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma dimidiata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma sp. cf. vestita	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma marinae	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma churchilliana	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma granulata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma simplex	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma kilburni	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma montistempli	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma omisssa	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma semidecussata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma ustulata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Cochlitoma zebra	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Metachatina kraussi	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
Rumina decollata	TTGGTGGTAG	TAGCAAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGA	GAAGGGTTCC
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	2410	2420	2430	2440	2450	2460		
Achatina fulica	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Achtina immaculata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Achatina reticulata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Achatina zanzibarica	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Euaethiopina loveridgei	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Achatina achatina	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Archachatina marginata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Achatina bisculpta	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Achatina damarensis	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Achatina stuhlmanni	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Atopocochlis exarata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Limicolaria kambeul	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Limicolaria martenssii	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Limicolariopsis sp.	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Limicolariopsis ruwenzori	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma varicosa	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma dimidiata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma sp. cf. vestita	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma marinae	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma churchilliana	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma granulata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma simplex	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma kilburni	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma montistempli	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma omissa	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma semidecussata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma ustulata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Cochlitoma zebra	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Metachatina kraussi	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
Rumina decollata	ATGTGAACAG	CAGTTGAACA	TGGGTCAGTC	GGTCCTAAGA	GATAGGAAAA	CTCCGTTCTG		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	2470	2480	2490	2500	2510	2520		
Achatina fulica	ACCCCGGGGC	ATTCTTTCTT	TCTTTCT---	-----	---AGAAGA	GTCTGCCCGC		
Achtina immaculata	ACCCCGGGGC	ATTCTTTCTT	TCTTTCT---	-----	---AGAAGA	GTCTGCCCGC		
Achatina reticulata	ACCCCGGGGC	ATTCTTTCTT	TCTTTCT---	-----	---AGAAGA	GTCTGCCCGC		
Achatina zanzibarica	ACCCCGGGGC	ATTCTTTCTT	TCTTTCT---	-----	---AGAAGA	GTCTGCCCGC		
Euaethiopina loveridgei	ACCCCGGGGC	ATTCTTTCTT	TCTTTCT---	-----	---AGAAGA	GTCTGCCCGC		
Achatina achatina	ACCCCGGGGC	ATTCTTTCTT	TCTTTCT---	-----	---AGCAA	GTCTGCCCGC		
Archachatina marginata	ACCCCGGGGC	ATTCTTTCTT	AGCAGT---	-----	---CAA	GTCTGCCCGC		
Achatina bisculpta	ACCCCGGGGC	ATTCTTTATT	A---TTCGTT	ATTTCGTT---	---AGA	GTCTGCCCGC		
Achatina damarensis	ACCCCGGGGC	ATTCTTTATT	ACTCTTCGTT	ATT-----	---AGA	GTCTGCCCGC		
Achatina stuhlmanni	ACCCCGGGGC	ATTCTTTATT	A---TTCGTT	ATT-----	---AGA	GTCTGCCCGC		
Atopocochlis exarata	ACCCCGGGGC	ATTCTTTATT	A---ATCGTT	TAA-----	---AGA	GTCTGCCCGC		
Limicolaria kambeul	ACCCCGGGGC	ATTCTTTATT	A---TTCGTT	-----	---AGA	GTCTGCCCGC		
Limicolaria martenssii	ACCCCGGGGC	ATTCTTTATT	A---TTCGTT	-----	---AGA	GTCTGCCCGC		
Limicolariopsis sp.	ACCCCGGGGC	ATTCTTTATT	ATA-TTCGTT	TT-----	---AGA	GTCTGCCCGC		
Limicolariopsis ruwenzori	ACCCCGGGGC	ATTCTTTATT	A---TTCGTT	-----	---AGA	GTCTGCCCGC		
Cochlitoma varicosa	ACCCCGGGGC	ATTCTTTCTT	T--CTTT---	-----	CGTGATTAAA	GTCTGCCCGC		
Cochlitoma dimidiata	ACCCCGGGGC	ATTCTTTCTT	T--CTTT---	-----	---A	GTCTGCCCGC		
Cochlitoma sp. cf. vestita	ACCCCGGGGC	ATTCTTTCTT	G-----	-----	---ATTAAA	GTCTGCCCGC		
Cochlitoma marinae	ACCCCGGGGC	ATTCTTTCTT	G-----	-----	---ATTAAA	GTCTGCCCGC		
Cochlitoma churchilliana	ACCCCGGGGC	ATTCTTTCTT	T--CTTTCT	T-----	---GATTAAA	GTCTGCCCGC		
Cochlitoma granulata	ACCCCGGGGC	ATTCTTTCTT	TTTCTT---	-----	G ATTAATTAAA	GTCTGCCCGC		
Cochlitoma simplex	ACCCCGGGGC	ATTCTTTCTT	T--CTTT---	-----	---A	GTCTGCCCGC		
Cochlitoma kilburni	ACCCCGGGGC	ATTCTTTCTT	T--CTTT---	---ATTA	ATTAAATAAA	GTCTGCCCGC		
Cochlitoma montistempli	ACCCCGGGGC	ATTCTTTCTT	TTTCTT---	-----	G ATTAATTAAA	GTCTGCCCGC		
Cochlitoma omissa	ACCCCGGGGC	ATTCTTTCTT	TTTCTT---	-----	G ATTAATTAAA	GTCTGCCCGC		
Cochlitoma semidecussata	ACCCCGGGGC	ATTCTTTCTT	TTTCTT---	-----	G ATTAATTAAA	GTCTGCCCGC		
Cochlitoma ustulata	ACCCCGGGGC	ATTCTTTCTT	G-----	-----	---ATTAAA	GTCTGCCCGC		
Cochlitoma zebra	ACCCCGGGGC	ATTCTTTCTT	G-----	-----	---ATTAAA	GTCTGCCCGC		
Metachatina kraussi	ACCCCGGGGC	ATTCTTTCTT	T--CTT---	-----	---AAA	GTCTGCCCGC		
Rumina decollata	ACCCCGGGGC	AATCTTTCTC	GAGCTA---	-----	---AGA	AGAAAAAGAA	GTCTGCCCGC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	-----	-----	-----	-----	mmmmmmmmmm		

	2530	2540	2550	2560	2570	2580
Achatina fulica	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Achatina immaculata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Achatina reticulata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Achatina zanzibarica	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Euaethiopina loveridgei	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Achatina achatina	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Archachatina marginata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Achatina bisculpta	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Achatina damarensis	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Achatina stuhlmanni	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Atopocochlis exarata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Limicolaria kambeul	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Limicolaria martenssii	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Limicolariopsis sp.	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Limicolariopsis ruwenzori	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma varicosa	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma dimidiata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma sp. cf. vestita	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma marinae	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma churchilliana	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma granulata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma simplex	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma kilburni	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma montistempli	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma omissa	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma semidecussata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma ustulata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Cochlitoma zebra	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Metachatina kraussi	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
Rumina decollata	AGCCTATCGA	AAGGGAATCG	GGTTAATATT	CCCGAACCTG	GACACGGAGA	TTGGTCCTCA	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	2590	2600	2610	2620	2630	2640
Achatina fulica	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Achatina immaculata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Achatina reticulata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Achatina zanzibarica	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Euaethiopina loveridgei	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Achatina achatina	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Archachatina marginata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Achatina bisculpta	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Achatina damarensis	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Achatina stuhlmanni	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Atopocochlis exarata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Limicolaria kambeul	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Limicolaria martenssii	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Limicolariopsis sp.	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Limicolariopsis ruwenzori	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma varicosa	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma dimidiata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma sp. cf. vestita	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma marinae	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma churchilliana	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma granulata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma simplex	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma kilburni	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma montistempli	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma omissa	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma semidecussata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma ustulata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Cochlitoma zebra	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Metachatina kraussi	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
Rumina decollata	GGGGCCACGT	GCGGCAACGC	AAACGAAGTG	GGGGACGTCG	GCGGGAGCCC	CGGGAAGAGT	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	2650	2660	2670	2680	2690	2700		
Achatina fulica	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Achatina immaculata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Achatina reticulata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Achatina zanzibarica	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Euaethiopina loveridgei	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Achatina achatina	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Archachatina marginata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Achatina bisculpta	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Achatina damarensis	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Achatina stuhlmanni	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Atopocochlis exarata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Limicolaria kambeul	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Limicolaria martenssii	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Limicolariopsis sp.	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Limicolariopsis ruwenzori	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma varicosa	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma dimidiata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma sp. cf. vestita	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma marinae	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma churchilliana	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma granulata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma simplex	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma kilburni	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma montistempli	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma omissa	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma semidecussata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma ustulata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Cochlitoma zebra	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Metachatina kraussi	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
Rumina decollata	TCTCTTTTCT	TTGTAAGGAG	CCACATCCCT	GGAATCGGCT	TGCCCCGAGA	TAGGGACCGC		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	2710	2720	2730	2740	2750	2760	
Achatina fulica	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Achatina immaculata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Achatina reticulata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Achatina zanzibarica	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Euaethiopina loveridgei	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Achatina achatina	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Archachatina marginata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Achatina bisculpta	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Achatina damarensis	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Achatina stuhlmanni	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Atopocochlis exarata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Limicolaria kambeul	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Limicolaria martenssii	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Limicolariopsis sp.	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Limicolariopsis ruwenzori	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma varicosa	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma dimidiata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma sp. cf. vestita	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma marinae	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma churchilliana	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma granulata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma simplex	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma kilburni	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma montistempli	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma omissa	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma semidecussata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma ustulata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Cochlitoma zebra	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Metachatina kraussi	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
Rumina decollata	GGCCCCGTAA	AGCACC GCGG	CTCTTGCGGT	GTCCGGTGCG	CTCCCGTCGG	CCCTTGAAAA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2770	2780	2790	2800	2810	2820	
Achatina fulica	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Achatina immaculata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Achatina reticulata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Achatina zanzibarica	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Euaethiopina loveridgei	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Achatina achatina	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Archachatina marginata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Achatina bisculpta	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Achatina damarensis	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Achatina stuhlmanni	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Atopocochlis exarata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Limicolaria kambeul	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Limicolaria martenssii	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Limicolariopsis sp.	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Limicolariopsis ruwenzori	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma varicosa	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma dimidiata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma sp. cf. vestita	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma marinae	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma churchilliana	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma granulata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma simplex	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma kilburni	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma montistempli	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma omisssa	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma semidecussata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma ustulata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Cochlitoma zebra	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Metachatina kraussi	CCCCACGNAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
Rumina decollata	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	2830	2840	2850	2860	2870	2880	
Achatina fulica	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Achatina immaculata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Achatina reticulata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Achatina zanzibarica	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Euaethiopina loveridgei	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Achatina achatina	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Archachatina marginata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Achatina bisculpta	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Achatina damarensis	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Achatina stuhlmanni	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Atopocochlis exarata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Limicolaria kambeul	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Limicolaria martenssii	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Limicolariopsis sp.	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Limicolariopsis ruwenzori	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma varicosa	AGGTGCACAG	TCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma dimidiata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma sp. cf. vestita	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma marinae	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma churchilliana	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma granulata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma simplex	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma kilburni	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma montistempli	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma omisssa	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma semidecussata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma ustulata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Cochlitoma zebra	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Metachatina kraussi	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
Rumina decollata	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	3010	3020	3030	3040	3050	3060		
Achatina fulica	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Achatina immaculata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Achatina reticulata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Achatina zanzibarica	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Euaethiopina loveridgei	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Achatina achatina	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Archachatina marginata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Achatina bisculpta	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Achatina damarensis	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Achatina stuhlmanni	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Atopocochlis exarata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Limicolaria kambeul	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Limicolaria martenssii	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Limicolariopsis sp.	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Limicolariopsis ruwenzori	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma varicosa	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma dimidiata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma sp. cf. vestita	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma marinae	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma churchilliana	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma granulata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma simplex	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma kilburni	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma montistempli	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma omissa	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma semidecussata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma ustulata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Cochlitoma zebra	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Metachatina kraussi	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG		
Rumina decollata	TCGGAACGCG	GCTGCAACCT	TCCCGTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCT-CCCCG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	3070	3080	3090	3100	3110	3120	
Achatina fulica	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina immaculata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina reticulata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina zanzibarica	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Euaethiopina loveridgei	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina achatina	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Archachatina marginata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina bisculpta	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina damarensis	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina stuhlmanni	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Atopocochlis exarata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Limicolaria kambeul	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Limicolaria martenssii	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Limicolariopsis sp.	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Limicolariopsis ruwenzori	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma varicosa	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma dimidiata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma sp. cf. vestita	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma marinae	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma churchilliana	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma granulata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma simplex	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma kilburni	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma montistempli	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma omissa	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma semidecussata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma ustulata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma zebra	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Metachatina kraussi	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Rumina decollata	CGTCGTTCGC	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	3130	3140	3150	3160	3170	3180		
Achatina fulica	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Achatina immaculata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Achatina reticulata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Achatina zanzibarica	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Euaethiopina loveridgei	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Achatina achatina	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Archachatina marginata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Achatina bisculpta	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Achatina damarensis	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Achatina stuhlmanni	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Atopocochlis exarata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Limicolaria kambeul	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Limicolaria martenssii	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Limicolariaopsis sp.	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Limicolariaopsis ruwenzori	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma varicosa	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma dimidiata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma sp. cf. vestita	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma marinae	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma churchilliana	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma granulata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma simplex	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma kilburni	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma montistempli	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma omissa	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma semidecussata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma ustulata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Cochlitoma zebra	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Metachatina kraussi	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
Rumina decollata	CCGACTGTCT	AATTAATAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	3190	3200	3210	3220	3230	3240		
Achatina fulica	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Achatina immaculata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Achatina reticulata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Achatina zanzibarica	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Euaethiopina loveridgei	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Achatina achatina	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Archachatina marginata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Achatina bisculpta	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Achatina damarensis	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Achatina stuhlmanni	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Atopocochlis exarata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Limicolaria kambeul	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Limicolaria martenssii	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Limicolariaopsis sp.	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Limicolariaopsis ruwenzori	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma varicosa	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma dimidiata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma sp. cf. vestita	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma marinae	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma churchilliana	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma granulata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma simplex	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma kilburni	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma montistempli	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma omissa	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma semidecussata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma ustulata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Cochlitoma zebra	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Metachatina kraussi	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
Rumina decollata	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAAACGG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	3250	3260	3270	3280	3290	3300		
Achatina fulica	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Achatina immaculata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Achatina reticulata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Achatina zanzibarica	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Euaethiopina loveridgei	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Achatina achatina	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Archachatina marginata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Achatina bisculpta	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Achatina damarensis	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Achatina stuhlmanni	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Atopocochlis exarata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Limicolaria kambeul	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Limicolaria martenssii	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Limicolariopsis sp.	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Limicolariopsis ruwenzori	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma varicosa	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma dimidiata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma sp. cf. vestita	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma marinae	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma churchilliana	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma granulata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma simplex	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma kilburni	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma montistempli	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma omissa	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma semidecussata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma ustulata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Cochlitoma zebra	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Metachatina kraussi	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
Rumina decollata	CGGGAGTAAC	TATGACTCTC	TTAAGGTAGC	CAAATGCCTC	GTCATCTAAT	TAGTGACGCG		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	3310	3320	3330	3340	3350	3360	
Achatina fulica	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Achatina immaculata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Achatina reticulata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Achatina zanzibarica	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Euaethiopina loveridgei	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Achatina achatina	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Archachatina marginata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Achatina bisculpta	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Achatina damarensis	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Achatina stuhlmanni	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Atopocochlis exarata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Limicolaria kambeul	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Limicolaria martenssii	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Limicolariopsis sp.	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Limicolariopsis ruwenzori	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma varicosa	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma dimidiata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma sp. cf. vestita	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma marinae	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma churchilliana	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma granulata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma simplex	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma kilburni	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma montistempli	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma omissa	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma semidecussata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma ustulata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Cochlitoma zebra	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Metachatina kraussi	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
Rumina decollata	CATGAATGGA	TTAACGAGAT	TCCCCTGTG	CCTATCTACT	ATCTAGCGAA	ACCACAGCCA	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	3370	3380	3390	3400	3410	3420		
Achatina fulica	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Achtina immaculata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Achatina reticulata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Achatina zanzibarica	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Euaethiopina loveridgei	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Achatina achatina	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Archachatina marginata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Achatina bisculpta	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Achatina damarensis	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Achatina stuhlmanni	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Atopocochlis exarata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Limicolaria kambeul	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Limicolaria martenssii	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Limicolariopsis sp.	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Limicolariopsis ruwenzori	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma varicosa	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma dimidiata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma sp. cf. vestita	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma marinae	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma churchilliana	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma granulata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma simplex	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma kilburni	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma montistempli	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma omissa	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma semidecussata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma ustulata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Cochlitoma zebra	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Metachatina kraussi	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
Rumina decollata	AGGGAACGGG	CTTGGTAGAA	TCAGCGGGGA	AAGAAGACCC	TGTTGAGCTT	GACTCTAGTC		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	3430	3440	3450	3460	3470	3480		
Achatina fulica	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Achtina immaculata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Achatina reticulata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Achatina zanzibarica	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Euaethiopina loveridgei	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Achatina achatina	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Archachatina marginata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Achatina bisculpta	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Achatina damarensis	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Achatina stuhlmanni	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Atopocochlis exarata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Limicolaria kambeul	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Limicolaria martenssii	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Limicolariopsis sp.	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Limicolariopsis ruwenzori	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma varicosa	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma dimidiata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma sp. cf. vestita	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma marinae	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma churchilliana	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma granulata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma simplex	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma kilburni	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma montistempli	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma omissa	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma semidecussata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma ustulata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Cochlitoma zebra	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Metachatina kraussi	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
Rumina decollata	CGACTTTGTG	AAGACACATG	AAGGGTGTAG	CATAGGTGGG	AGCGCAAGCG	CAATTGAAAT		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	3490	3500	3510	3520	3530	3540	
Achatina fulica	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Achatina immaculata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Achatina reticulata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Achatina zanzibarica	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Euaethiopina loveridgei	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Achatina achatina	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Archachatina marginata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Achatina bisculpta	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Achatina damarensis	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Achatina stuhlmanni	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Atopocochlis exarata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Limicolaria kambeul	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Limicolaria martenssii	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Limicolariopsis sp.	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Limicolariopsis ruwenzori	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma varicosa	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma dimidiata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma sp. cf. vestita	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma marinae	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma churchilliana	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma granulata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma simplex	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma kilburni	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma montistempli	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma omisa	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma semidecussata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma ustulata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Cochlitoma zebra	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Metachatina kraussi	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
Rumina decollata	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	3550	3560	3570	3580	3590	3600	
Achatina fulica	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Achatina immaculata	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Achatina reticulata	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Achatina zanzibarica	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Euaethiopina loveridgei	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Achatina achatina	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Archachatina marginata	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Achatina bisculpta	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Achatina damarensis	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Achatina stuhlmanni	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Atopocochlis exarata	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Limicolaria kambeul	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Limicolaria martenssii	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Limicolariopsis sp.	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Limicolariopsis ruwenzori	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma varicosa	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma dimidiata	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma sp. cf. vestita	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma marinae	TCGCTTCTGG	AATTAAGCGG	CAACAGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma churchilliana	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma granulata	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma simplex	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-TCCGCGATC	CGCTCTGAAG	
Cochlitoma kilburni	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma montistempli	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma omisa	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma semidecussata	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma ustulata	TCGCTTCTGG	AGTTAAGCGG	CAATCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Cochlitoma zebra	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Metachatina kraussi	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
Rumina decollata	TCGCTTCTGG	AATTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	-mmmmmmmm	mmmmmmmm	

	3610	3620	3630	3640	3650	3660	
Achatina fulica	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Achatina immaculata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Achatina reticulata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Achatina zanzibarica	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Euaethiopina loveridgei	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Achatina achatina	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Archachatina marginata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Achatina bisculpta	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Achatina damarensis	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Achatina stuhlmanni	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Atopocochlis exarata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Limicolaria kambeul	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Limicolaria martenssii	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Limicolariaopsis sp.	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Limicolariaopsis ruwenzori	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma varicosa	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma dimidiata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma sp. cf. vestita	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma marinae	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma churchilliana	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma granulata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma simplex	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma kilburni	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma montistempli	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma omissa	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma semidecussata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma ustulata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Cochlitoma zebra	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Metachatina kraussi	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
Rumina decollata	ACAGTGTCTAG	GCGGGGAGTT	TGACTGGGGC	GGTACATCTG	TCAAAAGGTA	ACGCAGGTGT	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3670	3680	3690	3700	3710	3720	
Achatina fulica	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Achatina immaculata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Achatina reticulata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Achatina zanzibarica	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Euaethiopina loveridgei	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Achatina achatina	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Archachatina marginata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Achatina bisculpta	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Achatina damarensis	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Achatina stuhlmanni	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Atopocochlis exarata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Limicolaria kambeul	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Limicolaria martenssii	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Limicolariaopsis sp.	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Limicolariaopsis ruwenzori	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma varicosa	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma dimidiata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma sp. cf. vestita	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma marinae	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma churchilliana	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma granulata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma simplex	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma kilburni	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma montistempli	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma omissa	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma semidecussata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma ustulata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Cochlitoma zebra	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Metachatina kraussi	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
Rumina decollata	CCTAAGGCGA	GCTCAGCGAG	GACGGAAACC	TCGCGTAGAG	CAAAAGGGCA	AAAGCTCGCT	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3730	3740	3750	3760	3770	3780	
Achatina fulica	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Achtina immaculata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Achatina reticulata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Achatina zanzibarica	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Euaethiopina loveridgei	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Achatina achatina	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Archachatina marginata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Achatina bisculpta	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Achatina damarensis	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Achatina stuhlmanni	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Atopocochlis exarata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Limicolaria kambeul	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Limicolaria martenssii	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Limicolariopsis sp.	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Limicolariopsis ruwenzori	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma varicosa	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma dimidiata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma sp. cf. vestita	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma marinae	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma churchilliana	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma granulata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma simplex	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma kilburni	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma montistempli	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma omisa	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma semidecussata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma ustulata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Cochlitoma zebra	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Metachatina kraussi	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
Rumina decollata	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	3790	3800	3810	3820	3830	3840	
Achatina fulica	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Achtina immaculata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Achatina reticulata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Achatina zanzibarica	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Euaethiopina loveridgei	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Achatina achatina	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Archachatina marginata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Achatina bisculpta	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Achatina damarensis	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Achatina stuhlmanni	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Atopocochlis exarata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Limicolaria kambeul	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Limicolaria martenssii	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Limicolariopsis sp.	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Limicolariopsis ruwenzori	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma varicosa	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma dimidiata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma sp. cf. vestita	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma marinae	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma churchilliana	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma granulata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma simplex	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma kilburni	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma montistempli	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma omisa	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma semidecussata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma ustulata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Cochlitoma zebra	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Metachatina kraussi	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
Rumina decollata	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	3970	3980	3990	4000	4010	4020		
Achatina fulica	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Achtina immaculata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Achatina reticulata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Achatina zanzibarica	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Euaethiopina loveridgei	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Achatina achatina	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Archachatina marginata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Achatina bisculpta	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Achatina damarensis	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Achatina stuhlmanni	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Atopocochlis exarata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Limicolaria kambeul	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Limicolaria martenssii	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Limicolariopsis sp.	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Limicolariopsis ruwenzori	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma varicosa	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma dimidiata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma sp. cf. vestita	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma marinae	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma churchilliana	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma granulata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma simplex	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma kilburni	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma montistempli	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma omisa	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma semidecussata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma ustulata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Cochlitoma zebra	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Metachatina kraussi	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
Rumina decollata	TTTAGACCGT	CGTGAGACAG	GTTAGTTTTA	CCCTACTGAT	GACAGGTCGT	TGCTACGGTA		
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm		

	4030	4040	4050	4060	4070	4080	
Achatina fulica	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Achtina immaculata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Achatina reticulata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Achatina zanzibarica	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Euaethiopina loveridgei	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Achatina achatina	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Archachatina marginata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Achatina bisculpta	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Achatina damarensis	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Achatina stuhlmanni	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Atopocochlis exarata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Limicolaria kambeul	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Limicolaria martenssii	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Limicolariopsis sp.	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Limicolariopsis ruwenzori	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma varicosa	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma dimidiata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma sp. cf. vestita	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma marinae	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma churchilliana	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma granulata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma simplex	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma kilburni	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma montistempli	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma omisa	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma semidecussata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma ustulata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Cochlitoma zebra	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Metachatina kraussi	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
Rumina decollata	ATCCTGCTCA	GTACGAGAGG	AACCGCAGGT	TCAGACATTT	GGTTCACGTG	CTTGCGTGAT	
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	4090	4100	4110		
Achatina fulica	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Achatina immaculata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Achatina reticulata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Achatina zanzibarica	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Euaethiopina loveridgei	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Achatina achatina	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Archachatina marginata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Achatina bisculpta	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Achatina damarensis	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Achatina stuhlmanni	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Atopocochlis exarata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Limicolaria kambeul	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Limicolaria martenssii	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Limicolariopsis sp.	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Limicolariopsis ruwenzori	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma varicosa	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma dimidiata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma sp. cf. vestita	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma marinae	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma churchilliana	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma granulata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma simplex	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma kilburni	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma montistempli	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma omissa	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma semidecussata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma ustulata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Cochlitoma zebra	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Metachatina kraussi	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
Rumina decollata	AAGCCAATGG	TGCGAGGCTA	CCATCTGAGG	GATTATGG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

Appendix 4.2: Alignment of the actin gene for the Achatinidae and the subulinid outgroup *Rumina decollata*. Note that sequencing was unsuccessful for *Limicolariopsis ruwenzoriensis*.

	10	20	30	40	50	60	
<i>Achatina fulica</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Achatina immaculata</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Achatina reticulata</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Achatina zanzibarica</i>	TCCAAGAGGG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Euaethiopina loveridgei</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Achatina achatina</i>	TCCAAGAGAG	GTATYCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Archachatina marginata</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Achatina bisculpta</i>	TCYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Achatina damarensis</i>	TYYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Achatina stuhlmanni</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Atopocochlis exarata</i>	TCYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Limicola kambeul</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Limicolaria martenssii</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Limicolariopsis sp.</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma varicosa</i>	TCCAAGAGAG	GYATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma dimidiata</i>	TCCAAGAGAG	GYATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma sp. cf. vestita</i>	TCCAAGAGAG	GYATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma marinae</i>	TCTAAGAGGG	GTATTCTCAC	ACTCAAGTAC	CCCATTGAAC	ATGGTATTGT	CACAAACTGG	
<i>Cochlitoma churchilliana</i>	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma granulata</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma simplex</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cpchlitoloma kilburni</i>	TCCAAGRGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma montistempli</i>	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma omisa</i>	TCCAAGAGAG	GYATCCTCAC	TCTCAAGTAT	CCYATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma semidecussata</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma ustulata</i>	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Cochlitoma zebra</i>	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Metachatina kraussi</i>	TYCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	
<i>Rumina decollata</i>	TCYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG	

	70	80	90	100	110	120	
<i>Achatina fulica</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTSAGAGT	TGCACCAGAA	
<i>Achatina immaculata</i>	GATGATATGG	AGAAGATCTG	GCATCACACY	TTCTACAATG	AGCTGAGAGT	TGCWCCAGAA	
<i>Achatina reticulata</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCRGAA	
<i>Achatina zanzibarica</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Euaethiopina loveridgei</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Achatina achatina</i>	GATGATATGG	AGAAGATCTG	GCATCACACY	TTCTACAATG	AGTTGAGAGT	TGCWCCAGAA	
<i>Archachatina marginata</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGYTGAGAGT	TGCACCAGAA	
<i>Achatina bisculpta</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Achatina damarensis</i>	GATGATATGG	AGAAGATCTG	GCATCACACY	TTCTACAATG	AGCTGAGAGT	TGCWCCAGAA	
<i>Achatina stuhlmanni</i>	GATGATATGG	AGAAAATATG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Atopocochlis exarata</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTAYAATG	ARCTGAGAGT	TGCACCAGAA	
<i>Limicola kambeul</i>	GATGAYATGG	AGAAGATCTG	GCATCACACC	TTCTAYAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Limicolaria martenssii</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTATAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Limicolariopsis sp.</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma varicosa</i>	GATGATATGG	AGAAGATCTG	GCAYCACACY	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma dimidiata</i>	GATGATATGG	AGAAGATCTG	GCAYCACACY	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma sp. cf. vestita</i>	GATGATATGG	AGAAGATCTG	GCAYCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma marinae</i>	GACGATATGG	AGAAGATCTG	GCATCACACT	TTCTACARCG	AGCTTAGAGT	TGCCCCCTGAA	
<i>Cochlitoma churchilliana</i>	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma granulata</i>	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma simplex</i>	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cpchlitoloma kilburni</i>	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma montistempli</i>	GATGATATGG	AGAAGATCTG	GCACCATACT	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma omisa</i>	GATGATATGG	AGAAGATCTG	GCACCATACT	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma semidecussata</i>	GATGATATGG	AGAAGATCTG	GCACCATACT	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma ustulata</i>	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Cochlitoma zebra</i>	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA	
<i>Metachatina kraussi</i>	GATGATATGG	AGAARATCTG	GCAYCACACC	TTCTACAATG	AGCTGAGAGT	TGCWCCAGAA	
<i>Rumina decollata</i>	GAYGATATGG	AGAAGATCTG	GCATCACACY	TTCTACAATG	AGCTGAGAGT	TGCWCCAGAA	

	130	140	150	160	170	180		
Achatina fulica	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Achatina immaculata	GAGCATCCAG	TTCTGCTTAC	AGAAGCTCCA	CTCAACCCCA	AGGCCAAYAG	AGAGAAGATG		
Achatina reticulata	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Achatina zanzibarica	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Euaethiopina loveridgei	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Achatina achatina	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Archachatina marginata	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Achatina bisculpta	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Achatina damarensis	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Achatina stuhlmanni	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Atopocochlis exarata	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Limicolaria kambeul	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Limicolaria martenssii	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Limicolariaopsis sp.	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma varicosa	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma dimidiata	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma sp. cf. vestita	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma marinae	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma churchilliana	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma granulata	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma simplex	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cpchlitoloma kilburni	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma montistempli	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma omissa	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma semidecussata	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma ustulata	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Cochlitoma zebra	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Metachatina kraussi	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		
Rumina decollata	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG		

	190	200	210	220	230	240		
Achatina fulica	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTCGCCAT	YCAAGCYGTG		
Achatina immaculata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Achatina reticulata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Achatina zanzibarica	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Euaethiopina loveridgei	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Achatina achatina	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Archachatina marginata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Achatina bisculpta	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Achatina damarensis	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Achatina stuhlmanni	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Atopocochlis exarata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Limicolaria kambeul	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Limicolaria martenssii	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Limicolariaopsis sp.	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma varicosa	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma dimidiata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma sp. cf. vestita	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma marinae	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma churchilliana	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma granulata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma simplex	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cpchlitoloma kilburni	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma montistempli	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma omissa	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma semidecussata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma ustulata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Cochlitoma zebra	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Metachatina kraussi	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		
Rumina decollata	ACCCAGATCA	TGTTTGA AAC	YTTCAAYTCT	CCAGCCATGT	ATGTYGCCAT	TCAAGCTGTG		

	250	260	270	280	290	300		
Achatina fulica	CTTTCTTTT	ATGCWTCAGG	TCGTACAAC	GGTATTGTC	TGGATTCTGG	TGATGGTGTA		
Achatina immaculata	CTTTCTTTT	AYGCATCAGG	TCGTACAAC	GGTATTGTGY	TGGATTCTGG	TGATGGTGTA		
Achatina reticulata	CTTTCCYYT	AYGCWTCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	TGATGGTGTA		
Achatina zanzibarica	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTAC	TGGATTCTGG	TGATGGTGTA		
Euaethiopina loveridgei	CTTTCTTTT	ACGCATCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	TGATGGTGTA		
Achatina achatina	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	WGATGGTGTM		
Archachatina marginata	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	TGATGGTGTM		
Achatina bisculpta	CTTTCTTTT	AYGCWTCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	TGATGGTGTR		
Achatina damarensis	CTTTCTTTT	ATGCWTCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	TGATGGTGTR		
Achatina stuhlmanni	CTTTCTTTT	ATGCATCGGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	TGATGGTGTA		
Atopocochlis exarata	CTTTCTTTT	AYGCWTCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	TGATGGTGTA		
Limicolaria kambeul	CTTTCCYYT	ATGCWTCAGG	TCGTACAAC	GGTATYGTGC	TGGATTCTGG	TGATGGTGTA		
Limicolaria martenssii	YTTTCYTTT	AYGCWTCAGG	TCGTACAAC	GGTATYGTGC	TGGATTCTGG	TGATGGTGTR		
Limicolariopsis sp.	CTTTCTTTT	AYGCWTCAGG	TCGTACAAC	GGTATYGTGC	TGGATTCTGG	TGATGGTGTA		
Cochlitoma varicosa	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	WGATGGTGTA		
Cochlitoma dimidiata	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	WGATGGTGTM		
Cochlitoma sp. cf. vestita	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTGC	TGGAYTCTGG	WGATGGTGTM		
Cochlitoma marinae	CTGTCACTGT	ATGCCTTCAGG	TCGAACAACA	GGCATTGTCC	TTGACTCAGG	AGATGGTGTC		
Cochlitoma churchilliana	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	TGATGGTGTA		
Cochlitoma granulata	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	TGATGGTGTA		
Cochlitoma simplex	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	TGATGGTGTA		
Cpchlitoloma kilburni	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	TGATGGTGTA		
Cochlitoma montistempli	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	TGATGGTGTA		
Cochlitoma omisa	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	TGATGGTGTA		
Cochlitoma semidecussata	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	TGATGGTGTA		
Cochlitoma ustulata	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTTC	TGGATTCTGG	TGATGGTGTA		
Cochlitoma zebra	CTTTCTTTT	ATGCATCAGG	TCGTACAAC	GGTATTGTGC	TGGATTCTGG	TGATGGTGTA		
Metachatina kraussi	CTTTCTTTT	ATGCWTCAGG	TCGYACAAC	GGTATTGTTC	TGGATTCTGG	WGATGGTGTV		
Rumina decollata	CTKTCYTTG	ATGCCTCTGG	TCGTACAAC	GGTATTGTGC	TAGATTCTGG	WGATGGTGTT		

	310	320	330	340	350	360		
Achatina fulica	ACCCACACTG	TCCCAATTTA	TGAAGGTTAT	GCTCTACCTC	ATGCCATCAT	GAGACTGGAC		
Achatina immaculata	ACCCACACTG	TCCCAATYTA	TGAAGGTTAT	GCTCTACCTC	ATGCCATCAT	GAGACTGGAC		
Achatina reticulata	ACCCACACTG	TCCCAATYTA	TGAAGGTTAT	GCTCTTCCCC	AYGCCATCAT	GAGACTGGAC		
Achatina zanzibarica	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Euaethiopina loveridgei	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Achatina achatina	ACYCAYACTG	TCCCAATMTA	TGARGGTTAT	GCTCTTCCYC	AYGCCATCAT	GAGACTGGAY		
Archachatina marginata	ACYCAYACTG	TCCCAATCTA	YGARGGTTAT	GCTCTTCCYC	ATGCCATYMT	GAGACTGGAC		
Achatina bisculpta	ACCCACACTG	TYCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTRGAY		
Achatina damarensis	ACCCACACTG	TYCCAATCTA	TGAAGGTTAT	GCTCTTCCYC	ATGCCATCAT	GAGACTGGAY		
Achatina stuhlmanni	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Atopocochlis exarata	ACYCAYACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCYC	ATGCCATCAT	GAGACTGGAC		
Limicolaria kambeul	ACYCAYACTG	TYCCAATCTA	TGAAGGTTAT	GCYCTTCCYC	ATGCCATCAT	GAGACTGGAC		
Limicolaria martenssii	ACCCACACTG	TYCCAATCTA	YGAAGGTTAT	GCYCTTCCYC	ATGCCATCAT	GAGACTKGAC		
Limicolariopsis sp.	ACCCACACTG	TYCCAATCTA	YGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma varicosa	ACCCACACTG	TCCCAATCTA	TGAAGGWTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma dimidiata	ACYCAYACTG	TCCCAATCTA	TGAAGGWTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAY		
Cochlitoma sp. cf. vestita	ACCCACACTG	TCCCAATCTA	TGAAGGWTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAY		
Cochlitoma marinae	ACTCACACTG	TGCCAATCTA	TGAAGGTTAT	GCCCTCCCTC	ATGCCATCAT	GAGACTGGAT		
Cochlitoma churchilliana	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma granulata	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma simplex	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cpchlitoloma kilburni	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma montistempli	ACCCATACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma omisa	ACCCAYACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma semidecussata	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma ustulata	ACTCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Cochlitoma zebra	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC		
Metachatina kraussi	ACCCACACTG	TCCCMATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCMW	GAGACTGGAC		
Rumina decollata	ACCCAYACTG	TCCCAATCTA	TGAAGGTTAT	GCCCTTCCCT	ATGCCATCAT	GAGAYTGGAC		

	370	380	390	400	410	420		
Achatina fulica	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Achatina immaculata	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Achatina reticulata	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Achatina zanzibarica	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Euaethiopina loveridgei	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAATATCC	TTACAGAGAG	AGGCTACAGC		
Achatina achatina	TTGGCTGGTC	GTGACCTCAC	AGATTATCTC	ATGAARATCC	TCACAGAGAG	AGGTTACAGC		
Archachatina marginata	TTGGCTGGWC	GTGACCTCAC	AGATTAYCTC	ATGAARATYC	TCACAGAGAG	AGGYTACAGY		
Achatina bisculpta	TTGGCTGGTC	GTGACCTYAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Achatina damarensis	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Achatina stuhlmanni	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Atopocochlis exarata	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Limicolaria kambeul	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGY		
Limicolaria martenssii	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCY	TCACAGAGAG	AGGCYACAGY		
Limicolariopsis sp.	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma varicosa	TTGGCTGGTC	GTGACCTCAC	AGATTATCTS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma dimidiata	TTGGCTGGTC	GTGACCTCAC	AGATTATCTS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma sp. cf. vestita	TTGGCTGGTC	GTGACCTCAC	AGATTATCTS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma marinae	CTGGCTGGTC	GTGACCTTAC	TGATTATTG	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma churchilliana	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma granulata	TTGGCTGGTC	GTGACCTCAC	AGATTAYCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma simplex	TTGGCTGGTC	GTGACCTCAC	AGATTATCTS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cpchlitoloma kilburni	TTGGCTGGTC	GTGACCTCAC	AGATTATCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma montistempli	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma omissa	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma semidecussata	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma ustulata	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Cochlitoma zebra	TTGGCTGGTC	GTGACCTCAC	AGATTATCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Metachatina kraussi	TTGGCTGGTC	GTGACCTYAC	AGATTAYCWS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		
Rumina decollata	TTGGCTGGTC	GTGACCTYAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC		

	430	440	450	460	470	480		
Achatina fulica	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Achatina immaculata	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Achatina reticulata	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAARCT	TTGCTATGTG		
Achatina zanzibarica	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Euaethiopina loveridgei	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Achatina achatina	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGAYATCA	ARGAGAAGCT	GTGCTATGTG		
Archachatina marginata	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AGGAGAAGCT	GTGCTATGTG		
Achatina bisculpta	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG		
Achatina damarensis	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG		
Achatina stuhlmanni	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG		
Atopocochlis exarata	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG		
Limicolaria kambeul	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG		
Limicolaria martenssii	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG		
Limicolariopsis sp.	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG		
Cochlitoma varicosa	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma dimidiata	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma sp. cf. vestita	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma marinae	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma churchilliana	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma granulata	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma simplex	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cpchlitoloma kilburni	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma montistempli	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma omissa	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma semidecussata	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma ustulata	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Cochlitoma zebra	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG		
Metachatina kraussi	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGAYATCA	AAGAGAAGCT	TTGCTATGTG		
Rumina decollata	TTCACCACCA	CTGCTGAGAG	AGAAATTGTT	CGAGACATCA	AGGAGAAGCT	GTSTATGTG		

	490	500	510	520	530	540	
Achatina fulica	GCCCTTGACT	TTGAGCAAGA	RATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Achatina immaculata	GCCCTTGACT	TTGAGCARGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Achatina reticulata	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCYACAT	CATCTTCATT	AGAGAAGAGT	
Achatina zanzibarica	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTGCTACAT	CATCTTCATT	AGAGAAGAGT	
Euaethiopina loveridgei	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTGCTACAT	CATCTTCATT	AGAGAAGAGT	
Achatina achatina	GCYCTTGACT	TTGAGCAAGA	GATGTCRACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGY	
Archachatina marginata	GCYCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
Achatina bisculpta	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
Achatina damarensis	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
Achatina stuhlmanni	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTGCCACAT	CATCTTYATT	AGAGAAGAGT	
Atopocochlis exarata	GCYCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
Limicolaria kambeul	GCCCTTGAYT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGARAAGAGT	
Limicolaria martenssii	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
Limicolariopsis sp.	GCCCTTGACT	TTGAGCAAGA	GATGGSWACA	GCTKCCACGT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma varicosa	GCCCTTGACT	TTGARCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma dimidiata	GCCCTTGACT	TTGARCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma sp. cf. vestita	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma marinae	GCACCTTGACT	TTGAGCAAGA	AATGGGTACA	GCAGCAACAT	CATCATCTTT	GGAGAAGAGC	
Cochlitoma churchilliana	GCCCTTGACT	TTGAACAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma granulata	GCCCTTGACT	TTGAGCARGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma simplex	GCCCTTGACT	TTGAACAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cpchlitoloma kilburni	GCCCTTGACT	TTGAACAAGA	GATGGCAACA	GCTTCAACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma montistempli	GCCCTTGACT	TTGAGCAGGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma omisa	GCCCTTGACT	TTGAGCARGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma semidecussata	GCCCTTGACT	TTGAGCAAGA	GATGGCCACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma ustulata	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Cochlitoma zebra	GCCCTTGACT	TTGAACAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Metachatina kraussi	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
Rumina decollata	GCTCTTGACT	TTGAACAAGA	AATGGCAACA	GCTGCCACTT	CATCTTCWCT	GGAGAAAAGT	

	550	560	570	580	590	600	
Achatina fulica	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	ATGTCCAGAA	
Achatina immaculata	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAATG	AGCGTTTCAG	ATGTCCAGAA	
Achatina reticulata	TATGAAYTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	RTGYCCAGAA	
Achatina zanzibarica	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGCAACG	AGCGTTTCAG	GTGCCCAGAA	
Euaethiopina loveridgei	TATGAACCTGC	CTGATGGACA	GGTCATCACC	ATTGGTAATG	AGCGTTTCAG	GTGCCCAGAA	
Achatina achatina	TATGAATTGC	CTGATGGACA	RGTCATYACT	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA	
Archachatina marginata	TATGAATTGC	CTGATGGACA	GGTCATCACY	ATTGGWAAAYG	AGCGCTTCAG	RTGYCCAGAA	
Achatina bisculpta	TATGAATTGC	CTGATGGACA	GGTCATCACY	ATTGGTAAYG	AGCGTTTCAG	ATGTCCAGAA	
Achatina damarensis	TATGAATTGC	CTGATGGACA	GGTCATCACY	ATTGGTAACG	AGCGTTTCAG	ATGTCCAGAA	
Achatina stuhlmanni	TATGAATTGC	CTGATGGACA	GGTCATCACT	ATTGGTAACG	AGCGATTTAG	GTGCCCAGAA	
Atopocochlis exarata	TATGAATTGC	CTGATGGACA	GGTCATCACY	ATTGGTAACG	AGCGTTTCAG	RTGTCCAGAA	
Limicolaria kambeul	TATGAATTGC	CTGATGGACA	GGTCATCACT	ATTGGTAACG	AGCGTTTCAG	RTGYCCAGAA	
Limicolaria martenssii	TATGAATTGC	CTGATGGACA	GGTCATCACT	ATTGGTAACG	AGCGTTTCAG	ATGTCCAGAA	
Limicolariopsis sp.	TATGAATTGC	CTGATGGACA	GGTCATCACY	ATTGGTAACG	AGCGTTTCAG	ATGYCCAGAA	
Cochlitoma varicosa	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA	
Cochlitoma dimidiata	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA	
Cochlitoma sp. cf. vestita	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA	
Cochlitoma marinae	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA	
Cochlitoma churchilliana	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGCTTCAG	GTGCCCAGAA	
Cochlitoma granulata	TATGAATTGC	CTGAYGGACA	GGTCATMACY	ATTGGTAAYG	AGCGCTTCAG	GTGCCCAGAA	
Cochlitoma simplex	TATGAATTGC	CTGATGGACA	GGTCATCACT	ATTGGTAACG	AGCGCTTCAG	GTGCCCAGAA	
Cpchlitoloma kilburni	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGCTTCAG	GTGCCCAGAA	
Cochlitoma montistempli	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGCTTCAG	GTGCCCAGAA	
Cochlitoma omisa	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGCTTCAG	GTGCCCAGAA	
Cochlitoma semidecussata	TATGAATTGC	CTGACGGACA	GGTCATAACT	ATTGGTAATG	AGCGCTTCAG	GTGCCCAGAA	
Cochlitoma ustulata	TATGAATTGC	CTGATGGACA	GGTCATAACT	ATTGGTAATG	AGCGCTTCAG	GTGCCCAGAA	
Cochlitoma zebra	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAATG	AGCGCTTCAG	GTGCCCAGAA	
Metachatina kraussi	TATGAATTGC	CTGATGGACA	GGTCATCACM	ATTGGTAAYG	AGCGTTTYAG	ATGTCCAGAA	
Rumina decollata	TATGAATTGC	CTGAYGGACA	GGTCATCACC	ATCGGCAAYG	AGCGTTTCAG	ATGTCCAGAA	

	610	620	630	640	650	660	
<i>Achatina fulica</i>	GCCATGTTCC	AGCCWTCTTT	CCTTGGTATG	GARWCCGCAG	GTATTCATGA	RACCACCTAC	
<i>Achatina immaculata</i>	GCCATGTTCC	AGCCATCTTT	CCTTGGTATG	GAAWCCGCAG	GTATTCATGA	AACCACCTAC	
<i>Achatina reticulata</i>	GCMRWRTTTC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GYATTCATGA	AACCACCTAC	
<i>Achatina zanzibarica</i>	GCAGAATTTC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GCATTCATGA	GACCACCTAC	
<i>Euaethiopina loveridgei</i>	GCAGAATTTC	AGCCCTCTTT	CCTTGGTATG	GAGTCTGCAG	GCATTCATGA	AACCACCTAC	
<i>Achatina achatina</i>	GCCATGTTCC	AGCCATCTTT	CCTTGGTATG	GARWCTGCAG	GTATTCATGA	AACCACCTAC	
<i>Archachatina marginata</i>	GCCATGTTCC	AGCCATCTTT	CCTTGGTATG	GARTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Achatina bisculpta</i>	GCCATGTTCC	AGCCATCTTT	CCTTGGYATG	GAGTCTGCAG	GTATTCATGA	GACCACSTAC	
<i>Achatina damarensis</i>	GCCATGTTCC	AGCCATCTTT	YCTTGGTATG	GARWCYGCAG	GTATTCATGA	GACCACCTAC	
<i>Achatina stuhlmanni</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCACGA	GACCACCTAC	
<i>Atopocochlis exarata</i>	GCMATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCYGCAG	GTATTCATGA	GACCACCTAC	
<i>Limicolaria kambeul</i>	GCCATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCAYGA	GACCACCTAY	
<i>Limicolaria martenssii</i>	GCCATGTTCC	AGCCMTCTTT	YCTTGGTATG	GAGTCHGCAG	GTATTCAYGA	GACCACCTAC	
<i>Limicolariopsis sp.</i>	GCCATGTTCC	AGCCATCTTT	YCTTGGTATG	GARWCWGCAG	GTATTCAYGA	GACCACCTAC	
<i>Cochlitoma varicosa</i>	GCMRWRTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma dimidiata</i>	GCMRWRTTCC	AGCCWTCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma sp. cf. vestita</i>	GCMRWRTTCC	AGCCATCTTT	CCTTGGYATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma marinae</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTAYTCATGA	AACAACATAC	
<i>Cochlitoma churchilliana</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma granulata</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma simplex</i>	GCAGAATTCC	AGCCTTCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cpchlitoloma kilburni</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma montistempli</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma omissa</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma semidecussata</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma ustulata</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Cochlitoma zebra</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Metachatina kraussi</i>	GCCATGTTCC	AGCCATCTTT	YCTTGGTATG	GARWCYGCAG	GTATTCATGA	RACCACCTAC	
<i>Rumina decollata</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATTCATGA	GACCACCTAC	

	670	680	690	700	710	720	
<i>Achatina fulica</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Achatina immaculata</i>	AACTCCATCA	TGAAGTGYGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Achatina reticulata</i>	AACTCCATCA	TGAAGTGYGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Achatina zanzibarica</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Euaethiopina loveridgei</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Achatina achatina</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTA	
<i>Archachatina marginata</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTA	
<i>Achatina bisculpta</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTACTG	
<i>Achatina damarensis</i>	AACTCCATCA	TGAAGTGYGA	YGTYGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTAYTG	
<i>Achatina stuhlmanni</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTACTT	
<i>Atopocochlis exarata</i>	AACTCCATCA	TGAAGTGYGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTACTG	
<i>Limicolaria kambeul</i>	AACTCCATCA	TGAARTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Limicolaria martenssii</i>	AACTCCATCA	TGAARTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTR	
<i>Limicolariopsis sp.</i>	AACTCCATCA	TGAAGTGYGA	CGTCGACATC	CGTAAAGACT	TRTATKCCAA	CACTGTMTTG	
<i>Cochlitoma varicosa</i>	AACTCCATCA	TGAAGTGYGA	CGTYGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma dimidiata</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma sp. cf. vestita</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma marinae</i>	AACTCCATCA	TGAAGTGCGA	TGTTGATATC	CGTAAAGACT	TGTATGCCAA	CACTGTCTTA	
<i>Cochlitoma churchilliana</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma granulata</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	YACTGTATTG	
<i>Cochlitoma simplex</i>	AACTCCATCA	TGAAGTGCGA	TGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cpchlitoloma kilburni</i>	AACTCCATCA	TGAAGTGCGA	CGTTGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma montistempli</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma omissa</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma semidecussata</i>	AACTCCATCA	TGAAGTGCGA	CGTTGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma ustulata</i>	AACTCCATCA	TGAAGTGCGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Cochlitoma zebra</i>	AACTCCATCA	TGAAGTGCGA	TGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Metachatina kraussi</i>	AACTCCATCA	TGAARTGCGA	YGTYGACATC	CGTAAAGACT	TGTATGCCAA	CWCTGTATTG	
<i>Rumina decollata</i>	AATTCCATCA	TGAARTGTGA	TGTYGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTCTTG	

	730	740	750	760	770	780	
Achatina fulica	TCTGGTGGAW	CTACCATGTA	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATYGYCAAC	
Achatina immaculata	TCTGGTGGAA	CTACCATGTA	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATYGYCAAC	
Achatina reticulata	TCTGGTGGAW	CYACMATGTW	YCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Achatina zanzibarica	TCTGGTGGAT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Euaethiopina loveridgei	TCTGGTGGAT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Achatina achatina	TCTGGTGGAW	CTACCATGTA	TCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACHAAY	
Archachatina marginata	TCTGGTGGAW	CYACCATGTA	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACYAAY	
Achatina bisculpta	TCTGGTGGMT	CYACMATGTW	YCCCGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAC	
Achatina damarensis	TCTGGTGGAT	CYACCATGTW	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACYAAY	
Achatina stuhlmanni	TCTGGTGGCT	CCACAATGTT	TCCAGGCATC	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Atopocochlis exarata	TCTGGTGGAT	CCACCATGTW	CCMYGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACYAAY	
Limicolaria kambeul	TCTGGTGGAT	CYACCATGTA	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAC	
Limicolaria martenssii	TCTGGTGGAT	CTACATGTA	TCCAGGCMTT	GCTGACCGCA	TGCAGAAGGA	AATCACCAC	
Limicolariopsis sp.	TCTGGTGGAT	CTACCATGTW	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCRYAAY	
Cochlitoma varicosa	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGY	
Cochlitoma dimidiata	TCTGGTGGMT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGY	
Cochlitoma sp. cf. vestita	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGY	
Cochlitoma marinae	TCTGGTGGTT	CCACAATGTT	CCCAGGCATC	GCTGACCGCA	TGCAGAAAGGA	AATCACTGCT	
Cochlitoma churchilliana	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAAGGA	AATCACCAGC	
Cochlitoma granulata	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Cochlitoma simplex	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Cpchlitoloma kilburni	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Cochlitoma montistempli	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Cochlitoma omisa	TCTGGTGGCT	CCACAATGTT	CCCKGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Cochlitoma semidecussata	TCTGGTGGCT	CCACAATGTT	CCCGGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Cochlitoma ustulata	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Cochlitoma zebra	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC	
Metachatina kraussi	TCTGGTGGAW	CYACCATGTT	CCCTGGCATT	GCTGACCGYA	TGCAGAAGGA	AATYRYCTSY	
Rumina decollata	TCTGGTGGHT	CAACAATGTT	CCCAGGCATC	GCTGACCGCA	TGCAGAAAGGA	AATCACCAMT	

	790	800	810	820	830	840	
Achatina fulica	CTGGCTCCTT	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Achatina immaculata	CTGGCTCCAT	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	GTACTCTGTC	
Achatina reticulata	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Achatina zanzibarica	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Euaethiopina loveridgei	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Achatina achatina	CTGGCTCCAS	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CAGAACGTAA	ATACTCTGTC	
Archachatina marginata	CTGGCTCCAC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CAGAACGTAA	ATACTCTGTC	
Achatina bisculpta	YTGCTCCWS	CSACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC	
Achatina damarensis	CTGGCTCCTC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATWCTCTGTC	
Achatina stuhlmanni	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC	
Atopocochlis exarata	CTGGCTCCTC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC	
Limicolaria kambeul	CTRGCTCCAC	CCACMATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC	
Limicolaria martenssii	CTRGCTCCAC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC	
Limicolariopsis sp.	CTRGCTCCAC	CCACAATGAA	GATCAAGATH	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC	
Cochlitoma varicosa	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTS	
Cochlitoma dimidiata	CTGGCTCCTS	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTG	
Cochlitoma sp. cf. vestita	CTGGCTCCTS	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTG	
Cochlitoma marinae	CTTGCTCCAC	CCACAATGAA	AATCAAGATT	ATTGCTCCAC	CAGAAAGAAA	ATACTCTGTT	
Cochlitoma churchilliana	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTG	
Cochlitoma granulata	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Cochlitoma simplex	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Cpchlitoloma kilburni	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Cochlitoma montistempli	CTGGCTCCTG	CCACAATGAA	GATCAAGATT	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC	
Cochlitoma omisa	CTGGCTCCTG	CCACAATGAA	GATCAAGATT	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Cochlitoma semidecussata	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Cochlitoma ustulata	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Cochlitoma zebra	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC	
Metachatina kraussi	CTGGCTCCAC	CCACAATGAA	GATCAAGATC	ATTGCTCCWC	CTGAGMGTA	ATACTCTGTC	
Rumina decollata	CTGGCTCCAC	CCACAATGAA	GATCAAAATC	ATTGCTCCTC	CTGAGAGAAA	ATACTCTGTC	

	850 860 870
<i>Achatina fulica</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Achatina immaculata</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Achatina reticulata</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Achatina zanzibarica</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Euaethiopina loveridgei</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Achatina achatina</i>	TGGATTGGAG GTTCCATTCT GGCCTCTCTG TCC
<i>Archachatina marginata</i>	TGGATTGGAG GTTCCATTCT GGCCTCTCTG TCC
<i>Achatina bisculpta</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Achatina damarensis</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Achatina stuhlmanni</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Atopocochlis exarata</i>	TGGATTGGAG GATCCATCCT WGCCTCTCTG TCC
<i>Limicolaria kambeul</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Limicolaria martenssii</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Limicolariopsis sp.</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma varicosa</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma dimidiata</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma sp. cf. vestita</i>	TGGATTGGAG GWTCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma marinae</i>	TGGATTGGAG GCTCCATTTT GGCCTCTCTA TCC
<i>Cochlitoma churchilliana</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma granulata</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma simplex</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cpchlitoma kilburni</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma montistempli</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma omissa</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma semidecussata</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma ustulata</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Cochlitoma zebra</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Metachatina kraussi</i>	TGGATTGGAG GATCCATCCT GGCCTCTCTG TCC
<i>Rumina decollata</i>	TGGATTGGAG GTTCCATCCT GGCCTCTCTG TCC

	130	140	150	160	170	180	
Achatina fulica	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGAGCTT	CTCATCCGCA	AACTGCCCTT	
Achatina immaculata	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGAGCTT	CTCATCCGCA	AACTGCCCTT	
Achatina reticulata	WCTKMGAGAG	ATCCGTCGKT	ACCAGAAGAG	CACKGAGCTK	CTCATYCGCA	ARCTGCCCTT	
Achatina zanzibarica	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGAGCTT	CTCATCCGCA	AACTGCCCTT	
Achatina achatina	TCTKCGAGAR	ATCCGTCGTT	ACCAGAAGAG	CACKGAGCTC	CTCATCCGCA	AGCTGCCCTT	
Archachatina marginata	CCTSMGAGAR	ATCCGTCGTT	ACCAGAAGAG	CASGGAGCTG	CTCATYCGCA	AGYTGCCCTT	
Achatina bisculpta	WCTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	YACGGAGCTT	CTCATCCGCA	ARCTGCCATT	
Achatina damarensis	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	TACGGAGCTT	CTCATCCGCA	AACTGCCATT	
Achatina stuhlmanni	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AACTGCCATT	
Atopocochlis exarata	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AACTGCCATT	
Limicolaria kambeul	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAAAG	CACGGAGCTT	CTCATCCGCA	AACTGCCATT	
Limicolaria martenssii	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAAAG	CACGGAGCTT	CTCATCCGCA	AACTGCCATT	
Limicolariaopsis sp.	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma varicosa	WCTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	ARCTGCCCTT	
Cochlitoma dimidiata	WCTKCGAGAR	ATCCGTCGTT	ACCAGAAGAG	CACWGAGCTT	CTCATCCGCA	ARCTGCCCTT	
Cochlitoma sp. cf. vestita	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAAAG	CACGGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma marinae	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAAAG	CACGGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma churchilliana	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAAAG	CACGGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma granulata	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma simplex	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma kilburni	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma montistempli	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma omissa	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma semidecussata	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma ustulata	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AACTGCCCTT	
Cochlitoma zebra	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AACTGCCCTT	
Metachatina kraussi	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AACTGCCCTT	
Rumina decollata	GCTTCGAGAG	ATCCGACGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	AGCTGCCCTT	

	190	200	210	220	230	240	
Achatina fulica	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTTAAGACC	GATCTGCGTT	TCCAGAGCTC	
Achatina immaculata	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTTAAGACC	GATCTGCGTT	TCCAGAGCTC	
Achatina reticulata	CCAGCGGCTG	GTSAGAGAGA	TYGCCCAGGA	CTTYAAGACC	GATCTGCGYT	TCCAGAGCTC	
Achatina zanzibarica	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTTAAGACC	GATCTGCGTT	TCCAGAGCTC	
Achatina achatina	CCAGCGGCTG	GTCAGAGAAA	TCGCCCAGGA	CTTCAAGACY	GACCTGCGCT	TCCAGAGCTC	
Archachatina marginata	CCAGCGCYTG	GTGAGAGARA	TCGCCCARGA	CTTCAAGACH	GAYCTGCGYT	TCCAGAGCTC	
Achatina bisculpta	CCAGCGGCTG	GTCAGAGAGA	TGCGMCAGGA	CTTCAAGACY	GATCTGCGYT	TCCAGAGCTC	
Achatina damarensis	CCAGCGGCTG	GTCAGAGAGA	TCGCACAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Achatina stuhlmanni	CCAGCGGCTG	GTCAGAGAGA	TCGCACAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Atopocochlis exarata	CCAGCGGCTG	GTCAGAGAGA	TCGCACAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Limicolaria kambeul	CCAGCGGCTG	GTCAGAGAGA	TCGCACAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Limicolaria martenssii	CCAGCGGCTG	GTCAGAGAGA	TCGCACAGGA	CTTCAAGACC	GATCTGCGTT	TYCAGAGCTC	
Limicolariaopsis sp.	CCAGCGGCTG	GTCAGAGAGA	TCGCACAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma varicosa	CCAGCGGYTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma dimidiata	CCAGCGGCTG	GTSMGAGARA	TCGCYCAGGA	CTTCAAGACC	GATCTGCGYT	TCCAGAGCTC	
Cochlitoma sp. cf. vestita	CCAGCGGCTG	GTCARARARA	TCGCCCAGGA	CTTCAAGACT	GATYTGCGTT	TCCAGAGCTC	
Cochlitoma marinae	CCAGCGGCTT	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma churchilliana	CCAGCGGCTG	GTCARAGAGA	TCGCCCAGGA	CTTCAARACC	GATYTGCGTT	TCCARAGCTC	
Cochlitoma granulata	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma simplex	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma kilburni	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACT	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma montistempli	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma omissa	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma semidecussata	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma ustulata	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Cochlitoma zebra	CCAGCGGCTG	GTCAGAGAGA	TCGCCCAGGA	CTTCAAGACT	GATCTGCGTT	TCCAGAGCTC	
Metachatina kraussi	CCAACGGCTG	GTCAGAGAGA	TCGCCCAAGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
Rumina decollata	CCAGCGCCTT	GTCAGAGAGA	TCGCCCAGGA	TTTCAAGACT	GACCTGCGTT	TTCAGAGCTC	

	250	260	270	280	290	300			
Achatina fulica	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Achatina immaculata	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Achatina reticulata	VGCTGTCATG	GCWCTGCAAG	AGGCYAGCGA	RGCMTACCTG	GTGGGTCTGT	TYGAGGACAC			
Achatina zanzibarica	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Achatina achatina	GGCYGTTCATG	GCCCTGCAAG	AGGCYAGCGA	RGCCTACCTG	GTGGGTCTGT	TYGAGGACAC			
Archachatina marginata	GGCTGTCATG	GCYCTGCAAG	AGGCYAGCGA	GGCMTACCTG	GTGGGCCTGT	TYGAGGACAC			
Achatina bisculpta	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Achatina damarensis	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Achatina stuhlmanni	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Atopocochlis exarata	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Limicolaria kambeul	AGCTGTCATG	GCACTGCAAG	AAGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Limicolaria martenssii	AGCTGTCATG	GCACTGCAAG	AAGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGARGACAC			
Limicolariopsis sp.	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma varicosa	GGCTGTCATG	GCACTGCAAG	AGGCYAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma dimidiata	GGCTGTCAWG	GCACTGCAAG	AGGCYAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma sp. cf. vestita	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma marinae	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma churchilliana	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma granulata	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma simplex	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma kilburni	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma montistempli	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma omissa	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma semidecussata	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma ustulata	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Cochlitoma zebra	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC			
Metachatina kraussi	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTA	GTGGGTCTGT	TTGAGGACAC			
Rumina decollata	RGCYGTTCATG	GCACTGCAAG	AGGCTAGCGA	GGCMTACYTR	GTGGGCCTGT	TTGAGGACAC			

	310	320	
Achatina fulica	CAACCTGTGC	GCCATCCACG	CCAAACGT
Achatina immaculata	CAACCTGTGC	GCCATCCACG	CCAAACGT
Achatina reticulata	CAACCTGTGC	GCCATCCAYG	CCAARMGT
Achatina zanzibarica	CAACCTGTGC	GCCATCCACG	CCAAACGT
Achatina achatina	YAACYTGTGC	GCYATCCACG	CCAARCGT
Archachatina marginata	CMACCTKTGC	GCCATCCAYG	CCARVCGT
Achatina bisculpta	CAACCTGTGC	GCCATCCACG	CCAAACGT
Achatina damarensis	CAACCTGTGC	GCCATCCACG	CCAAACGT
Achatina stuhlmanni	CAACCTGTGC	GCCATYCACG	CCAAACGT
Atopocochlis exarata	CAACCTGTGC	GCCATCCACG	CCAAACGT
Limicolaria kambeul	CAACCTGTGC	GCCATCCACG	CCAAACGT
Limicolaria martenssii	CAACCTGTGC	GCCATCCACG	CCAAACGT
Limicolariopsis sp.	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma varicosa	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma dimidiata	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma sp. cf. vestita	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma marinae	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma churchilliana	CAACTTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma granulata	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma simplex	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma kilburni	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma montistempli	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma omissa	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma semidecussata	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma ustulata	CAACCTGTGC	GCCATCCACG	CCAAACGT
Cochlitoma zebra	CAACCTGTGC	GCCATCCACG	CCAAACGT
Metachatina kraussi	CAACCTGTGC	GCCATCCACG	CCAAACGT
Rumina decollata	CAACCTGTGC	GCCATCCACG	CCAAACGT

	10	20	30	40	50	60
<i>Achatina fulica</i>	TTTTTGGTGT	ATGATGTGGG	TTAGTGTGGCA	CAGGCTTGTC	ACTCTTAATT	CGGTTAGAGC						
<i>Achatina immaculata</i>	TTTTGGGGRT	TTGATGTGGC	TTGGTAGGAA	CCGGCCTTTC	ATTATTAATT	CGATTGGAGC						
<i>Achatina reticulata</i>	TCCTCGGGGT	ATGATGTGGT	TTGGTTGGTA	CTGGTTTATC	ATTATTAATT	CGTCTAGAGC						
<i>Achatina zanzibarica</i>	TATTTGGTGT	TTGGTGCGGC	TTGGTGGGGA	CAGGCTTATC	ATTACTAATT	CGACTAGAGT						
<i>Achatina achatina</i>	TTTTTGGGGT	TTGATGTGGC	CTAGTAGGTA	CAGGCTATGC	ATTGTTAATT	CGATTAGAAT						
<i>Archachatina marginata</i>	TTTTTGGGGT	GTGATGTGGC	CTTGTGGGAA	CAGGTCTATC	TTTTTTAATT	CGGTTAGAAT						
<i>Achatina bisculpta</i>	TTTTTGGTGT	TTGATGTGGT	TTGGTAGGAA	CTGGGCTATC	TTTACTAATT	CGGTTAGAGT						
<i>Achatina damarensis</i>	TTTTTGGTGT	GTGGTGTGGG	TTGGTGCGGA	CTGGGCTATC	ATTACTAATT	CGGTTAGAGT						
<i>Achatina stuhlmanni</i>	TTTTTGGGGT	CTGGTGTGGA	TTAGTTGGTA	CCGGCTTGTC	TTTACTTATC	CGGCTAGAAC						
<i>Atopocochlis exarata</i>	TCTTTGGGGT	TTGATGTGGG	CTAGTGGGTA	CTGGCTTATC	TATTTTAATT	CGATTAGAAC						
<i>Limicolaria kambeul</i>	TTTTTGGAAAT	TTGATGTGGG	TTGGTTGGGA	CAGGCCATAT	GTTTTAAATT	CGTTTAGAAC						
<i>Limicolaria martenssii</i>	TCTTTGGAGT	ATGATGTGGT	TTAGTGGGGA	CAGGGTTATC	ATTACTGATT	CGTTTAGAGT						
<i>Limicolariopsis sp.</i>	TCTTAGGGGT	ATGATGTGGG	TTAGTGGGAA	CAGGCCATAT	ATTATTAATT	CGACTAGAAC						
<i>Cochlitoma varicosa</i>	TTTTTCGGGGT	GTGATGTGGT	TTAGTTGGGA	CTGGGCTATC	TTTACTTATT	CGATTAGAGT						
<i>Cochlitoma dimidiata</i>	TTTTTGGTGT	TTGATGTGGT	TTGGTTGGGA	CCGGGCTATC	TCTACTAATC	CGTTTAGAGC						
<i>Cochlitoma sp. cf. vestita</i>	TTTTTGGTGT	GTGATGTGGT	TTGGTTGGGA	CTGGGCTATC	CTTACTAATC	CGTCTAGAGT						
<i>Cochlitoma marinae</i>	TCTTCGGGAC	CTGGTGTGGC	TTGGTGGGCA	CTGGGCTATC	TTTACTAATC	CGTTTGGAGT						
<i>Cochlitoma churchilliana</i>	TTTTTCGGGGT	CTGGTGTGGG	TTAGTGGGGA	CTGGTTTGTC	CCATTAATT	CGATTAGAGC						
<i>Cochlitoma granulata</i>	TCTTTGGATT	CTGATGCGGG	TTAGTTGGTA	CTGGGTTATC	GTTATTGATC	CGGTTAGAAC						
<i>Cochlitoma simplex</i>	TTTTTGGAGT	TTGATGTGGA	CTAGTGGGAA	CTGGGTTATC	CTTACTAATC	CGACTAGAAC						
<i>Cochlitoma kilburni</i>	TTTTTGGAGT	GTGATGTGGC	TTGGTTGGGA	CAGGACTATC	ACTGCTTATC	CGTTTAGAGT						
<i>Cochlitoma montistempli</i>	TTTTTGGGTT	TTGGTGCGGA	TTGGTTGGAA	CAGGGCTATC	ATTATTAATT	CGATTAGAGC						
<i>Cochlitoma omissa</i>	TTTTTCGGGTT	CTGATGCGGG	TTAGTTGGTA	CTGGGTTATC	ATTATTGATC	CGGTTAGAGC						
<i>Cochlitoma semidecussata</i>	TTTTTCGGGTT	TTGGTGCGGG	TTGGTTGGTA	CAGGGCTATC	GTTATTAATT	CGATTAGAGC						
<i>Cochlitoma ustulata</i>	TTTTTGGGGT	ATGGTGTGGT	CTAGTGGGCA	CAGGCTATCT	TTTTATTGATC	CGACTAGAGC						
<i>Cochlitoma zebra</i>	TTTTTCGGAGT	GTGATGTGGA	CTGGTTGGGA	CWGRTTTATC	TTTAYTKATY	CGGYTAGAGT						
<i>Metachatina kraussi</i>	TTTTTGGGGT	ATGATGTGGC	TTAGTTGGCA	CAGGCTTATC	TCTTTTAAAT	CGACTAGAAC						
<i>Rumina decollata</i>	TTTTTGGGGT	CTGATGCGGT	TTAGTTGGTA	CTGGTTTATC	TCTTCTTATT	CGTTTAGAAC						

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	130	140	150	160	170	180	
Achatina fulica	CTTTTGTGAT	AATTTTCTTT	ATGGTTATAC	CAATTATAAT	TGGCGGATTT	GGAAACTGGA	
Achatina immaculata	CGTTTGTGAT	AATTTTCTTT	ATGGTTATGC	CAATTATGAT	TGGGGGATTT	GGTAATTGAA	
Achatina reticulata	CTTTTGTGAT	AATTTTCTTT	ATAGTTATAC	CAATTATAAT	CGGAGGATTT	GGGAACTGAA	
Achatina zanzibarica	CATTGTGTTAT	AATTTTCTTT	ATAGTGATAC	CAATTATGAT	TGGGGGGTTT	GGGAACTGAA	
Achatina achatina	CTTTTGTGAT	AATTTTCTTT	ATAGTCATAC	CAATTATGAT	TGGGGGGTTT	GGTAATTGAA	
Archachatina marginata	CATTGTGTAAT	AATTTTCTTT	ATGGTTATGC	CAATTATAAT	TGGGGGGTTT	GGAAACTGAA	
Achatina bisculpta	CTTTTGTGTAAT	GATTTTCTTT	ATAGTAATGC	CTATTATAAT	TGGGGGGTTT	GGTAATTGAA	
Achatina damarensis	CTTTTGTGTAAT	AATCTTCTTC	ATAGTTATAC	CTATTATAAT	TGGAGGATTT	GGTAAC TGAA	
Achatina stuhlmanni	CTTTTGTGAT	AATTTTCTTT	ATAGTGATAC	CAATTATAAT	TGGTGGGTTT	GGAAACTGGA	
Atopocochlis exarata	CTTTTCGTAAT	AATTTTCTTT	ATAGTTATAC	CAATCATAAT	TGGCGGGTTT	GGGAACTGAA	
Limicolaria kambeul	CTTTTGTGTAAT	AATCTTCTTC	ATAGTTATAC	CTATTATAAT	TGGAGGTTTC	GGAAAT TGGA	
Limicolaria martenssii	CTTTTGTGTTAT	AATTTTCTTT	ATAGTTATAC	CCATTATAAT	TGGAGGATTT	GGAAAT TGAA	
Limicolariopsis sp.	CCTTCGTAAT	AATTTTCTTT	ATAGTTATAC	CTATTATAAT	TGGTGGATTT	GGTAAT TGAA	
Cochlitoma varicosa	CATTGTGTAAT	AATTTTCTTT	ATAGTGATAC	CCATTATAAT	TGGTGGTTT	GGGAACTGGA	
Cochlitoma dimidiata	CTTTTCGTTAT	AATTTTCTTC	ATAGTAATAC	CCATTATAAT	TGGGGGGTTT	GGGAACTGAA	
Cochlitoma sp. cf. vestita	CATTTCGTTAT	AATTTTCTTT	ATGGTAATAC	CAATTATAAT	TGGGGGGTTT	GGTAAT TGAA	
Cochlitoma marinae	CATTGTGAT	GATTTTCTTC	ATAGTAATAC	CAATTATGAT	TGGGGGGTTT	GGGAACTGGA	
Cochlitoma churchilliana	CGTTTGTGAT	GATCTTCTTC	ATGGTAATGC	CTATTATAAT	TGGTGGTTT	GGAAAT TGAA	
Cochlitoma granulata	CATTTCGTTAT	AATTTTCTTC	ATAGTAATAC	CAATTATAAT	TGGGGGGTTT	GGTAAC TGAA	
Cochlitoma simplex	CTTTTGTGTTAT	GATTTTCTTC	ATGGTGATGC	CCATTATAAT	CGGGGGCTTT	GGAAAT TGGA	
Cochlitoma kilburni	CATTTCGTTAT	AATTTTCTTC	ATAGTAATAC	CTATTATAAT	TGGGGGGTTT	GGCAACTGGA	
Cochlitoma montistempli	CGTTTGTGTTAT	AATTTTCTTT	ATAGTAATAC	CTATTATAAT	TGGGGGGTTT	GGGAACTGGA	
Cochlitoma omissa	CATTTCGTTAT	AATTTTCTTT	ATAGTAATAC	CTATTATAAT	CGGGGGTTT	GGTAAC TGAA	
Cochlitoma semidecussata	CATTGTGTTAT	AATTTTCTTT	ATAGTAATAC	CTATTATAAT	TGGGGGGTTT	GGGAAT TGAA	
Cochlitoma ustulata	CTTTTGTGTAAT	AATTTTCTTC	ATAGTAATAC	CCATTATAAT	TGGGGGGTTT	GGTAAT TGAA	
Cochlitoma zebra	CTTTTGTGTTAT	AATCTTCTTC	ATGGTAATAC	CCATTATAAT	TGGGGGATTC	GGAAAT TGRA	
Metachatina kraussi	CATTGTGTTAT	GATTTTCTTT	ATAGTTATGC	CTATTATAAT	TGGTGGATTT	GGAAAT TGGA	
Rumina decollata	CATTGTGAT	AATTTTCTTT	ATAGTAATGC	CTATTATAAT	CGGTGGTTT	GGAAAT TGAA	

	190	200	210	220	230	240	
Achatina fulica	TGGTCCCAAT	ACTTATTGGT	GTCCTGATA	TAAGATTTC	ACGAATAAAT	AATATAAGGT	
Achatina immaculata	TGGTTCCCT	TTTAATTGGG	GTCCTGATA	TAAGTTTTC	CCGGATGAAT	AATATGAGGT	
Achatina reticulata	TGGTTCCCAT	ACTTATTGGG	GTCCTGATA	TAAGTTTTC	ACGTATAAAC	AATATAAGAT	
Achatina zanzibarica	TGGTACCACT	ATTAATTGGA	GCACCTGACA	TAAGATTCCC	ACGCATAAAT	AATATAAGAT	
Achatina achatina	TAGTCCCACT	ATTAATTGGG	GTCCTGATA	TAAGATTCCC	TCGGATGAAT	AATATAAGGT	
Archachatina marginata	TAGTTCCTCT	ATTAATTGGG	GTCCTGATA	TAAGTTTTC	TCGATAAAT	AATATAAGGT	
Achatina bisculpta	TGGTCCCACT	TTTAATTGGG	GCACCTGATA	TAAGATTTC	TCGAATAAAT	AATATAAGAT	
Achatina damarensis	TGGTGCCCT	TTTGATTGGT	GCACCTGACA	TAAGATTTC	TCGTATAAAC	AACATAAGGT	
Achatina stuhlmanni	TGGTCCCAT	ACTGATTGGT	GTCCTGATA	TAAGTTTTC	GCGGATAAAT	AACATAAGAT	
Atopocochlis exarata	TAGTTCCATT	ATTAATTGGG	GCCCCGACA	TAAGTTTCCC	TCGAATAAAT	AACATGAGAT	
Limicolaria kambeul	TAGTTCCCTT	ATTAATTGGT	GTCCTGATA	TAAGATTCCC	CCGAATGAAT	AATATAAGAT	
Limicolaria martenssii	TAGTACCATT	ATTGATTGGT	GCACCTGATA	TAAGCTTTC	TCGCATGAAT	AATATAAGAT	
Limicolariopsis sp.	TAGTACCATT	GTTAATTGGG	GCACCTGACA	TAAGATTTC	TCGTATAAAC	AATATAAGGT	
Cochlitoma varicosa	TAGTCCCTT	GCTAATTGGT	GCCCCGATA	TAAGCTTTC	CCGAATGAAT	AATATAAGTT	
Cochlitoma dimidiata	TAGTTCCCTT	ATTGATTGGG	GCACCTGATA	TGAGATTTC	TCGTATAAAC	AATATAAGCT	
Cochlitoma sp. cf. vestita	TGGTCCCTT	ACTGATCGGG	GCCCCGATA	TAAGTTTTC	CCGAATGAAT	AATATAAGTT	
Cochlitoma marinae	TAGTACCCTT	ATTGGTTGGT	GCCCCGATA	TAAGATTTC	TCGAATAAAT	AATATAAGTT	
Cochlitoma churchilliana	TGATTCCCT	ACTAATTGGG	GTCCTGATA	TAAGTTTTC	GCGAATAAAC	AACATAAGGT	
Cochlitoma granulata	TAGTACCAAT	GTTAATCGGG	GCCCCGATA	TAAGTTTTC	TCGTATAAAT	AATATAAGCT	
Cochlitoma simplex	TAGTTCCCTT	GCTAATTGGG	GCACCTGACA	TGAGATTTC	TCGTATAAAC	AATATAAGCT	
Cochlitoma kilburni	TGGTTCCCTT	GTTGATTGGG	GTCCTGATA	TAAGTTTTC	CCGAATGAAT	AATATAAGCT	
Cochlitoma montistempli	TAGTACCATT	GCTTATTGGG	GTCCTGATA	TAAGATTTC	CCGTATGAAT	AATATAAGCT	
Cochlitoma omissa	TAGTACCATT	GTTAATCGGG	GCCCCGATA	TAAGTTTTC	TCGTATAAAT	AATATAAGCT	
Cochlitoma semidecussata	TAGTTCCCAT	GTTAATTGGG	GCCCCGATA	TAAGATTTC	TCGTATAAAT	AATATAAGCT	
Cochlitoma ustulata	TAAC TCCCT	TTTAATCGGG	GTCCTGATA	TAAGCTTTC	TCGTATAAAT	AACATGAGCT	
Cochlitoma zebra	TAATYCCCTT	ACTTATTGGG	GTCCTGATA	TAAGCTTTC	CCGGATAAAT	AACATRAGYT	
Metachatina kraussi	TAGTTCCACT	ATTAATTGGG	GTCCTGATA	TAAGCTTTC	ACGAATAAAT	AATATAAGCT	
Rumina decollata	TAGTCCCTT	ATTAATTGGG	GCACCTGATA	TAAGTTTTC	TCGAATAAAT	AATATGAGAT	

	
	250 260 270 280 290 300	
<i>Achatina fulica</i>	TTTGACTTTT ACCACCTTCA TTTACTTTGT TAATCTGTTT AAGTATAGTG GAAGGAGGGG	
<i>Achatina immaculata</i>	TTTGGCTTCT CCCACCATCG TTTACTTTAT TAATTAGGTC AAGCCTGGTT GAAGGTGGTG	
<i>Achatina reticulata</i>	TTTGACTTTT ACCCCCTGCG TTTACATTAT TAATCTGCTC AAGTATAGTC GAAGGTGGGG	
<i>Achatina zanzibarica</i>	TTTGACTTCT ACCACCTTCT TTTTGTCTTT TAATTAGATC GAGAATGGTA GAGGGGGGGG	
<i>Achatina achatina</i>	TCTGATTATT ACCACCTTCA TTTACCTTAC TTATTCTTTC TAGTATAGTT GAGGGAGGAG	
<i>Archachatina marginata</i>	TTTGATTGCT TCCACCTTCA TTTGTCTGCG TAATCTCTCT GAGGATGGTC GAGGGTGGAG	
<i>Achatina bisculpta</i>	TCTGACTTTT ACCCCCATCT TTTACTCTTT TAATCCTATC GAGTATAGTT GAAGGTGGAG	
<i>Achatina damarensis</i>	TTTGACTTTT ACCACCGTCT TTTACTTTAC TAATTTTATC AAGTATGGTT GAAGGTGGAG	
<i>Achatina stuhlmanni</i>	TTTGACTACT TCCCCCATCA TTTTCTTTGC TAATCTCTCT AAGTATAGTA GAAGGTGGGG	
<i>Atopocochlis exarata</i>	TTTGGTTACT TCCACCATCA TTTACCCTTT TAATTTTATC TAGAATAGTA GAAGGAGGGG	
<i>Limicolaria kambeul</i>	TTTGATTACT ACCACCTTCT TTTACTTTAC TAATCTTATC AAGCATAGTA GAGGGTGGGG	
<i>Limicolaria martenssii</i>	TCTGGTTACT ACCCCAGCT TTTACTTTAT TAATCTCTCT TAGGATAGTA GAAGGAGGCG	
<i>Limicolariaopsis sp.</i>	TCTGATTACT TCCTCCTGCA TTTACTTTAC TAATTTTATC AAGGATAGTA GAAGGAGGCG	
<i>Cochlitoma varicosa</i>	TCTGACTATT ACCACCTTCG TTTCCCTGT TAATCTATC AAGGATAGTT GAGGGGGGTG	
<i>Cochlitoma dimidiata</i>	TTTGATTGCT CCCACCTTCA TTTTCTCTTT TGATTTTATC AAGAATAGTG GAAGGGGGAG	
<i>Cochlitoma sp. cf. vestita</i>	TTTGATTACT TCCACCTTCG TTTTCTCTAT TAATTTTATC AAGAATGGTA GAAGGTGGTG	
<i>Cochlitoma marinae</i>	TCTGATTATT ACCACCTTCA TTCTCACTAC TAATCTTATC TAGTATAGTA GAAGGGGGAG	
<i>Cochlitoma churchilliana</i>	TCTGATTACT TCCCCCTCT TTTTCGTTAC TGATTTTATC TAGTATAGTG GAAAGTGGTG	
<i>Cochlitoma granulata</i>	TTTGACTCCT ACCCCCTCA TTCTCTCTTC TAATCTTATC AAGAATGGTT GAGGGTGGTG	
<i>Cochlitoma simplex</i>	TCTGACTGCT TCCTCCTCA TTTTCTCTTC TAATCTTATC AAGACTAGTG GAGGGTGGGG	
<i>Cochlitoma kilburni</i>	TTTGATTACT CCCACCTTCG TTTTCACTTC TAATCTTATC AAGAATGGTA GAGGTGGTG	
<i>Cochlitoma montistempli</i>	TCTGGCTCCT ACCCCCTTCT TTTTCCCTCT TAATTTTATC AAGAATAGTA GAGGGCGGAG	
<i>Cochlitoma omissa</i>	TTTGACTCCT GCCCCCTCG TTCTCTCTTT TAATCTTATC TAGAATGGTT GAGGGTGGTG	
<i>Cochlitoma semidecussata</i>	TCTGGCTCCT ACCCCCTTCG TTTTCCCTCT TAATTTTATC AAGAATAGTA GAGGGTGGGG	
<i>Cochlitoma ustulata</i>	TTTGATTGTT ACCCCCTTCC TTTTCACTTC TTATCTTGTC AAGAATAGTG GAGGGGGGTG	
<i>Cochlitoma zebra</i>	TTTGATTCTT CCCMCCTTCG TTCTCTCTAC TAATTTTCTC AAGRATAGTA GAGGGAGGTG	
<i>Metachatina kraussi</i>	TTTGGTTATT ACCGCGTCT TTTACATTAT TGATTTTGTC AAGAATAGTA GAAGGAGGTG	
<i>Rumina decollata</i>	TTTGACTACT TCCTCCTTCT TTTATTTTAT TAATTATATC TAGGATAGTA GAAGGTGGGG	

	
	310 320 330 340 350 360	
<i>Achatina fulica</i>	CTGGAAGTGG GTGGACTGTG TACCCGCCCT TAAGTTCTTG CTTAGGACAC AGAGGGGCTT	
<i>Achatina immaculata</i>	CTGGGACTGG TTGAAGTGTG TACCCGCCGT TAAGTTCTTA TATGGGTCAC AGGGGGGCCCT	
<i>Achatina reticulata</i>	CTGGCACAGG TTGAAGTGTG TACCCACCTT TAAGCTCTTA TTAGGGGCAT AGTGGGGCAT	
<i>Achatina zanzibarica</i>	CAGGTACAGG TTGGACTGTG TACCCACCAT TGAGCTCAAT TATTGGGCAT AGGGGCCCAT	
<i>Achatina achatina</i>	CTGGTACTGG GTGAACAGTT TATCCACCTT TAAGATCTAC TTTAGGTCAT AGAGGTGCCT	
<i>Archachatina marginata</i>	CAGGTACTGG GTGGACAGTT TACCCCCCCC TTAGGTCAGT CTTAGGACAT AGTGGAGCTT	
<i>Achatina bisculpta</i>	CGGGAAGTGG CTGAACAGTT TATCCACCAT TTAGTTCATT AATTGGACAT AGTGGAGCAT	
<i>Achatina damarensis</i>	CAGGAAGTGG TTGGACAGTT TACCCACCTC TTAGTTCTCT GATTGGTCAT AGGGGAGCAT	
<i>Achatina stuhlmanni</i>	CTGGGACAGG ATGGACAGTT TACCTCCTC TAAGTTTCAAT AATTGGCCAT AGTGGAGCAT	
<i>Atopocochlis exarata</i>	CAGGGACTGG TTGAACGGTG TACCCCCCTC TTAGCTCTAT GCTTGGCCAC AGGGGTGCCT	
<i>Limicolaria kambeul</i>	CTGGAAGTGG TTGAACAGTT TACCTCCTC TTAGAGCTAT TACAGGCCAT AGAGGTGCTT	
<i>Limicolaria martenssii</i>	CGGGAACAGG GTGAAGTGTG TATCCTCCTT TAAGATCCCT AGTGGGTCAT AGAGGGGCCA	
<i>Limicolariaopsis sp.</i>	CAGGAAGTGG TTGAAGTGTG TACCCCCCTC TAAGTTCTAT AATTGGACAT AGAGGAGCCT	
<i>Cochlitoma varicosa</i>	CAGGGACTGG TTGAACAGTT TACCCCCCTC TGAGTTCTAT AATAGGACAT AGAGGAGCGT	
<i>Cochlitoma dimidiata</i>	CAGGGACAGG CTGGACGGTA TACCCGCCTC TAAGTTCGAT TATAGGTCAC AGAGGGGCTT	
<i>Cochlitoma sp. cf. vestita</i>	CAGGTACTGG GTGAAGTGTG TACCCACCTC TCAGGTCTAT GTTAGGGCAT AGAGGTGCGT	
<i>Cochlitoma marinae</i>	CAGGCACCGG TTGGACTGTG TACCCGCCTT TAAGCTCTAT TATTGGTCAT AGGGGCGCTT	
<i>Cochlitoma churchilliana</i>	CTGGGACAGG GTGAACAGTA TACCTCCCC TGAGGTCAAC TATTGGTCAT AGGGGCGCAT	
<i>Cochlitoma granulata</i>	CAGGTACAGG TTGAAGTGTG TACCTCCTT TGAGTTCTGT AATAGGCCAT AGAGGTGCAT	
<i>Cochlitoma simplex</i>	CAGGAACAGG CTGAACAGTA TACCTCCCC TTAGTTCTAT TATTGGTCAT AGAGGAGCTT	
<i>Cochlitoma kilburni</i>	CAGGTACTGG TTGAAGTGTG TACCCACCTC TTAGATCTAT AATTAGGCAT AGGGGTGCAT	
<i>Cochlitoma montistempli</i>	CAGGTACTGG TTGAACAGTT TACCCCCCTT TAAGTTCAAT TATTGGCCAT AGAGGTGCAT	
<i>Cochlitoma omissa</i>	CAGGTACTGG TTGAACAGTT TACCCCCCTT TGAGCTCTAT AATAGGTCAT AGAGGTGCAT	
<i>Cochlitoma semidecussata</i>	CAGGTACTGG TTGAACGGTT TACCTCCTT TAAGTTCAAT TATTGGCCAT AGAGGAGCAT	
<i>Cochlitoma ustulata</i>	CAGGTACAGG TTGAAGTGTG TATCCACCTT TAAGGTCTAT CATAGGTCAT AGGGGAGCAT	
<i>Cochlitoma zebra</i>	CWGGTACWGG CTGRACAGTG TATCCCCCTT TAAGTTCCAT GCTWGGACAT AGRGGWGCAT	
<i>Metachatina kraussi</i>	CCGGGACTGG TTGAAGTGTG TACCCACCGT TAAGTTCTAT TCTTGGGCAT AGGGGAGCAT	
<i>Rumina decollata</i>	CAGGAAGTGG TTGAACAGTG TATCCACCTT TAAGCAGAAT CATAGGACAT AGTGGGGCAT	

	370	380	390	400	410	420
<i>Achatina fulica</i>	CAGTTGATTT	AGCTATTTTT	TCTTTACATT	TGGCTGGTGT	ATCCTCAATT	TTAGGGGCAA	
<i>Achatina immaculata</i>	CAGTTGATTT	AGCAATTTTT	TCCCTTCATC	TTGCTGGTGT	CTCATCGATT	TTAGGGGCAA	
<i>Achatina reticulata</i>	CAGTAGATTT	AGCTATCTTC	TCTCTCCACT	TAGCTGGGGT	ATCCTCAATC	TTAGGGGCTA	
<i>Achatina zanzibarica</i>	CTGTTGATTT	AGCGATTTTT	TCACCTCATT	TGGCAGGAGT	TTCTGCAATT	TTAGGGGCAA	
<i>Achatina achatina</i>	CGGTAGATTT	AGCGATTTTT	TCATTACATT	TGGCTGGTAT	ATCATCTATC	TTAGGGGCTA	
<i>Archachatina marginata</i>	CTGTGGACCT	GGCTATTTTT	TCTTTGCACT	TAGCCGGTAT	ATCTTCTATT	CTTGGTGCAA	
<i>Achatina bisculpta</i>	CAGTGGATTT	AGCAATTTTT	TCTTTACACT	TAGCTGGTAT	ATCCTCAATC	CTTGGAGCCA	
<i>Achatina damarensis</i>	CTGTAGATTT	AGCAATTTTT	TCTTTACATC	TAGCTGGGAT	ATCCTCAATC	CTTGGTGCTA	
<i>Achatina stuhlmanni</i>	CAGTGGACTT	GGCAATTTTT	TCTCTACACT	TAGCCGGAAT	ATCCTCTATT	CTCGGAGCTA	
<i>Atopocochlis exarata</i>	CGGTTGACTT	AGCAATTTTT	TCACTTCATC	TAGCAGGTAT	ATCCTCGATT	CTTGGGGCTA	
<i>Limicolaria kambeul</i>	CAGTAGACTT	AGCAATCTTC	TCTCTTCACC	TTGCAGGAAT	GTCTTCAATC	CTAGGAGCAA	
<i>Limicolaria martenssii</i>	GGGTTGACTT	AGCTATTTTT	TCCTTACACT	TAGCTGGAAT	ATCTTCCATT	CTAGGGGCTA	
<i>Limicolariopsis</i> sp.	CAGTAGATTT	AGCTATTTTT	TCATTACATC	TAGCAGGTAT	GTCTTCCATT	TTAGGGGCTA	
<i>Cochlitoma varicosa</i>	CTGTAGATCT	GGCTATCTTC	TCTCTTCATC	TTGCTGGGAT	GTCTTCAATC	TTAGGCGCCA	
<i>Cochlitoma dimidiata</i>	CTGTTGATTT	GGCCATTTTT	TCCTTACATC	TGGCAGGAAT	ATCGTCAATT	CTAGGTGCTA	
<i>Cochlitoma</i> sp. cf. <i>vestita</i>	CTGTGATTT	AGCTATCTTC	TCACTACATC	TTGCAGGAT	ATCTTCGATT	TTAGGAGCCA	
<i>Cochlitoma marinae</i>	CAGTGGATCT	AGCTATTTTT	TCTCTTCACC	TAGCTGGAAT	ATCTTCTATT	TTAGGTGCTA	
<i>Cochlitoma churchilliana</i>	CAGTGGATTT	AGCAATTTTT	TCCTTTCACC	TGGCGGGGAT	ATCTTCAATC	CTAGGTGCTA	
<i>Cochlitoma granulata</i>	CAGTAGATTT	AGCTATTTTT	TCCCTTCACC	TCGCAGGTCT	GTCTTCAATC	TTGGGTGCTA	
<i>Cochlitoma simplex</i>	CTGTAGATCT	GGCTATTTTT	TCCTTACATC	TAGCAGGAAT	GTCTTCAATC	TTAGGCGCTA	
<i>Cochlitoma kilburni</i>	CTGTGATTT	AGCAATCTTC	TCATTACATC	TTGCAGGAT	ATCCTCAATT	ATCGGCTA	
<i>Cochlitoma montistempli</i>	CTGTAGATCT	GGCTATTTTT	TCGCTACATC	TTGCGGGTAT	ATCCTCAATT	CTAGGTGCTA	
<i>Cochlitoma omisa</i>	CTGTGGATTT	AGCTATTTTT	TCTCTTCACC	TTGCGGGTCT	GTCTTCAATC	CTAGGTGCTA	
<i>Cochlitoma semidecussata</i>	CGGTGGACCT	TGCAATCTTC	TCACTTCATC	TTGCGGGTAT	ATCCTCAATT	CTTGGTGCTA	
<i>Cochlitoma ustulata</i>	CAGTGGATTT	GGCTATCTTC	TCTCTACACT	TAGCTGGTAT	ATCTTCTATT	CTCGGGGCTA	
<i>Cochlitoma zebra</i>	CCGTSGATCT	RGCTATCTTC	TCTCTACATY	TMGCTGGCAT	ATCCTCTATT	TTAGGKGCCA	
<i>Metachatina kraussi</i>	CAGTTGACTT	AGCTATTTTT	TCTTTACATT	TAGCTGGAAT	ATCATCTATT	TTAGGAGCTA	
<i>Rumina decollata</i>	CTGTTGACTT	AGCTATTTTT	TCTCTTCATT	TAGCAGGAT	ATCTTCTATT	TTAGGTGCAA	

	430	440	450	460	470	480
<i>Achatina fulica</i>	TCAATTTTAT	TACGACGGTG	TATAATATAC	GAGCCTCTGG	GCTAACTATA	GAACGGGTAA	
<i>Achatina immaculata</i>	TTAATTTTAT	TACAACAGTA	TATAATATAC	GTTCTTCAGG	GTTAACCATA	GAACGAGTAA	
<i>Achatina reticulata</i>	TTAATTTTAT	TACTACAGTT	TACAATATAC	GTGCTTCTGG	GTTAACTATA	GAACGAGTAA	
<i>Achatina zanzibarica</i>	TTAATTTTAT	TACCACAGTC	TATAATATGC	GGGCTTCGGG	CTTAGAGATA	GAACGGATGA	
<i>Achatina achatina</i>	TTAATTTTAT	TACTACAATT	TTTAATATAC	GAGCATCAGG	GATATCTATA	GAACGGGTAA	
<i>Archachatina marginata</i>	TTAATTTTAT	CACAACATAT	TTCAACATGC	GAGCTCCCGG	TCTTACAATG	GAGCGGGTAA	
<i>Achatina bisculpta</i>	TTAATTTTAT	TACTACTATC	TTTAATATAC	GGGTGTCTGG	TTTGTCTATG	GAACGGGTAA	
<i>Achatina damarensis</i>	TTAATTTTAT	TACTACTATC	TTCAATATGC	GGGTGTCTGG	GCTAACTATA	GAACGGGTAA	
<i>Achatina stuhlmanni</i>	TCAATTTTAT	TACTACAATT	TTTAACATAC	GAGTATCTGG	ACTATCTATG	GAACGGGTGA	
<i>Atopocochlis exarata</i>	TCAATTTTAT	TACCACAATT	TATAATATAC	GGGTAGGGG	ATTGACAATA	GAACGAGTAA	
<i>Limicolaria kambeul</i>	TTAATTTTAT	TACTACAATT	TTTAATATAC	GAGCATCAGG	GTTATCTTTA	GAGCGTATAA	
<i>Limicolaria martenssii</i>	TTAATTTTAT	TACTACTATT	TTTAATATAC	GAGTAAAGG	CTTATCTATA	GAACGTCTAA	
<i>Limicolariopsis</i> sp.	TTAATTTTAT	TACAACGTGA	TTTAATATGC	GAGCATCAGG	GTTAACTATA	GAACGAGTAA	
<i>Cochlitoma varicosa</i>	TTAATTTTAT	TACCACAGTA	TTTAATATAC	GAGCGCAGGG	GTTATCTATG	GAGCGAGTCA	
<i>Cochlitoma dimidiata</i>	TTAATTTTAT	TACAACAGTA	TTTAATATAC	GGGTGCAAGG	ACTCTCAATG	GAACGGGTAA	
<i>Cochlitoma</i> sp. cf. <i>vestita</i>	TTAATTTTAT	TACTACAGTA	TTTAATATAC	GAGCACAAGG	CTTATCAATA	GAACGTGTCA	
<i>Cochlitoma marinae</i>	TTAATTTTAT	CACCACTATT	TTTAATATAC	GGGCGCAAGG	TTTAACATATA	GAACGAGTGA	
<i>Cochlitoma churchilliana</i>	TTAATTTTAT	TACCACAGTC	TTTAATATAC	GTACTCCTGG	TTTAACATATA	GAACGGGTAA	
<i>Cochlitoma granulata</i>	TTAATTTTAT	TACTACAGTG	TTTAACATAC	GGGCACAAGG	ATTATCTATG	GAACGGGTGA	
<i>Cochlitoma simplex</i>	TTAATTTTAT	TACCACAGTG	TTCAATATGC	GGGTGAAAGG	CCTGTCAATA	GAACGAGTGA	
<i>Cochlitoma kilburni</i>	TTAATTTTAT	TACTACAGTA	TTCAATATAC	GAGCACAAGG	ACTATCAATA	GAACGAGTCA	
<i>Cochlitoma montistempli</i>	TTAATTTTAT	TACTACAGTG	TTTAATATAC	GGGCCAGGG	ATTATCAATG	GAACGAGTAA	
<i>Cochlitoma omisa</i>	TTAATTTTAT	TACAACAGTG	TTTAATATAC	GGGCACAAGG	GTTAACTATG	GAACGAGTCA	
<i>Cochlitoma semidecussata</i>	TCAACTTTCAT	TACTACAGTG	TTTAATATAC	GGGCCAGGG	AATATCTATA	GAACGAGTAA	
<i>Cochlitoma ustulata</i>	TTAATTTTAT	CACAACATATC	TTTAATATAC	GAGCTCAAGG	CTTATCTATG	GAGCGTATAA	
<i>Cochlitoma zebra</i>	TTAATTTTAT	YACTACTGTA	TTTAATATAC	GAGCACAGGG	GTTGTCAATG	GAACGAGTCA	
<i>Metachatina kraussi</i>	TTAATTTTAT	TACTACCGTC	TTTAATATAC	GAACCCCGGG	GTTATCTATA	GAGCGGGTAA	
<i>Rumina decollata</i>	TTAATTTTAT	TACTACTATT	TATAATATAC	GGTCCCCAGG	TCTTTCAATA	GAACGGGTAA	

	490	500	510	520	530	540	
<i>Achatina fulica</i>	GCTTATTTCGT	GTGGTCAATT	TTAGTTACAG	TGTTTCTACT	ATTGTTATCA	TTGCCAGTTC	
<i>Achatina immaculata</i>	GGCTATTTCGT	CTGGTCTATT	TTAGTAACCG	TTTTTTTATT	ACTGCTATCC	CTACCAGTTC	
<i>Achatina reticulata</i>	GATTATTTCGT	GTGATCAATT	TTGGTTACCG	TGTTTCTTTT	ACTACTATCA	TTACCTGTAC	
<i>Achatina zanzibarica</i>	GTTTATTTCGT	TTGATCAATT	TTAGTCACCG	TATTTTACT	ACTACTCTCA	TTACCTGTAT	
<i>Achatina achatina</i>	GACTGTTTCGT	ATGATCTATT	TTAGTTACAG	TATTTTACT	GCTTTTATCT	CTCCCAGTAT	
<i>Archachatina marginata</i>	GTCTTTTCGT	GTGGTCTATT	TTGGTGACAG	TTTTTCTGTT	GTGTTGTCT	TTACCAGTGT	
<i>Achatina bisculpta</i>	GTTTATTTCGT	TTGATCTATT	CTTGTAACCTG	TATTTCTTTT	ACTTCTCTCA	TTGCCAGTCC	
<i>Achatina damarensis</i>	GTTTATTTCGT	TTGGTCAATT	CTTGTCACAG	TGTTTCTTTT	ACTTCTTTCT	TTGCCCTGTCT	
<i>Achatina stuhlmanni</i>	GGTTATTTCGT	ATGATCAATT	CTTGTTACTG	TTTTCTTTT	ACTTTTATCT	CTACCAGTCT	
<i>Atopocochlis exarata</i>	GGTTATTTCGT	GTGGTCAATT	TTGGTTACAG	TATTCCTTTT	ACTCTTATCA	TTACCAGTAT	
<i>Limicolaria kambeul</i>	GCTTATTTCGT	TTGGTCTATT	CTTGTTACAG	TATTTTACT	TCTTCTCTCT	CTCCCTGTTT	
<i>Limicolaria martenssii</i>	GTCTGTTTCGT	ATGATCTATC	TTAGTAACCG	TCTTTTATT	ACTCCTATCT	CTTCCAGTAC	
<i>Limicolariopsis sp.</i>	GGTTATTTCGT	ATGATCTATT	TTAATTACTG	TATTTCTTTT	ACTCTTATCT	CTACCAGTCC	
<i>Cochlitoma varicosa</i>	GCTTATTTCGT	GTGGTCAATC	CTTGTCACCTG	TATTTTACT	TCTTTTATCT	CTACCTGTTT	
<i>Cochlitoma dimidiata</i>	GCCTATTTCGT	GTGGTCTATC	TTAGTTACCG	TGTTTCTTCT	ACTCCTCTCC	CTTCCCTGTCT	
<i>Cochlitoma sp. cf. vestita</i>	GGTTATTTCGT	GTGGTCAATT	TTAGTAACCTG	TATTTTACT	ACTTCTCTCT	CTCCCAGTTC	
<i>Cochlitoma marinae</i>	GGTTGTTTCGT	GTGATCAATC	CTAGTAACCG	TGTTTCTTACT	ATTATTATCC	TTACCAGTTT	
<i>Cochlitoma churchilliana</i>	GTTTGTTTCGT	ATGGTCTGTT	CTAGTGACAG	TATTCCTTACT	ACTTTTGTCT	TTACCTGTAT	
<i>Cochlitoma granulata</i>	GTTTATTTCGT	TTGGTCTATT	TTGATTACTG	TTTTTTTACT	CCTTCTCTCT	CTTCCCTGTTT	
<i>Cochlitoma simplex</i>	GCCTCTTTCGT	GTGGTCAATC	TTAGTCACCG	TGTTTCTTACT	ACTTCTCTCC	CTCCCTGTCT	
<i>Cochlitoma kilburni</i>	GGTTATTTCGT	ATGGTCAATT	TTAGTAACCTG	TATTTTACT	ACTTCTCTCT	CTACCTGTTT	
<i>Cochlitoma montistempli</i>	GCTTGTTTCGT	ATGGTCAATC	TTGGTAACAG	TATTTTACT	TCTTCTCTCC	CTTCCGGTTT	
<i>Cochlitoma omissa</i>	ATTTATTTCGT	ATGGTCTATT	TTAGTTACTG	TATTTTACT	CCTTCTCTCT	CTTCCCTGTTT	
<i>Cochlitoma semidecussata</i>	GCTTATTTCGT	GTGGTCCATC	TTGGTAACAG	TTTTTTTACT	TCTTCTCTCC	CTTCCCTGTTT	
<i>Cochlitoma ustulata</i>	GATTATTTCGT	ATGGTCTATT	TTAGTTACTG	TTTTCTTACT	GCTCTTATCT	CTACCAGTAC	
<i>Cochlitoma zebra</i>	GMCTATTTCGT	MTGRTCCATT	YTAGTAACAG	TTTTCTTACT	SCTCTTATCA	TTACCCGTCT	
<i>Metachatina kraussi</i>	GATTATTTCGT	TTGATCTATT	TTAGTGACCG	TGTTCTTATT	ACTTCTTTCT	TTACCAGTTT	
<i>Rumina decollata</i>	GCCTATTTCGT	GTGATCTATT	TTAGTAACCG	TTTTTCTATT	GCTTTTATCC	CTACCAGTAC	

	550	560	570	580	590	600	
<i>Achatina fulica</i>	TAGCGGGGGC	AATTACTATA	CTATTAACCG	ATCGTAACCT	TAACACATCT	TTTTTTGACC	
<i>Achatina immaculata</i>	TTGCCGGGAGC	TATTACTATA	CTGCTAACAG	ACCGTAATTT	TAATACCTCA	TTTTTTGACC	
<i>Achatina reticulata</i>	TGGCTGGGGC	TATTACTATA	TTATTAACAG	ACCGTAATTT	TAATACATCT	TTTTTTGATC	
<i>Achatina zanzibarica</i>	TAGCAGGGGC	AATTACTATA	TTATTAACAG	ACCGAAACTT	TAACACATCA	TTTTTTGACC	
<i>Achatina achatina</i>	TAGCGGGTGC	AATTACTATA	CTTCTGACCG	ATCGTAACCT	TAATACATCT	TTTTTTGATC	
<i>Archachatina marginata</i>	TAGCTGGTGC	TATTACTATA	CTTTAACTG	ATCGAAATTT	TAATACATCT	TTTTTTGACC	
<i>Achatina bisculpta</i>	TAGCTGGGGC	AATTACAATA	TTATTAACAG	ATCGAAATTT	TAATACATCA	TTTTTTGATC	
<i>Achatina damarensis</i>	TAGCTGGAGC	AATTACAATG	TTATTAACCTG	ATCGAAATTT	TAATACATCG	TTTTTTGACC	
<i>Achatina stuhlmanni</i>	TAGCTGGGGC	GATTACTATA	CTATTAACCTG	ATCGAAATTT	TAATACATCT	TTTTTTGATC	
<i>Atopocochlis exarata</i>	TAGCAGGGGC	AATTACTATA	CTATTAACAG	ATCGAAATTT	TAATACTTCA	TTTTTTGACC	
<i>Limicolaria kambeul</i>	TAGCTGGAGC	TATTACTATA	TTATTAACCTG	ATCGTAATTT	TAATACATCT	TTTTTTGACC	
<i>Limicolaria martenssii</i>	TGGCAGGGGC	TATTACTATA	TTATTAACAG	ATCGTAATTT	TAATACCTCT	TTTTTTGACC	
<i>Limicolariopsis sp.</i>	TTGCAGGTGC	AATCACTATA	CTATTAACCTG	ATCGTAACCT	TAATACGTCA	TTTTTTGATC	
<i>Cochlitoma varicosa</i>	TAGCAGGGGC	TATCACTATA	CTATTAACAG	ATCGAAATTT	TAACACTTCT	TTTTTTGATC	
<i>Cochlitoma dimidiata</i>	TAGCGGGGGC	TATCACTATA	CTACTTACCG	ACCGAAATTT	TAACACTTCA	TTTTTTGACC	
<i>Cochlitoma sp. cf. vestita</i>	TAGCAGGGGC	TATTACTATG	CTTTTAACTG	ATCGTAATTT	TAATACTTCC	TTTTTTGACC	
<i>Cochlitoma marinae</i>	TAGCAGGTGC	AATCACAAATG	CTCTTGACTG	ATCGGAATTT	TAATACTTCC	TTTTTTGATC	
<i>Cochlitoma churchilliana</i>	TGGCAGGGGC	GATTACTATA	CTGCTTACTG	ACCGGAATTT	TAATACCTCT	TTTTTCGACC	
<i>Cochlitoma granulata</i>	TAGCTGGGGC	TATTACTATA	CTTTTAACTG	ATCGCAATTT	TAACACTTCC	TTCTTTGACC	
<i>Cochlitoma simplex</i>	TGGCCGGGGC	TATTACTATG	CTTCTAACAG	ACCGAAATTT	TAATACTTCA	TTCTTTGACC	
<i>Cochlitoma kilburni</i>	TAGCAGGGGC	TATTACCATA	CTTCTGACAG	ACCGTAATTT	TAATACTTCT	TTTTTTGATC	
<i>Cochlitoma montistempli</i>	TGGCAGGAGC	TATTACTATG	CTTTTAACTG	ATCGAAATTT	TAACACTTCC	TTCTTTGATC	
<i>Cochlitoma omissa</i>	TAGCAGGGGC	TATTACTATA	CTTTTAACTG	ACCGAAATTT	TAACACTTCC	TTCTTCGACC	
<i>Cochlitoma semidecussata</i>	TAGCAGGGGC	TATTACTATA	CTTTTAACTG	ATCGGAATTT	CAACACTTCA	TTTTTTGATC	
<i>Cochlitoma ustulata</i>	TAGCAGGGGC	TATTACAATA	CTCCTTACTG	ATCGTAATTT	TAACACATCC	TTTTTCGACC	
<i>Cochlitoma zebra</i>	TAGCGGGGGC	TATTACCATA	TTATTAACAG	ATCGAAATTT	TAATACATCT	TTTTTTGATC	
<i>Metachatina kraussi</i>	TAGCTGGAGC	TATTACTATA	CTTCTTACTG	ATCGAAATTT	TAATACTTCT	TTTTTTGACC	
<i>Rumina decollata</i>	TAGCAGGAGC	TATTACAATA	CTACTAACCTG	ACCGTAACCT	TAACACTTCA	TTCTTTGATC	

	610	620	630	640	
<i>Achatina fulica</i>	CAGCTGGAGG	GGGGGACCCA	GTCTGTACC	AGCACCTATT	T
<i>Achatina immaculata</i>	CAGCAGGGGG	TGGTGATCCA	ATTTTATACC	AACATTTATT	T
<i>Achatina reticulata</i>	CTGCTGGAGG	TGGGGATCCG	GTCTTATATC	AACACTTGTT	T
<i>Achatina zanzibarica</i>	CAGCGGGAGG	GGGAGACCCA	ATTTTGTATC	AGCACTTGTT	T
<i>Achatina achatina</i>	CAGCTGGAGG	TGGTGATCCA	GTGCTTTACC	AGCACCTATT	C
<i>Archachatina marginata</i>	CGGCAGGGAGG	TGGGGACCCCT	ATTCTTTATC	AGCATTTGTT	T
<i>Achatina bisculpta</i>	CTGCTGGAGG	GGGTGATCCT	GTCTTGATC	AACACTTGTT	T
<i>Achatina damarensis</i>	CTGCCGGTGG	TGGTGACCCG	ATTTTATATC	AACATTTATT	T
<i>Achatina stuhlmanni</i>	CGGCAGGGGG	TGGTGATCCA	ATCCTATACC	AGCAYCTATT	C
<i>Atopocochlis exarata</i>	CGGCAGGGGG	TGGTGACCCCT	GTGCTATATC	AACATTTGTT	T
<i>Limicolaria kambeul</i>	CTGCAGGCGG	CGGGGATCCT	ATTTTGTATC	AGCACTTATT	C
<i>Limicolaria martenssii</i>	CAGCTGGAGG	GGGGGATCCT	ATTCTTTATC	AGCATTTATT	C
<i>Limicolariopsis</i> sp.	CTGCAGGAGG	AGGGGATCCA	GTCTTATATC	AACATTTATT	T
<i>Cochlitoma varicosa</i>	CGGCCGGGGG	GGGTGACCCCT	ATTCTATATC	AGCATCTATT	C
<i>Cochlitoma dimidiata</i>	CAGCTGGAGG	TGGGGACCCCT	ATCTTGATACC	AGCACCTCTT	C
<i>Cochlitoma</i> sp. cf. <i>vestita</i>	CGGCTGGGGG	AGGCGATCCT	ATTTTGTATC	AACATCTCTT	T
<i>Cochlitoma marinae</i>	CTGCAGGAGG	TGGGGACCCCT	ATTTTATACC	AGCATTTATT	T
<i>Cochlitoma churchilliana</i>	CTGCAGGAGG	GGGAGACCCG	ATTTTGTACC	AACACCTTTT	C
<i>Cochlitoma granulata</i>	CAGCTGGTGG	TGGGGACCCCT	ATTTTATATC	AGCACCTTTT	T
<i>Cochlitoma simplex</i>	CAGCTGGAGG	TGGAGACCCCT	ATTCTATACC	AGCACCTCTT	T
<i>Cochlitoma kilburni</i>	CAGCTGGTGG	TGGTGACCCCT	ATTTTATACC	AACATCTCTT	T
<i>Cochlitoma montistempli</i>	CGGCTGGTGG	AGGTGATCCC	ATTTTATATC	AACATCTTTT	T
<i>Cochlitoma omissa</i>	CAGCTGGTGG	TGGGGACCCCT	ATTTTATATC	AGCACCTTTT	T
<i>Cochlitoma semidecussata</i>	CTGCTGGGGG	TGGTGATCCT	ATTTTATATC	AACACCTTTT	C
<i>Cochlitoma ustulata</i>	CGGCAGGGGG	AGGAGATCCT	ATCTTGATC	AGCATCTCTT	C
<i>Cochlitoma zebra</i>	CGGCTGGAGG	GGGWGAYCCA	ATCTTATACC	AACATCTATT	T
<i>Metachatina kraussi</i>	CTGCAGGAGG	CGGAGATCCT	ATTTTATATC	AGCATCTTTT	T
<i>Rumina decollata</i>	CTGCAGGGGG	AGGTGATCCA	ATTCTATATC	AGCATTTATT	T

Appendix 4.5: Alignment of the 16S rRNA gene for the Achatinidae and the subulinid outgroup *Rumina decollata*. Positions included for the phylogeny analyses were marked by “m” within “NUCEOTIDES INCLUDED”. Note that sequencing was unsuccessful for *Limicolariopsis ruwenzoriensis*.

	10	20	30	40	50	60
Achatina fulica	TAGCTTAAAG	A-GTGC GTAA	CACAGATT TT	AAGTAACCCC	TGCCCGGTGA	GTTATAGA--
Achatina immaculata	TAGTTTAAAG	AA----ATTA	AAAAGATT TT	AAGTGATCCC	TGCCCGGTGA	GTAAAA----
Achatina reticulata	TAGCTTAAAG	CG----TWCC	TAAAGATT TT	AAGTAATCTC	TGCCCGGKGA	GTTAC-----
Achatina zanzibarica	TAGCTTAAAG	GA-----CC	AAAMCATT TT	AAGTAACCCC	TGCCCGGTGG	GCTGT-----
Euaethiopina loveridgei	CAGCTTGAAG	TT-----TAA	AAAAGATT TT	AAGTGATCCC	TGCCCGGTGA	GATTTGTT--
Achatina achatina	TAGCTTTAGG	A-----	TTTAAATCTT	TAGTACCTCC	TGCCCAGTGA	TAGGTTAC--
Archachatina marginata	TAGCTCTAGG	A-----	CTTAAATCTT	GAGTACACCC	TGCCCAGTGA	CAATTT----
Achatina bisculpta	TRGKTARTAG	T-----	TTATGATTAT	TAGTGGATCC	TGCCCAGTGG	AATTTTAA--
Achatina damarensis	TAGCCAGTAG	C-----TAG	CTAGGATTGC	TGGTGGATCC	TGCCCAGTGG	AAAATTCTC-
Achatina stuhlmanni	TAGCTAATAG	G---CTCTTT	CCATAATTAT	TAGTGGACCC	TGCCCGGTGA	AGTTAA----
Atopocochlis exarata	TAGCTAAAAG	T-----	-AGAAATT TT	TAGTGTGTCC	TGCCCACTGG	GTTCC-----
Limicolaria kambeul	TAGCTAATAG	A-----	-ATAGATTAT	TAGTATTCCC	TGCCCAGTGA	AAATATTCTCA
Limicolaria martenssii	TAGCTAATAG	C-----ATTT	TATATATTGT	TAGTGTGTCC	TGCCCAGTGG	AGACAATA-
Limicolariopsis sp.	TAGYTGAAC	G-----TT	AGARATTTT	CAGTATATTC	TGCCCAGTGA	GATAAT----
Cochlitoma varicosa	TAGCTTAAAG	T-----	AATTATCTT	AAGTGGTCCC	TGCCCGGTGA	AAGTT-----
Cochlitoma dimidiata	TAGCTKGAAG	A-----A	GTTTGATT TT	AAGTGACCCC	TGCCCAGTGG	TAAATC----
Cochlitoma sp. cf. vestit	TAGCTTGAAG	T-----	TTTTCTTTT	AAGTGGTCCC	TGCCCGGTGG	ATTAA-----
Cochlitoma marinae	TAGCTTAAAG	G-----T	ATTAGGT YTT	AAGTAGTCCC	TGCCCGGTGG	TAATAA----
Cochlitoma churchilliana	TAGCTTAAAG	T-----TCTA	AGTCTATTTT	AAGTAGTCCC	TGCCCGGTGA	GTAGAGAAA-
Cochlitoma granulata	TAGCTTGAGA	G-----T	ACTTTATTTT	AAGTGTCCCC	TGCCCAGTGA	ACAATAA----
Cochlitoma simplex	TAGCTTGAAG	C-----AC	ATTTGATT TT	AAGTGACCCC	TGCCCGGTGG	CGCAGTAGCT
Cochlitoma kilburni	TAGCTTGAAG	T-----	TTTTCGTTT	AAGTGATCCC	TGCCCGGTGG	TGTAA-----
Cochlitoma montistempli	TAGCTTGAGA	G-----T	ATTTTATTTT	AAGTGTCCCC	TGCCCAGTGA	TAATTA----
Cochlitoma omissa	TAGCTTGRGA	G-----T	ACTTCATTTT	AAGTGTCCCC	TGCCCAGTGA	TCAATAA----
Cochlitoma semidecussata	TAGCCTAAGA	G-----T	CGTTTATTTT	AGGTGTCCCC	TGCCCACTGA	TAATTA----
Cochlitoma ustulata	TAGCCTGCTG	T-----A	GATTTATTAC	AGGTGATCCC	TGCCCGGTGG	GGCCTATT--
Cochlitoma zebra	TAGCTTTAAG	T-----	AATTCATTTT	AAGTGGTCCC	TGCCCGGTGA	GAATC-----
Metachatina kraussi	TTGCTTGAAG	T-----AGT	ATTTTATTTT	AAGTAATCCC	TGCCCGGTGG	GATAATA----
Rumina decollata	-----	-----	-----TTT	AGGTAACCCC	TGCCCAGCGA	TATAT-----
NUCLEOTIDES INCLUDED	-----	-----	-----mmmm	mmmmmmmm	mmmmmmmm	m-----

	70	80	90	100	110	120
Achatina fulica	---AACTTTA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Achatina immaculata	---TATTTTA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Achatina reticulata	---TACTTTA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Achatina zanzibarica	---GGCTTAA	ACGGCTGCAG	TACCCTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Euaethiopina loveridgei	---TTCTTTA	ACGGCCGCAG	TACCTTGACT	GTGCTAAGGT	AGCATAATAA	TTTGTCTCTT
Achatina achatina	---CTATTTA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Archachatina marginata	---TGACTTA	ACGGCCGCAG	TACCTTGACT	GTGCTAAGGT	AGCATAATAA	TTTGTCTCTT
Achatina bisculpta	---ATTCTTA	ACGGCCGCAG	TACCTTGACT	GTGCTAAGGT	AGCATAATAA	TTTGTCTCTT
Achatina damarensis	---ATTCTTA	ACGGCCGCAG	TACCCTGACT	GTGCTAAGGT	AGCATAATAA	TTTGTCTCTT
Achatina stuhlmanni	---CCTTTTA	ACGGCCGCAG	TACCCTGACT	GTGCTAAGGT	AGCATAATAA	TTTGTCTCTT
Atopocochlis exarata	---CTCCTTA	ACGGCCGCAG	TACCTTGACT	GTGCTAAGGT	AGCATAATAA	CTTGTCTCTT
Limicolaria kambeul	TAGATTTTAA	ACGGCCGCAG	TACCCTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Limicolaria martenssii	---TACCTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATCA	TTTGTCTCTT
Limicolariopsis sp.	---ATCTTTA	ACGGCCGCAG	TACCCTGACT	GTGCTAAGGT	AGCATAATAA	TTTGTCTCTT
Cochlitoma varicosa	---AGTTTAA	ACGGCCGCAG	TACCCTGACT	GTGCAAAGGT	AGCATAATAA	CTTGTCTCTT
Cochlitoma dimidiata	---ACCCTAA	ACGGCCGCAG	TACCTTGACT	GTGCTAAGGT	AGCATAATAA	TTTGTCTCTT
Cochlitoma sp. cf. vestit	---AACCTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Cochlitoma marinae	---ACTTTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	CTTGTCTCTT
Cochlitoma churchilliana	---TACTTCA	ACGGCCGCAG	TACCCTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Cochlitoma granulata	---ACCTTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Cochlitoma simplex	---GCCCTAA	ACGGCCGCAG	TACCTTGACT	GTGCTAAGGT	AGCATAATAA	CTTGTCTCTT
Cochlitoma kilburni	---AACCTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	CTTGTCTCTT
Cochlitoma montistempli	---ACCTTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	CTTGTCTCTT
Cochlitoma omissa	---ACCTTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Cochlitoma semidecussata	---ACCTTTA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	CTTGTCTCTT
Cochlitoma ustulata	---ACCCTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATCA	CTTGTCTCTT
Cochlitoma zebra	---AGTTTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	CTTGTCTCTT
Metachatina kraussi	---AACCTAA	ACGGCCGCAG	TACCTTGACT	GTGCAAAGGT	AGCATAATAA	TTTGTCTCTT
Rumina decollata	---AAGTTAA	ACGGCCGCAG	TACCTTGACT	GTGCTAAGGT	AGCATAATAA	CTTGTCTCTT
NUCLEOTIDES INCLUDED	-----mmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	130	140	150	160	170	180
Achatina fulica	AATTAAGGTC	TGGAATGAAG	GGGGACA--C	AGGGGAG-AG	CTGTCTCCAA	TAAGGTTAAT
Achatina immaculata	AATTGAGGTC	TAGTATGAAT	GGGTTCA--C	AGGGGAA-AG	CTGTCTCAAG	GGAGATTAAT
Achatina reticulata	AATTTAGGTC	TGGAATGAAT	GAGAACAA--C	AGGGGAG-AG	CTGTCTCTAG	WARGGTTAAC
Achatina zanzibarica	AATTGAGGTC	TGGTATGAAT	GGGTTCA--C	GGGGGAA-AC	CTGTCTTTAT	TAAGGTAACC
Euaethiopina loveridgei	AATTGAGGTC	TGGTATGAAT	GGGGGTA--C	GGGCGGG-TG	CTGTCTTTAC	TAAAGGTTAC
Achatina achatina	AATTAGGGTC	TGGTATGAAA	GGATTTT--C	GGGAGTT-TA	CTGTCTTAGT	GAGGTTAATA
Archachatina marginata	AATTGAGGTC	TGGAATGAAT	GGGCTAC--C	GGGGGTT-GG	CTGTCTGCTA	-AAGGATAAT
Achatina bisculpta	AATTAAGGTC	TGTATGAAA	GGGGGAA--C	GGGGGGT-AA	CTGTCTCTAA	GGGGTTTCAG
Achatina damarensis	AATTAAGGTC	TGTATGAAA	GGGAGAA--C	GGGGGAT-AA	CTGTCTCTTA	TGGGGTTTTAA
Achatina stuhlmanni	AATTAAGGTC	TGCAATGAAT	GGGTGTT--C	GAGGAGT-CG	CTGTCTTTAT	TAGGGTTTAA
Atopocochlis exarata	GATTGGGGTC	TAGAATGAAT	GGATGAT--T	GGGGGAA-AA	CTGTTTCTTT	CATGTTAATA
Limicolaria kambeul	AAATGAGGTC	TGTATGAAT	GGGATAA--C	GGGGGGT-AG	CTGTCTTATT	ATAGTTTCTT
Limicolaria martenssii	AAATGAGGTC	TGTATGAAA	GGATTAA--C	GGGGGAG-AG	CTGTCTTATC	TATAAAAAAC
Limicolariopsis sp.	AATTAGGGTC	TAGAATGAAA	GAACGCA--C	GATATAA-AG	CTGTCTCAAA	ATGGGTATT
Cochlitoma varicosa	AATTGGGGTC	TAGTATGAAT	GGGTTAA--C	GGGTATT-AG	CTGTCTCAGG	GGGGTCTTAT
Cochlitoma dimidiata	AATTGAGGTC	TGGAATGAAC	GGGTTAGA-C	GAATGTT-AG	CTGTCTTATG	TTAGGTCCTA
Cochlitoma sp. cf. vestit	AATTGAGGTC	TGGAATGAAT	GGGTAAAA-C	GAGTATT-AG	CTGTCTTATG	TTGGTTAATG
Cochlitoma marinae	AATTGAGGTC	TAGAATGAAT	GGGAAAAG-C	GGGTGAT-AG	CTGTCTTAAG	TCAGTTTTTA
Cochlitoma churchilliana	AATTAAGGTC	TGGTATGAAT	GGGTTAA--C	GGGTGTT-GA	CTGTCTCAGG	GATTGATATT
Cochlitoma granulata	AATTGAGGTC	TGGTATGAAT	GGGTTAGA-C	GGGTATA-AG	CTGTCTTAAT	TTGGTTATCT
Cochlitoma simplex	AATTGAGGTC	TAGAATGAAT	GGGTTAAAA	GGGTGTT-AG	CTGTCTCGCG	TTGGGTTTTT
Cochlitoma kilburni	AATTGAGGTC	TGGAATGAAT	GGGTTTAA-C	GGGTGTT-AG	CTGTCTCATG	CTGGTTAATG
Cochlitoma montistempli	AATTGAGGTC	TGGAATGAAT	GGGTTAGA-C	GGGTATA-AG	CTGTCTTAAT	TTGGTTATCT
Cochlitoma omisa	AATTGAGGTC	TGGAATGAAT	GGGTTAGA-C	GGGTATG-AG	CTGTCTTAAT	TTGGTTATTT
Cochlitoma semidecussata	AATTGAGGTC	TGGAATGAAT	GGGTTAGA-C	GGGTATA-AG	CTGTCTTAAT	TTGGTTATCC
Cochlitoma ustulata	AATTGAGGAC	TGGAATGAAA	GGGAAGAAAC	GGGTATG-AG	CTGTCTCTAG	TTAGCTCTTT
Cochlitoma zebra	AATTGAGGTC	TGGAATGAAT	GGGTTTAA-C	GGGTAAA-AG	CTGTCTTAGG	AAGGCTTTAT
Metachatina kraussi	AATTGAGGTC	TGGAATGAAT	GGGAAAA--C	GGGTATA-AG	CTGTCTCAAG	CTATTTTTGT
Rumina decollata	AATTGAGGTC	TGGAATGAAC	GGGTTTA--C	GAAAAATGTT	CTGTCTCCTG	TACAAATTTT
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	190	200	210	220	230	240
Achatina fulica	T--TAACCTT	C--TTATCAGG	TGAAAATTCC	TGAGCCT-CC	GATGAAAGAC	GAGAAGACCC
Achatina immaculata	C--TAATTTA	C--TTATCAGG	TGAAAATTCC	TGAGGAT-AA	AAGGAAAGAC	GAGAAGACCC
Achatina reticulata	T--TAACCTT	C--TTATCAGG	TGAAAATTCC	TGAGCTT-CT	TTGGAAAGAC	GAGAAGACCC
Achatina zanzibarica	T--AAATTTG	C--TTATCAGG	TGAAAATTCC	TGAATTTATA	AAGGAAAGAC	GAGAAGACCC
Euaethiopina loveridgei	CT-AAACTTA	C--TTATCAGG	TGAAAATTCC	TGAACCT-TT	AAGGAAAGAC	GAGAAGACCC
Achatina achatina	T--GAATTTA	C--TCAATAAG	GTGAAAATTC	CTTAAAAATTA	GATAAAAGAC	GAGAAGACCC
Archachatina marginata	TT-GAATTTG	C--TTATAAGG	TGAAAATTCC	TTTAATTATA	GATAAAAGAC	GAGAAGACCC
Achatina bisculpta	T--GAATTTT	C--TCAGGAGG	TGAAAATTCC	TCTGATATTT	GACAAAAGAC	GAGAAGACCC
Achatina damarensis	T--AAATTTA	C--TTAAGAGG	TGAAAATTCC	TCTAATT-GT	GACAAAAGAC	GAGAAGACCC
Achatina stuhlmanni	TT-AAATTTA	C--TCAAGAGG	TGAAAATTCC	TCTAAATTAA	GATAAAAGAC	GAGAAGACCC
Atopocochlis exarata	T--TAACCTA	C--TTAAAAGG	TGAAAATTCC	TTTAGTT-AA	GAAAAAAGAC	GAGAAGACCC
Limicolaria kambeul	T--GAACCTA	CTAAATTAGG	TGAAAATTCC	TACACTTATA	TACAAAAGAC	GAGAAGACCC
Limicolaria martenssii	---GAAATTA	CTTAATTAGG	TGAAAATTCC	TATAAAT-AA	GAAAAAAGAC	GAGAAGACCC
Limicolariopsis sp.	TT-TAATTTG	C--TAAAGAGG	TGAAAATTCC	TTTAATTTT	TATAAAAGAC	GAGAAGACCC
Cochlitoma varicosa	T--TAATTTA	T--TTAGAAGG	TGAAAATACC	TTTAAATTTA	TATAGTAGAC	GAGAAGACCC
Cochlitoma dimidiata	T--TAACCTA	A--TTAGAAGG	TGAAAATTCC	TTTATGTATT	ATTAGAAGAC	GAGAAGACCC
Cochlitoma sp. cf. vestit	C--TAACCTA	A--TTAGAAGG	TGAAAATGCC	TTCATATTTT	AATAGAAGAC	GAGAAGACCC
Cochlitoma marinae	T--TAATTTA	A--TTAGGGGG	KGAAAATTCC	CCTATTCCTA	AACAGTAGAC	GAGAAGACCC
Cochlitoma churchilliana	ATTGAATTTA	A--TCATAAGG	TGAAAATGCC	TTTACTTATT	AACAGAAGAC	GAGAAGACCC
Cochlitoma granulata	T--AAATTTA	A--TTAAAAGG	TGAAAATTCC	TTTAAATATA	GATAGAAGAC	GAGAAGACCC
Cochlitoma simplex	T--TAACCTA	ATCTAGAAGG	TGAAAATTCC	TTCAAGAATA	GACAGAAGAC	GAGAAGACCC
Cochlitoma kilburni	T--CAACTTA	A--TTAGAAGG	TGAAAATACC	TTTATGTATT	AATAGAAGAC	GAGAAGACCC
Cochlitoma montistempli	T--AAACTTA	A--TTAGAAGG	TGAAAATTCT	TTTAAATATAT	TACAGAAGAC	GAGAAGACCC
Cochlitoma omisa	T--AAATTTA	A--TTAAAAGG	TGAAAATTCC	TTTAAAGTGA	AATAGAAGAC	GAGAAGACCC
Cochlitoma semidecussata	T--GAACCTA	A--TGAAAAGG	TGAAAATTCC	TTTAAAGT-GT	TACAGAAGAC	GAGAAGACCC
Cochlitoma ustulata	T--AAATTTA	A--TTGTTAGG	TGAAAATACC	TATACCT-TT	AACAGAAGAC	GAGAAGACCC
Cochlitoma zebra	T--TAACCTA	A--TTAGAAGG	TGAAAATCCC	TTTAAAGTATA	AAAAGTAGAC	GAGAAGACCC
Metachatina kraussi	T--GAACCTA	CTTTAGAAGG	TGAAAATACC	TTTGTTT-TA	GTTAGAAGAC	GAGAAGACCC
Rumina decollata	ACTGAACCTA	C--TTAAGAGG	TGAAAATTCC	TTTAGTA-AT	GAAAAAAGAC	GAGAAGACCC
NUCLEOTIDES INCLUDED	---mmmmmm	m-mmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	250 260 270 280 290 300
Achatina fulica	TTAGAGTTTT TATTATAACT TGATA----- TATTAAGATT CTGTGTCTGT TTTTGTGGG
Achatina immaculata	TTAGAGCTTT TATGAAAATT TGAGA----- AGTTACTAAT CTAATATCGT TTTTGTGGG
Achatina reticulata	TTAGAGTTTT AATAATTATT TAGAT----- AAATAAAATT CTAATATCGT TTTTGTGGG
Achatina zanzibarica	TTAGAGTTTT CATTAAAGCC TAGGA----- --GTAATAGC ATGGTTATGT TTTTGTGGG
Euaethiopina loveridgei	TTAGAGTTTT TACTACAAC T AGAAA----- -TGTCATATT CTAGTATTGT TTTTGTGGG
Achatina achatina	TTGGAGTTTT TATTATATAG ATAGA----- ----- TTCTATTCTT TTTTCGTTGGG
Archachatina marginata	TTGGAATTTT AAGGAATTG TAAGC----- -----T TTACATAAAT TTTTCGTTGGG
Achatina bisculpta	TTAGAGCTTT AATAAAAAGT AAATC----- ----ACTATT TTACTTAAGT TTTTGTGGG
Achatina damarensis	TTAGAGCTTT TACAAAAAGT AGACAAGTAG TGTACTATT CTACTTAAGT TTTTGTGGG
Achatina stuhlmanni	TTAGAGTTTT AATGAAAAGT AAAC----- -----T TTACTTAAGT TTTTGTGGG
Atopocochlis exarata	TTAGAAAATT GTAGAAAATG CAAAT----- ----TCAAT TTGCTTACTT TTTTGTGGG
Limicolaria kambeul	TATGAGTTTT TATGAATAAA TGAAA----T CATATTTTAT CATACTAAGT TTTTGTGGG
Limicolaria martenssii	TAAGAGTTTT AATGAGAAATA CTATT----- AGGTAATTAA ATAGAGAAGT TTTTGTGGG
Limicolariaopsis sp.	TAAGAGCTTT TAATAATCTA AATAT----- -AAAAATTTA TATTTAAAGT TTTTGTGGG
Cochlitoma varicosa	TTAGAGTTTC CATGAAAAGAG TATAA----- --TCTTTAT ATATCTAAGT TTTTCGTTGGG
Cochlitoma dimidiata	TTAGAGTTTT AATGAAACAA TATAT----- TTTACAATTG TATTTTATAGT TTTTCGTTGGG
Cochlitoma sp. cf. vestit	TTAGAGTTTA AAATGAATAA ATATT----A ATCTTACGAT TATTTTATAGT TTTTCGTTGGG
Cochlitoma marinae	TTAGAGTTTT TATGAAAAGTA GTCTA--AGG TTATCTAGTA TACTACTAGT TTTTCGTTGGG
Cochlitoma churchilliana	TTAGAGTTTT TATGAAGAAG TTATA--TAT ATATCAAATA AGCTTTAAGT TTTTCGTTGGG
Cochlitoma granulata	TTAGAGTTTT TATAAATAAA TATAG---AA CAGATGAGTT ATATTTTATAGT TTTTGTGGG
Cochlitoma simplex	TTAGAGTTTT AATGAAATAA TATAG----- TTTAAAGTAG TGTTTTTATAGT TTTTCGTTGGG
Cochlitoma kilburni	TTAGAGTTTT AAATGAATAA ATACT---A ATATTATAAT TATTTTATAGT TTTTCGTTGGG
Cochlitoma montistempli	TTAGAGTTTT TATAAATAAA TATAG---A ACAATAAGTT GTATTTTATAGT TTTTGTGGG
Cochlitoma omissa	TTAGAGTTTT TATAAACAAA TGTAG---A ACAATGAGTT ATATTTTATAGT TTTTGTGGG
Cochlitoma semidecussata	TTAGAGTTTT TATAAATAAA TGTAG---A ACAATGCGTT TGATTTTATAGT TTTTCGTTGGG
Cochlitoma ustulata	TTAGAGTTTT TATGATAAGC ATTCT----- ----ACATA TGCTGTCAGT TTTTCGTTGGG
Cochlitoma zebra	TTAGAGTTTT TATAAGAATA TATAA----- --CTTTTTT ATATTTTATAGT TTTTCGTTGGG
Metachatina kraussi	TTAGAGTTTT TATAAAAATA TATGG---TA ATTATTATAA TATATTGAGT TTTTCGTTGGG
Rumina decollata	TTAGAATTTT AATAAAAAGT GTCAT----- --TGCGTG ATTGTTTATAGT TTTTCGTTGGG
NUCLEOTIDES INCLUDED	mmmmmmmmmm m-----m -----m -----mmmmmmmmmm

	310 320 330 340 350 360
Achatina fulica	GCGACAGGGT TACA----- -----A TAGATAAAGT ACCCTACCAT A--TTTTTAT
Achatina immaculata	GCGACAGGGT AGCA----- ----- TATTTAAAGT ACCTGATCTT T--AAAGTAT
Achatina reticulata	GCGACAGGGT AACA----- -----AT AGGTAAAGT ACCCATTCAT ATATTTTAT
Achatina zanzibarica	GCGACGGGGT TACA----- ----- GTTTGAAGT ACCTAGAATA C-ITCCTGTT
Euaethiopina loveridgei	GCGACAGGGT GACA----- -----A TAATTAAGT ACCAACACAA TATTATATAT
Achatina achatina	GCAGCGAGAT TACA----- ----- -TCTTTA CTTTTAATTA ATCTTTACTT
Archachatina marginata	GCGGCGAGAC TACA----- ----AGGTT TCATTAACTA GTCTTACATT ATAATTGTT
Achatina bisculpta	GCAACAAGAT GGCA----- -AATGAAAAT AAACCCATCT AATATTCTTA TTTTATTTGA
Achatina damarensis	GCAACAAGAT GGCA----- ----AAGTT TAACCCATCC A-TATTCTAT GTTAATTGTA
Achatina stuhlmanni	GCAACAAGAT GGCAAAAATT AGTAATACTT AAACCCATCT AATTTTCTAA AGTGAATCAG
Atopocochlis exarata	GCAACAGAAT GGCA----- ----AATATA TAACCCATTT AAAACTTAAG CTTAAATCTA
Limicolaria kambeul	GCAACAAGAT ATCA----- ----- AGATTAAATAT ATCTTTATTA TAGTAAATAA
Limicolaria martenssii	GCAACAGAAT TCCA----- ----- GATTAAATGA ATTTATCTAT ATTATATTAT
Limicolariaopsis sp.	GCAACAAGAT ATCA----- ----AA AAGTTAAAGT ATCTGTTTTT CTGGAATACA
Cochlitoma varicosa	GCAACGGGGT CGTA----- ----- AAGTTAACC GCTGTATAG ATGGTAAATG
Cochlitoma dimidiata	GCAACGGAAT CGCA----- -----G TGTTTAACCG GTTTAA--AA GTAATACACA
Cochlitoma sp. cf. vestit	GCAACGGAGT CGCA----- -----A TATTTAACCG GCTAACGAAA TAAACTAAGA
Cochlitoma marinae	GCAACGGATC TGCA----- ----AT TAGTTAAACA GATTAAATTT TTTAGGGTTT
Cochlitoma churchilliana	GCAACGGGGT AGCA----- ----- ATATTAAAGT ACTTGTTAGG GAAGTAATAA
Cochlitoma granulata	GCAACAGGTT CGCA----- -----A GCATAAACCG AATTATTTAG TAACTAACAT
Cochlitoma simplex	GCAACGGAAC CGCA----- ----GA GCTTGAACCG GTTTAATAAA TAATGCATAT
Cochlitoma kilburni	GCAACGGAAT CGCA----- ----TACTTAAGT GTTAACGAAA TAAACTAATA
Cochlitoma montistempli	GCAACGGAAT CGCA----- ----A ATACTAACCG ATTTATCAAA TAATTAATGA
Cochlitoma omissa	GCAACAGTTT CGCA----- ----A GCATAAACCG AATTATTTAG TAACTAACAT
Cochlitoma semidecussata	GCAACAGAAT CGCA----- ----A ATATTAAAGT ATCTAT-TAA TACCTAGTGG
Cochlitoma ustulata	GCAACGAGAT TGCA----- ----TG AAATAAACCA ATCAT--ACG TTAATAGTAG
Cochlitoma zebra	GCAACGGGGT CGAA----- ---- AAATTAACCG ATCAAAAAAT TTTTAAATAA
Metachatina kraussi	GCAACGAGAT TTCA---TTA TTTTAAATATA TCTGTATAAA ATTTTATC--
Rumina decollata	GCGACAGGGT AGCA----- ----AT CAAATAACCT ACCATTAAAT ATTAAATACA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmm-----m -----m -----m-----m

	370 380 390 400 410 420
Achatina fulica	ATGGCGATCA TTATACTTT- ----- ---TAATTAA GCTACCTAAG GGATAACAGC
Achatina immaculata	ATACCGATGG TAATATTAA- ----- ---CACGAAA GCTACCTAAG GGATAACAGC
Achatina reticulata	CGGTGCGATAG TTATACTAA- ----- ---TAATTAA ACTACCTAAG GGATAACAGC
Achatina zanzibarica	TTGACGACTT TCTTACCTA- ----- ---TAATAAA ACTACCTAAG GGATAACAGC
Euaethiopina loveridgei	CCAGCGATTA TTATACTAG- ----- ---TAAATAA GCTACCTAAG GGATAACAGC
Achatina achatina	ATCTAAACAT TTGCCAAGTA TTTATCTTAA GAAAAACTAA ACTACCCAAG GGATAACAGC
Archachatina marginata	CAAGTTTATT AAGAA----- ---GAATTAA ATTACCCAAG GGATAACAGC
Achatina bisculpta	AGAGG-TTTC TAAGAG---- ----- ---TAGTTAA GCTACCTAAG GGATAACAGC
Achatina damarensis	AGAGTTGTTT TAAGAG---- ----- ---TAGATAA GCTACCTAAG GGATAACAGC
Achatina stuhlmanni	TCAATTATTA TAAGAA----- ---CAGAAAA ACTACCTAAG GGATAACAGC
Atopocochlis exarata	CTAGTTAATT TTAATAAGAA ----- ---TGAAGAA TTTACCTAAG GGATAACAGC
Limicolaria kambeul	ATCGATTATG ATAATGG--- ----- ---AAACTAA ACTACCTTAG GGATAACAGC
Limicolaria martenssii	AAGAAGAAAG GTAATAAGAA ----- ---AAAATAA GCTACCTTAG GGATAACAGC
Limicolariopsis sp.	TACCGACTAG TTAGTATAAA ----- ---TGATCAA GCTACCTTAG GGATAACAGC
Cochlitoma varicosa	TATATGCCAA TTTTATAAG AA----- ---TAATTAA ACTACCTAAG GGATAACAGC
Cochlitoma dimidiata	TATATGACCA TTTTAAAAAG AA----- ---TAAGTAA ACTACCTAAG GGATAACAGC
Cochlitoma sp. cf. vestit	GATTTGCCAA TATTAATAAG AA----- ---TAAGAAA ACTACCTAAG GGATAACAGC
Cochlitoma marinae	TATCTGCCAA TTTATTATAA GGG----- ---TAAATAA ACTACCTAAG GGATAACAGC
Cochlitoma churchilliana	ACA-TGCGTC AATATTTTAT TCGTT----- ---TAACAAA ACTACCTAAG GGATAACAGC
Cochlitoma granulata	ATA-TGACAA TTCCTATAAG AA----- ---TAAGTAA ACTACCTAAG GGATAACAGC
Cochlitoma simplex	ATT-TGTCCA CTTTCAAAAG AA----- ---TAAGTAA ACTACCTAAG GGATAACAGC
Cochlitoma kilburni	CATTTGCCAA TATTAATAAG AA----- ---TAAGAAA ACTACCTAAG GGATAACAGC
Cochlitoma montistempli	ATA-TGACAA TTCGTATGAG AG----- ---TAAATAA ACTACCTAAG GGATAACAGC
Cochlitoma omisa	ATA-TGACAA TTTATATAAG TA----- ---TAAGTAA ACTACCTAAG GGATAACAGC
Cochlitoma semidecussata	ATA-AGACAA TTCGAATAAG AA----- ---TAAGCAA ACTACCTAAG GGATAACAGC
Cochlitoma ustulata	ATA-AGTCAA TCTTAATAGG TA----- ---TAATTAA ACTACCTAAG GGATAACAGC
Cochlitoma zebra	AATTTGATAA TCTTTATAAG AA----- ---TAATTAA ACTACCTAAG GGATAACAGC
Metachatina kraussi	----TGCCAA TTAATTCTATA AA----- ---CAAGTAA GCTACCTAAG GGATAACAGC
Rumina decollata	CCGAAATCTA TTAAACA--- ----- ---TAATTAA ATTACCTAAG GGATAACAGC
NUCLEOTIDES INCLUDED	----- mmmmmmm mmmmmmmmm mmmmmmmmm

	430 440 450 460 470 480
Achatina fulica	GTAATCTTTT T-TTTTGGT TTGCGACCTC GATGTTGGAC TAGGGGCGCTA CTGGTTAGTT
Achatina immaculata	GTAATCTTTG ---TTATGGT TTGCGACCTC GATGTTGGAC TAGGAACCCCT TTGGTTAGTT
Achatina reticulata	GTAATCTTTT T-TTTTGGT TTGCGACCTC GATGTTGGAC TAGGGGCGCTA CTGGTTAGCA
Achatina zanzibarica	GTAATCTAAA ---AGTAGT TTGCGACCTC GATGTTGGAC TAGGGTCCAC TTGGTTAGCC
Euaethiopina loveridgei	GTAATCTTTT ---TTAAGGC TTGCGACCTC GATGTTGGAC TAGGGACCTG TTGGTTAGTT
Achatina achatina	ATAATCTCCT ---AGGGAGA TTGTGACCTC GATGTTGGAC TAGGGCTCTA TTGGTTAACT
Archachatina marginata	ATAATCTCCA ---ATGGAGT TTGTGACCTC GATGTTGGAC TAGGGCCCTT ATGATTAAGT
Achatina bisculpta	GTAATCTAAA ATGTTTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCAA ATGATTAGAT
Achatina damarensis	GTAATCTAAT AG-TTTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCAA ATGATTAGAT
Achatina stuhlmanni	GTAATCTAAT ---ATTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA ATGATGAGAT
Atopocochlis exarata	GTAATCTAGC ---ATTAGGA TTACGACCTC GATGTTGGAC TAGGGACCAA ATGATAAGAT
Limicolaria kambeul	GTAATCTAAA ---TTTAGGT TTGCGACCTC GATGTTGGAC TAGGGCCCTA ATGATTAGTC
Limicolaria martenssii	GTAATCTATT A-GTTAGGT TTGCGACCTC GATGTTGGAC TAGGGACCGA ATGATTAGC
Limicolariopsis sp.	GTAATCTATT ---AATAGGA TTGCGACCTC KATGTTGGAC TAGGGWCCAA ATGATTWAGT
Cochlitoma varicosa	GTAATCTTCT ---TTTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA TTGGTTAKTA
Cochlitoma dimidiata	GTAATCTAYT T-TTTAGGY TTGCGACCTC GATGTTGGAC TAGGGACCTA TTGGTTAGCA
Cochlitoma sp. cf. vestit	GTAATTTATT ---TTTAAGA TTGCGACCTC GATGTTGGAC TAGGGACCTT TTGGTTAGTA
Cochlitoma marinae	GTAATCTTAC T-ATTGGGA TTGCGACCTC GATGTTGGAC TAGGGACCCA TTGGTTAGTA
Cochlitoma churchilliana	GTAATCTCTC ---TTTTGGA TTGCGACCTC GATGTTGGAC TAGGGACTGA ATGGTTAACA
Cochlitoma granulata	GTTATCTACT ---ATTAGGA TTGCGACCTC GATGTTGGAC TAGGGACTTA TCGGTAATCA
Cochlitoma simplex	GTGATCTATT ---CATAGGT TTGCGACCTC GATGTTGGAC TAGGGACCCA TTGGTTAACA
Cochlitoma kilburni	GTAATTTATT ---TTTAAGA TTGCGACCTC GATGTTGGAC TAGGGACCTT TTGGTTAGTA
Cochlitoma montistempli	GTAATCTACT A-TTTGGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA TTGGTTAAGCA
Cochlitoma omisa	GTTATCTACT ---ATTAGGA TTGCGACCTC GATGTTGGAC TAGGGACTTA TTGGTTAACA
Cochlitoma semidecussata	GTAATCTACT ---ATTGGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA TTGGTTAAGTA
Cochlitoma ustulata	GTAATCTTAT ---TTATGGA TTGCGACCTC GATGTTGGAC TAGGGACCCA GTGGTTAGAA
Cochlitoma zebra	GTAATCTTCT ---TTTTGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA ATGGTTAGTA
Metachatina kraussi	GTAATCTTT- ---TGAAGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA TCGGTTAGCA
Rumina decollata	ATAATACTAA ---GTAGTGT TTGTGACCTC GATGTTGGAC TAGGGACTAA TTGGTTAACC
NUCLEOTIDES INCLUDED	mmmmmmmm-----mmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

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      ....|....| ....|....| ....|....| ....|....| ....|..
      490      500      510      520
Achatina fulica      ACCATTA--- --TGGG-GCG GTTCTGTTTCG AACTTTTCCT ACCCTAC
Achatina immaculata  ACCATTT--- --CTGG-ATA GTTCTGTTTCG AACTT--TTT TTCCTAC
Achatina reticulata  ACCATCA--- --ATGG-AGG GTTCTGTTTCG AACTT--AAT CCCCTAC
Achatina zanzibarica ACCATTT--- --GTGG-GTG GTTCTGTTTCG AACTT--ATC TCCCTAC
Euaethiopina loveridgei ACCATCA--- --TGGG-GTG GTTCTGTTTCG AACTT--TTT TCCCTAC
Achatina achatina    ACCAAGA--- --ATGT-AAG GTTCTGTTTCG AACTTT-TTA TCCCTAC
Archachatina marginata ATCAATT--- ---TGG-ACA GTTCTGTTTCG AACTC--TCG CCCCTAC
Achatina bisculpta   ATCAAGA--T AACTGG-ATT GTTCTGTTTCG AACAC--TGG ACCCTAC
Achatina damarensis  ATCAAGT--T TTCTGG-ATT GTTCTGTTTCG AACAC--TTG ACCCTAC
Achatina stuhlmanni  ATCAAGGTGA CTATGG-ATT GTTCTGTTTCG AACAC--TAG ACCCTAC
Atopocochlis exarata ATTAAGA--T TCACGG-ATT GTTCTGTTTCG AACACT-TAG ACCCTAC
Limicolaria kambeul  GTCAGGC--- ---TGG-ATT AAGTTCTGTT CGAACTTCGT ACCCTAC
Limicolaria martenssii ATCAAGT--- ---TGGAGT GTTCTGTTTCG AACTT--AAT ACCCTAC
Limicolariopsis sp. ATCATCT--- ---AGG-AYT GTTCTGTTTCG AACAT--TAA MCCCTAM
Cochlitoma varicosa  ACCAGCT--- ---TGG-CAG GTTCTGTTTCG AACCC--TTA CCCCTAC
Cochlitoma dimidiata ACCAGAT--- ---TGTCCAG GATCTGTKCR MACMM--YKK AMYCTAC
Cochlitoma sp. cf. vestit ACCAGGC--- ---TGG-CGG GTTCTGTTTCG AACCT--TTT CCCCTAC
Cochlitoma marinae  ACCAGTA--- ---TAG-CGA GTTCTGTTTCG AACTAA-TTT CCCCTAC
Cochlitoma churchilliana ACCAGGC--- ---TAG-CAG GTTCTGTTTCG AACTT--TTT ACCCTAC
Cochlitoma granulata ACCAGAA--- ---TAGCCAG GTTCTGTTTCG AACTATTTT CCCCTAC
Cochlitoma simplex  ACCAGGA--- ---TGG-CAG GTTCTGTTTCG AACCC--TCT CCCCTAC
Cochlitoma kilburni  ACCAGGC--- ---TGG-CGG GTTCTGTTTCG AACCT--TTT CCCCTAC
Cochlitoma montistempli ATCTAAA--- ---GGG-CTG GTTCTGTTTCG AACCTC-TTA CCCCTAC
Cochlitoma omissa   ACCAGAA--- ---TAG-CAG GTTCTGTTTCG AACTAT-TTT CCCCTAC
Cochlitoma semidecussata ATCAGGA--- ---GGG-CTG GTTCTGTTTCG AACCT--TTT CCCCTAC
Cochlitoma ustulata  ACCAGCT--- ---AGG-CAA GTTCTGTTTCG AACTTT-CCA CCCCTAC
Cochlitoma zebra     ATCAGCT--- ---TGG-CAG GTTCTGTTTCG AACCC--CTT CCCCTAC
Metachatina kraussi  ACCAGAT--- ---TGG-CGA GTTCTGTTTCG AACTTC-TTA CCCCTAC
Rumina decollata     ACCAGGA--- ---TAG-ATG ATTCTGTTTCG AATCC--TTC ACCCTAC
NUCLEOTIDES INCLUDED mmmmmmm--- ---mm--- mmmmmmmmm mmm----- -mmmmmm

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Appendix 4.6: Log likelihood scores of the different models used for the Achatinidae using (A) 3648 unambiguously aligned nucleotides from the rRNA gene cluster; (B) 873 unambiguously aligned nucleotides of actin; (B.1) 582 unambiguously aligned nucleotides of the 1st and 2nd codon positions of actin; (B.2) 291 unambiguously aligned nucleotides of the 3rd codon position of actin; (C) 328 unambiguously aligned nucleotides of histone 3; (C.1) 218 unambiguously aligned nucleotides of the 1st and 2nd codon positions of histone 3; (C.2) 110 unambiguously aligned nucleotides of the 3rd codon position of histone 3; (D) 641 unambiguously aligned nucleotides of CO1; (D.1) 427 unambiguously aligned nucleotides of the 1st and 2nd codon positions of CO1; (D.2) 214 unambiguously aligned nucleotides of the 3rd codon position of CO1; and (E) 310 unambiguously aligned nucleotides of the 16S rRNA gene. Scores in **bold** belong to optimal models as determined by Likelihood Ratio Test. Note that the K2P and F81 models are non-nested and therefore cannot be compared for significant difference using the LRT.

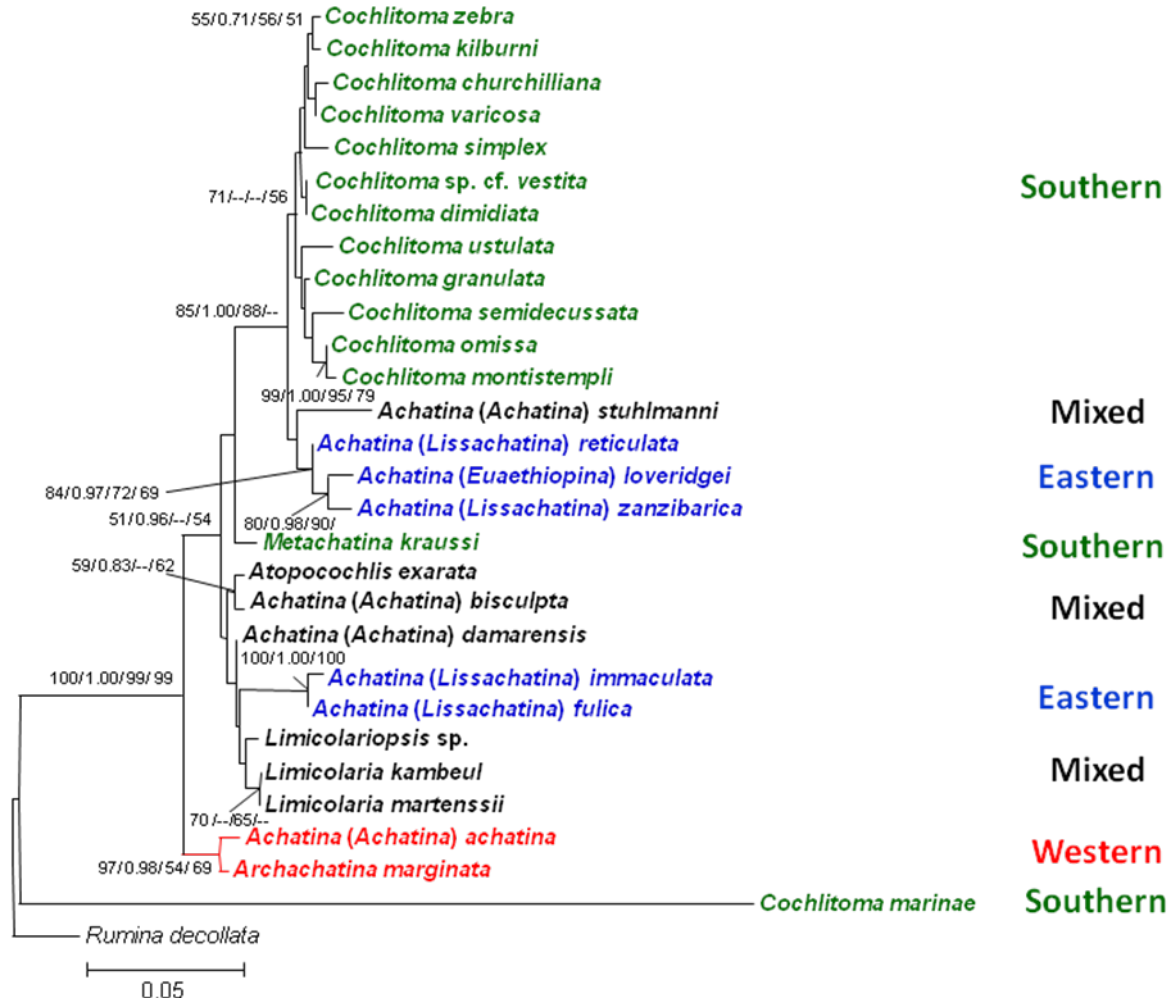
	(A)	(B)	(B.1)	(B.2)	(C)	(C.1)	(C.2)	(D)	(D.1)	(D.2)	(E)
Model	rRNA gene cluster	Actin-combined	Actin-1 st +2 nd codons	Actin-3 rd codons	Histone 3	Histone 3-1 st +2 nd codons	Histone 3-3 rd codons	CO1	CO1-1 st +2 nd codons	CO1-3 rd codons	16S rRNA
JC69	6248.60194	3079.30195	1033.47821	1770.38462	961.35579	330.69952	547.08597	10577.13076	2639.75972	6467.22043	3410.68577
JC69+Γ	6274.56535	2941.42677	1019.80247	1748.03013	944.88406	330.69952	545.05987	9044.70208	2207.91549	6415.07916	3057.86503
F81	6187.62669	3067.34670	1028.44871	1747.01512	951.97785	325.87231	537.40127	10519.45591	2597.94022	6379.86740	3392.31335
F81+Γ	6213.92744	2928.60315	1015.11409	1722.19734	938.63868	325.87231	535.75438	8289.79099	2137.55284	6334.75953	3022.41771
K2P	6217.14469	2999.90791	1029.51509	1690.58148	942.44107	330.63592	520.04898	10305.12794	2494.32907	6291.12771	3332.43652
K2P+Γ	6233.51676	2856.89829	1018.80719	1657.31863	927.23046	330.63592	525.37382	8654.44378	2053.68656	6219.17666	2961.52886
HKY85	6155.43566	2987.60266	1024.45322	1677.30224	932.61470	325.81310	514.06374	10214.37634	2447.65021	6026.06510	3311.76468
HKY85+Γ	6173.49112	2847.31250	1013.84964	1646.32805	909.37626	325.81310	513.02241	8294.44610	2020.38852	5887.87774	2902.18513
TrN93	6146.48364	2976.74300	1019.97904	1677.26346	932.53700	324.88259	513.02953	10210.89277	2422.04372	6023.81427	3308.08745
TrN93+Γ	6164.98831	2837.76616	1013.43024	1644.81901	909.25171	324.88259	512.04887	8289.71414	2015.28913	5879.85256	2900.05462
GTR	6143.87015	2967.76023	1018.79267	1663.08220	931.83493	321.54407	512.77514	9954.78203	2405.51236	6018.45857	3243.11777
GTR+Γ	6157.24483	2821.80122	1011.67014	1634.16756	908.90524	321.54420	511.78485	8289.46582	1995.68125	5878.05759	2876.32253

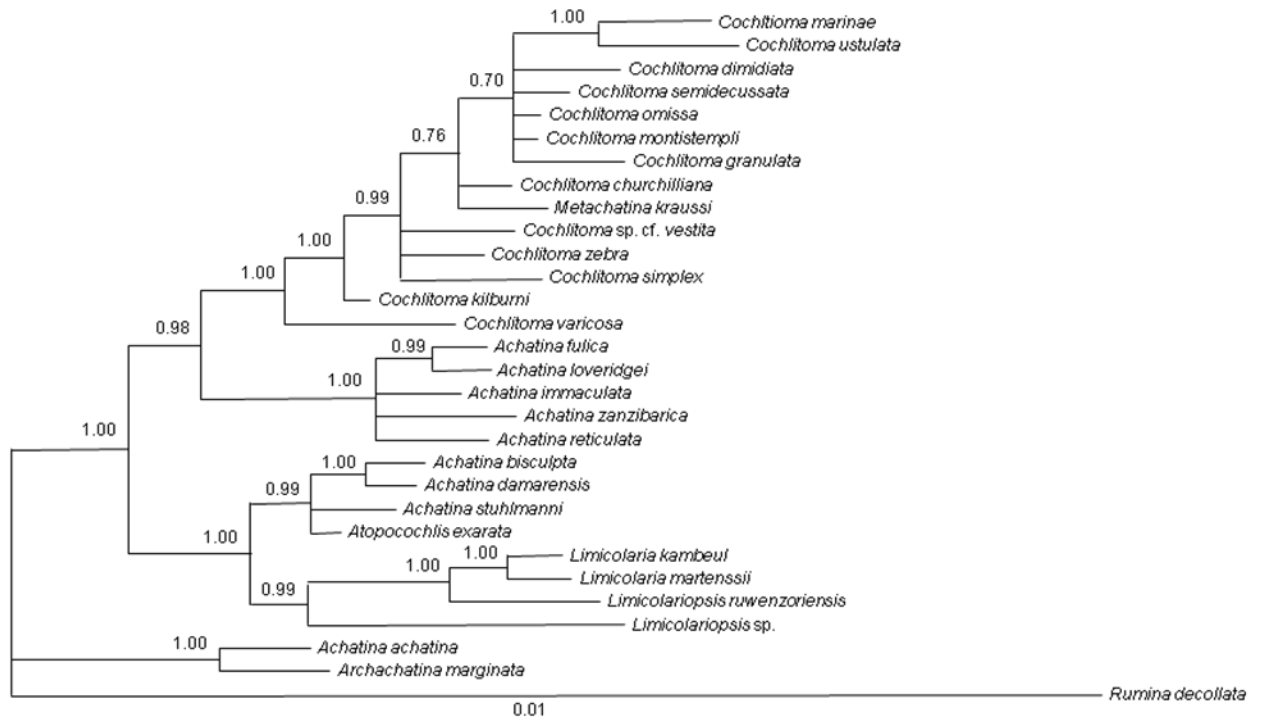
Appendix 4.7: Summary of ambiguous sites for the actin gene for the Achatinidae

Species	Total ambiguous sites	% ambiguous sites (out of 873 total)	Total ambiguous 3 rd codon positions	% ambiguous 3 rd codon positions relative to all ambiguous sites	Number of RY sites at ambiguous 3 rd codon positions	% RY sites relative to all ambiguous 3 rd codon positions	Number of ambiguous amino acids	% ambiguous amino acids (out of 291 total)
WEST AFRICAN								
acha	45	5.15	40	88.89	31	77.50	4	1.37
arcm	27	3.09	23	85.19	21	91.30	3	1.03
EAST AFRICAN								
achf	18	2.06	16	88.89	12	75.00	2	0.69
achi	15	1.72	12	80.00	11	91.67	1	0.34
achr	33	3.78	26	78.79	23	88.46	4	1.37
achz	1	0.11	0	0	0	0	1	0.34
euel	0	0	0	0	0	0	0	0
OTHERS								
achb	32	3.67	26	81.25	17	65.38	4	1.37
achd	34	3.89	27	79.41	25	92.59	6	2.06
achs	2	0.23	1	50.00	1	100.00	1	0.34
atcx2	29	3.32	25	86.21	21	84.00	3	1.03
lim	30	3.44	27	90.00	25	92.59	2	0.69
limm	33	3.78	27	81.82	23	85.19	7	2.41
lip	40	4.58	32	80.00	27	84.38	7	2.41
SOUTHERN AFRICAN								
achv	27	3.09	24	88.89	18	75.00	2	0.69
arcd	31	3.55	27	87.10	20	74.07	3	1.03
arcv	30	3.32	26	86.67	19	73.08	3	1.03
coia	4	0.46	2	50.00	1	50.00	2	0.69
coic	0	0	0	0	0	0	0	0
coik	1	0.11	0	0	0	0	0	0
coig	9	1.03	9	100.00	8	88.89	0	0
coii	2	0.23	2	100.00	0	0	0	0
coim	0	0	0	0	0	0	0	0
coio	6	0.69	6	100.00	5	83.33	0	0
cois	0	0	0	0	0	0	0	0
coiu	0	0	0	0	0	0	0	0
coiz	0	0	0	0	0	0	0	0
metk	52	5.96	38	73.08	29	76.32	9	3.09

NOTE: primers used: ActF1 and ActR

Appendix 4.8. Maximum likelihood phylogenetic tree of the Achatinidae based on 873 unambiguously aligned nucleotide sites of the actin gene. The phylogenies were rooted on the subulinid *Rumina decollata*. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap supports for NJ (1000 replicates) and MP (69 replicates only after the PAUP* version 4.0b10 was terminated after running for 135 hours). The scale bar represents 5 substitutional changes per 100 nucleotide positions.

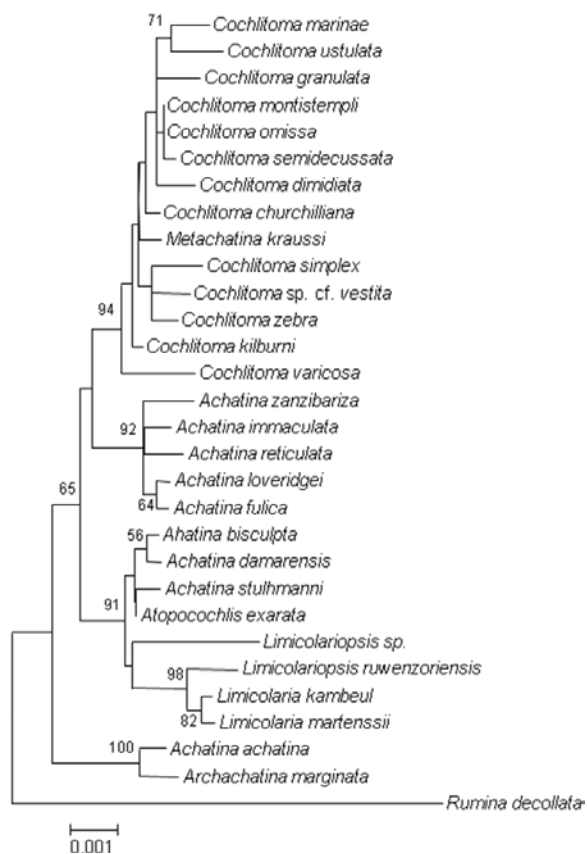




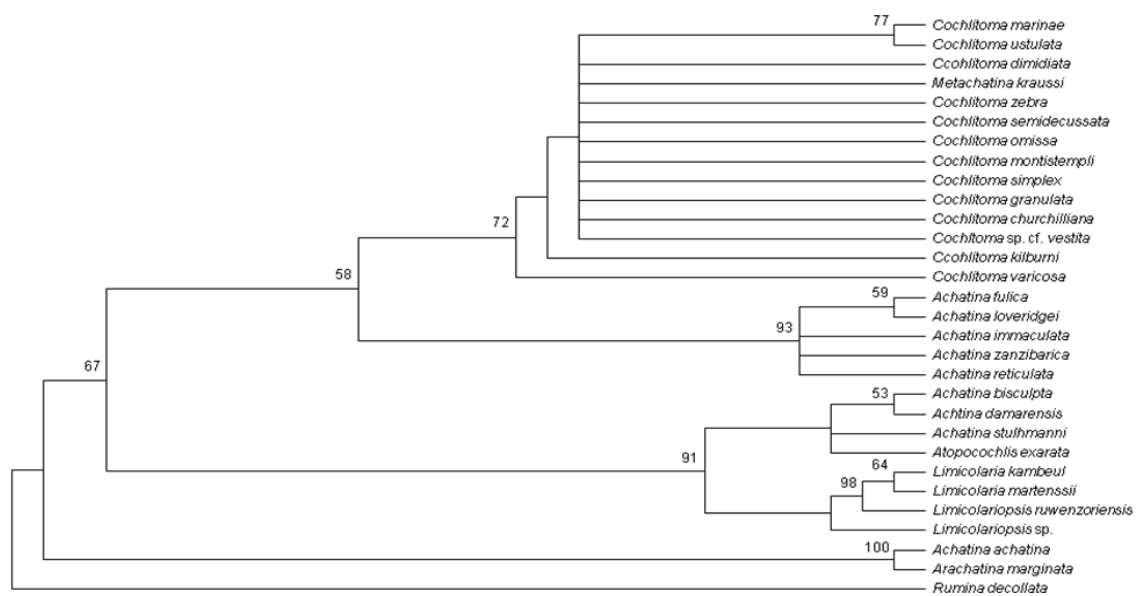
(A) BI (LSU rRNA)

Appendix 4.9: Phylogenetic trees of the Achatinidae based on the large subunit rRNA gene using (A) Bayesian analysis, (B) neighbor-joining (p. 512) and (C) maximum parsimony based on 25 equally parsimonious trees (p. 512). The phylogenies were constructed from 3648 unambiguously aligned nucleotide sites and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 100 nucleotides and that of (B) represents 1 substitutional change per 1000 nucleotides.

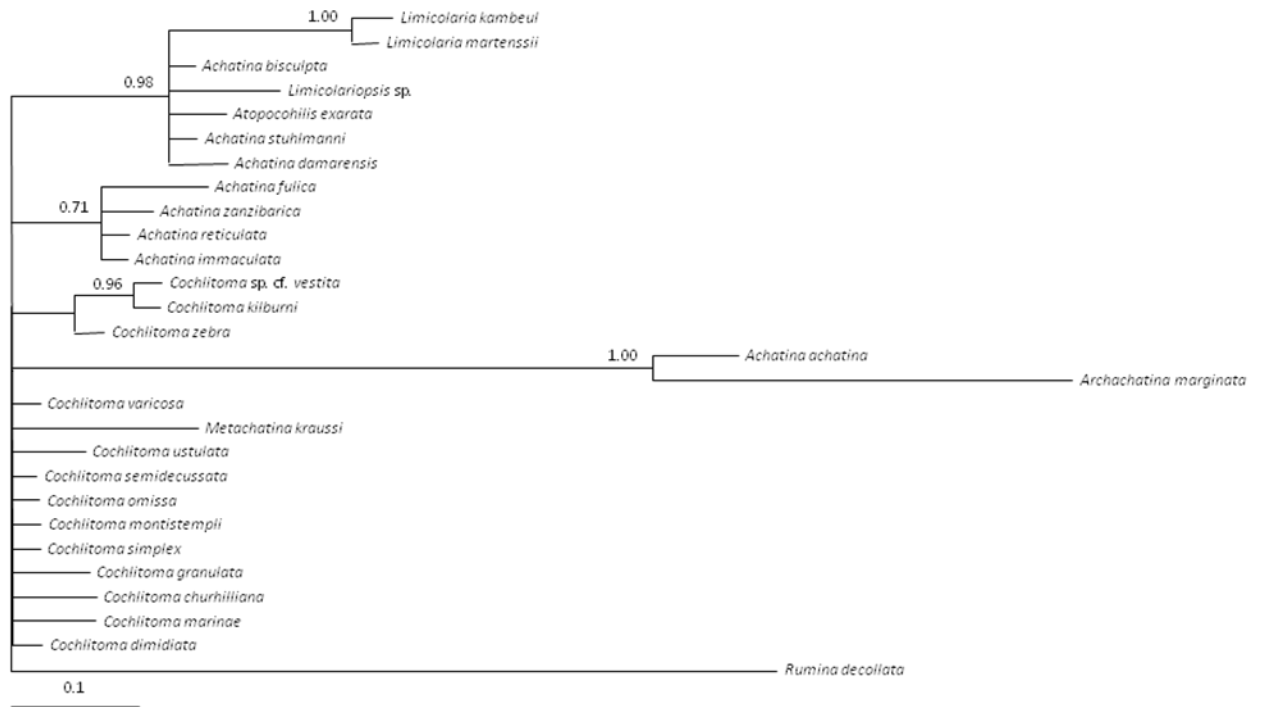
Appendix 4.9 (contd.)



(B) NJ (LSU rRNA)



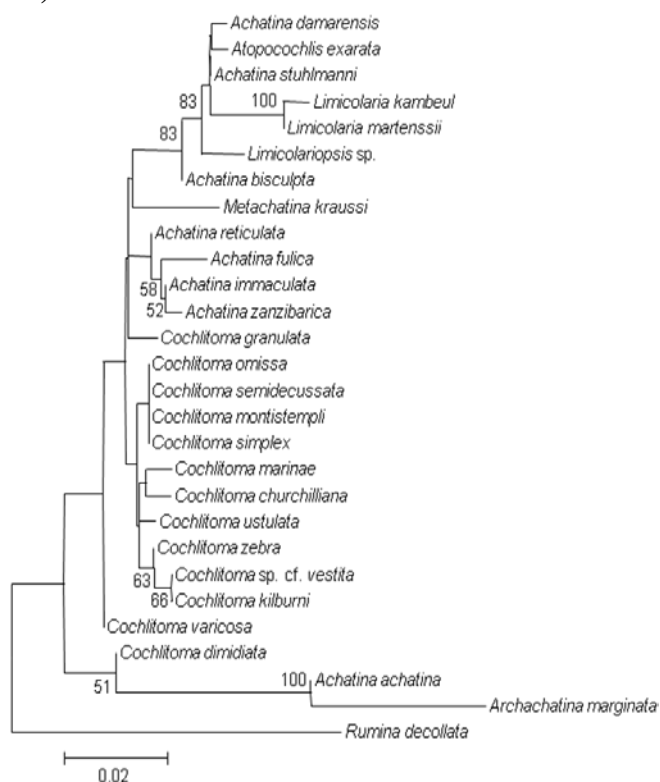
(C) MP (LSU rRNA)



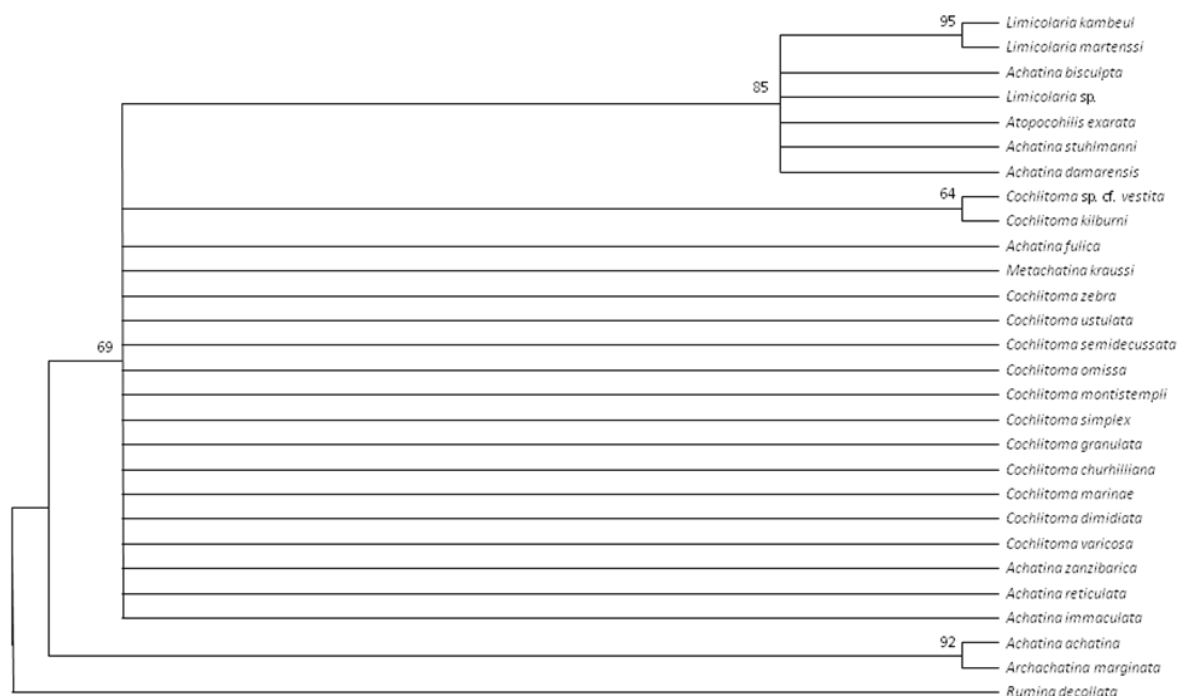
(A) BI (histone 3)

Appendix 4.10: Phylogenetic trees of the Achatinoidea based on the histone 3 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 514) and (C) maximum parsimony based on 8404 equally parsimonious trees (p. 514). The phylogenies were constructed from 328 unambiguously aligned nucleotide sites and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 10 nucleotides while that of (B) represents 2 substitutional changes per 100 nucleotides. Note that *Achatina loveridgei* and *Limicolaria ruwenzoriensis* were not included in the phylogenies as they were not successfully sequenced.

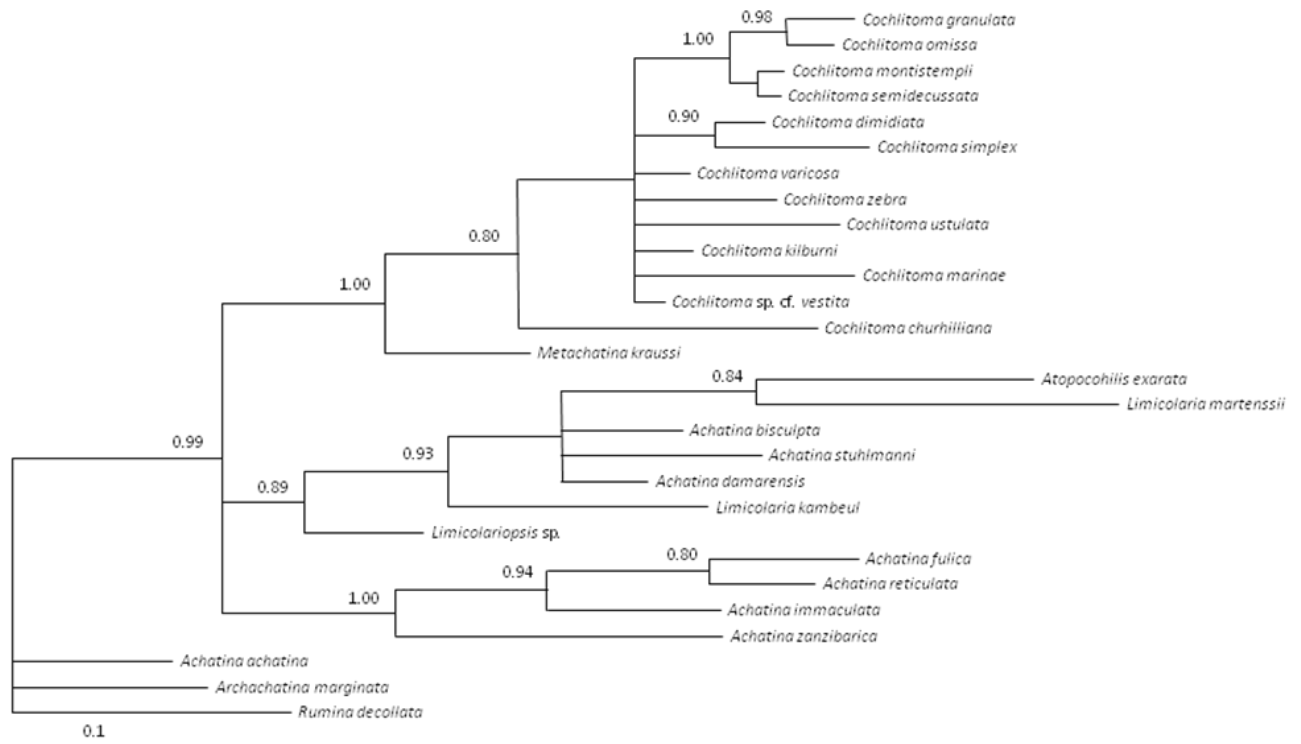
Appendix 4.10 (contd.)



(B) NJ (histone 3)



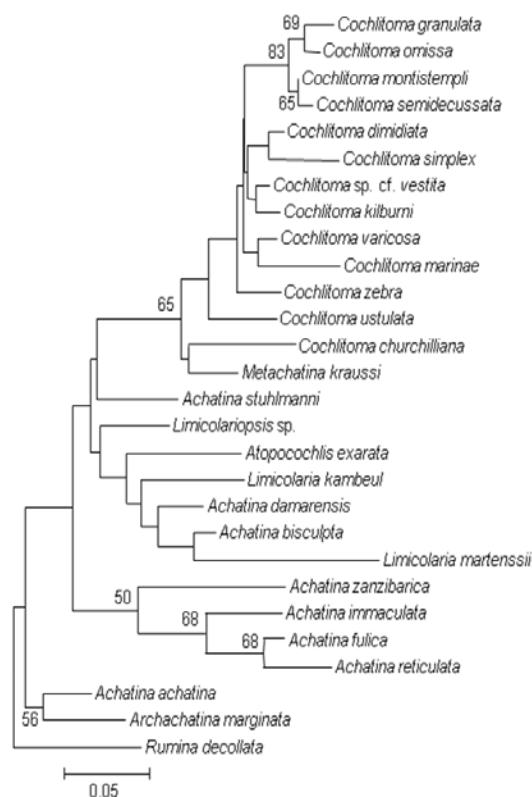
(C) MP (histone 3)



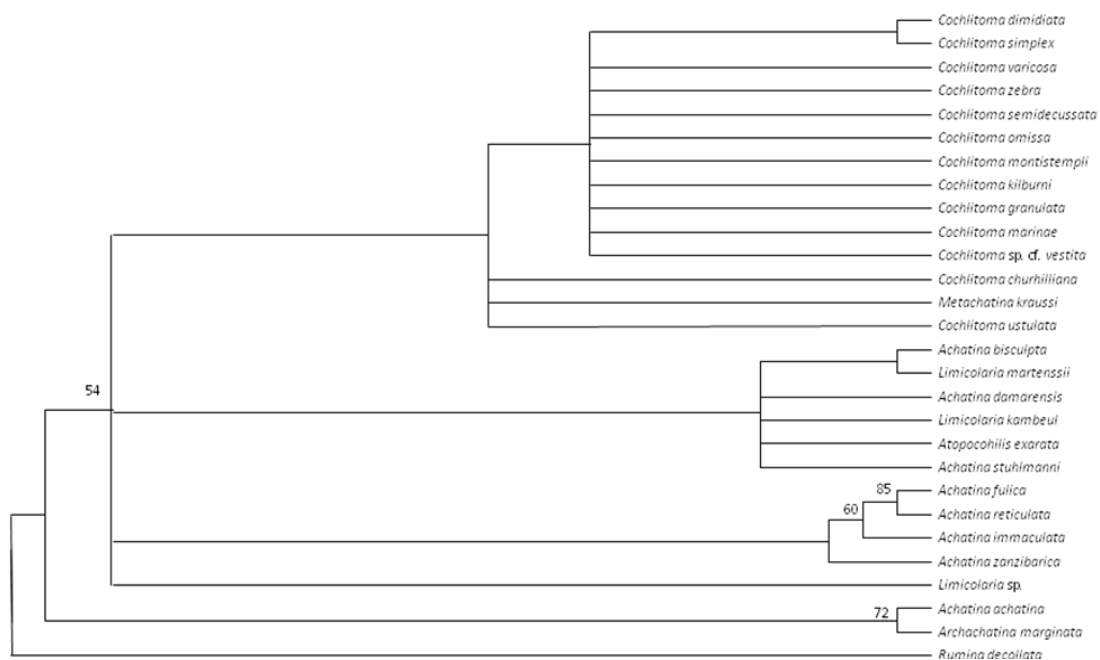
(A) BI (1st and 2nd codon positions of CO1 gene)

Appendix 4.11: Phylogenetic trees of the Achatinidae based on the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 516) and (C) maximum parsimony based on 127 equally parsimonious trees (p. 516). The phylogenies were constructed from 427 unambiguously aligned nucleotide sites and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 10 nucleotides and that of (B) represents 5 substitutional changes per 100 nucleotides. Note that *Achatina loveridgei* and *Limicolariopsis ruwenzoriensis* were not included in the phylogenies as they were not successfully sequenced.

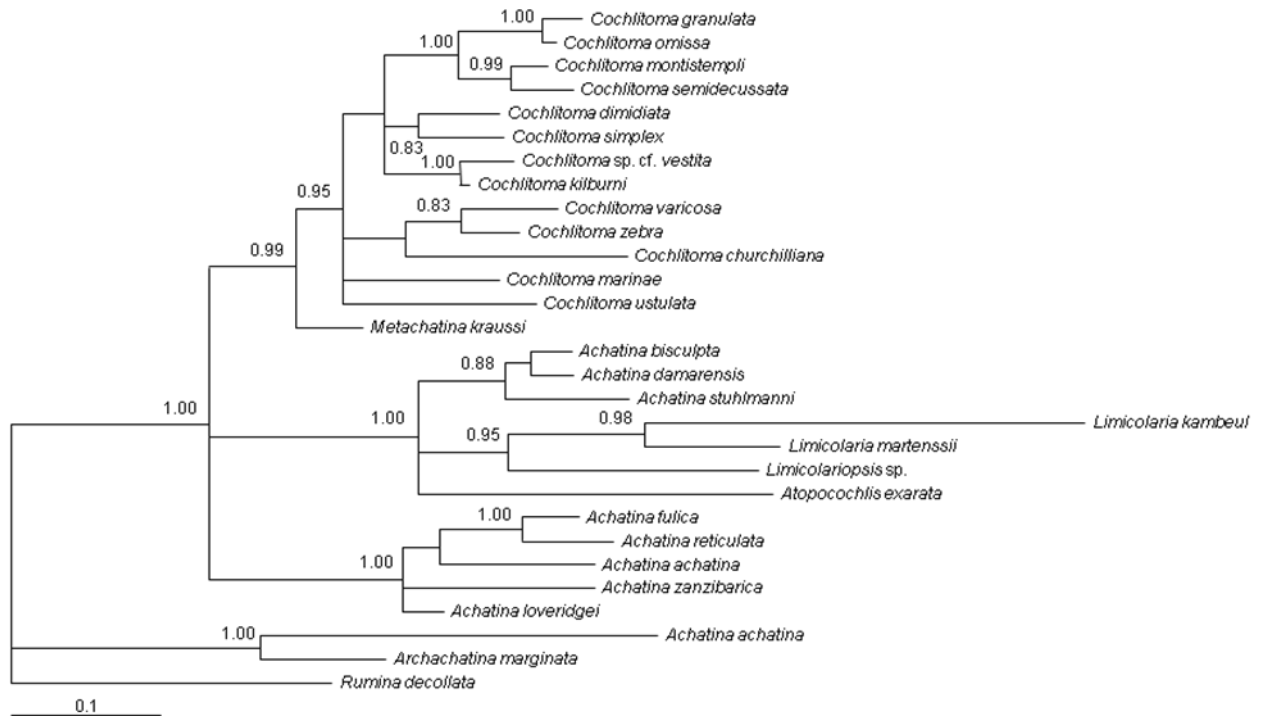
Appendix 4.11 (contd.)



(B) NJ (1st and 2nd codon positions of the CO1 gene)



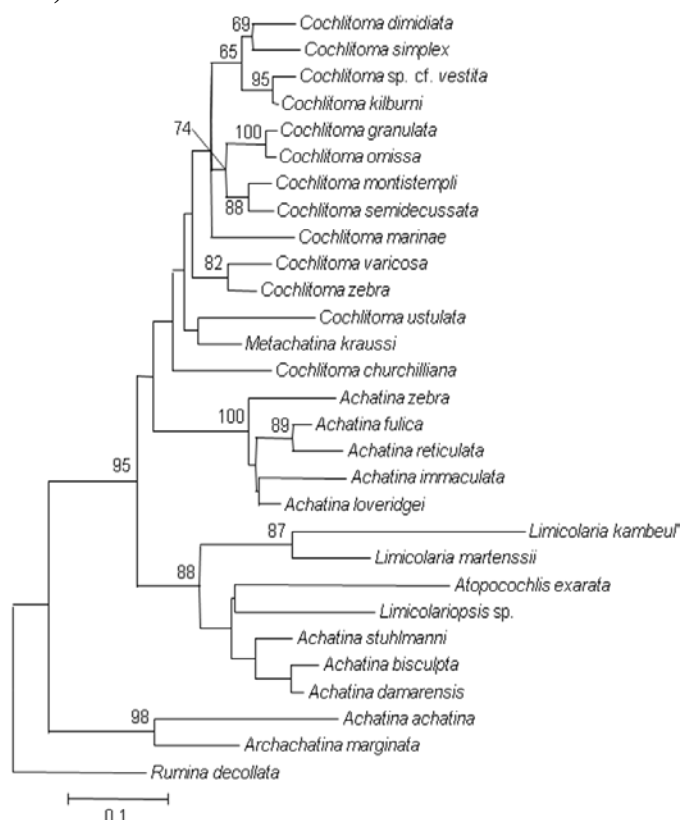
(C) MP (1st and 2nd codon positions of the CO1 gene)



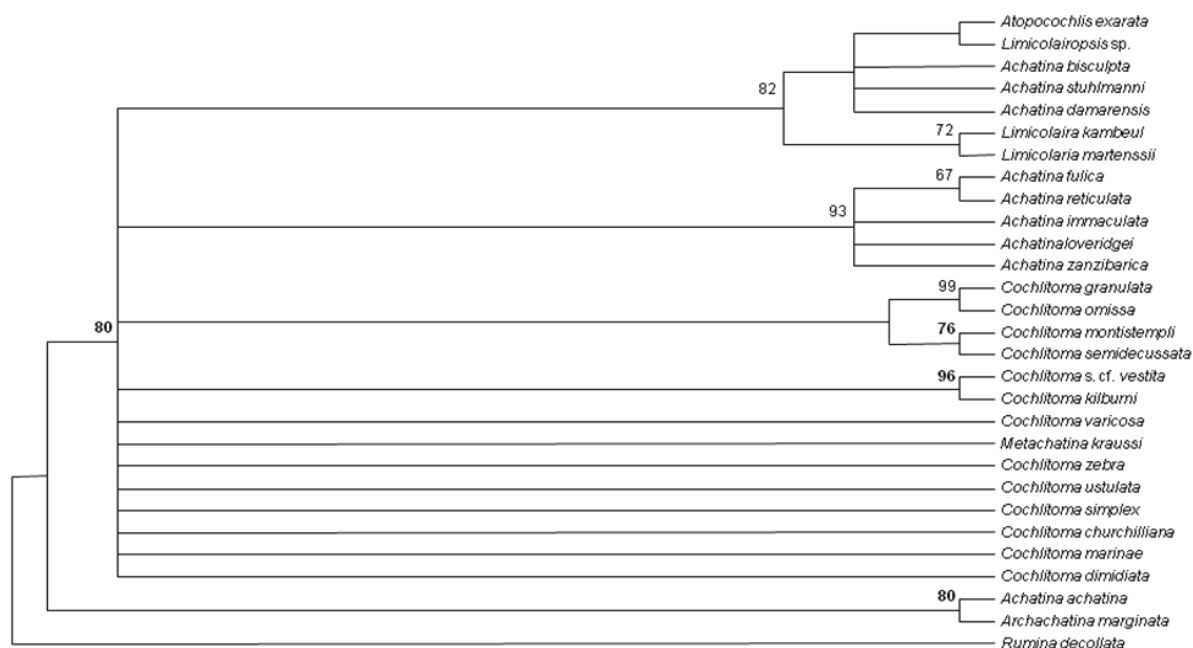
(A) BI (16S rRNA gene)

Appendix 4.12: Phylogenetic trees of the Achatinidae based on the 16S rRNA gene using (A) Bayesian analysis, (B) neighbor-joining (p. 518) and (C) maximum parsimony based on 11 equally parsimonious trees (p. 518). The phylogenies were constructed from 310 unambiguously aligned nucleotide sites and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) and (B) represents 1 substitutional change per 10 nucleotides. Note that *Limicolariopsis ruwenzoriensis* was excluded in the analyses as it was not successfully sequenced.

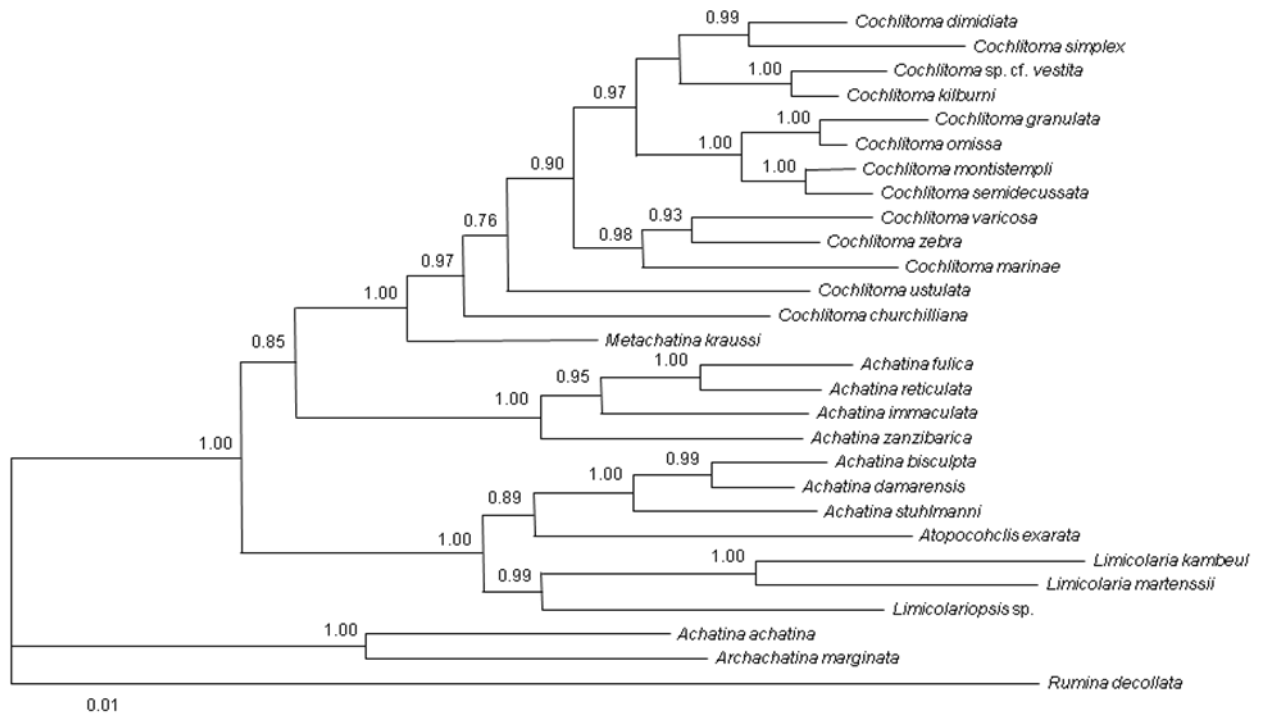
Appendix 4.12 (contd.)



(B) NJ (16S rRNA gene)



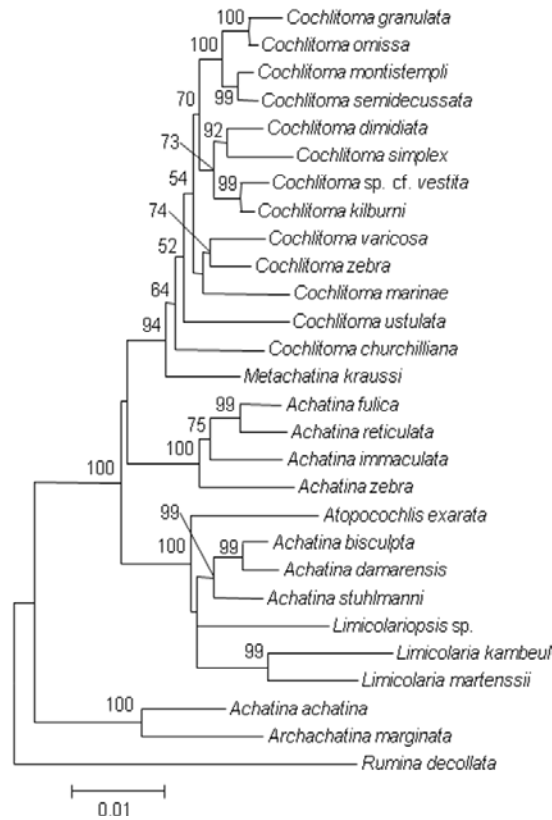
(C) MP (16S rRNA gene)



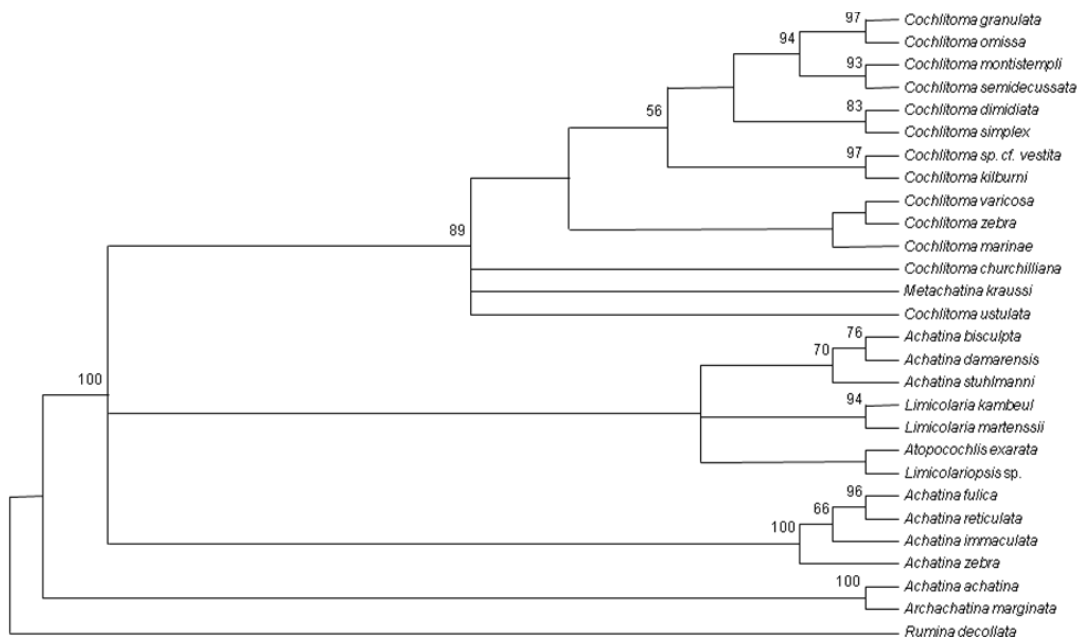
(A) BI (concatenated sequence – taxa with complete datasets only)

Appendix 4.13: Phylogenetic trees of the Achatinidae (taxa with complete datasets only) based on the combined dataset of the LSU rRNA, histone 3, the 1st and 2nd codon positions of the CO1 gene and the 16S rRNA using (A) Bayesian analysis, (B) neighbor-joining (p. 520) and (C) maximum parsimony based on four equally parsimonious trees (p. 520). The phylogenies were constructed from a concatenated sequence of 4713 nucleotides and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bars for (A) and (B) represent 1 substitutional change per 100 nucleotides. Note that *Achatina loveridgei* and *Limicolariopsis ruwenzoriensis* were not included in the phylogenies as they lack at least one of the genes used for the combined analyses.

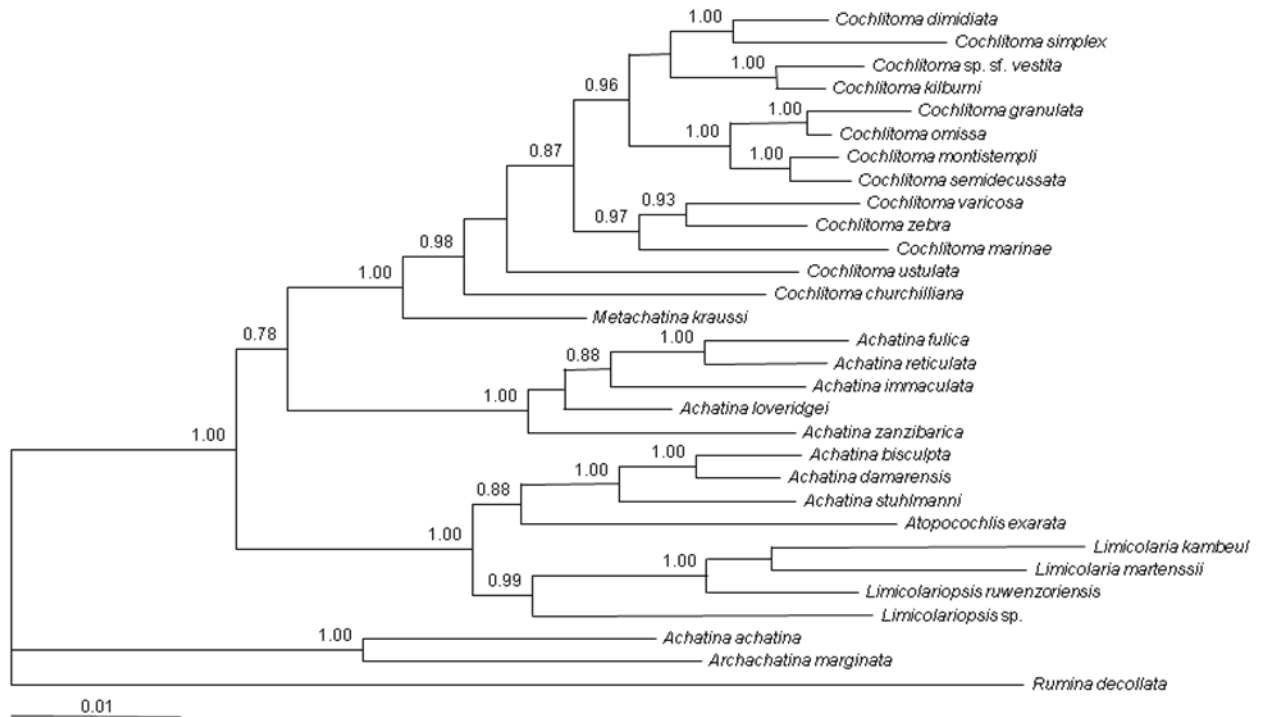
Appendix 4.13 (contd.)



(B) NJ (concatenated sequence – taxa with complete datasets only)



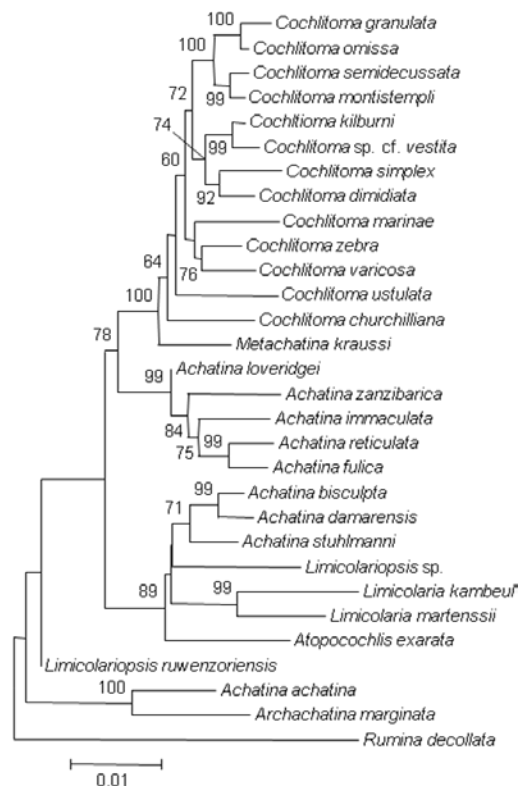
(C) MP (concatenated sequence – taxa with complete datasets only)



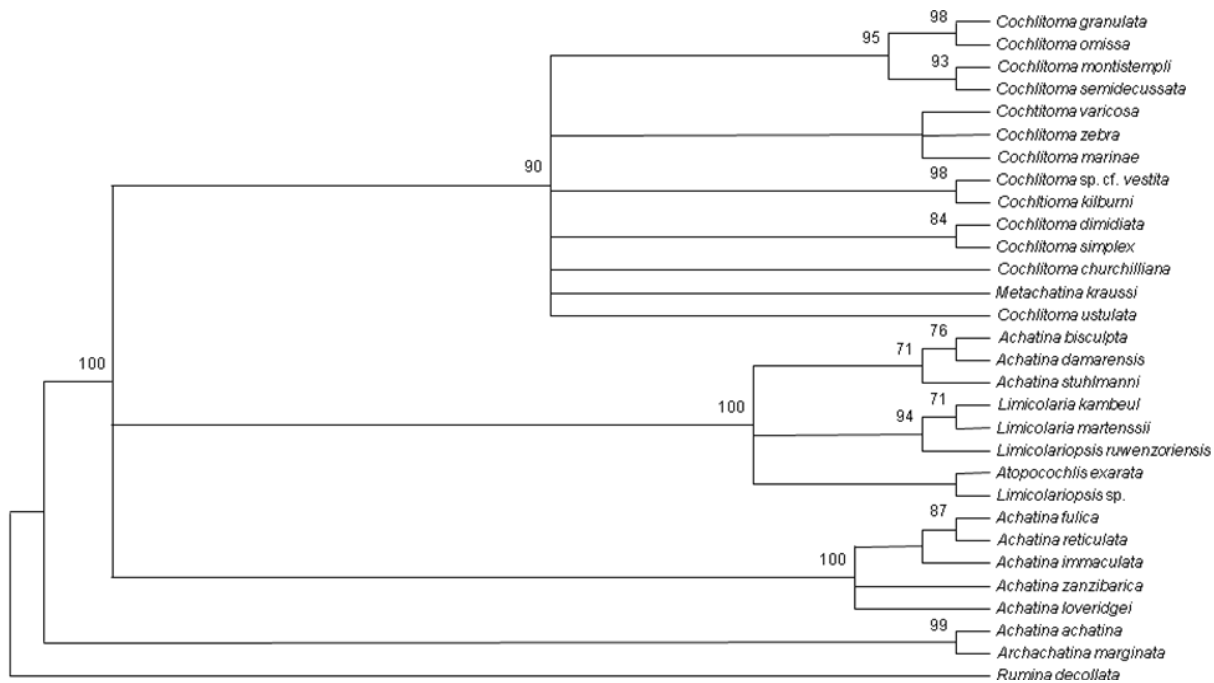
(A) BI (concatenated sequence – all taxa)

Appendix 4.14: Phylogenetic trees of the Achatinidae (all taxa) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 522) and (C) maximum parsimony based on eight equally parsimonious trees (p. 522). The phylogenies were constructed from a concatenated sequence of 4713 nucleotides and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bars for (A) and (B) represent 1 substitutional change per 100 nucleotides.

Appendix 4.14 (contd.)



(B) NJ (concatenated sequence – all taxa)



(C) MP (concatenated sequence – all taxa)

Appendix 5.1: Pair-wise uncorrected distances of the East African *Achatina* species using the SSCP fragment of the 16S rRNA gene (293 nucleotides for *A. fulica*).

	<i>A. fulica</i>	<i>A. zanzibarica</i>	<i>A. immaculata</i>
<i>A. fulica</i>	--		
<i>A. zanzibarica</i>	0.24518		
<i>A. immaculata</i>	0.22864	0.26893	
<i>A. reticulata</i>	0.14414	0.24952	0.22459

Appendix 5.2: Alignment of the 15 haplotypes for the global *Achatina fulica* populations based on 293 nucleotides of the 16S rRNA gene. Sequence for *Achatina reticulata* was used as outgroup.

	10 20 30 40 50 60
Haplotype A	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype B	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype C	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype D	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype E	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype F	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype G	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype H	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype I	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype J	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype K	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype L	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype M	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype N	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
Haplotype O	TAATTTGTCC TCTAATTAGG GTCTGGAATG AAGGGGGACA CAGGGGAGAG CTGTCTCCAA
<i>A. reticulata</i>	TAATTTGTCC TTTAATTTAG GTCTGGAATG AATGAGAACA CAGGGGAGAG CTGTCTCTAG

	70 80 90 100 110 120
Haplotype A	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
Haplotype B	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
Haplotype C	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
Haplotype D	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
Haplotype E	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
Haplotype F	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
Haplotype G	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCTGATG AAAGACGAGA
Haplotype H	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
Haplotype I	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GTCTCCGATG AAAGACGAGA
Haplotype J	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GTCTCCGATG AAAGACGAGA
Haplotype K	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
Haplotype L	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GGCTCCGATG AAAGACGAGA
Haplotype M	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GTCTCCGATG AAAGACGAGA
Haplotype N	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GTCTCCGATG AAAGACGAGA
Haplotype O	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCCTCCGATG AAAGACGAGA
<i>A. reticulata</i>	TAAGGTTAAT TTAACCTTCT TATCAGGTGA AAATTCCTGA GCTTCTTTGG AAAGACGAGA

	130	140	150	160	170	180	
Haplotype A	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATTAAAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype B	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATTAAAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype C	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATTAAAGAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype D	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATTAAAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype E	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATTAAAGAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype F	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATCAAGAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype G	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATTAAAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype H	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATTAAAGAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype I	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	GTATTAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype J	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	GTATTAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype K	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	ATATTAAAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype L	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	GTATTAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype M	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	GTATTAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype N	AGACCCTTAG	AGTTTTTATT	ATAACTTGAT	GTATTAAAT	TCTTGTTCTG	TTTTTGTTGG	
Haplotype O	AGACCCTTAG	AGTTTTTATT	ATAATTGAT	ATATTAAAAAT	TCTTGTTCTG	TTTTTGTTGG	
A. reticulata	AGACCCTTAG	AGTTTAAATA	ATTATTAGA	TAAATAAAAT	TCTACTATG	TTTTTGTTGG	

	190	200	210	220	230	240	
Haplotype A	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype B	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype C	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype D	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype E	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype F	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype G	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype H	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype I	GGCGACAGGG	TTACAACAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype J	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype K	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype L	GGCGACAGGG	TTACAACAGA	TAA-CCTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype M	GGCGACAGGG	TTACAACAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
Haplotype N	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATG--	TTTTTATATG	GCGATCATT	A
Haplotype O	GGCGACAGGG	TTACAATAGA	TAA-CTTACC	CTACCATA--	TTTTTATATG	GCGATCATT	A
A. reticulata	GGCGACAGGG	TAACAATAGG	TAAACTTACC	CATTCAATATA	TTTTTATGCG	TCGATAGTTA	

	250	260	270	280	290	
Haplotype A	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTTG	GTTTGC
Haplotype B	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTTG	GTTTGC
Haplotype C	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype D	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype E	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype F	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype G	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype H	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype I	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype J	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype K	TACTTTTAAT	TAAGCTACCT	GAGGGATAAC	AGCGTAATCT	TTTTTTTTT--G	GTTTGC
Haplotype L	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT--G	GTTTGC
Haplotype M	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype N	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
Haplotype O	TACTTTTAAT	TAAGCTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC
A. reticulata	TACTAATAAT	TAAACTACCT	AAGGGATAAC	AGCGTAATCT	TTTTTTTTT-G	GTTTGC

Appendix 5.3: Log likelihood scores of the different models used for comparison for the 16S haplotypes of the global *A. fulica* populations using 293 nucleotides of the 16S rRNA gene. Likelihood Ratio Test revealed that GTR and TrN93 models were not significantly better than HKY85; the latter was therefore used for the phylogenetic analysis. Note that the K2P and F81 models are non-nested and therefore cannot be compared using LRT.

Model	Log-Likelihood Score
JC69	697.00855
JC69+ Γ	696.73625
F81	676.85077
F81+ Γ	676.50788
K2P	692.34383
K2P+ Γ	691.96730
HKY85	670.85793
HKY85+ Γ	670.25753
TN93	670.82611
TN93+ Γ	670.23779
GTR	666.27323
GTR+ Γ	666.03208

Appendix 6.1: Nematode species used for the *Angiostrongylus cantonensis* survey as found in GenBank Release No. 157 (22 December 2006) or GenBank Release No. 158 (15 February 2007) as indicated by (*).

Species	Accession No.	Taxonomy
<i>Gordius aquaticus</i>	X87985	Nematomorpha; Gordioida
<i>Chordodes morgani</i>	AF036639	Nematomorpha; Gordioida
<i>Priapulius caudatus</i>	Z38009	Priapulida; Priapulidae
<i>Brachionus plicatilis</i>	U49911	Rotifera; Monogononta
<i>Axonolaimus helgolandicus</i>	AY854232	Nematoda; Chromadorea; Araeolaimida; Axonolaimoidea; Axonolaimidae
<i>Ascolaimus elongatus</i>	AY854231	Nematoda; Chromadorea; Araeolaimida; Axonolaimoidea; Axonolaimidae
<i>Odontophora rectangula</i>	AY854233	Nematoda; Chromadorea; Araeolaimida; Axonolaimoidea; Axonolaimidae
<i>Cylindrolaimus</i> sp. 202149	AF202149	Nematoda; Chromadorea; Araeolaimida; Cylindrolaimidae
<i>Tylocephalus auriculatus</i>	AF202155	Nematoda; Chromadorea; Araeolaimida; Plectoidea; Plectidae
<i>Plectus acuminatus</i> strain BS9	AF037628	Nematoda; Chromadorea; Araeolaimida; Plectoidea; Plectidae
<i>Plectus aquatilis</i>	AF036602	Nematoda; Chromadorea; Araeolaimida; Plectoidea; Plectidae
<i>Anaplectus</i> sp. PDL-2005	AJ966473	Nematoda; Chromadorea; Araeolaimida; Plectoidea; Plectidae
<i>Anisakis</i> sp. WKT	U81575	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Anisakis</i> sp. Nadler	U94365	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Contraecum multipapillatum</i>	U94370	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Pseudoterranova decipiens</i>	U94380	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Raphidascaris acus</i> *	DQ503460	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Terranova caballeroi</i>	U94382	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Ascaris suum</i>	U94367	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Ascaris lumbricoides</i>	U94366	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Baylisascaris procyonis</i>	U94368	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Baylisascaris transfuga</i>	U94369	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Parascaris equorum</i>	U94378	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Porrocaecum depressum</i>	U94379	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Toxascaris leonine</i>	U94383	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Heteroncheilus tunicatus</i>	U94373	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Heteroncheilidae
<i>Goezia pelagia</i>	U94372	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Hysterothylacium fortalezae</i>	U94374	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Hysterothylacium pelagicum</i>	U94375	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Hysterothylacium reliquens</i>	U94376	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Iheringascaris iniquus</i>	U94377	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Toxocara canis</i>	U94382	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Toxocaridae
<i>Nemhelix bakeri</i>	DQ118537	Nematoda; Chromadorea; Ascaridida; Cosmocercidae; Cosmocercidae
<i>Raillietnema</i> sp. V3060*	DQ503461	Nematoda; Chromadorea; Ascaridida; Cosmocercidae; Cosmocercidae
<i>Cruzia Americana</i>	U94371	Nematoda; Chromadorea; Ascaridida; Cosmocercidae; Kathliniidae
<i>Paraspidodera</i> sp. 21303	AF083005	Nematoda; Chromadorea; Ascaridida; Heterakoidea; Aspidoderidae

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<i>Heterakis gallinarum</i> *	DQ503462	Nematoda; Chromadorea; Ascaridida; Heterakoidea; Heterakidae
<i>Heterakis</i> sp.14690	AF083003	Nematoda; Chromadorea; Ascaridida; Heterakoidea; Heterakidae
<i>Chromadoropsis vivipara</i>	AF047891	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Chromadora nudicapitata</i>	AY854205	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Chromadora</i> sp. BHMM_2005	AY854206	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Atrochromadora microlaima</i>	AY854204	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Chromadorita tentabundum</i>	AY854208	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Chromadorina germanica</i>	AY854207	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Dichromadora</i> sp. BHMM_2005	AY854209	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Neochromadora</i> BHMM_2005	AY854210	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Spilophorella paradoxa</i>	AY854211	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Paracanthonchus caecus</i>	AF047888	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Paracyatholaimus intermedius</i>	AJ966495	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Praeacanthonchus punctatus</i>	AY854214	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Praeacanthonchus</i> sp.	AF036612	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Cyatholaimus</i> sp. BHMM_2005	AY854213	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Spirinia parasitifera</i>	AY854217	Nematoda; Chromadorea; Desmodorida; Desmodoridae
<i>Acanthopharynx micans</i>	Y16911	Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Desmodora ovigera</i>	Y16913	Desmodorinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Desmodora communis</i>	AY854215	Desmodorinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Xyzzors</i> sp.	Y16923	Desmodorinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Metachromadora</i> sp.	AF036595	Desmodorinae Nematoda; Chromadorea; Desmodorida; Desmodoridae; Spiriniinae
<i>Metachromadora remanei</i>	AY854216	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Spiriniinae
<i>Catanema</i> sp.	Y16912	Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Eubostrichus diana</i>	Y16915	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Eubostrichus topiarus</i>	Y16917	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Eubostrichus parasitiferus</i>	Y16916	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Laxus oneistus</i>	Y16919	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Laxus cosmopolites</i>	Y16918	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Leptonemella</i> sp.	Y16920	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Robbea hypermnestra</i>	Y16921	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Stilbonema majum</i>	Y16922	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Desmodoridae;
<i>Monoposthia costata</i>	AY854221	Stilbonematinae Nematoda; Chromadorea; Desmodorida; Monoposthiidae
<i>Nudora bipapillata</i>	AY854222	Nematoda; Chromadorea; Desmodorida; Monoposthiidae
<i>Calomicrolaimus parahonestus</i>	AY854218	Nematoda; Chromadorea; Desmodorida; Richtersioidea;
<i>Calomicrolaimus</i> sp. BHMM_2005	AY854219	Microlaimidae Nematoda; Chromadorea; Desmodorida; Richtersioidea;
<i>Molgolaimus demani</i>	AY854220	Microlaimidae Nematoda; Chromadorea; Desmodorida; Richtersioidea;
<i>Diplogaster lethieri</i>	AF036643	Microlaimidae Nematoda; Chromadorea; Diplogasterida; Diplogasteridae

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<i>Aduncospiculum halicti</i>	U61759	Nematoda; Chromadorea; Diplogasterida; Diplogasteridae
<i>Pristionchus lheritieri</i>	AF036640	Nematoda; Chromadorea; Diplogasterida; Neodiplogasteridae
<i>Pristionchus pacificus</i>	U81584	Nematoda; Chromadorea; Diplogasterida; Neodiplogasteridae
<i>Pristionchus pacificus</i> PS312	AF083010	Nematoda; Chromadorea; Diplogasterida; Neodiplogasteridae
<i>Sabatieria punctata</i> Strain 343	AY854237	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria</i> sp. 355 BHMM_2005	AY854239	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria celtica</i>	AY854234	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria punctata</i> Strain 200	AY854235	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria punctata</i> Strain 223	AY854236	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria</i> sp. 210 BHM_2005	AY854238	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Setosabatieria hilarula</i>	AY854240	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Desmolaimus zeelandicus</i>	AY854229	Nematoda; Chromadorea; Monhysterida; Linhomoeidae
<i>Terschellingia longicaudata</i>	AY854230	Nematoda; Chromadorea; Monhysterida; Linhomoeidae
<i>Cyartonema elegans</i>	AY854203	Nematoda; Chromadorea; Monhysterida; Monhysterida incertae sedis
<i>Tridentulus</i> sp. PDL-2005	AJ966507	Nematoda; Chromadorea; Monhysterida; Monhysteridae
<i>Diplolaimelloides meylli</i>	AJ966507	Nematoda; Chromadorea; Monhysterida; Monhysteridae
<i>Diplolaimella dievengatensis</i>	AJ966482	Nematoda; Chromadorea; Monhysterida; Monhysteridae
<i>Geomonhystera disjuncta</i>	AJ966485	Nematoda; Chromadorea; Monhysterida; Monhysteridae
<i>Sphaerolaimus hirsutus</i>	AY854228	Nematoda; Chromadorea; Monhysterida; Sphaerolaimidae
<i>Theristus acer</i>	AJ966505	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema procerus</i>	AF047889	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema hirsutum</i>	AY854223	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema normandicum</i>	AY854224	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema oxycerca</i>	AY854225	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema setosum</i>	AY854226	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Metadesmolaimus</i> sp. PDL-2005	AJ966491	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Dentostomella</i> sp.	AF036590	Nematoda; Chromadorea; Oxyurida; Oxyuroidea; Heteroxynematidae
<i>Bunonema franzi</i>	AJ966477	Nematoda; Chromadorea; Rhabditida; Bunonematoidea; Bunonematidae
<i>Bunonema</i> sp.	U81582	Nematoda; Chromadorea; Rhabditida; Bunonematoidea; Bunonematidae
<i>Seleborca complexa</i>	U81577	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobida
<i>Cephaloboides</i> sp. SB227	AF083027	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobelloides nanus</i>	DQ102707	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobelloides bodenheimeri</i> PS1158	AF202159	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobelloides</i> sp. PS1146	AF034391	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobeles ciliates</i>	AF202148	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobeles</i> sp. PS1156	U81576	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobeles complexus</i> WCUG2	U81577	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Cephalobus cubaensis</i>	AF202161	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Cephalobus</i> sp. PS1143	AF202158	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae

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<i>Cephalobus</i> sp. PS1196	AF202160	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Cephalobus oryzae</i> PS1165	AF034390	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Cervidellus alutus</i>	AF202152	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Pseudacrobeles variabilis</i>	AF202150	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Triligulla alluta</i>	AF331911	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Zeldia punctata</i>	U61760	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Myolaimus</i> sp U81585	U81585	Nematoda; Chromadorea; Rhabditida; Myolaimoidea; Myolaimidae
<i>Rhabditophanes</i> sp. KR3021	AF202151	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Alloionematidae
<i>Brevibucca</i> sp. SB261	AF202163	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Brevibuccidae
<i>Halicephalobus gingivalis</i>	AF202156	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrobelus stammeri</i>	AF202153	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Plectonchus</i> sp. PDL0025	AF202154	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Turbatrix aceti</i>	AF202165	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrellus redivivus</i>	AF036599	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrellus redivivus</i> PS1163	AF083007	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus subelongatus</i>	AY284681	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus davidi</i>	AJ567385	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus cf rigidus</i> AF40	DQ285636	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm7	AF430487	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm5	AF430485	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm6	AF430486	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm4	AF430484	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm3	AF430483	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Steinernema carpocapsae</i>	AF036604	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Steinernematidae
<i>Strongyloides stercoralis</i>	M84229	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Strongyloididae
<i>Strongyloides ratti</i>	U81581	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Strongyloididae
<i>Diploscapter</i> sp PS1897	AF083009	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Diploscapteridae
<i>Diploscapter</i> sp PS2017	U81586	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Diploscapteridae
<i>Heterorhabditis bacteriophora</i>	AF036593	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Heterorhabditidae
<i>Heterorhabditis hepialus</i>	AF083004	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Heterorhabditidae
<i>Heterorhabditis zelandica</i>	AJ920368	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Heterorhabditidae
<i>Phasmarhabditis hermaphrodita</i> *	DQ639981	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Phasmarhabditis neopapillosa</i> *	DQ639982	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Parasitorhabditis</i> sp SB281	AF083028	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Rhabditoides inermiformis</i>	AF083017	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae

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<i>Rhabditoides inermis</i> DF5001	AF082996	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Rhabditoides regina</i>	AF082997	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Poikilolaimus oxycerca</i> SB200	AF083023	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Poikilolaimus regenfussi</i> SB199	AF083022	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Choriorhabditis dudichi</i>	AF083012	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Distolabrellus veechi</i>	AF083011	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Distolabrellus veechi</i> DF5024	AF082999	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Protorhabditis</i> sp.	AF083024	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Protorhabditis</i> sp DF5055	AF083001	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Cruznema tripartitum</i> DF5015	U73449	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Mesorhabditis</i> sp. PS1179	U73452	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Mesorhabditis spiculigera</i> SB157	AF083016	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Mesorhabditis anisomorpha</i> SB123	AF083013	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Teratorhabditis palmarum</i> DF5019	U13937	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Teratorhabditis synpapillata</i> SB131	AF083015	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Caenorhabditis briggsae</i> PB102	U13929	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis elegans</i> N2	X03680	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp CB5161	U13930	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. PS1010	AF083006	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis japonica</i>	AY602182	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. DF5170	AY602181	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. SB341	AY602180	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis plicata</i>	AY602178	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. CB5161	U13930	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis vulgaris</i>	U13931	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis sonoreae</i>	AF083026	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis drosophilae</i>	AF083025	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Pellioditis marina</i>	AF083021	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Pellioditis mediterranea</i>	AF083020	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Prodontorhabditis wirthi</i>	AY602179	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Protorhabditinae
<i>Crustorhabditis scanica</i> SB125	AF083014	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Dolichorhabditis</i> sp CEW1 (=Oscheius)	AF036591	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Oscheius</i> sp. DF5000	AF082995	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Oscheius</i> sp. BW282	AF082994	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Oscheius myriophila</i> EM435	U81588	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae

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<i>Oscheius insectivora</i>	AF083019	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Oscheius dolichuroides</i>	AF082998	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis blumi</i> DF5010	U13935	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis sp</i> PS1191	AF083008	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis sp</i> PS1010	AF083006	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis myriophila</i> EM435	U13936	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis colombiana</i> *	AY751546	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditella axei</i> DF5006	U13934	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditella sp.</i> DF5044	AF083000	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Cuticularia sp.</i> PS2083	U81583	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Necator americanus</i>	AY295811	Nematoda; Chromadorea; Rhabditida; Strongylida; Ancylostomatoidea; Ancylostomatidae; Bunostominae
<i>Kalicephalus cristatus</i>	AJ920349	Nematoda; Chromadorea; Rhabditida; Strongylida; Diaphanocephaloidea; Diaphanocephalidae
<i>Ancylostoma caninum</i>	AJ920347	Nematoda; Chromadorea; Rhabditida; Strongylida; Ancylostomatoidea; Ancylostomatidae; Ancylostomatinae
<i>Angiostrongylus cantonensis</i>	AY295804	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Angiostrongylus costaricensis</i>	This study	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Angiostrongylus malaysiensis</i>	This study	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Angiostrongylus dujardini</i>	This study	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Angiostrongylus vasorum</i>	This study	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Aelurostrongylus abstrusus</i>	AJ920366	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Didelphostrongylus hayesi</i>	AY295806	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Crenosoma mephitidis</i>	AY295805	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Crenosomatidae
<i>Crenosoma vulpis</i>	AJ920367	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Crenosomatidae
<i>Otostrongylus circumlitus</i>	AY295813	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Crenosomatidae
<i>Troglostrongylus wilsoni</i>	AY295820	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Crenosomatidae
<i>Filaroides martis</i>	AY295807	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Filaroididae
<i>Oslerus osleri</i>	AY295812	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Filaroididae
<i>Parafilaroides decorus</i>	AY295814	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Filaroididae
<i>Metastrongylus salmi</i>	AY295809	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Metastrongylidae
<i>Metastrongylus elongatus</i>	AJ920363	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Metastrongylidae
<i>Skrjabinogylus chitwoodrum</i>	AY295819	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Metastrongylidae
<i>Parelaphostrongylus odocoilei</i>	AY295815	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Protostrongylidae; Elaphostrongylinae
<i>Muellerius capillaries</i>	AY295810	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Protostrongylidae; Elaphostrongylinae
<i>Protostrongylus rufescens</i>	AJ920364	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Protostrongylidae; Protostrongylinae
<i>Halocercus invaginatus</i>	AY295808	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Pseudaliidae

Species	Accession No.	Taxonomy
<i>Pseudalius inflexus</i>	AY295816	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Pseudaliidae
<i>Stenurus minor</i>	AY295817	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Pseudaliidae
<i>Syngamus trachaea</i>	AJ920344	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Syngamidae
<i>Torynurus convolutus</i>	AY295818	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Unclassified Metastrongyloidea
<i>Nematodirus battus</i>	U01230	Nematoda; Chromadorea; Rhabditida; Strongylida; Molineoidea; Molineidae; Nematodirinae
<i>Petrovinema poculatum</i>	AJ920343	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Strongylidae
<i>Strongylus equines</i>	DQ094176	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Strongylidae
<i>Cylicocyclus insignis</i>	AJ920342	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Strongylidae
<i>Zoniolaimus mawsonae</i>	AJ920338	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Cloacinidae
<i>Labiostrongylus bipapillosus</i>	AJ920337	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Cloacinidae
<i>Hypodontus macropi</i>	AJ920339	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Cloacinidae
<i>Chabertia ovina</i>	AJ920341	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Chabertiidae
<i>Cyclodontostomum purvisi</i>	AJ920340	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Chabertiidae
<i>Stephanurus dentatus</i>	AJ920345	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Stephanuridae
<i>Deletrocephalus dimidiatus</i>	AJ920346	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Deletrocephalidae
<i>Dictyocaulus eckerti</i> P7B8	AY168857	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus capreolus</i> P3B2	AY168859	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus</i> sp. P6A1	AY168860	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus filarial</i>	AY168861	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus capreolus</i> P2C10	AY168862	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus viviparus</i>	AY168856	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus filarial</i>	AJ920362	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus viviparus</i>	AJ920361	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Haemonchus</i> sp. V3091*	DQ503465	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Haemonchinae
<i>Haemonchus contortus</i>	L04153	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Haemonchinae
<i>Haemonchus placei</i>	L04154	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Haemonchinae
<i>Haemonchus similis</i>	L04152	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Haemonchinae
<i>Ostertagia ostertagi</i>	AJ920352	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Ostertagiinae
<i>Ostertagia leptospicularis</i>	AJ920351	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Ostertagiinae
<i>Nippostrongylus brasiliensis</i>	AJ920356	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Heligmonellidae; Nippostrongylinae
<i>Heligmosomoides polygyrus</i>	AJ920355	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Heligmosomatidae
<i>Trichostrongylus colubriformis</i>	AJ920350	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Trichostrongylidae
<i>Tetrabothriostrongylus mackerrasae</i>	AJ920359	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Mackerrastrongylidae
<i>Herpetostrongylus pythonis</i>	AJ920358	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Herpetostrongylidae

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<i>Nicollina cameroni</i>	AJ920357	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Nicollinidae
<i>Filarinema flagrifer</i>	AJ920354	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dromaeostrongylidae
<i>Amidostomum cygni</i>	AJ920353	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Amidostomatidae
<i>Teratocephalus lirellus</i>	AF036607	Nematoda; Chromadorea; Rhabditida; Teratocephaloidea; Teratocephalidae; Teratocephalinae
<i>Brumptaemilius justini</i>	AF036589	Nematoda; Chromadorea; Rhigonematida
<i>Anguillicola crassus</i>	DQ118535	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Anguillicolidae
<i>Dracunculus medinensis</i>	AY852268	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Dracunculidae
<i>Dracunculus oesophageus</i>	AY852269	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Dracunculidae
<i>Dracunculus</i> sp. V3104*	DQ503457	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Dracunculidae
<i>Philonema</i> sp. A	U81574	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Philometridae
<i>Philometra obturans</i>	AY852267	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Philometridae
<i>Camallanus oxycephalus</i> *	DQ503463	Nematoda; Chromadorea; Spirurida; Camallanida; Camallanidae
<i>Acanthocheilonema viteae</i>	DQ094171	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Loa loa</i>	DQ094173	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Onchocerca cervicalis</i>	DQ094174	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Dirofilaria immitis</i>	AF036638	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Brugia malayi</i>	AF036588	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Wuchereria bancrofti</i>	AF227234	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Litomosoides sigmodontis</i>	AF227233	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Setaria digitata</i>	DQ094175	Nematoda; Chromadorea; Spirurida; Filarioidea; Setariidae
<i>Gnathostoma turgidum</i>	Z96948	Nematoda; Chromadorea; Spirurida; Gnathostomatoidea; Gnathostomatidae
<i>Gnathostoma neoprocyonis</i>	Z96947	Nematoda; Chromadorea; Spirurida; Gnathostomatoidea; Gnathostomatidae
<i>Gnathostoma binuncleatum</i>	Z96946	Nematoda; Chromadorea; Spirurida; Gnathostomatoidea; Gnathostomatidae
<i>Physaloptera alata</i>	AY702703	Nematoda; Chromadorea; Spirurida; Physalopteroidea; Physalopteridae
<i>Physaloptera turgid</i> *	DQ503459	Nematoda; Chromadorea; Spirurida; Physalopteroidea; Physalopteridae
<i>Ascarophis arctica</i>	DQ094172	Nematoda; Chromadorea; Spirurida; Spiruroidea; Cystidicolidae
<i>Spinitectus carolini</i> *	DQ503565	Nematoda; Chromadorea; Spirurida; Spiruroidea; Cystidicolidae
<i>Thelazia lacrymalis</i> *	DQ503458	Nematoda; Chromadorea; Spirurida; Thelazioidea; Thelaziidae
<i>Aphelenchus avenae</i>	AF036586	Nematoda; Chromadorea; Tylenchida; Aphelenchoidea; Aphelenchidae
<i>Aphelenchoides fragariae</i>	AJ966475	Nematoda; Chromadorea; Tylenchida; Aphelenchoidea; Aphelenchidae
<i>Bursaphelenchus</i> sp.	AF037369	Nematoda; Chromadorea; Tylenchida; Aphelenchoidoidea; Aphelenchoididae
<i>Deladenus</i> sp.	AJ966481	Nematoda; Chromadorea; Tylenchida; Sphaerulariidae
<i>Criconema</i> sp.	AJ966480	Nematoda; Chromadorea; Tylenchida; Tylenchina; Criconematoidea; Criconematidae; Criconematinae
<i>Hemicyclophora conida</i>	AJ966471	Nematoda; Chromadorea; Tylenchida; Tylenchina; Criconematoidea; Criconematidae; Hemicyclophorinae
<i>Pratylenchus thornei</i>	AJ966499	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Pratylenchinae
<i>Paratylenchus dianthus</i>	AJ966496	Nematoda; Chromadorea; Tylenchida; Tylenchina; Criconematoidea; Tylenchulidae; Paratylenchidae
<i>Tylenchulus semipetrans</i>	AJ966511	Nematoda; Chromadorea; Tylenchida; Tylenchina; Criconematoidea; Tylenchulidae; Tylenchulinae

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<i>Ditylenchus angustus</i>	AJ966483	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Anguinidae
<i>Subanguina radicola</i>	AF202164	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Anguinidae; Anguininae
<i>Tylenchorhynchus maximus</i>	AY993979	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Belonolaimidae
<i>Geocenamus quadrifer</i>	AY993977	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Belonolaimidae
<i>Globodera pallid</i>	AFO36592	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Heteroderidae; Heteroderinae
<i>Scutellonema bradys</i>	AJ966504	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Hoplolaimidae; Hoplolaiminae
<i>Helicotylenchus dihystra</i>	AJ966486	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Hoplolaimidae; Hoplolaiminae
<i>Rotylenchus robustus</i>	AJ966503	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Hoplolaimidae; Rotylenchulinae
<i>Meloidogyne arenaria</i>	U42342	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne incognita</i>	U81578	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne javanica</i>	AF442193	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne artiellia</i>	AF442192	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne duytsi</i>	AF442197	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne exigua</i>	AF442200	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne hapla</i>	AF442194	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne ichinohei</i>	AF442191	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne maritima</i>	AF442199	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne microtyla</i>	AF442198	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Nacobbus aberrans</i>	AJ966494	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Nacobbiniae
<i>Pratylenchoides ritteri</i>	AJ966497	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Pratylenchinae
<i>Pratylenchoides magnicauda</i>	AF202157	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Pratylenchinae
<i>Hirschmanniella</i> sp. JH_2003	AY284616	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Pratylenchinae
<i>Radopholus similis</i>	AJ966502	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Radopholinae
<i>Pratylenchus goodyeri</i>	AJ966498	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Radopholinae
<i>Boleodorus thylactus clone2</i>	AY993976	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Tylenchidae; Boleodorinae
<i>Paractinolaimus macrolaimus</i>	AY993978	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Actinolaimoidea; Paractinolaiminae
<i>Aporcelaimellus obtusicaudatus</i>	DQ141212	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Aporcelaimidae; Aporcelaiminae
<i>Wilsonema schuurmansstekhoveni</i>	AJ966513	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Dorylaimidae
<i>Mesodorylaimus</i> cf. <i>nigritulus</i> AV-2005	AJ966490	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Dorylaimidae; Laimyodorinae
<i>Mesodorylaimus japonicus</i>	AJ966489	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Dorylaimidae; Laimyodorinae
<i>Mesodorylaimus bastiani</i>	AJ966488	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Dorylaimidae; Laimyodorinae
<i>Pungentus</i> sp. PDL-2005	AJ966501	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Nordiidae; Pungentinae
<i>Allodorylaimus</i> sp. PDL-2005	AJ966472	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Qudsianematidae
<i>Eudorylaimus carteri</i>	AJ966484	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Qudsianematidae; Qudsianematinae

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<i>Microdorylaimus</i> sp. PDL-2005	AJ966492	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Qudsianematidae; Qudsianematinae
<i>Longidorus elongates</i>	AF036594	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Longidoroidea; Longidoridae
<i>Xiphinema rivesi</i>	AF036610	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Longidoroidea; Longidoridae
<i>Tylencholaimus</i> sp. PDL-2005	AJ966510	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Tylencholaimoidea; Tylencholaimidae; Tylencholaiminae
<i>Anoplostoma</i> sp. BHMM_2005	AY854194	Nematoda; Enoplea; Enoplida; Enoploidea; Anoplostomatidae
<i>Adoncholaimus fuscus</i>	AY854195	Nematoda; Enoplea; Enoplida; Enoploidea; Enopliidae
<i>Enoplus meridionalis</i>	Y16914	Nematoda; Enoplea; Enoplida; Enoploidea; Enopliidae
<i>Enoplus brevis</i>	U88336	Nematoda; Enoplea; Enoplida; Enoploidea; Enopliidae
<i>Enoplus communis</i>	AY854192	Nematoda; Enoplea; Enoplida; Enoploidea; Enopliidae
<i>Enoploides brunettii</i>	AY854193	Nematoda; Enoplea; Enoplida; Enoploidea; Thoracostomopsidae
<i>Syringolaimus striatocaudatus</i>	AY854200	Nematoda; Enoplea; Enoplida; Ironoidea; Ironidae
<i>Ironus dentifurcatus</i>	AJ966487	Nematoda; Enoplea; Enoplida; Ironoidea; Ironidae
<i>Calyptronema maxweberi</i>	AY854199	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Enchelidiidae
<i>Viscosia viscosa</i>	AY854198	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Oncholaimidae
<i>Viscosia</i> sp. BHMM_2005	AY854197	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Oncholaimidae
<i>Pontonema</i> sp.	AF047890	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Oncholaimidae
<i>Oncholaimus</i> sp. BHMM_2005	AY854196	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Oncholaimidae
<i>Alaimus</i> sp. PDL-2005	AJ966514	Nematoda; Enoplea; Enoplida; Oxystominoidea; Alaimidae
<i>Prismatolaimus intermedius</i>	AF036603	Nematoda; Enoplea; Enoplida; Tripyloidea; Prismatolaimidae
<i>Tobrilus gracilis</i>	AJ966506	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripylidae
<i>Tripyla</i> cf. <i>filicaudata</i> JH-2004 isolate TripFil2Z	AY284731	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripylidae
<i>Bathylaimus</i> sp. PB-2005	AM234619	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Bathylaimus assimilis</i>	AJ966476	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Bathylaimus</i> sp. BHMM_2005	AY854201	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Tripyloides</i> sp. BHMM_2005	AY854202	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Trischistoma monohystera</i>	AJ966509	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Mermis nigrescens</i>	AF036641	Nematoda; Enoplea; Mermithida; Mermithoidea; Mermithidae
<i>Mylonchulus arenicolus</i>	AF036596	Nematoda; Enoplea; Mononchida; Mononchina; Mononchoidea; Mylonchulidae; Mylonchulinae
<i>Anatonchus tridentatus</i>	AJ966474	Nematoda; Enoplea; Mononchida; Mononchina; Anatonchoidea; Anatonchidae
<i>Mononchus truncatus</i>	AJ966493	Nematoda; Enoplea; Mononchida; Mononchina; Mononchoidea; Mononchidae; Mononchinae
<i>Prionchulus muscorum</i>	AJ966500	Nematoda; Enoplea; Mononchida; Mononchina; Mononchoidea; Mononchidae; Prionchulinae
<i>Clarkus</i> sp. PDL-2005	AJ966479	Nematoda; Enoplea; Mononchida; Mononchina; Mononchoidea; Mononchidae; Prionchulinae
<i>Trichinella spiralis</i>	TSU60231	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella papuae</i>	AY851263	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella britovi</i>	AY851257	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella murelli</i>	AY851259	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella pseudospiralis</i>	AY851258	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella native</i>	AY851256	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella zimbabwensis</i>	AY851264	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella nelson</i>	AY851261	Nematoda; Enoplea; Trichocephalida; Trichinellidae

Species	Accession No.	Taxonomy
<i>Trichuris suis</i>	AY851265	Nematoda; Enoplea; Trichocephalida; Trichiuridae
<i>Trichuris trichiura</i>	DQ118536	Nematoda; Enoplea; Trichocephalida; Trichiuridae
<i>Trichuris muris</i>	AF036637	Nematoda; Enoplea; Trichocephalida; Trichiuridae
<i>Tylolaimophorus minor</i>	AJ966512	Nematoda; Enoplea; Triplonchida; Diphtherophorina; Diphtherophoroidea; Diphtherophoridae
<i>Paratrichodorus pachydermus</i>	AF036601	Nematoda; Enoplea; Triplonchida; Diphtherophorina; Trichodoroidea; Trichodoridae
<i>Paratrichodorus anemones</i>	AF036600	Nematoda; Enoplea; Triplonchida; Diphtherophorina; Trichodoroidea; Trichodoridae
<i>Trichodorus primitivus</i>	AF036609	Nematoda; Enoplea; Triplonchida; Diphtherophorina; Trichodoroidea; Trichodoridae

Appendix 6.2: Alignment of the 5' end of the SSU rRNA gene for the Nematoda. Positions included for the phylogeny analyses were marked by “m” within “NUCEOTIDES INCLUDED”. This alignment included new nematode sequences from Chapter 6 and Chapter 7.

	10	20	30	40	50	60				
Gordius_aquaticus	---CTGGTTG	ATYCTGCCAG	TAGTCATATG	CTT-----	-----	-----				
Priapulius_caudatus	AACCTGGTTG	ATCCTGCCAG	TAGTCATATG	CTT-----	-----	-----				
Brachionus_plicatilis	-ACCTGGTTG	ATCCTGCCAG	TAGTCATATG	CTT-----	-----	-----				
Chordodes_morgani	-----	-----	---AAGCTTG	CTT-----	-----	-----				
Paractinolaimus_macrolaimus	-----	-----	-----	-----	-----	-----				
Aporcelaimellus_obtusicaudatus	-----	-----	-----	-----	-----	-----				
Wilsonema_schuermansstekhoveni	-----	-----	-----	-----	-----	-----				
Mesodorylaimus_sp_cf_nigritul	-----	-----	-----	-----	-----	-----				
Mesodorylaimus_bastiani	-----	-----	-----	-----	-----	-----				
Mesodorylaimus_japonicus	-----	-----	-----	-----	-----	-----				
Pungentus_sp._PDL_2005	-----	-----	-----	-----	-----	-----				
Allodorylaimus_sp	-----	-----	-----	-----	-----	-----				
Eudorylaimus_carteri	-----	-----	-----	-----	-----	-----				
Microdorylaimus_sp	-----	-----	-----	-----	-----	-----				
Longidorus_elongatus	-----	-----	-----	-----	-----	-----				
Xiphinema_rivesi	-----	-----	-----	-----	-----	-----				
Tylencholaimus_sp	-----	-----	-----	-----	-----	-----				
Anoplostoma_sp._BHMM_2005	-----	-----	-----	-----	-----	-----				
Adoncholaimus_fuscus	-----	-----	-----	-----	-----	-----				
Enoplus_meridionalis	-----	-----	GAATTCGTCG	ACAACCTGGT	TGATCCTGCC	AGTAGTCATA				
Enoplus_brevis_U88336	-----	-----	-----	--AGCCTGGT	TGATCCTGCC	AGTAGTCATA				
Enoplus_communis	-----	-----	-----	-----	-----	-----				
Enoploides_brunettii	-----	-----	-----	-----	-----	-----				
Syringolaimus_striatocaudatus	-----	-----	-----	-----	-----	-----				
Ironus_dentifurcatus	-----	-----	-----	-----	-----	-----				
Calyptronema_maxweberi	-----	-----	-----	-----	-----	-----				
Viscosia_sp._BHMM_2005	-----	-----	-----	-----	-----	-----				
Viscosia_viscosa	-----	-----	-----	-----	-----	-----				
Pontonema_vulgare	-----	-----	-----	--AACCTGGT	TGATCCTGCC	AGTAGTCATA				
Oncholaimus_sp._BHMM_2005	-----	-----	-----	-----	-----	-----				
Alaimus_sp._PDL_2005	-----	-----	-----	-----	-----	-----				
Prismatolaimus_intermedius	-----	-----	-----	-----	-----	-----				
Tobrilus_gracilis	-----	-----	-----	-----	-----	-----				
Tripyla_cf_filicaudata_JH_2004	-----	-----	-----	-----	-----	-----				
Bathylaimus_sp	-----	-----	-----	-----	-----	-----				
Bathylaimus_assimilis	-----	-----	-----	-----	-----	-----				
Bathylaimus_sp._BHMM_2005	-----	-----	-----	-----	-----	-----				
Tripyloides_sp._BHMM_2005	-----	-----	-----	-----	-----	-----				
Trischistoma_monohystera	-----	-----	-----	-----	-----	-----				
Mermis_nigrescens	-----	-----	-----	-----	-----	-----				
Mylonchulus_arenicolus	-----	-----	-----	-----	-----	-----				
Anatonchus_tridentacus	-----	-----	-----	-----	-----	-----				
Mononchus_truncatus	-----	-----	-----	-----	-----	-----				
Prionchulus_muscorum	-----	-----	-----	-----	-----	-----				
Clarkus_sp	-----	-----	-----	-----	-----	-----				
Trichinella_spiralis	-----	-----	-----	-----	-----	-----				
Trichinella_papuae	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA				
Trichinella_britovi	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA				
Trichinella_murrelli	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA				
Trichinella_pseudospiralis	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA				
Trichinella_nativa	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA				
Trichinella_zimbabwensis	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA				
Trichinella_nelsoni	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA				
Trichuris_suis	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA				
Trichuris_trichiura	-----	-----	-----	-----	-----	-----				
Trichuris_muris	GCTTGCATGC	CTGCAGGTCG	ACTCTAGAGG	ATCCCCGGGT	ACCGAGCTCG	AATTCAAGCT				
Tylolaimophorus_minor	-----	-----	-----	-----	-----	-----				
Paratrichodorus_pachydermus	-----	-----	-----	-----	-----	-----				
Paratrichodorus_anemones	-----	-----	-----	-----	-----	-----				
Trichodorus_primitivus	-----	-----	-----	-----	-----	-----				
Axonolaimus_helgolandicus	-----	-----	-----	-----	-----	-----				
Ascolaimus_elongatus	-----	-----	-----	-----	-----	-----				
Odontophora_rectangula	-----	-----	-----	-----	-----	-----				
Cylindrolaimus_sp._202149	-----	-----	-----	-----	-----	-----				
Tylocephalus_auriculatus	-----	-----	-----	-----	-----	-----				
Plectus_acuminatus_BS9	-----	-----	-----	-----	-----	-----				
Plectus_aquatilis	-----	-----	-----	-----	-----	-----				
Anaplectus_sp	-----	-----	-----	-----	-----	-----				
Anisakis_sp_WKT	-----	-----	-----	-----	-----	-----				
Anisakis_sp_Nadler	-----	-----	-----	-----	-----	-----				
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	GGTTATA				

[illegible]

	10 20 30 40 50
Daptonema_hirsutum	-----
Daptonema_normandicum	-----
Daptonema_oxycerca	-----
Daptonema_setosum	-----
Desmodora_communis	-----
Desmodora_ovigera	-----GA ATTCG--TCG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Metadesmolaimus_sp	-----
Dentostomella_sp	-----
Bunonema_franzi	-----T
Bunonema_sp	-----
Seleborca_complexa	-----
Acrobeloides_nanus	-----
Acrobeloides_bodenheimeri	-----
Acrobeloides_sp_PS1146	-----
Acrobeles_ciliatus	-----
Acrobeles_sp_PS1156	-----
Acrobeles_complexus_WCUG2	-----
Cephaloboides_sp_SB227	-----CTGA TTGATTCTGT CAGCGATGTA
Cephalobus_cubaensis	-----
Cephalobus_sp_PS1143	-----
Cephalobus_sp_PS1196	-----
Cephalobus_oryzae_PS1165	-----
Cervidellus_alutus	-----
Pseudacrobeles_variabilis	-----
Triligulla_aluta	-----
Zeldia_punctada	-----
Myolaimus_sp_U81585	-----
Rhabditophanes_sp_KR3021	-----
Brevibucca_sp_SB261	-----
Halicephalobus_gingivalis	-----
Panagrobelus_stammeri	-----
Plectonchus_sp_PDL0025	-----
Turbatrix_aceti	-----
Panagrellus_redivivus	-----
Panagrellus_redivivus_PS1163	-----
Panagrolaimus_subelongatus	-----
Panagrolaimus_davidi	-----
Panagrolaimus_cf_rigidus_AF40	-----T
Panagrolaimus_sp_Sourhope_ED2	-----
Panagrolaimus_sp_Sourhope_ED2	-----
Panagrolaimus_sp_Sourhope_ED2	-----
Panagrolaimus_sp_Sourhope_ED2	-----
Panagrolaimus_sp_Sourhope_ED2	-----
Steinernema_carpocapsae	-----
Strongyloides_stercoralis	-----ACCTGGT TGATCCTGCC AGTAGTCATA
Strongyloides_ratti	-----
Diploscapter_sp_PS1897	-----CGGTATA
Diploscapter_sp_PS2017	-----
Heterorhabditis_bacteriophora	-----
Heterorhabditis_hepialus	-----CGCTATA
Heterorhabditis_zelandica	-----CGCTATA
Parasitorhabditis_sp_SB281	-----GACGATA
Rhabditoides_inermiformis	-----
Rhabditoides_inermis_DF5001	-----GATCATA
Rhabditoides_regina_DF5012	-----GGTGATA
Poikilolaimus_oxycerca_SB200	-----
Poikilolaimus_regenfussi_SB199	-----
Distolabrellus_veechi_DWF1604	-----GGTCTGTC AGTGGCGATA
Distolabrellus_veechi_DF5024	-----GATTCTGTC AGTGGCGATA
Choriorhabditis_dudichi	-----
Protorhabditis_sp	-----CGGTATA
Protorhabditis_sp_DF5055	-----
Cruznema_tripartitum_DF5015	-----
Mesorhabditis_sp_PS1179	-----
Mesorhabditis_spiculigera_SB15	-----
Mesorhabditis_anisomorpha_SB12	-----
Teratorhabditis_palmarum_DF501	-----
Teratorhabditis_synpapillata_S	-----GCC GCTACCTGAT TGATTCTGTC AGTGGCGATA
Caenorhabditis_briggsae_PB102	-----
Caenorhabditis_elegans_N2	-----ATACCTGA TTGATTCTGT CAGCGCGATA
Caenorhabditis_sp_CB5161	-----
Caenorhabditis_sp_PS1010	-----CGCGATA
Caenorhabditis_japonica	-----CGCGATA
Caenorhabditis_sp_DF5170	-----GCGCGATA
Caenorhabditis_sp_SB341	-----CGCTATA
Caenorhabditis_plicata	-----CGCGATA
Caenorhabditis_vulgaris	-----
Caenorhabditis_sonorae	-----CGCTATA
NUCLEOTIDES INCLUDED	-----

	10	20	30	40	50
Caenorhabditis_drosophilae	-----	-----	-----	-----	--CGCGATA
Phasmarhabditis_hermaphrodita	-----	-----	-----	-----	-----
Phasmarhabditis_neopapillosa	-----	-----	-----	-----	-----
Phasmarhabditis_neopapillosa	-----	-----	-----	-----	--CGCTGTA
Pellioditis_mediterranea_SB173	-----	-----	-----	-----	--CGCTGTA
Pellioditis_marina	-----	-----	-----	-----	--CGCTGTA
Prodontorhabditis_wirthi	-----	-----	-----	-----	-T
Crustorhabditis_scanica	-----	-----	-----	-----	-----
Dolichorhabditis_sp_CEW1	-----	-----	-----	-----	-----
Oscheius_sp_DF5000	-----	-----	-----	-GA TTGATTCTGT	CAGCGCTGTA
Oscheius_sp_BW282	-----	-----	-----	-----	--CGCTGTA
Oscheius_myriophila_EM435	-----	-----	-----	-----	-----
Oscheius_insectivora	-----	-----	-----	-----	--CGCTGTA
Oscheius_dolichuroides	-----	-----	-----	-----	--CGCTGTA
Rhabditis_blumi_DF5010	-----	-----	-----	-----	-----
Rhabditis_sp_PS1191	-----	-----	-----	-----	--CGCTGTA
Rhabditis_sp_PS1010	-----	-----	-----	-----	-----
Rhabditis_myriophila_EM435	-----	-----	-----	-----	-----
Rhabditis_colombiana	-----	-----	-----	-----	-----
Rhabditella_axei_DF5006	-----	-----	-----	-----	-----
Rhabditella_sp_DF5044	-----	-----	-----	-----	--CGATGTA
Cuticularia_sp_PS2083	-----	-----	-----	-----	-----
Necator_americanus	-----	-----	-----	-----	-----
Kalicephalus_cristatus	-----	-----	--GTACCTGA	TTGATTCTGT	CAGCGCTATA
Ancylostoma_caninum	-----	-----	-----	-----	--CGCTATA
Angiostrongylus_cantonensis	-----	-----	-----	-----	-----
Angiostrongylus_costaricensis	-----	-----	-----	-----	-----
Angiostrongylus_malaysiensis	-----	-----	-----	-----	-----
Angiostrongylus_dujardini	-----	-----	-----	-----	-----
Angiostrongylus_vasorum	-----	-----	-----	-----	-----
Aulurostrongylus_abstrusus	-----	-----	-----	-----	--CGATATA
Didelphostrongylus_hayesi	-----	-----	-----	-----	---TATAT
Crenosoma_mephitidis	-----	-----	-----	-----	---TATAT
Crenosoma_vulpis	-----	-----	-----	-----	--CGCTATA
Otostrongylus_circumlitus	-----	-----	-----	-----	-----
Troglostrongylus_wilsoni	-----	-----	-----	-----	-----
Filaroides_martis	-----	-----	-----	-----	-----
Oslerus_osleri	-----	-----	-----	-----	-----
Parafilaroides_decorus	-----	-----	-----	-----	-----
Metastrongylus_salmi	-----	-----	-----	-----	-----
Metastrongylus_elongatus	-----	-----	-----	-----	--CGCTATA
Skrjabinigylus_chitwoodrum	-----	-----	-----	-----	---TATA
Parelaphostrongylus_odocoilei	-----	-----	-----	-----	-----
Muelerius_capillaris	-----	-----	-----	-----	-----
Protostrongylus_rufescens	-----	-----	-----	-----	--CGCTATA
Halocercus_invaginatus	-----	-----	-----	-----	-----
Pseudalius_inflexus	-----	-----	-----	-----	CAGCGCTATA
Stenurus_minor	-----	-----	-----	-----	-----
Torynurus_convolutus	-----	-----	-----	-----	-----
Syngamus_trachea	-----	-----	--TACCTGA	TTGATTCTGT	CAGCGCTATA
Stephanurus_dentatus	-----	-----	-----	-----	--CGCTATA
Nematodirus_battus	-----	-----	-----	-----	--CGCTATA
Strongylus_equinus	-----	-----	-----	-----	---TAAGCC
Labiostromgylus_bipapillosus	-----	-----	--TACCTGA	TTGATTCTGT	CAGCGCTATA
Petrovinema_poculatum	-----	-----	-----	-----	--CGCTATA
Cylicocyclus_insignis	-----	-----	-----	-----	--CGCTATA
Chabertia_ovina	-----	-----	-----	-----	--CGCTATA
Cyclodontostomum_purvisi	-----	-----	-----	-----	--CGCTATA
Zoniolaimus_mawsonae	-----	-----	-----	-----	--CGCTATA
Hypodontus_macropi	-----	-----	--TACCTGA	TTGATTCTGT	CAGCGCTATA
Deletrocephalus_dimidiatus	-----	-----	-----	-----	--CGCTATA
Dictyocaulus_eckerti_P7B8	-----	-----	-----	-----	-----
Dictyocaulus_capreolus_P3B2	-----	-----	-----	-----	-----
Dictyocaulus_sp._P6A1	-----	-----	-----	-----	-----
Dictyocaulus_filaria	-----	-----	-----	-----	-----
Dictyocaulus_capreolus_P2C10	-----	-----	-----	-----	-----
Dictyocaulus_viviparus	-----	-----	-----	-----	-----
Haemonchus_sp._V3091	-----	-----	-----	-----	-----
Haemonchus_contortus	-----	-----	--TACCTGA	TTGATTCTGT	CAGCGCTATA
Haemonchus_placeii	-----	-----	--TACCTGA	TTGATTCTGT	CAGCGCTATA
Haemonchus_similis	-----	-----	--TACCTGA	TTGATTCTGT	CAGCGCTATA
Ostertagia_ostertagi	-----	-----	-----	-----	--CGCTATA
Ostertagia_leptospicularis	-----	-----	-----	-----	--CGCTATA
Nippostrongylus_brasileinsis	-----	-----	-----	-----	--CGCTATA
Heligmosomoides_polygyrus	-----	-----	-----	-----	--CGCTATA
Trichostrongylus_colubriformis	-----	-----	-----	-----	--CGCTATA
Tetrabothriostrongylus_mackerr	-----	-----	-----	-----	--CGCTATA
Herpestrongylus_pythonis	-----	-----	-----	-----	--CGCTATA
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----

	10	20	30	40	50	
Nicollina_cameroni	-----	-----	-----	-----	-----	---CGCTATA
Filarimena_flagrifer	-----	-----	-----	-----	-----	---CGCTATA
Amidostomum_cygni	-----	-----	-----	-----	-----	---CGCTATA
Teratocephalus_lirellus	-----	-----	-----	-----	-----	
Brumptaemilius_justini	-----	-----	-----	-----	-----	
Anguillicola_crassus	-----	-----	-----	-----	-----	
Dracunculus_medinensis	-----	-----	-----	-----	-----	
Dracunculus_oesophageus	-----	-----	-----	-----	-----	
Dracunculus_sp._V3104	-----	-----	-----	-----	-----	
Philonema_sp_A	-----	-----	-----	-----	-----	
Philometra_obturans	-----	-----	-----	-----	-----	
Camallanus_oxycephalus	-----	-----	-----	-----	-----	
Acanthocheilonema_viteae	-----	-----	-----	-----	-----	
Loa_loa	-----	-----	-----	-----	-----	
Onchocerca_cervicalis	-----	-----	-----	-----	-----	
Dirofilaria_immitis	-----	-----	-----	-----	-----	
Brugia_malayi	-----	-----	-----	-----	-----	
Wuchereria_bancrofti	-----	-----	-----	-----	-----	
Litomosoides_sigmodontis	-----	-----	-----	-----	-----	
Setaria_digitata	-----	-----	-----	-----	-----	
Gnathostoma_turgidum	-----	-----	---TCTCCGAT	TGATTCTGTC	GGCGATTATA	
Gnathostoma_neoprocyonis	-----	-----	---TCTCCGAT	TGATTCTGTC	GGCGATTATA	
Gnathostoma_binucleatum	-----	-----	---TCTCCGAT	TGATTCTGTC	GGCGATTATA	
Physaloptera_alata	-----	-----	-----	-----	-----	
Physaloptera_turgida	-----	-----	-----	-----	-----	
Ascarophis_arctica	-----	-----	-----	-----	-----	
Spinitectus_carolini	-----	-----	-----	-----	-----	
Thelazia_lacrymalis	-----	-----	-----	-----	-----	
Aphelenchus_avenae	-----	-----	-----	-----	-----	
Aphelenchoides_fragariae	-----	-----	-----	-----	-----	
Bursaphelelenchus_sp	-----	-----	-----	-----	-----	
Deladenus_sp	-----	-----	-----	-----	-----	
Criconema_sp	-----	-----	-----	-----	-----	
Hemicycliophora_conida	-----	-----	-----	-----	-----	
Paratylenchus_dianthus	-----	-----	-----	-----	-----	
Pratylenchus_thornei	-----	-----	-----	-----	-----	
Tylenchulus_semipetrans	-----	-----	-----	-----	-----	
Ditylenchus_angustus	-----	-----	-----	-----	-----	
Subanguina_radicola	-----	-----	-----	-----	-----	
Tylenchorhynchus_maximus	-----	-----	-----	-----	-----	
Geocenamus_quadriifer	-----	-----	-----	-----	-----	
Globodera_pallida	-----	-----	-----	-----	-----	
Scutellonema_bradys	-----	-----	-----	-----	-----	
Helicotylenchus_dihystera	-----	-----	-----	-----	-----	
Rotylenchus_robustus	-----	-----	-----	-----	-----	---CA
Meloidogyne_arenaria	-----	-----	---ATATCTGGT	TGATCTGCCC	TGAACTGATG	
Meloidogyne_incognita_KT	-----	-----	-----	-----	-----	
Meloidogyne_javanica	-----	-----	-----	-----	-----	
Meloidogyne_artiellia	-----	-----	-----	-----	-----	
Meloidogyne_duytsi	-----	-----	-----	-----	-----	
Meloidogyne_exigua	-----	-----	-----	-----	-----	
Meloidogyne_hapla	-----	-----	-----	-----	-----	
Meloidogyne_ichinohei	-----	-----	-----	-----	-----	---G
Meloidogyne_maritima	-----	-----	-----	-----	-----	
Meloidogyne_microtyla	-----	-----	-----	-----	-----	
Nacobbus_aberrans	-----	-----	-----	-----	-----	
Pratylenchoides_ritteri	-----	-----	-----	-----	-----	
Pratylenchoides_magnicauda	-----	-----	-----	-----	-----	
Hirschmanniella_sp._JH_2003	-----	-----	-----	-----	-----	
Radopholus_similis	-----	-----	-----	-----	-----	
Pratylenchus_goodeyi_VF	-----	-----	-----	-----	-----	
Boleodorus_thylactus_clone2	-----	-----	-----	-----	-----	---T
Philippine Sequence 1	-----	-----	-----	-----	-----	
Philippine Sequence 2	-----	-----	-----	-----	-----	
Philippine Sequence 3	-----	-----	-----	-----	-----	
Philippine Sequence 4	-----	-----	-----	-----	-----	
Philippine Sequence 5	-----	-----	-----	-----	-----	
Philippine Sequence 6	-----	-----	-----	-----	-----	
Philippine Sequence 7	-----	-----	-----	-----	-----	
Tahiti Sequence 1	-----	-----	-----	-----	-----	
Tahiti Sequence 2	-----	-----	-----	-----	-----	
Thailand Sequence 1	-----	-----	-----	-----	-----	
Ogasawara Sequence 1	-----	-----	-----	-----	-----	
Ogasawara Sequence 2	-----	-----	-----	-----	-----	
Ivory Coast Sequence 1	-----	-----	-----	-----	-----	
Ivory Coast Sequence 2	-----	-----	-----	-----	-----	
Ivory Coast Sequence 3	-----	-----	-----	-----	-----	
Ivory Coast Sequence 4	-----	-----	-----	-----	-----	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	

	60	70	80	90	100	110	120
Gordius aquaticus	----	GTCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-TA-	--ACTTA-AA
Priapulius caudatus	----	GTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C-AGAC-GG-	--ATACA-AA
Brachionus plicatilis	----	GTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C-ATAC-CT-	--TAGCA-CG
Chordodes morgani	----	GTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AAAC-TT-	--ACTAC-AA
Paractinolaimus macrolaimus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG	
Aporcelaimellus obtusicaudatus	-----	---	GAT-AAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG
Wilonema schuurmansstekhoveni	-----	CTC	AAAGATTAAG	CCATGCATGT	CTAT-GC--A	C-AAGC-CG-	--ATTAA-TG
Mesodorylaimus sp cf nigritul	-----	---	GATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG
Mesodorylaimus bastiani	-----	TCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TTATA-AG
Mesodorylaimus japonicus	-----	CTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CA-	--TTTTA-TG
Pungentus sp. PDL 2005	-----	TTT	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAAAA-CG
Allodorylaimus sp	-----	CTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG
Eudorylaimus carteri	-----	TCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG
Microdorylaimus sp	-----	TC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG
Longidorus elongatus	--GCTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG	
Xiphinema rivesi	-----	---	TAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG
Tylencholaimus sp	-----	CC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CT-	--TAATA-CG
Anoplostoma sp. BHMM 2005	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CG-	--ATTAA-TG
Adoncholaimus fuscus	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	CTAG-GC--A	T-AAAT-GA-	--ATCAA-TT
Enoplus meridionalis	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C-ATAC-TG-	--ATTAA-TA	
Enoplus brevis U88336	T-CTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C-ACAC-TG-	--ATTAA-TA	
Enoplus communis	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C-ACAC-TG-	--ATTAA-TA
Enoploides brunettii	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	CTCA-GT--A	C-ATAC-TG-	--ATTAA-TA
Syringolaimus striatocaudatus	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	T-GAGC-CT-	--CAAAA-TG
Ironus dentifurcatus	-----	CTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-CG-	--ATTAA-AG
Calyptronema maxweberi	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-TA-	--TTTAA--
Viscosia sp. BHMM 2005	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	CTCA-GC--A	T-AAAT-AA-	--ATTCCATT
Viscosia viscosa	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAAT-GA-	--ATTCA-TA
Pontonema vulgare	TGCTTGTCTC	AAAGACTTAAG	CCATGCATGT	CTAA-GC--A	C-AAAC-TA-	--TT-AA-TT	
Oncholaimus sp. BHMM 2005	----	TGCTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-GAAC-TA-	--TCTAG--
Alaimus sp PDL 2005	-----	---	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ACGC-CG-	--ATTAAA-TG
Prismatolaimus intermedius	-----	---	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-CT-	--AAGAA-TG
Tobrilus gracilis	-----	CTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-CC-	--TTGAA-TG
Tripyla cf. filicaudata JH 2004	-----	CTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--	--AGAGC-CC-	--TTGAA-TG
Bathylaimus sp	-----	TCTC	AAAGATTAAG	CCATGCAAGT	CTTA-GC--A	C-AAGC-CG-	--TTAGTATG
Bathylaimus assimilis	-----	TCTC	AAAGATTAAG	CCATGCAAGT	CTTA-GC--A	C-AAGC-CG-	--TTAGTATG
Bathylaimus sp. BHMM 2005	----	TGCTCTC	AAAGATTAAG	CCATGCAAGT	CTAA-GC--A	C-AAGC-CG-	--TCAGTATG
Tripyloides sp. BHMM 2005	----	TGCTCTC	AAAGATTAAG	CCATGCAAGT	CTAA-GC--A	C-AAGC-CG-	--TTAAA-TG
Trischistoma monohystera	-----	TCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-CG-	--AA-CAATG
Mermis nigrescens	-GCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-C--	--TTAAAACG	
Mylonchulus arenicolus	-----	---	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	C-ATGC-C--	--TTAAAACG
Anatonchus tridentatus	-----	---	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	C-ATGC-C--	--TTAAAATG
Mononchus truncatus	-----	TCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-ATGC-C--	--TTAAAATG
Prionchulus muscorum	-----	---	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	C-ATGC-C--	--TTAAAATG
Clarkus sp	-----	TCTC	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	C-ATGC-C--	--TTAAAACG
Trichinella spiralis	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	C-ACAC-TGT	T-TTAAAA--	
Trichinella papuae	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	C-ACAC-TGT	T-TTAAAA--	
Trichinella britovi	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	C-ACAC-TGT	T-TTAAAA--	
Trichinella murrelli	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	GTAA-GC--A	C-ACAC-TGT	T-TTAAAA--	
Trichinella pseudospiralis	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	GTAA-GC--A			

	70	80	90	100	110	120	
Odontophora_rectangula	-----	-----	-----	-----	--aagc-ct-	--aata--ag	
Cylindrolaimus_sp._202149	TGGCTGTCTC	AAAGATTAAG	CCATGCATGT	CTAT-GC--A	C-AAGC-CA-	--AATTA-AG	
Tylocephalus_auriculatus	-----	-----	-----CATGT	CTAA-GC--A	C-AAGC-CG-	--ATTAA-TG	
Plectus_acuminatus_BS9	-----	-----	-----	-----	--AAGC-CG-	--TTTAA-TG	
Plectus_aquatilis	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAT-GC--A	C-AAGC-CG-	--ATAA--TG	
Anaplectus_sp	-----CTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-CG-	--ATAA--CG	
Anisakis_sp_WKT	-----	-----	-----TGT	CTAA-GTTCA	AATGGC-CT-	--ATAGA-AG	
Anisakis_sp_Nadler	TGCTTATCTC	GAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAGA-AG	
Contracaecum_multipapillatum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAT-GTTCA	AATGGC-CT-	--ATAA--AG	
Pseudoterranova_decipiens	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAGA-AG	
Raphidascaris_acus	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	--ATAAAA--G	
Terranova_caballeri	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAA--AG	
Ascaris_suum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAA--AG	
Ascaris_lumbricoides	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--AAAA--AG	
Baylisascaris_procyonis	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAA--AG	
Baylisascaris_transfuga	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAA--AG	
Parascaris_equorum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAA--AG	
Porrocaecum_depressum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAA--AG	
Toxascaris_leonina	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--ATAA--AG	
Heterocheilus_tunicatus	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTCA-GTTCA	AATGGC-CT-	--TAA--AG	
Goezia_pelagia	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	--TTAA--AG	
Hysterothylacium_fortalezae	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	--AAA--AG	
Hysterothylacium_pelagicum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	--ATAA--AG	
Hysterothylacium_reliquens	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	--ATAA--AG	
Iheringascaris_inquies	TGCTTATCTG	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	--ATAA--AG	
Toxocara_canis	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	--TTAA--AG	
Nemhelix_bakeri	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GTTCA	AATAAC-CT-	--ATAA--TG	
Raillietnema_sp._V3060	-----	--AGGCTAAG	CCATGCATGT	CTAA-GTTCA	TATGAC-CT-	--TTAA--AG	
Cruzia_americana	TGCTTGTCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAAC-CT-	--ATAA--TG	
Heterakis_sp_14690	TGCTTGTCTC	AAAGGCTAAG	CCATGCATGT	GTAT-GTTCA	AATAAC-CT-	--ATAA--AG	
Heterakis_gallinarum	TGCTTGTCTC	AAAGGCTAAG	CCATGCATGT	GTAT-GTTCA	AATAAC-CT-	--ATAA--AG	
Paraspidodera_sp_21303	TGCT--GTCTC	AAAGGCTAAG	CC--TGCAATGT	CTAA-GTTCA	AATAAC-CT-	--TTAA--TG	
Chromodora_nudicapitata	----TGCTC	AAAGATTAAG	CCATGCATGT	GTAA-GA--A	T-AAAC-CG-	--ATAA--TG	
Chromadora_sp._BHMM_2005	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	T-AAAC-CG-	--ATAA--TG	
Atrochromadora_microalima	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	T-AAAC-CG-	--ATAA--TG	
Chromadorina_germanica	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	T-AAAC-CG-	--ATAA--TG	
Chromadorita_tentabundum	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--ATAA--TG	
Dichromadora_sp._BHMM_2005	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	T-AAAC-CG-	--ATAA--TG	
Neochromadora_BHMM_2005	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAAC-CG-	--ATAA--TG	
Spilophorella_paradoxa	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--ATAA--TG	
Paracanthonchus_caecus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--ATTAA-TG	
Paracyatholaimus_intermedius	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-CA-	--TAGAA-TG	
Praeacanthonchus_punctatus	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--ATTAA-TG	
Praeacanthonchus_sp	-----YMC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--ATAAA-TG	
Cyatholaimus_sp._BHMM_2005	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--ATTAA-TG	
Spirinia_parasitifera	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--AATAA-TG	
Acanthopharynx_micans	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CA-	--AATAA-TG	
Xyzzors_sp	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--ATTAA-TG	
Metachromadora_sp	----CTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--AATTA-TG	
Metachromadora_remanei	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--AATTA-TG	
Catanema_sp	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--AATTA-TG	
Eubostrichus_dianae	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAAAGC--A	T-GAGC-CG-	--ATTTA-TG	
Eubostrichus_topiarus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--ATAAA-TG	
Eubostrichus_parasitiferus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--AATAA-TG	
Laxus_oneistus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--AATTA-TG	
Laxus_cosmopolitus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--AATTA-TG	
Leptonemella_sp	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AGCC--G	--AATTA-TG	
Robbea_hypermnestra	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--AATTA-TG	
Stilbonema_majum	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	--AATTA-TG	
Monoposthia_costata	----TGCTC	AAAGATTAAG	CCATGCATGT	GCAA-GT--A	T-GAAC-CA-	--ATTGAACT	
Nudora_bipapillata	----TGCTC	AAAGATTAAG	CCATGCATGT	TTAA-GC--A	C-AAGC-CG-	--ATTAA-TG	
Calomicrolaimus_parahonestus	----TGCTC	AAAGATTAAG	CCATGCACGT	TTAA-AT--A	C-AAGC-CT-	--CAAAA-TG	
Calomicrolaimus_sp._BHMM_2005	----TGCTC	AAAGATTAAG	CCATGCACGT	TTAA-AT--A	C-AAGC-CT-	--CAAAA-TG	
Molgolaimus_demani	----TGCTC	AAAGATTAAG	CCATGCACGT	TTAT-AT--A	C-AAAC-CT-	--TCAAA-TG	
Diplogaster_lethieri	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	C-ATAT-GT-	--GTAAA-C	
Aduncospiculum_haicti	-----	-----	-----	-----	G-ATAG-TT-	--CTTGA--A	
Pristionchus_lheritieri	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	C-ATAT-GT-	--GTAAA-C	
Pristionchus_pacificus	-----	-----	-----TGT	CTAA-GA--A	C-ATAT-GT-	--GTAAA-C	
Pristionchus_pacificus	-----A	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	C-ATAT-GT-	--GTAAA-C	
Sabatieria_punctata_STRAIN_343	----TGCTC	AAAGATTAAG	CCATGCATGT	TTAA-GC--A	G-AAGC-CG-	--CACAA-TG	
Sabatieria_sp._355_BHMM_2005	----TGCTC	AAAGATTAAG	CCATGCATGT	TTAA-GC--A	G-AAGC-CG-	--CACAA-TG	
Sabatieria_celtica	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	--TACAC--G	
Sabatieria_punctata_STRAIN_200	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	--AACAA-CG	
Sabatieria_punctata_STRAIN_223	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	--AACAA-TG	
Sabatieria_sp._210_BHM_2005	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	--AACAA-CG	
Setosabatieria_hilarula	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	--TACTA-CG	
Desmolaimus_zeelandicus	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-TG-	--AATA--CA	
Terschellingia_longicaudata	----TGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AAGC-TA-	--AATTA-AA	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	70	80	90	100	110	120
Cyartonema_elegans	---TGTCTC	AAAGATTAAG	CCATGCATGT	CTCA-GT--A	C-AAGC-TG-	--AAATA-AA
Tridentulus_sp	-----TCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-A-GC-TG-	--GAAA--CA
Diplolaimelloides_meyli	-----	-----	---CATGCATGT	CTAA-GC--A	T-GCGC-TG-	--AAGTA-AA
Diplolaimella_diavengatensis	-----	-----	---TGCATGT	CTAA-GC--A	T-GCGT-TG-	--AATTT-AA
Geomonhystera_disjuncta	-----CTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-TA-	--AATTA-AA
Sphaerolaimus_hirsutus	-----TGTCTC	AAAGATTAAG	CCATGCACGT	GCAA-GC--A	C-AAGC-TG-	--AATTA-AA
Theristus_acer	-----C	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAAC-TA-	--AAGTA-AA
Daptonema_procerus	TGCTTGCTC	AAAGATTAAG	CCATGCATGT	CTAC-GC--A	T-AAAC-GA-	--ATCTA-AA
Daptonema_hirsutum	-----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAC-GC--A	T-AAAC-GA-	--ATCTA-AA
Daptonema_normandicum	-----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAAC-TG-	--AACTA-AA
Daptonema_oxycerca	-----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAC-GC--A	T-AAAC-GA-	--ATCTA-AA
Daptonema_setosum	-----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAC-GC--A	T-AAAC-GA-	--ATCTA-AA
Desmodora_communis	-----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--TATAA-TG
Desmodora_ovigera	TGCTTGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	--AATAA-TG
Metadesmolaimus_sp	-----TTT	AAAGATTAAG	CCATGCATGT	CTAC-GC--A	T-AAAC-GA-	--ATCTA-AA
Dentostomella_sp	-----	-----TTAAG	CCATGCATGT	-TAA-GT--T	T-AAAT-AC-	--ACATT-AA
Bunonema_franzi	GCTT-GTCTC	AAAGATTAAG	CCATGCATGT	TATA-AA--A	C-TTGC-CC-	--TAGAA-TG
Bunonema_sp	-----	-----	-----	-----	-----	-----G
Seleborca_complexa	-----	-----	-----TGT	CTAA-GT--A	T-AAAC-GA-	--TTTAA-TC
Acrobeloides_nanus	-----	--AGATTAAG	CCATGCATGT	CTAA-GT--A	T-AAAC-GA-	--TTTAA-TC
Acrobeloides_bodenheimeri	-----	-----	-----	-----	--AAAC-GG-	--CTTGT-CC
Acrobeloides_sp_PS1146	-----	-----	-----TGT	CTAA-GT--A	T-AAAC-GA-	--TTTAA-TC
Acrobeles_ciliatus	-----	-----	-----	-----	--AAAC-GA-	--TTTAA-TC
Acrobeles_sp_PS1156	-----	-----	-----	-----	--AAAC-GA-	--TTTAA-TC
Acrobeles_complexus_WCUG2	-----	-----	-----	-----	--AAAC-GA-	--TTTAA-TC
Cephaloboides_sp_SB227	TGCTCAAGTA	AAAGATTAAG	CCATGCATGC	ATA--GT--T	C-AAAT-TT-	--T-----TG
Cephalobus_cubaensis	-----	-----	-----	-----	--AAAC-GA-	--CTCGA-TC
Cephalobus_sp_PS1143	-----	-----	-----	-----	--AAAC-GA-	--TTTAA-TC
Cephalobus_sp_PS1196	-----	-----	-----	-----	--AAAC-GA-	--TTTAA-TC
Cephalobus_oryzae_PS1165	-----	-----	-----TGT	CTAA-GT--A	T-AAAC-GA-	--TTTAA-TC
Cervidellus_alutus	-----	-----	-----	-----	--AAAC-GA-	--TTTAA-TC
Pseudacrobeles_variabilis	-----	-----	-----	-----	--AAAC-GA-	--TTTAA-TC
Triligulla_aluta	-----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AAAC-GA-	--TTTAA-TC
Zeldia_punctada	-----	-----	-----	-----	-----GA-	--TTTAA-TC
Myolaimus_sp_U81585	-----	-----	-----TGT	CTCA-GT--A	C-AAAC-CA-	--CTTGT-TG
Rhabditophanes_sp_KR3021	-----	-----	-----	-----	-----AA-	--CGTTC-AC
Brevibucca_sp_SB261	TGCTTGCTC	AAAGATTAAG	CCATGCATGT	CTCA-GT--G	C-AGAC-TA-	--TTTGA-AA
Halicephalobus_gingivalis	-----C	AAAGATTAAG	CCATGCATGT	CTAA-GT--	---ACA-TG-	--TGAGT-AC
Panagrobilus_stammeri	-----TC	AAAGATTAAG	CCATGCATGT	TTAA-GT--G	C---A-TG-	--GCTTGC-AC
Plectonchus_sp_PDL0025	-----	-----	--ATGCATGT	CTAA-GT--G	C---A-TG-	--GTTTAT-AC
Turbatrix_aceti	-----TG	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C---TT-GC-	--ATATAA-AT
Panagrellus_redivivus	-----	-----TAG	CCATGCATGT	CTAA-GT--A	C--ATG-TA-	--GCTTGC-CT
Panagrellus_redivivus_PS1163	-----	-----	--ATGCATGT	CTAA-GT--A	C--ATG-TA-	--GCTTGC-CT
Panagrolaimus_subelongatus	-----CTC	AAAGATTAAG	CCATGCATGN	CTAA-GT--A	C--ATG-TG-	--TAATTTAAAC
Panagrolaimus_davidi	-GCTTGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C--ATG-TG-	--TAATTTAAAC
Panagrolaimus_cf_rigidus_AF40	GCTTGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C--ATG-TG-	--TAATTTAAAC
Panagrolaimus_sp_Sourhope_ED2	-----TCC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	C--ATG-TG-	--TATATAAC
Panagrolaimus_sp_Sourhope_ED2	-----	-----	-----	-----	-----	-----AAC
Panagrolaimus_sp_Sourhope_ED2	-----	-----	-----	-----	-----	-----ATATAAC
Panagrolaimus_sp_Sourhope_ED2	-----	-----	-----	-----	-----	-----AAC
Steinernema_carpocapsae	-----	-----	-----G	TTAA-GT--T	T---AAA-C-	--GAATCA-AA
Strongyloides_stercoralis	TGCTTGCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T---AAA-CA-	--AATTCATACT
Strongyloides_ratti	-----	-----	-----TGT	GTAA-GT--A	C--AAT-GT-	--TTTAA-AC
Diploscapter_sp_PS1897	TGCTCAAGTA	AAAGATTAAG	CCATGCATGC	TC---GC--A	C-AACT-CG-	--ATTAT-GA
Diploscapter_sp_PS2017	-----	-----	-----TGC	TC---GC--A	C-AACT-CG-	--ATTAT-TA
Heterorhabditis_bacteriophora	-----	AAAGATTAAG	CCATGCATGT	ACT--GT--T	-----CA-	--ACTAA-CG
Heterorhabditis_hepialus	TGCTTACGTA	AAAGACTAAG	CCATGCATGT	ACTG-TT--C	-----	--AGCTTACG
Heterorhabditis_zelandica	TGCTTACGTA	AAAGACTAAG	CCATGCATGT	ACT--GT--T	C-AGCC-AA-	-----CG
Parasitorhabditis_sp_SB281	TGCTTATGTA	GAAGATTAAG	CCATGCATAT	GTCA-GT--T	C-CTGC-CC-	--TCGCA-CG
Rhabditoides_inermiformis	-----	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AGCG-TG-	--CAA---AC
Rhabditoides_inermis_DF5001	TGCTCAGTTA	GAAGATTAAG	CCATGTACGT	CTAA-GT--T	C-ATCT-CT-	--CG---AA
Rhabditoides_regina_DF5012	CGCTTATGTC	AAAGATTAAG	CCATGCATGT	ATTC-GT--A	C-GACC-AT-	--CTCA--TG
Poikilolaimus_oxycerca_SB200	-----GTA	AAAGATTAAG	CCATGCACGT	GTAA-GT--T	C-ACAT-CT-	--TTTA--TG
Poikilolaimus_regenfussi_SB199	-----	---GATTAAG	CCATGCACGT	TTAA-GT--T	CACGCT-GA-	--TTAA--CG
Distolabrellus_veechi_DWF1604	TGCTTATGTA	-GAGATTAAG	CCATGCATGT	GTAA-TT--A	TTAAC-CAT-	--CTAA--TG
Distolabrellus_veechi_DF5024	TGCTTATGTA	TGAGATTAAG	CCATGCATGT	ATAT-GT---	TCAAC-CAT-	--CTAA--TG
Choriorhabditis_dudichi	-----	-----	---TTAAGCCAT	GCAT-GT-AA	WAGTA-CAT-	--CGRA--TA
Protorhabditis_sp	TGCTTAAGTA	AAAGATTAAG	CCATGCATGT	TCGT-TC--A	TCT---CGA-	--TTAT--GA
Protorhabditis_sp_DF5055	-----	-----	--ATGCATGC	ATAA-GT---	T-CAG-CTT-	--TATC--AA
Cruznema_tripartitum_DF5015	-----	-----	-TGC-GT--G	T-TCA-ACG-	T-----	-----AA
Mesorhabditis_sp_PS1179	-----	-----	-----TAT	CTTA-GT--T	C-AGCC-CT-	--TTTA--CG
Mesorhabditis_spiculigera_SB15	-----AAGTA	AAAGATTAAG	CCATGCATAT	CTAA-GT--T	C-CCAC-CC-	--TCGCT-CG
Mesorhabditis_anisomorpha_SB12	-----	AAAGATTAAG	CCATGCATAT	GTAA-GT--C	C-AACC-AAT	--TTAT--CG
Teratorhabditis_palmarum_DF501	-----	---GATTAAG	CCATGCATGT	ATTC-GA--A	C-GGTC-ATT	--TTAT--G
Teratorhabditis_sympyallata_S	TGCTTATGTT	AAAGATTAAG	CCATGCATGT	ATTA-GA--A	C-GGTC-TC-	--AC-----G
Caenorhabditis_briggsae_PB102	-----	---GATTAAG	CCATGCATG-	CTTT-G---A	T-TCAT-CA-	--A-----
Caenorhabditis_elegans_N2	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	CTTT-G---A	T-TCAT-CA-	--A-----
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	70	80	90	100	110	120
Caenorhabditis_sp_CB5161	-----	---GATTAAG	CCATGCATG-	CTTT-G---	A T-TCAT-CA-	--A-----
Caenorhabditis_sp_PS1010	TGCTCAAGTA	AA-GATTAAG	CCATGCATG-	CTT--G---	A T-TCAT-CA-	--A-----
Caenorhabditis_japonica	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	CTTT-G---	A T-TCAT-AA-	--A-----
Caenorhabditis_sp_DF5170	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	CTA--G---	A T-TCAT-CA-	--A-----
Caenorhabditis_sp_SB341	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	TTG--G---	T--CAT-CG-	--TAAG----
Caenorhabditis_plicata	TGCTTAAGTA	AAAGATTAAG	CCATGCATG-	CTTT-G---	A T-TCAT-AA-	--A-----
Caenorhabditis_vulgaris	-----	---GATTAAG	CCATGCATG-	CTTT-G---	A T-TCAT-CA-	--A-----
Caenorhabditis_sonorae	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	TTG--G---	T-CATC-GT-	--AA-----
Caenorhabditis_drosophilae	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	CTG--G---	A T-TCAT-CA-	--A-----
Phasmarhabditis_hermaphrodita	-----GCT	TGCTCTCAAAG	ATTAAGCCAT	GCA--TG---	---TCGAGTT	TATATATAAA
Phasmarhabditis_neopapillosa	-----GCT	TGCTCTCAAAG	ATTAAGCCAT	GCA--TG---	---TCGAGTT	CATTTT---A
Phasmarhabditis_neopapillosa	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--	T C-AACT-TT-	-----T
Pellioiditis_mediterranea_SB173	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--	T C-AACT-TT-	-----A
Pellioiditis_marina	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--	T C-AACT-TT-	-----T
Prodontorhabditis_wirthi	TGCTCAAGTA	AAAGATTAAG	CCATGCATGC	A--CAGT--	C C-AATC-GA-	---CTTTTCGA
Crustorhabditis_scanica	-----	---GATTAAG	CCATGCATGA	TGTCAGT--	G C-GACC-AC-	--ACGTATTG
Dolichorhabditis_sp_CEW1	-----	---CGCATGT	AAAAAGT--	T C-AACT-TT-	--AT-----	A
Oscheius_sp_DF5000	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--	T C-AACC-TT-	--AT-----G
Oscheius_sp_BW282	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--	T C-AACT-TT-	--TT-----
Oscheius_myriophila_EM435	-----	---GATTAAG	CCATGCATGT	AAAA-GT--	T C-AACT-TT-	--T-----
Oscheius_insectivora	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	ATAAAGT--	A C-AGCT-TT-	--TA-----
Oscheius_dolichuroides	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	GAAAAAGT--	T C-AACC-TT-	--AT-----G
Rhabditis_blumi_DF5010	-----	---GACTAAG	CCATGCATGT	CTCA-G---	A T-CCCC-AT-	--TT-----CA
Rhabditis_sp_PS1191	TGCTCACGCT	AAAGATTAAG	CCATGCAAGC	TTGAGAT--	C-ACCC-TT-	--TT-----
Rhabditis_sp_PS1010	-----	---TG-	CTT--G---	A T-TCAT-CA-	--A-----	
Rhabditis_myriophila_EM435	-----	---GATTAAG	CCATGCATGT	A-AAAAAGT--	T C-AACT-TT-	--T-----
Rhabditis_colombiana	-----	---AAAGT--	T A-GCTT-TT-	A-A-----	AA	
Rhabditella_axei_DF5006	-----	---GATTAAG	CCATGCATGT	AT--AGT--	T C-ATTT-TT-	--T-----
Rhabditella_sp_DF5044	TGCTCACGTA	AAAGATTAAG	CCATGCATGT	AT--AGT--	T C-ATTT-TT-	--T-----
Cuticularia_sp_PS2083	-----	---CGT	GT-AAGT--	T C-ACAT-CT-	---TTTA--	TG
Necator_americanus	-----	---ATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT--A-	-----AA
Kalicephalus_cristatus	TGCTCAGTTT	AAAGACTAAG	CCATGCATGT	CGA--GT--	T C-ATCTTT--	-----GA
Ancylostoma_caninum	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT--A-	-----AA
Angiostrongylus_cantonensis	-----	---ATTAAG	CCATGCATGA	GGA--GT--	T C-AGCT-TT-	-----AA
Angiostrongylus_costaricensis	-----	-----A	GGA--GT--	T C-AGCT-TC-	-----AA	
Angiostrongylus_malaysiensis	-----	-----A	GGA--GT--	T C-AGCT-TC-	-----AA	
Angiostrongylus_dujardini	-----	-----A	GGA--GT--	T C-AGCT-TC-	-----AA	
Angiostrongylus_vasorum	-----	-----A	GGA--GT--	T C-AGCT-TC-	-----AA	
Aulurostrongylus_abstrusus	TGTTCAAGTA	AAAGGTTAAG	CCATGCACGA	GGA--GT--	T C-AACT-TC-	-----AA
Didelphostrongylus_hayesi	GTCTCAGTTAA	AAGATT--AAG	CCATGCATGT	GGA--GT--	T C-AGCT-TT-	--A-----AA
Crenosoma_mephitidis	GTCTCAGTTAA	AAGATTGAAG	CCATGCATGT	GGA--GT--	T C-AGCT-TC-	-----AA
Crenosoma_vulpis	TGCTCGGTTA	AAAGATTACA	AGCCATGCAT	GT--GT--A	G-TTCAGTA	-----AA
Otostrongylus_circumlitus	-----	---GATTAAG	CCATGCATGT	CGA--GT--	T C-AGCT-TC-	-----AA
Troglostrongylus_wilsoni	-----	---GATTAAG	CCATGCATGT	TGA--GT--	T C-AGCT-TC-	-----AA
Filaroides_martis	-----	---GATTAAG	CCATGCATGT	CGA--GT--	T C-AGCT-TC-	-----AA
Oslerus_osleri	-----	---GATTAAG	CCATGCATGT	CGA--GT--	T C-AACT-TC-	-----AA
Parafilaroides_decorus	-----	---ATTAAG	CCATGCATGT	CGA--GT--	T C-AGCT-TC-	-----AA
Metastrongylus_salmi	-----	---TTAAG	CCATGCATGT	CGA--GT--	T C-AACT-TC-	-----AA
Metastrongylus_elongatus	TGCTCAGTTA	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-AACT-TC-	-----AA
Skrjabinstrongylus_chitwoodrum	TGCTCAGTTA	AAAGATTAAG	CCATGCAATG	TGGA--GT--	T C-AGCT-TC-	-----AA
Parelaphostrongylus_odocolei	-----	---ATTAAG	CCATGCATGT	GGA--GT--	T C-AACT-TC-	--A-----AA
Muelerius_capillaris	-----	-----TGT	GGA--GT--	T C-AATT-TC-	--T-----AT	
Protostrongylus_rufescens	TGCTCAGTTA	AAAGATTAAG	CCATGCATGT	GGA--GT--	T C-AACT-TC-	-----AA
Halocercus_invaginatus	-----	-----TGT	CGA--GT--	T C-AGCT-TT-	--G-----AA	
Pseudalius_inflexus	-----	---CATGCATGT	CGA--GT--	T C-AACT-TC-	-----AA	
Stenurus_minor	-----	---ATTAAG	CCATGCATGT	CGA--GT--	T C-AGCT-TC-	-----AA
Torynurus_convolutus	-----	---ATTAAG	CCATGCATGT	CGA--GT--	T C-AGCT-TC-	-----AA
Syngamus_trachea	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCTT---	-----GA
Stephanurus_dentatus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT--A-	-----TG
Nematodirus_battus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT-TT-	-----GA
Strongylus_equinus	ATGCTTGTC	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT-T-	-----AA
Labiostrongylus_bipapillosus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCTA---	-----AA
Petrovinema_poculatum	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCA--C-	-----AA
Cylicocyclus_insignis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT--C-	-----AA
Chabartia_ovina	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT--A-	-----AA
Cyclodontostomum_purvisi	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT--A-	-----AA
Zoniolaimus_mawsonae	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT--A-	-----AA
Deletrocephalus_dimidiatus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT--A-	-----AA
Dictyocaulus_eckerti_P7B8	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--	T C-AACC-TG-	--TA-----TA
Dictyocaulus_capreolus_P3B2	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--	T C-AACT-TG-	--TG-----AA
Dictyocaulus_sp._P6A1	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--	T C-AACT-TT-	--AA-----A
Dictyocaulus_filaria	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--	T C-AACT-AA-	--AA-----AA
Dictyocaulus_capreolus_P2C10	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--	T C-AACT-TG-	--TG-----AA
Dictyocaulus_viviparus	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--	T C-AACT-TT-	--TC-----AA
Haemonchus_sp._V3091	-----	---CATGCATGT	CGA--GT--	T C-ATCT-TT-	--G-----AA	
Haemonchus_contortus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT-TT-	--G-----AA
Haemonchus_placeii	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--	T C-ATCT-TT-	--G-----AA
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	70	80	90	100	110	120
Haemonchus similis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-TT-	--G-----AA
Ostertagia ostertagi	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-AA-	-----AA
Ostertagia leptospicularis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-AA-	-----AA
Nippostrongylus brasileinsis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A-	-----TA
Heligmosomoides polygyrus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A-	-----AA
Trichostrongylus colubriformis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	GGA--GT--T	C-ATCT-AA-	-----AA
Tetrabothriostromylus mackerr	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-TT-	-----GA
Herpestrostrongylus pythonsis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--C-	-----AA
Nicollina cameroni	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--C-	-----AA
Filarimena flagrifer	TGCTCAGTTA	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCC-CT-	-----GA
Amidostomum cygni	TGCTCAGTTT	AAAGACTAAG	CCATGCATGT	CGA--GT--T	C-ATCT----	-----AA
Teratocephalus lirellus	-----TC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CA-	--ATTAA-TG
Brumptaemilius justini	-----	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	C-AAAT-AA-	--CCTAA-AG
Anguillicola crassus	-----C	AAAGATTAAG	CCATGCATGT	CTAT-GT--T	T-AAAT-AT-	-CCTATAAAG
Dracunculus medinensis	-----	-----	-----	-----	--ATTG-CC-	--TATAA-TG
Dracunculus oesophageus	-----	-----	-----	-----	--TTG-CC-	--TATAA-TG
Dracunculus sp. V3104	-----	-----	-----	--A-GT--T	C-ATAC-TG-	-CCTTAAATG
Philonema sp. A	-----	-----	-----TGT	CCAA-GT--T	C-AAAT-TG-	-CCTATAATG
Philometra obturans	-----	-----	-----	-----	--ATTG-CC-	--TATAA-TG
Camallanus oxycephalus	-----	-----	-----TG	TCAAAGT--T	C-AAATTGC-	--TTATAATG
Acanthocheilonema viteae	-----	-----TTAAG	CCATGCATGT	GTAA-GT--T	C-AAATAAAA-	-CCTATAAAG
Loa loa	-GCTTGTCCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	C-AAATAAAA-	-CCTATAATG
Onchocerca cervicalis	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	C-AAATAAAA-	-CCTATAATG
Dirofilaria immitis	-----	-----	-----	--GCCATGCA	TGTCTAATTT	C-AAATAAAA-
Brugia malayi	-----	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	C-AAATAAAA-	-CCTATAATG
Wuchereria bancrofti	-----	-----AG	CCATGCATGT	CTAA-GT--T	C-AAATAAAA-	-CCTATAATG
Litomosoides sigmodontis	-----	-----	-----	-----GT-CT	C-AAATAAAA-	-CCTATAAAG
Setaria digitata	-----TC	AAAGATTAAG	CCATGCATGT	GTAA-GT--T	C-AAATAAAA-	-CCTAAAATG
Gnathostoma turgidum	TGCTTGTCCTC	AAAGGTTAAG	CCATGCATGT	CTGC-GT--G	C-CAAC-TC-	--TCGAA-AA
Gnathostoma neoprocyonis	TGCTTGTCCTC	AAAGGTTAAG	CCATGCATGT	CTGC-GT--G	C-CAAC-TC-	--TCGAA-AA
Gnathostoma binucleatum	TGCTTGTCCTC	AAAGGTTAAG	CCATGCATGT	CTGC-GT--G	C-CTAC-TC-	--TTGAA-AA
Physaloptera alata	-----	-----	--ATGCATGT	CTAA-GT--T	C-AAAT-AA-	-CCTATAATG
Physaloptera turgida	-----	-----G	CCATGCATGT	CTAT-GT--T	C-AAAT-AA-	-CCTATAATG
Ascarophis arctica	-----	-----TTAAG	CCATGCATGT	CTAA-GT--T	C-AAAT-AA-	-CCTATAATG
Spinitectus carolini	-----	-----	-----	CTAA-GT--T	C-AAAC-AA-	-CCTATAATG
Thelazia lacrymalis	-----	-----AG	CCATGCATGT	CTAA-GT--T	C-AAAT-AA-	-CCTAAAATG
Aphelenchus avenae	-----	-----	-----	-----	-----	-----a
Aphelenchoides fragariae	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	GTAA-GT--A	G-AGTT-GA-	--TATAA--A
Bursaphelenchus sp	-----	-----	-----	-----	--GGAG-TA-	--TTATA-CA
Deladenus sp	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AANC-GA-	--TTTTA-TC
Criconema sp	-----	--AGATTAAG	CCATGCATGT	ATAA-GT--A	C-ACGC-CT-	--TGAAA-AG
Hemicyclophora conida	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATTA-GT--A	C-AAGC-CT-	--TAACA-AG
Paratylenchus dianthus	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	C-AAAC-CT-	--TCATA-AG
Pratylenchus thornei	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GC-	--TTTAA-GC
Tylenchulus semipentrans	-----CTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AATC-GT-	--CTTGA-AC
Ditylenchus angustus	-----C	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	T-AAAC-GA-	--TTAAT--C
Subanguina radicola	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA-	--TTTTA-TC
Tylenchorhynchus maximus	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA-	--CTTGA-TC
Geocenamus quadrifer	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA-	--TTTTA-TC
Globodera pallida	-----	AAAGATTAAG	CCATGCATGT	ATAA-GT--G	T-AACC-TG-	--CCAGACAG
Scutellonema bradys	-GCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AACC-TG-	--CCAGA-CA
Helicotylenchus dihystra	-----TCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AACC-TG-	--CCAGA-CA
Rotylenchus robustus	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AACC-TG-	--CCAGA-CA
Meloidogyne arenaria	TGTTTATTTT	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT-	--TTA---TC
Meloidogyne incognita_KT	-----	-----	-----TGT	ATAA-GT--T	T-AATC-GT-	--TTA---TC
Meloidogyne javanica	-----	-----C	CCAGGCMSSM	AWAA-GT--T	T-ATTC-GT-	--TTA---TC
Meloidogyne artiellia	---TTGTTTT	AAAGATTAAG	CCATGCATGT	ACAA-GT--T	T-AATC-GC-	--ATTA--GC
Meloidogyne duytsi	-GCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT-	--TTTC--AC
Meloidogyne exigua	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GA-	--TTTT--TC
Meloidogyne hapla	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT-	--ATTT--AC
Meloidogyne ichinohei	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATCA-GT--T	T-AATC-GC-	--ATCA--GC
Meloidogyne maritima	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT-	--TTT---AC
Meloidogyne microtyla	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT-	--TTTT--AC
Nacobbus aberrans	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA-	--CTTGA-TC
Pratylenchoides ritteri	-----TTT	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA-	--TTTTA-TC
Pratylenchoides magnicauda	-----	-----	-----	-----	--aaac-ga-	--tttta-tc
Hirschmanniella sp. JH_2003	-----CTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA-	-CTTTGA-TC
Radopholus similis	-----	AAAGATTAAG	CCATGCATGT	C-AA-GT--A	TAAATC-GA-	--AA---CG
Pratylenchus goodeyi_VF	-----C	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GC-	--ATA---GC
Boleodorus thylactus_clone2	GCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	C-AAAC-GA-	--CTCGA-TC
Philippine Sequence 1	-----	-----	-----	-----	-----	-----
Philippine Sequence 2	-----	AAGGATTAAG	CCATGCATGA	GGA--GT--T	C-AGCT-TT-	-----AA
Philippine Sequence 3	-----	-----	-----T	ATAA-AT--T	C-AACG-T-	-----AA
Philippine Sequence 4	-----	-----	-----T	CTAA-GT--A	C--ATG-TG-	----TTTAAC
Philippine Sequence 5	-----	-----	-----T	CGA--GT--T	C-ATCT--A-	-----AA
Philippine Sequence 6	-----	-----	-----T	ATAAAGT--T	C-ATCA----	-----TTTT
Philippine Sequence 7	-----	-----	-----	CTTT-G---A	T-TCAT-CA-	--A-----
Tahiti Sequence 1	-----	-----	-----	-----	-----	-----
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	70	80	90	100	110	120
Tahiti Sequence 2	-----				GGA--GT--T	C-AGCT-TT-	-----AA
Thailand Sequence 1	-----	-AGATTTAAG	CCATGCATGT	CTAT-GTTCA	AATAAC-CT-	--ATAA--AG	
Ogasawara Sequence 1	-----				CTAA-GT--A	C--ATG-TG-	--TATTTAAC
Ogasawara Sequence 2	-----	----ATTAAAG	CCATGCATGT	CGA--GT--A	C-ATTG-TA-	-----A	
Ivory Coast Sequence 1	-----				CGA--GT--T	C-AACT-TC-	-----AA
Ivory Coast Sequence 2	-----				CGA--GT--T	C-AACT-TT-	-----AAA
Ivory Coast Sequence 3	-----				CGA--GT--T	C-AACT-TC-	-----AA
Ivory Coast Sequence 4	-----				CGA--GT--T	C-AACT-TC-	-----AA
Ivory Coast Sequence 5	-----				CTAT-GTTCA	AATAAC-CT-	--ATAA--AG
Ivory Coast Sequence 6	-----				CGA--GT--T	C-AACT-TC-	-----AA
Ivory Coast Sequence 7	-----				CGA--GT--T	C-AACT-TC-	-----AAA
Singapore Sequence 1	-----				TATAAAT--T	C-AACG-TA-	-----A
Singapore Sequence 2	-----				CTAT-GTTCA	AATAAC-CT-	--ATAA--AG
Singapore Sequence 3	-----				CTTT-G--A	T-TCAT-CA-	--A-----
Singapore Sequence 4	-----				CGA--GT--T	C-AACT-TC-	-----AA
Nigerian Sequence 1	-----						
Nigerian Sequence 2	-----				CGA--GT--T	C-ATCT--A-	-----AA
Nigerian Sequence 3	-----				CGA--GT--T	C-AACT-TC-	-----AAA
NUCLEOTIDES INCLUDED	-----						

	130	140	150	160	170	180
Gordius_aquaticus	GTG-AAA-CC	GCGAATGGCT	CATTAA-ATC	AGTTA-TGGT	-TTATTAGAT	CGT-AC-AA-	
Priapulius_caudatus	GTG-AAA-CC	GCGAATGGCT	CATTAA-ATC	AGTTA-TGGT	-TCCTTAGAT	CTT-AC-TA-	
Brachionus_plicatilis	GTG-AAA-CC	GCGAATGGCT	CATTAA-ATC	AGTTA-TGGT	-TCCTTAGAT	CGT-TC-AT-	
Chordodes_morgani	GTG-AAA-CC	GCGAATGGCT	CATTAA-ATC	AGTTA-TGGT	-TTATTAGAT	CGT-CC-CA-	
Paractinolaemus_macrolaimus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-CCGT	-TTACTAGAA	AAT-AG-TT-	
Aporcelaimellus_obtusicaudatus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-CCGT	-TTACTAGAA	TAT-AG-TT-	
Wilsonema_schuurmansstekhoveni	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTTAGAT	CTT-GA-CT-	
Mesodorylaimus_sp_cf_nigritul	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-CCGT	-TTACTAGAA	TAT-AG-TT-	
Mesodorylaimus_bastiani	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-AAAT	-TTACTGGAA	TAT-AG-TT-	
Mesodorylaimus_japonicus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-CCGT	-TTACTAGAA	TAT-AG-TT-	
Pungentus_sp._PDL_2005	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TCGT	-TTATTAGAA	TAT-CG-TT-	
Allodorylaimus_sp	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TCGT	-TTATTAGAA	TAT-CG-TT-	
Eudorylaimus_carteri	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TCGT	-TTATTAGAA	TAT-CG-TT-	
Microdorylaimus_sp	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TCGT	-TTATTAGAA	TAT-CG-TT-	
Longidorus_elongatus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TCGT	-TTACTAGAA	AAT-AT-TT-	
Xiphinema_rivesi	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-CCGT	-TTACTAGAA	AAT-CA-GT-	
Tylencholaemus_sp	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-CCGT	-TTATTAGAA	TAT-CG-TT-	
Anoplostoma_sp._BHMM_2005	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TAGT	-TTACAGGAT	GAT-GC-TG-	
Adoncholaimus_fuscus	GTG-AAG-CC	GCGAATGGCT	CATTAC-ACC	AGCTG-AAAT	-TTACTGGAT	ATA-TC-CT-	
Enoplus_meridionalis	GTG-AAG-CT	GTGAATGGCT	CATTAC-AAC	AGCCG-TAGT	-TTATTTGAT	TCA-TA-GA-	
Enoplus_brevis_U88336	GTG-AAG-CT	GTGAATGGCT	CATTAC-AAC	AGCCG-TAGT	-TTATTTGAT	TTA-TA-GA-	
Enoplus_communis	GTG-AAG-CT	GTGAATGGCT	CATTAC-AAC	AGCCG-TAGT	-TTATTTGAT	TTA-TA-GA-	
Enoploides_brunettii	GTG-AAA-CC	GCAATGGCT	CATTAC-AAC	AGCTA-TAGT	-TTATTAGAT	CTT-AA-CT-	
Syringolaimus_striatocaudatus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TTGT	-TTATTTGAT	CTA-AT-GG-	
Ironus_dentifurcatus	GTA-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCT-CTGT	-TTATTAGAT	AAT-TC-TT-	
Calyptronema_maxweberi	GTGTAAG-CC	GCGAATGGCT	CATTAT-AAC	AGCCA-TTGT	-TTACTGGAT	ATA-TC-TT-	
Viscosia_sp._BHMM_2005	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TCACTGGAT	ATA-TT-AC-	
Viscosia_viscosa	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TCGT	-TTAATGGAT	ATA-TT-TT-	
Pontonema_vulgare	GTG-AAG-CT	GCTAATGGCT	CATTAT-AAC	AGCCA-TAGT	-TTACTGGTT	ATA-TT-CC-	
Oncholaimus_sp._BHMM_2005	TGT-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTACTGGAT	ATA-TT-CC-	
Alaimus_sp_PDL_2005	GCA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTAGAT	CTT-AC-TT-	
Prismatolaimus_intermedius	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTACTGGAT	CTT-AC-TT-	
Tobrilus_gracilis	GCA-AAG-CC	GCACATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAT	CGT-AC-TT-	
Tripyla_cf_filicaudata_JH_2004	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAAT	-TTACTAGGT	CTC-CT-TT-	
Bathylaimus_sp	GTA-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTACTAGAT	AGT-TC-AT-	
Bathylaimus_assimilis	GTA-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTACTAGAT	AGT-TC-AT-	
Bathylaimus_sp._BHMM_2005	GTA-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTAGAT	AGT-TC-AT-	
Tripyloides_sp._BHMM_2005	GTA-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTAGAT	AGT-TC-CT-	
Trischistoma_monohystera	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TCGT	-TTATTAGAT	CTT-AA-TT-	
Mermis_nigrescens	GTA-AAG-CC	GCGAATGGCT	CGGTAT-AAC	AGCTA-CCGT	-TTATTAGAT	GTT-TG-TT-	
Mylonchulus_arenicolus	GTA-AAG-CC	GCGAATGGCT	CGGTAT-AAC	AGCTG-TTGT	-TTACTAGAC	CAT-AG-TT-	
Mononchus_tridentatus	GTA-AAG-CC	GCGAATGGCT	CGGTAT-AAC	AGCCA-CCGT	-TTATTAGAT	CTT-AG-TT-	
Mononchus_truncatus	GTA-AAG-CC	GCGAATGGCT	CGGTAT-AAC	AGCCG-CTGT	-TTATTAGAT	TTT-AG-TT-	
Prionchulus_muscorum	GTA-AAG-CC	GCGAATGGCT	CGGTAT-AAC	AGCCA-CCGT	-TTATTAGAT	CTT-AG-TT-	
Clarkus_sp	GTA-AAG-CC	GCGAATGGCT	CGGTAT-AAC	AGCCA-CTGT	-TTATTAGAT	GTT-AG-TT-	
Trichinella_spiralis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAA	CTT-AC-A--	
Trichinella_papuae	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAA	CTT-AC-A--	
Trichinella_britovi	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAA	CTT-AC-A--	
Trichinella_murrelli	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAA	CTT-AC-A--	
Trichinella_pseudospiralis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAA	CTT-AC-A--	
Trichinella_nativa	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAA	CTT-AC-A--	
Trichinella_zimbabwensis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAA	CTT-AC-A--	
Trichinella_nelsoni	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CAGT	-TTATTAGAA	CTT-AC-A--	
Trichuris_suis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AGC	AGTCA-TTGT	-TCGCAAGAA	CTG-AT-GT-	
Trichuris_trichiura	GTT-AAG-CC	GCGAATGGCT	CATTAC-AGC	AGTCG-TTGT	-TCGCTAGAA	CTG-AT-GT-	
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmmmm	mmmmmm-mm	mmmmmm-mm	mmmmmmmmmm	mmmm-----	

	130	140	150	160	170	180
Trichuris muris	GTG-AAG-CC	GCGAATGGCT	CATTAT-AAC	AGCCA-TTGT	-TCGCAAGAA	CTG-AT-AT-
Tylolaimophorus minor	GCG-AAG-CC	GCGAATGGCT	CATTAT-AAC	AGCCG-TTGT	-TTACTAGAT	GAT-CC-AT-
Paratrichodorus pachydermus	GTG-AAG-CC	GCGAAAAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-TA-CC-
Paratrichodorus anemones	GCG-AAG-CC	GCGAAAAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-TC-CT-
Trichodorus primitivus	GTG-AAG-CC	GCGAAAAGCT	CATTAC-AAC	ASCCR-TAGT	-TTATTGGAT	GTT-TA-CG-
Axonolaimus helgolandicus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TTGT	-TTACTGGAT	CTT-AA-TA-
Ascolaimus elongatus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TTGT	-TTACTGGAT	CTT-AA-TA-
Odontophora rectangula	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TTGT	-TTACTGGAT	CTT-AA-AA-
Cylindrolaimus_sp._202149	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTACTTGAT	CTT-AA-AA-
Tylocephalus auriculatus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTTGAT	CTT-GA-CT-
Plectus acuminatus_BS9	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTTGAT	CTT-GA-CT-
Plectus aquatilis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTTGAT	CTT-GA-TT-
Anaplectus_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTTGAT	CTT-GA-TT-
Anisakis_sp_WKT	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Anisakis_sp_Nadler	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Contracaecum multipapillatum	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Pseudoterranova decipiens	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Raphidascaris acus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Terranova caballeroi	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Ascaris suum	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Ascaris lumbricoides	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Baylisascaris procyonis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Baylisascaris transfuga	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Parascaris equorum	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Porrocaecum depressum	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Toxascaris leonina	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Heterocheilus tunicatus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Goezia pelagia	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Hysterothylacium fortalezae	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Hysterothylacium pelagicum	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Hysterothylacium reliquens	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Iheringascaris iniques	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TA-
Toxocara canis	GTG-AAG-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TG-
Nemhelix bakeri	GTG-AAG-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TTAT	-TTACTTGAT	CTT-GA-TA-
Raillietnema_sp._V3060	GTG-AAG-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TTAT	-TTACTTGAT	CTT-GA-TA-
Cruzia americana	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	CTT-GA-TA-
Heterakis_sp_14690	GTG-AAG-CC	GCGAATGGCT	CATTAT-ACC	AGCTA-TAAT	-ATACTTGAT	CTT-GACTA-
Heterakis gallinarum	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TAAT	-ATACTTGAT	GTT-GATTA-
Paraspidodera_sp._21303	GTG-AAG-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TTAT	-ATACTTGAT	CTT-GA-TA-
Chromodora nudicapitata	GTA-AAT-CC	GCGAATGGCT	CATTAT--TC	AGCCA-CAAA	-TCATTGGAT	CTA-AT-CA-
Chromadora_sp._BHMM_2005	GTA-AAT-CC	GCGAATGGCT	CATTAC-AAC	AGCTT-TAGT	-TTGTTGGAT	CTA-AT-CC-
Atrochadora_sp._microlaima	GTA-AAT-CC	GCGAATGGCT	CATTAC-AAC	AGCCT-TAGT	-TTGTTGGAT	CTA-AA-TC-
Chromadorina germanica	GTA-AAT-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAAT	-TTATTGGAT	CTA-AC-AT-
Chromadorita tentabundum	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-AG-AG-
Dichromadora_sp._BHMM_2005	GTA-AAT-CC	GCGAATGGCT	CATTAC-AAC	AGCCT-TAGT	-TTGTTGGAT	CTA-AA-TC-
Neochromadora_BHMM_2005	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-AG-AG-
Spilophorella paradoxa	GTT-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTGTTGGAT	CTA-AA-AT-
Paracanthonchus caecus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Paracyatholaimus intermedius	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Praeacanthonchus punctatus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Praeacanthonchus_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Cyatholaimus_sp._BHMM_2005	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Spirinia parasitifera	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GA-TT-
Acanthopharynx micans	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GT-TT-
Xyzzors_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCT-TTGT	-TTCTTGGAT	CTT-GA-AC-
Metachromadora_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GA-TT-
Metachromadora remanei	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GA-TT-
Catanema_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Eubostriechus dianae	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AG-TT-
Eubostriechus topiarus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Eubostriechus parasitiferus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Laxus oneistus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Laxus cosmopolitus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Leptonemella_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Robbea hypermnestra	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Stilbonema majum	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Monoposthia costata	GTG-AAG-CT	STSAATGGCT	CATTAA-ATC	AGTTA-TAGT	-TTGTTTGGAT	GGT-AC-GT-
Nudora bipapillata	GTG-AAG-CC	GCGGACAGCT	CATTAA-TAC	ACTCC-TAAT	-CCTAGCAAT	GCT-TA-CC-
Calomicrolaimus parahonestus	GTG-AAG-TC	GCGAATAGCT	CATTAC-AAC	AGCCA-TTGT	-TTCTTGGAT	CTT-AC-TT-
Calomicrolaimus_sp._BHMM_2005	GTG-AAG-TC	GCGAATAGCT	CATTAC-AAC	AGCCA-TTGT	-TTCTTGGAT	CTT-AC-TT-
Molgolaimus demani	GTG-AAG-TC	GCGAATAGCT	CATTAC-AAC	AGCCA-TTGT	-TTCTTGGAT	CTT-AC-TT-
Diplogaster lethieri	ATG-AAT-CT	GCGAACGGCT	CATTATTAAC	ACCCG-TAAT	-CTACCCAGT	TTT-CG-TA-
Aduncospiculum halicti	CTG-AAT-CT	GCGAACGGCT	CATTAA-AAC	GGCTA-TAAT	-TTACTCGAA	TTA-TC-A--
Pristionchus_lheritieri	ATG-AAT-CT	GCGAACGGCT	CATTATTAAC	ACCCG-TAAT	-CTACCCAGT	TTT-CG-TA-
Pristionchus pacificus	ATG-AAT-CT	GCGAACGGCT	CATTATTAAC	ACCCA-TAAT	-CTACCCAGT	TTT-CG-TA-
Pristionchus pacificus	ATG-AAT-CT	GCGAACGGCT	CATTATTAAC	ACCCA-TAAT	-CTACCCAGT	TTT-CG-TA-
Sabatieria punctata_STRAIN_343	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-TA-AT-
Sabatieria_sp._355_BHMM_2005	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-TA-AT-
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmm	mmmmmm-mm	mmmm-mm	mmmmmmmm	mmmm-----

	130	140	150	160	170	180	
Sabatieria_celtica	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTTGGAT	CTC-TC-GA-	
Sabatieria_punctata_STRAIN_200	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTTGGAT	CTC-TT-AT-	
Sabatieria_punctata_STRAIN_223	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTTGGAT	CTC-TC-AT-	
Sabatieria_sp._210_BHM_2005	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTTGGAT	CTC-TT-AT-	
Setosabatieria_hilarula	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTTGGAT	CTC-TC-GA-	
Desmolaimus_zeelandicus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	CTACTTGGT-	CTT-GA-AA-	
Terschellingia_longicaudata	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	TCCCTTGAGA	CTA-TG-AA-	
Cyartonema_elegans	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	TCCCTAGAGA	CTA-CG-A--	
Tridentulus_sp	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-TT-CT-	
Diplolaimelloides_meyli	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-TT-CT-	
Diplolaimella_diavengatensis	ATG-AAG-CC	GCGAATAGCT	CATTAT-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-CT-CT-	
Geomonhystera_disjuncta	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAT	AGCCG-TTGT	-TTCTTGGAT	CCT-GA-AT-	
Sphaerolaimus_hirsutus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-AT-CC-	
Theristus_acer	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTTAGAT	CTC-CG-AT-	
Daptonema_procerus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-CG-TT-	
Daptonema_hirsutum	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-CG-CT-	
Daptonema_normandicum	GTA-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-CG-CA-	
Daptonema_oxycerca	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-CG-TT-	
Daptonema_setosum	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-CG-CT-	
Desmodora_communis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GT-TT-	
Desmodora_ovigera	GTG-AAG-CT	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GT-TT-	
Metadesmolaimus_sp	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-CG-CT-	
Dentostomella_sp	GTG-AAA-CC	GCGAAGAGCT	CATTAT-AAC	AGCTA-TTAT	-TTACTTGAT	ATT-GA-TA-	
Bunonema_franzi	GCG-AAT-TC	GCGGACGGCT	CATTAC-AAC	AGAGA-TAAT	-TTGCTTGAT	TTA-GA-TT-	
Bunonema_sp	GCA-AAT-CC	GCGGACGGCT	CATTAC-AAC	AGAGC-TAAT	-TTTCTTGAT	TTT-GA-CG-	
Seleborca_complexa	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTT-TGA-	-TTCTTGGA	CGT-AG-AC-	
Acrobeloides_nanus	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Acrobeloides_bodenheimeri	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Acrobeloides_sp_PS1146	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Acrobeles_ciliatus	GTG-AAA-CC	GCGAATGGCT	CATTAA-TAC	AGTAA-AAAC	-CTACAAGAT	GTT-GA-CA-	
Acrobeles_sp_PS1156	GTG-AAA-CC	GCGAATGGCT	CATTAA-TAC	AGTAA-AAAC	-CTACAAGAT	GTT-GA-CA-	
Acrobeles_complexus_WCUG2	GTG-AAA-CC	GCGAATGGCT	CATTAA-TAC	AGTAA-AAAC	-CTACAAGAT	GTT-GA-CA-	
Cephaloboides_sp_SB227	TTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTT-TTGTCT	G-----	
Cephalobus_cubaensis	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-	
Cephalobus_sp._PS1143	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Cephalobus_sp._PS1196	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Cephalobus_oryzae_PS1165	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Cervidellus_alutus	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Pseudacrobelus_variabilis	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	TTT-GA-CT-	
Triligulla_aluta	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Zeldia_punctada	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Myolaimus_sp_U81585	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	TTT-GA-TA-	
Rhabditophanes_sp._KR3021	GTG-AAA-CC	GCGGAAAGCT	CATTAT-AAC	AGCTA-TAAT	-TTACACGGC	AAA-TT-CC-	
Brevibucca_sp._SB261	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TTAT	-TTACTTGAC	GAG-AC-AA-	
Halicephalobus_gingivalis	ACG-AAA-CC	GCGCAAGAGCT	CATTAC-AAC	AGCTA-AAAT	-GTATACGGT	GAG-AT-AC-	
Panagrolaelus_stammeri	ACG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTT-TAAC	-CTACTTGAT	TTA-GA-CA-	
Plectonchus_sp._PDL0025	ACG-AAA-CC	GCGAACGGCT	CATTAC-AAC	GGCTT-TACT	-TTACTTGAT	TTT-GA-CT-	
Turbatrix_aceti	GCG-AAA-CC	GCGCATAGCT	CATTAT-AAC	AGTCC-TAAT	-TTACAAGTA	TTT-GA-TT-	
Panagrellus_redivivus	ACG-AAA-CC	GCGTATGGCT	CATTAC-AAC	AGCTA-TGAT	-TCTATTGAT	CAT-GA-CA-	
Panagrellus_redivivus_PS1163	ACG-AAA-CC	GCGTATGGCT	CATTAC-AAC	AGCTA-TGAT	-TCTATTGAT	CAT-GA-CA-	
Panagrolaimus_subelongatus	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGAT	TTT-GA-CT-	
Panagrolaimus_davidi	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGAT	TTT-GA-CT-	
Panagrolaimus_cf_rigidus_AF40	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGAT	TTT-GA-CT-	
Panagrolaimus_sp._Sourhope_ED2	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGAT	TTT-GA-CT-	
Panagrolaimus_sp._Sourhope_ED2	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGAT	TTT-GA-CT-	
Panagrolaimus_sp._Sourhope_ED2	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGAT	TTT-GA-CT-	
Panagrolaimus_sp._Sourhope_ED2	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGAT	TTT-GA-CT-	
Steinernema_carpocapsae	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCGA-TAAT	-TGACTAAAC	GAT-GT-TA-	
Strongyloides_stercoralis	GTG-AAA-CT	GCGAATGGCT	CATTAA-ATC	AGTTA-TAGT	-TTATTTGAT	GGT-TT-CT-	
Strongyloides_ratti	ATG-AAA-CC	GCGGAAAGCT	CATTAT-AAC	AGCTA-TAGA	-CTACACGGT	AAA-TA-TT-	
Diploscapter_sp_PS1897	GTG-AAG-CT	GCGCACGGCT	CATTAG-CAC	GGTTT-CGAC	CTTTTTCGGA	TAT-CT----	
Diploscapter_sp_PS2017	GTG-AAG-CT	GCGCACGGCT	CATTAG-CAC	GGTTT-CGAC	CTTTTTCGGA	TAT-CT----	
Heterorhabditis_bacteriophora	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAT	-TTATTCGGT	AAT-A-----	
Heterorhabditis_hepialus	GTG-AAA-C-	GCGAACGGCT	CATTAG-AGC	AGATA-TCAT	-TTATTCGGT	AAT-A-----	
Heterorhabditis_zelandica	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAT	-TTATTCGGT	AAT-A-----	
Parasitorhabditis_sp_SB281	GCA-GTA-CT	GTGAACGGCT	CATTAT-AAC	AGTTA-AAAT	-TTGCATGAA	GCT-TT-TC-	
Rhabditoides_inermiformis	GTG-AAA-CT	GCGAACGGCT	AATTAT-ACC	AGCTA-TAAT	-TTACTAGAT	TTT-GA-CA-	
Rhabditoides_inermis_DF5001	GTG-AAA-CT	GCGGGCGGCT	CATTAT-ATC	AGTTA-TGAT	-TTACTTGAT	TTT-GA-TT-	
Rhabditoides_regina_DF5012	GTT-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGCCA-AAAT	-CAGCATGAG	GTT-GA-C--	
Poikilolaimus_oxycerca_SB200	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TAAT	-TTACTTGAT	GTT-GA-CT-	
Poikilolaimus_regenfussi_SB199	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TAAT	-TTACTTGAT	GTT-GA-CA-	
Distolabrellus_veechi_DWF1604	GTG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGTTA-AAAT	-CAGCGTGAA	GCT-TA-C--	
Distolabrellus_veechi_DWF5024	GTG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGTTA-AAAT	-CAGCGTGAA	GCT-TA-C--	
Choriorhabditis_dudichi	GAG-AAA-CT	GCGTACGGCT	CATTAG-AGC	AGATA-TGCA	-CTTGTCGAG	GTT-Y-----	
Protorhabditis_sp	GTG-AAA-CT	GCGCACGGCT	CATTAG-AAC	GGTTT-ATA-	-CCTATTTCG	GAT-GT----	
Protorhabditis_sp_DF5055	GCG-AAA-CT	GCGAATGGCT	CATTAG-AGC	AGTTA-ATAG	-CCTTCCGT	TGA-TC----	
Cruzanema_tripartitum_DF5015	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TATT	-TTATTCGG	TGA-TC-C--	
NUCLEOTIDES INCLUDED	-mm-mmm-mmm	mmmmmmmmmm	mmmmmm-mmm	mmmmmm-mmmmm	mmmmmmmmmm	mmmm-----	

	130	140	150	160	170	180	
Mesorhabditis_sp_PS1179	GGT-AGA-CT	GTGAACGGCT	CATTAT-AAC	AGTTA-AAAT	-CCTCGCGAA	GTT-AG-C--	
Mesorhabditis_spiculigera_SB15	GTG-GTA-CT	GTGAACGGCT	CATTAC-AAC	AGTTG-AAAT	-TCGCGTGAA	ATC-CT-TC-	
Mesorhabditis_anisomorpha_SB12	GTT-AGA-CT	GTGAACGGCT	CATTAC-AAC	GGTCA-AAAT	-CGGCACGAA	GTA-GT-C--	
Teratorhabditis_palmarum_DF501	GCC-CAT-CT	GCGAACGGCT	CATTAC-AAC	AGTTA-AAAT	-TCGCATGAC	GAT-TG-T--	
Teratorhabditis_synpapillata_S	ACC-TAT-CT	GCGAACGGCT	CATTAC-AAC	AGTTA-AAAT	-CCGCATGAC	GAT-TG-T--	
Caenorhabditis_briggsae_PB102	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTGTTCCG	AAT-CC-T--	
Caenorhabditis_elegans_N2	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTATCCGG	GAT-CC-G--	
Caenorhabditis_sp_CB5161	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTATTCGG	AAT-CC----	
Caenorhabditis_sp_PS1010	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAT	-CTCGTTCGG	AAT-TC----	
Caenorhabditis_japonica	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTTCTCGG	AAT-CC----	
Caenorhabditis_sp_DF5170	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAT	-TTTACTCGG	AAT-TC----	
Caenorhabditis_sp_SB341	-AG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAC	-ATCCTTCGG	CAT-TC----	
Caenorhabditis_plicata	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATG-TATT	-CTCATTTCGG	CGG-TC----	
Caenorhabditis_vulgaris	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTGTTTCGG	AAT-CC----	
Caenorhabditis_sonorae	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAC	-TTCCCTTCGG	CGA-TA----	
Caenorhabditis_drosophilae	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAT	-TTTACTCGG	AAT-TC----	
Phasmarhabditis_hermaphrodita	TAG-AAA-CT	GCGTACGGCT	CATTAG-AGC	AGATTTAAAT	-GATTCCGGG	TAT-C-----	
Phasmarhabditis_neopapillosa	AAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATTTAAAT	-GATTCCAGG	TAT-C-----	
Phasmarhabditis_neopapillosa	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTTCGG	CTA-TC----	
Pellioditis_mediterranea_SB173	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTTCGG	GTA-TC----	
Pellioditis_marina	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTTCGG	CTA-TC----	
Prodontorhabditis_wirthi	ATG-AAA-CT	GCGGACGGCT	CATTAT-AGC	ATTGA-AACG	-TTATTCTCGG	CAC-CA-T--	
Crustorhabditis_scanica	GTT-CGA-CT	GTGAACGGCT	CATTAT-AAC	AGTTA-AAAT	-CAGCATGAA	GTA-TA-T--	
Dolichorhabditis_sp_CEW1	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TAAT	-ATACCCGAG	ATA-TC-T--	
Oscheius_sp_DF5000	GTG-AAA-CT	GCGAACGGCT	CATTAT-AGC	AGATA-TAAT	-GTACCCGAG	AAT-AT-C--	
Oscheius_sp_BW282	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTTCGG	ATA-TC-C--	
Oscheius_myriophila_EM435	CTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTTCGG	ATA-TC-C--	
Oscheius_insectivora	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTTCGG	ATA-TC-C--	
Oscheius_dolichuroides	GTG-AAA-CT	GCGAACGGCT	CATTAT-AGC	AGATA-TAAT	-GTACCCGAG	ATA-TT-C--	
Rhabditis_blumi_DF5010	CGG-AAT-CG	GCGAACGGCT	CAATAT-TAC	AGCTA-TCAC	-TTATT-CGG	TGA-T-----	
Rhabditis_sp_PS1191	GGG-AAATCA	GCGGCTAGCT	CATTAC-AAC	AGATC-TTAC	-TTATTTTCA	AGA-GA-T--	
Rhabditis_sp_PS1010	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAT	-CTCGTTTCGG	AAT-TC-T--	
Rhabditis_myriophila_EM435	CTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTTCGG	ATA-TC-C--	
Rhabditis_colombiana	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTTCGG	ATA-TC-C--	
Rhabditella_axei_DF5006	CTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATT-TTAC	-CTATTCCGA	AAT-CT----	
Rhabditella_sp_DF5044	CTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATT-TTAC	-CTATTCCGA	AAT-CT----	
Cuticularia_sp_PS2083	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TAAT	-TTACTGGAT	GTT-GA-CT-	
Necator_americanus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Kalicephalus_cristatus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ATG-TC----	
Ancylostoma_caninum	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Angiostrongylus_cantonensis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTTCGGA	AAA-TC----	
Angiostrongylus_costaricensis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTTCGGA	AAA-TC----	
Angiostrongylus_malaysiensis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTTCGGA	AAA-TC----	
Angiostrongylus_dujardini	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Angiostrongylus_vasorum	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Aulurostrongylus_abstrusus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAG-TT----	
Didelphostrongylus_hayesi	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TATT	-TTAATCGGT	AAT-TT----	
Crenosoma_mephitidis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Crenosoma_vulpis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Otostrongylus_circumlitus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Troglostrongylus_wilsoni	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Filaroides_martis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Oslerus_osleri	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAG-AC----	
Parafilaroides_decorus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Metastrongylus_salmi	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Metastrongylus_elongatus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Skrjabinstrongylus_chitwoodrum	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAT	-TTATTTCGGA	AAT-TC----	
Parelaphostrongylus_odocoeilei	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC-C--	
Muelerius_capillaris	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TG----	
Protostrongylus_rufescens	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TT----	
Halocercus_invaginatus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Pseudalius_inflexus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC-T--	
Stenurus_minor	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Torynurus_convolutus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----	
Syngamus_trachea	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	ACG-TG----	
Stephanurus_dentatus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAG-TC----	
Nematodirus_battus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC-C--	
Strongylus_equinus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Labiostrongylus_bipapillosus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAA-TC----	
Petrovinema_poculatum	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Cylicocyclus_insignis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Chabartia_ovina	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Cyclodontostomum_purvisi	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Zoniolaimus_mawsonae	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Hypodontus_macropi	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----	
Deletrocephalus_dimidiatus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAA-TC----	
Dictyocaulus_eckerti_P7B8	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTTCGGA	ATT-AT-T--	
Dictyocaulus_capreolus_P3B2	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTTCGGA	ATT-AT-T--	
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmmmm	mmmmmm-mmmm	mmmmmm-mmmm	mmmmmmmmmm	mmmm-----	

	130	140	150	160	170	180	
Dictyocaulus_sp._P6A1	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTCGGA	ATT-AT-T--	
Dictyocaulus_filaria	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTCGGA	AAA-TC-C--	
Dictyocaulus_capreolus_P2C10	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTCGGA	ATT-AT-t--	
Dictyocaulus_viviparus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTCGGA	ATT-AT-T--	
Haemonchus_sp._V3091	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Haemonchus_contortus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Haemonchus_placei	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Haemonchus_similis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Ostertagia_ostertagi	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	AAA-TC----	
Ostertagia_leptospicularis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACA-TC----	
Nippostrongylus_brasileinsis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Heligmosomoides_polygyrus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Trichostrongylus_colubriformis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Tetrabothriostromylus_mackerr	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Herpestrostrongylus_pythonsis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Nicollina_cameroni	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	AAG-TC----	
Filarimena_flagrifer	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	ACG-TC----	
Amidostomum_cygni	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCGGA	AAA-TC----	
Teratocephalus_lirellus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TTGT	-TTACTTGAT	CTT-GA-TA-	
Brumptaemilius_justini	GTC-AAA-CC	GCGTACGGCT	CATTAT-AAC	AGCCA-TAAT	-ATACTTGAC	GTT-GA-CT-	
Anguillicola_crassus	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGGT	CTT-GA-TA-	
Dracunculus_medinensis	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	CTT-GG-AT-	
Dracunculus_oesophageus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	CTT-GA-TT-	
Dracunculus_sp._V3104	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	GTT-GA-TT-	
Philonema_sp_A	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	TCT-GA-TT-	
Philometra_obturus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-TTACTTGAT	TTT-GA-TT-	
Camallanus_oxyccephalus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCCA-TAAT	-ATACTTGAT	GTT-GA-TT-	
Acanthocheilonema_viteae	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-	
Loa_loa	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-	
Onchocerca_cervicalis	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-	
Dirofilaria_immitis	GTG-AAA-CC	GCGAACGGCT	CAT--T-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-	
Brugia_malayi	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-	
Wuchereria_bancrofti	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-	
Litomosoides_sigmodontis	GTG-AAA-CC	GCGAACGGCT	CATNAT-NAC	AGCCTCTAAT	-GTACCTGAT	GCT-GA-CT-	
Setaria_digitata	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-	
Gnathostoma_turgidum	GTG-AAA-CC	GCGAATGGCT	CATTAC-GAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-TA-	
Gnathostoma_neoprocyonis	GTG-AAA-CC	GCGAATGGCT	CATTAC-GAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-TA-	
Gnathostoma_binucleatum	GTG-AAA-CC	GCGAATGGCT	CATTAC-GAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-TA-	
Physaloptera_alata	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	CTT-GA-TC-	
Physaloptera_turgida	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-CAAT	-GTACTTGAT	CTT-GA-TT-	
Ascarophis_arctica	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	CTT-GA-TA-	
Spinitectus_carolini	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	CTT-GA-TA-	
Thelazia_lacrymalis	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-	
Aphelenchus_avenae	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGGTA-TAAT	-TTACTTGAT	CTT-GA-AA-	
Aphelenchoides_fragariae	GCG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGATA-TAAT	-TTACTTGT	CTA-TT-CC-	
Bursaphelenchus_sp	GCG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGATA-TAAT	-TTACTAGTT	CTT-GA-TC-	
Deladenus_sp	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	TTT-CA-CT-	
Criconema_sp	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TAAT	-CTACTCGGT	CTT-GA-AC-	
Hemicycliophora_conida	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCCA-TAAT	-TTACTCGAT	CTT-GA-AC-	
Paratylenchus_dianthus	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTAGAC	CTT-GA-GC-	
Pratylenchus_thornei	GTG-AAA-CT	GCGTACGGCT	CATTAC-AAC	AGCAA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Tylenchulus_semipentrans	GAG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-AC-	
Ditylenchus_angustus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-CC-	
Subanguina_radicola	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-	
Tylenchorhynchus_maximus	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Geocenamus_quadriker	GTG-AAA-CC	GCGAACAGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-	
Globodera_pallida	GGG-AAA-CT	GCGTACGGCT	CATTAC-ACC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-	
Scutellonema_bradys	GGG-AAA-CT	GCGTACGGCT	CATTAC-ACC	AGCTG-TAAT	-TTACTTGAT	CTT-GA-CA-	
Helicotylenchus_dihystera	GGG-AAA-CT	GCGTACGGCT	CATTAC-ACC	AGCCT-TAAT	-TTACTTGAT	CTT-GA-CA-	
Rotylenchus_robustus	GGG-AAA-CT	GCGTACGGCT	CATTAC-ACC	AGCCT-TAAT	-TTACTTGAT	CTT-GA-CA-	
Meloidogyne_arenaria	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TTAT	-TTACTTGAT	CTT-GA-TT-	
Meloidogyne_incognita_KT	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TTAT	-TTACTTGAT	CTT-GA-TT-	
Meloidogyne_javanica	GAG-AAA-CC	GCGAACGGCT	MAWWAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TT-	
Meloidogyne_artiellia	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TTAT	-TTACTTGAT	CTT-GA-CA-	
Meloidogyne_duytsi	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TA-	
Meloidogyne_exigua	GAG-AAA-CC	GCGTACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TT-	
Meloidogyne_hapla	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TA-	
Meloidogyne_ichinohei	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	AGCCA-TTAT	-TTACTTGAT	CTT-GA-TCA	
Meloidogyne_maritima	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TTA	
Meloidogyne_microtyla	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TA-	
Nacobbus_aberrans	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Pratylenchoides_ritteri	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-	
Pratylenchoides_magnicauda	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-	
Hirschmanniella_sp._JH_2003	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Radopholus_similis	ATG-AAA-CC	GCAAACGGCT	CATTAC-ACC	AGCAG-TGAT	-CTATTTGAC	CTT-GA-TA-	
Pratylenchus_goodeyi_VF	GTG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGCTG-TAAT	-TTACTTGAT	CTT-GA-AA-	
Boleodorus_thylactus_clone2	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-	
Philippine Sequence 1	-TG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTCGGA	AAA-TC----	
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmmmm	mmmmmm-mm	mmmmmm-mm	mmmmmmmmmm	mmmm-----	

	130	140	150	160	170	180
Philippine Sequence 2	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTTCGGA	AAA-TC----
Philippine Sequence 3	GTG-AAA-TT	GCGAACGGCT	CATTAG-AGC	AGCTA-TTAT	TCTCTTCGGT	ATCAT-----
Philippine Sequence 4	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	TTT-GA-CA-
Philippine Sequence 5	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTTCGGA	AAG-TC----
Philippine Sequence 6	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TTAT	-TTTCTTCGA	ATA-T-----
Philippine Sequence 7	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTGTTCCG	AAT-AT----
Tahiti Sequence 1	-TG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTTCGGA	AAA-TC----
Tahiti Sequence 2	GTG-AAA-CT	GCGAACGGCT	TATTAG-AGC	AGATG-TGAT	-TTATTTCGGA	AAA-TC----
Thailand Sequence 1	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TAAT	-ATACTTGAT	CTT-GACTA-
Ogasawara Sequence 1	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	TTT-GA-CA-
Ogasawara Sequence 2	ATG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATATTAAT	GATTGCAGAG	-TA-TC----
Ivory Coast Sequence 1	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----
Ivory Coast Sequence 2	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAG-AC----
Ivory Coast Sequence 3	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-AT----
Ivory Coast Sequence 4	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTACTCGGT	AAGAC-----
Ivory Coast Sequence 5	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TAAT	-ATACTTGAT	CTT-GACTA-
Ivory Coast Sequence 6	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTACTCGGT	AAA-GAC----
Ivory Coast Sequence 7	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AA--GAC----
Singapore Sequence 1	GTG-AAA-TT	GCGAACGGCT	CATTAG-AGC	AGCTA-TTAT	-TCTCTTCGG	-TA-TC----
Singapore Sequence 2	GTG-AAR-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TAAT	-ATACTTGAT	CTT-GACTA-
Singapore Sequence 3	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTGTTCCG	AAT-CC-T--
Singapore Sequence 4	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAG-AC----
Nigerian Sequence 1	-TG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TTAT	-TTATTTCGGA	AAA-TA----
Nigerian Sequence 2	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCWC	-TTATTTCGGA	AAAG-TC----
Nigerian Sequence 3	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAG-ATGTA-
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmmmm	mmmmmm-mm	mmmmmm-mm	mmmmmmmmmm	mmmm-----

	190	200	210	220	230	240
Gordius_aquaticus	-----CC--	-----CACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCT-A
Priapulius_caudatus	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
Brachionus_plicatilis	-----AC--	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA--
Chordodes_morgani	-----AC--	-----CACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCT-G
Paractinolaimus_macrolaimus	-----ATCC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Aporcelaimellus_obtusicaudatus	-----ATCC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Wilsonema_schuermansstekhoveni	-----ATCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
Mesodorylaimus_sp_cf_nigritul	-----ATCT-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Mesodorylaimus_bastiani	-----ATCT-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Mesodorylaimus_japonicus	-----TTCC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Pungentus_sp._PDL_2005	-----ATCC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Allodorylaimus_sp	-----ATCT-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Eudorylaimus_carteri	-----ATCC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Microdorylaimus_sp	-----ATCT-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCG-A
Longidorus_elongatus	-----ATCC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Xiphinema_rivesi	-----ATCC-	-----TACT	CGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACGTCG-A
Tylencholaimus_sp	-----TTCC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Anoplostoma_sp._BHMM_2005	-----T---	-----TGAT	AGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Adoncholaimus_fuscus	-----T---	-----TACA	TGGATAAC--	TGTGGTAATT	CTACAGCTAA	TACACGCA-T
Enoplus_meridionalis	-----GT---	-----TACA	TGGATACC--	TGTGGTAACC	TAAGAGCTAA	TACGCGCA-A
Enoplus_brevis_U88336	-----GT---	-----TACA	CGGATACC--	TGTGGTAACC	TAAGAGCTAA	TACGCGCA-A
Enoplus_communis	-----GT---	-----TACA	CGGATACC--	TGTGGTAACC	TAAGAGCTAA	TACGCGCA-A
Enoploides_brunettii	-----C---	-----TACT	TGGATACC--	TGTGGTAACC	TAAGAGCTAA	TACATGCA-A
Syngnathoides_striotocaudatus	-----TTT-	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-T
Ironus_dentifurcatus	-----TCC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-A
Calyptronema_maxweberi	-----TAC-	-----TACA	TGGATAAC--	TGTGGTAATT	CTACAGCTAA	TACACGCT-C
Viscosia_sp._BHMM_2005	-----TCC-	-----TACA	TGGATAAC--	CGTGGTAATT	CTACGCTAA	TACACGCT-T
Viscosia_viscosa	-----T---	-----TACA	TGGATAAC--	TGTGGTAATT	CTACAGCTAA	TACACGCA-T
Pontonema_vulgare	-----AAT--	-----TACA	TGGATAAC--	TGTGGTAATT	CTACAGCTAA	TACACGCA-T
Oncholaimus_sp._BHMM_2005	-----AT---	-----TACA	TGGATAAC--	TGTGGTAATT	CTACAGCTAA	TACACGCA-T
Alaimus_sp_PDL_2005	-----TCC-	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCC-T
Prismatolaimus_intermedius	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTA-	TACACGCA-T
Tobrilus_gracilis	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-G
Tripyla_cf_filicaudata_JH_2004	-----T---	-----TACT	TGGATAAC--	TGAGCTAATT	GTTGAGCTAA	TACATGCA-C
Bathylaimus_sp	-----TAC-	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
Bathylaimus_assimilis	-----TAC-	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
Bathylaimus_sp._BHMM_2005	-----ACC-	-----TACA	TGGATAAC--	TGAGGTAATT	CTTGAGCTAA	TACATGCT-A
Tripyloides_sp._BHMM_2005	-----TAC-	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCT-A
Trischiostoma_monohystera	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCC-T
Mermis_nigrescens	-----ATC-	-----TACT	TGGATAAC--	TGCGGTAATT	CTAGAGCTAA	TACATGCA-A
Mylonchulus_arenicolus	-----TCT-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Anatonchus_tridentatus	-----ATC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Mononchulus_truncatus	-----ATC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Prionchulus_muscorum	-----ATC-	-----TACT	CGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACGTGCA-C
Clarkus_sp	-----ATC-	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Trichinella_spiralis	-----ATCA	-----TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C	
Trichinella_papuae	-----TACA	-----TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C	
Trichinella_britovi	-----ATCA	-----TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C	
NUCLEOTIDES INCLUDED	-----mmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmm--

	190	200	210	220	230	240
<i>Trichinella murrelli</i>	-----	-----ATCA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Trichinella pseudospiralis</i>	-----	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Trichinella nativa</i>	-----	-----ATCA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Trichinella zimbabwensis</i>	-----	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Trichinella nelsoni</i>	-----	-----CTCA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Trichuris suis</i>	-----	-----CACT	TGGATAAC--	TGTGGAAATT	CTAGAGCTAA	TAC-TGCC-T
<i>Trichuris trichiura</i>	-----C----	-----CACT	TGGATAAC--	TATGGAAATG	CTAGAGCTAA	TACATGCC-T
<i>Trichuris muris</i>	-----C----	-----CACT	TGGATAAC--	TGTGGAAATT	CTAGAGCTAA	TACATGCC-T
<i>Tyololaimophorus minor</i>	-----	-----TACA	TGGATAAC--	TGTGTGAATT	CAAGAGCTAA	TACATGCG--
<i>Paratrichodorus pachydermus</i>	-----TTCT-	-----TACA	CGGATAAC--	TGCGGTAATT	CTGGAGCTAC	TACGTGCA-T
<i>Paratrichodorus anemones</i>	-----TCT--	-----TACA	CGGATAAC--	TGCGGTAATT	CTGGAGCTAC	TACGTGCA-T
<i>Trichodorus primitivus</i>	-----TTCC-	-----TACA	AGGACACC--	TGCGGTAATT	CTGGAGCTAA	TACTTGCG--
<i>Axonolaimus helgolandicus</i>	-----TCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA--
<i>Ascolaimus elongatus</i>	-----TCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA--
<i>Odontophora rectangula</i>	-----TCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Cylindrolaimus sp._202149</i>	-----TCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Tylocephalus auriculatus</i>	-----ATCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Plectus acuminatus_BS9</i>	-----ATCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Plectus aquatilis</i>	-----ATCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Anaplectus sp</i>	-----ATCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Anisakis sp_WKT</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Anisakis sp_Nadler</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Contracaecum multipapillatum</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Pseudoterranova decipiens</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Raphidascaris acus</i>	-----tcc--	-----tacg	tggataac--	tgtggttaatt	ctagagctaa	tacatgca-c
<i>Terranova caballeri</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Ascaris suum</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Ascaris lumbricoides</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Baylisascaris procyonis</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Baylisascaris transfuga</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Parascaris equorum</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Porrocaecum depressum</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Toxascaris leonina</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Heterocheilus tunicatus</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Goezia pelagia</i>	-----TCCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Hysterothylacium fortalezae</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Hysterothylacium pelagicum</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Hysterothylacium reliquens</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Iheringascaris iniquus</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Toxocara canis</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Nemhelix bakeri</i>	-----TCC--	-----TACT	TGGATAAC--	TGAGGTAATT	CTTGAGCTAA	TACAATGCAG
<i>Raillietnema sp._V3060</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Cruzia americana</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Heterakis sp_14690</i>	-----TCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Heterakis gallinarum</i>	-----TCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Paraspidodera sp_21303</i>	-----TCC--	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Chromodora nudicapitata</i>	-----GT--	-----TACT	TGGATAAC--	TGTTCAAAAG	GAAGAGCTAA	GACATGCC-T
<i>Chromadora sp._BHMM_2005</i>	-----	-----CACA	GCAATAAC--	TGTGGAAAAG	CCAGAGCTAA	CTC-TGCA-T
<i>Atrochromadora microlaima</i>	-----	-----CACA	GCAATAAC--	TGTGGAAAAG	CCAGAGCTAA	CTC-TGCA-T
<i>Chromadorina germanica</i>	-----CC--	-----TACT	TGGATAAC--	TGTGGAAAAT	CCAGAGCTAA	TACATGCACC
<i>Chromadorita tentabundum</i>	-----TCC--	-----TACT	TGGATAAC--	TGTGGAAAAT	CTAGAGCTAA	TACACGCA-A
<i>Dichromadora sp._BHMM_2005</i>	-----	-----CACA	GCAATAAC--	TGTGGAAAAG	CCAGAGCTAA	CTCTGCAT--
<i>Neochromadora BHMM_2005</i>	-----TCC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Spilophorella paradoxa</i>	-----CC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Paracanthonchus caecus</i>	-----CA--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Paracyatholaimus intermedius</i>	-----CA--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-G
<i>Praeacanthonchus punctatus</i>	-----C--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Praeacanthonchus sp</i>	-----C--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Cyatholaimus sp._BHMM_2005</i>	-----CA--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Spirinia parasitifera</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGTTG-A
<i>Acanthopharynx micans</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Xyzzors sp</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Metachromadora sp</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Metachromadora remanei</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Catanema sp</i>	-----	-----TACT	CGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACGTGCC-A
<i>Eubostrichus diana</i>	-----T--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Eubostrichus topiarus</i>	-----T--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Eubostrichus parasitiferus</i>	-----T--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Laxus oneistus</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACGTGCA-A
<i>Laxus cosmopolitus</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Leptonemella sp</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Robbea hypermnestra</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGTA-T
<i>Stilbonema majum</i>	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Calomicrolaimus parahonestus</i>	-----TC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Calomicrolaimus sp._BHMM_2005</i>	-----TC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Molgolaimus demani</i>	-----TC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA--
NUCLEOTIDES INCLUDED	-----	-----nnnnn	nnnnnnnnnn--	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn--

	190	200	210	220	230	240
Diplogaster lethieri	-----TCC-	-----AAAA	CGGATATC--	TGCGTTAATT	TTGGAGCTAA	TACGTGCA-C
Aduncospiculum halicti	-----TCC-	-----TAAA	CGGATATC--	TGCGATAATT	TTGGAGCTAA	TACGTGCC-C
Pristionchus lheritieri	-----TCC-	-----AAAA	CGGATATC--	TGCGTTAATT	TTGGAGCTAA	TACGTGCA-C
Pristionchus pacificus	-----TCC-	-----AAAA	CGGATATC--	TGCGTTAATT	TTGGAGCTAA	TACGTGCA-C
Pristionchus pacificus	-----TCC-	-----AAAA	CGGATATC--	TGCGTTAATT	TTGGAGCTAA	TACGTGCA-C
Sabatieria punctata STRAIN_343	-----TT--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Sabatieria_sp._355_BHMM_2005	-----TT--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Sabatieria celtica	-----TC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCC-T
Sabatieria punctata STRAIN_200	-----T--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Sabatieria punctata STRAIN_223	-----TT--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Sabatieria_sp._210_BHM_2005	-----TT--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Setosabatieria hilarula	-----TC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCC-T
Desmolaimus zeelandicus	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA--
Terschellingia longicaudata	-----	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACACGCA-C
Cyartonea elegans	-----	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACACGCC-C
Tridentulus_sp	-----C---	-----TACT	TGGATATC--	TCTGGTAATT	CTAGAGCTAA	TACATGCA--
Diplolaimelloides meylli	-----C---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCAA-
Diplolaimella diavengatensis	-----C---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCCT--
Geomonhystrera disjuncta	-----T---	-----TACT	TGGATACC--	TGTGGTAATT	CTAGAGCTAA	TACATGCGA-
Sphaerolaimus hirsutus	-----T---	-----TACT	TGGATAAC--	TGTCGAAATG	CGAGAGCTAA	TACATGCAA-
Theristus acer	-----T---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCAT-
Daptonema procerus	-----T---	-----TACT	TGGATATG--	TGCTGCAATT	CAGGACTTAA	TACATGCAA-
Daptonema hirsutum	-----TT--	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCAA-
Daptonema normandicum	-----A---	-----TACT	TGGATAAC--	TGAGGTAATT	CTTGAGCTAA	TACACGCAA-
Daptonema oxycerca	-----T---	-----TACT	TGGATATG--	TGCTGCAATT	CAGGACTTAA	TACATGCAA-
Daptonema setosum	-----TT--	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCAA-
Desmodora communis	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCAA-
Desmodora ovigera	-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
Metadesmolaimus_sp	-----TT--	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCAA-
Dentostomella_sp	-----TCC-	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Bunonema franzi	-----CC--	-----CAAG	TGGATAAC--	TTTGGTAATT	CTGGAGCTAA	TACATGCCT-
Bunonema_sp	-----TCC-	-----CAAT	TGGACAAC--	TTTGGTAATT	CTGGAGCTAA	TACATGCCT-
Seleborca complexa	-----ACAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Acrobeloides nanus	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Acrobeloides bodenheimeri	-----CTAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Acrobeloides_sp_PS1146	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Acrobeles ciliatus	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Acrobeles_sp_PS1156	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Acrobeles_complexus_WCUG2	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Cephaloboides_sp_SB227	-----TCCT	C-----TATG	AGGATAAC--	TGCGATAAAT	TTGGAGCTAA	TACATATT-T
Cephalobus cubaensis	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Cephalobus_sp._PS1143	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Cephalobus_sp._PS1196	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Cephalobus_oryzae_PS1165	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Cervidellus alutus	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Pseudacrobeles variabilis	-----TCAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Triligulla aluta	-----CAAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Zeldia punctada	-----TCAT	CC-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Myolaimus_sp_U81585	-----TCC-	-----TACT	TGGACATATC	TGAGGAAAT	CTTGAGCTAA	TACATGCA-C
Rhabditophanes_sp._KR3021	-----C---	-----TAGT	TGGATAAC--	TGAGGTAATT	CTTGAGCTAA	TACACGCAAT
Brevibucca_sp._SB261	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Halicephalobus gingivalis	-----CAT-	-----TAAA	TGGATATC--	TGCGGTAAT	CTGGAGCTAA	TACATGCA-A
Panagrobelus stammeri	-----TNTA	TCT---TACA	TGGATAAC--	NGTGGTAATT	CTGGAGCTAA	TACATGCACT
Plectonchus_sp._PDL0025	-----CTAT	CC-----TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-C
Turbatrix aceti	-----TATC	-----TACA	CGGATAAC--	TTTGGCAATT	CTGGAGCTAA	TACGTGCA-T
Panagrellus redivivus	-----CTTT	CC-----TATA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrellus_redivivus_PS1163	-----CTTT	CC-----TATA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus subelongatus	-----TTTA	TCC---TACA	TGGATAAC-C	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus davidi	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus_cf_rigidus_AF40	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus sp. Sourhope ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus sp. Sourhope ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTCATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus sp. Sourhope ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus sp. Sourhope ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus sp. Sourhope ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Steinernema carpocapsae	-----ATCC	TT---TAAA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGGT-T
Strongyloides stercoralis	-----TGC-	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCTKA
Strongyloides ratti	-----T---	-----TAGT	TGGATAAC--	TGAGGTAATT	CTTGAGCTAA	TACACGCT-T
Diploscapter_sp_PS1897	-----	-----CAAT	GGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACCTGCA-C
Diploscapter_sp_PS2017	-----	-----CAAT	GGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACCTGCA-C
Heterorhabditis bacteriophora	-----	-----TTTT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Heterorhabditis hepialus	-----	-----TTTT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Heterorhabditis zelandica	-----	-----TCTT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Parasitorhabditis_sp_SB281	-----C---	-----CGTA	TGGATACT--	TCTAGTAATT	CTGGAGCTAA	TACATACT-A
Rhabditoides inermiformis	-----ATCC	-----TACA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-C
Rhabditoides inermis_DF5001	-----A---	-----TCTA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	AACATGCA-T
Rhabditoides regina_DF5012	-----	-----CTTA	TGGATACT--	TGAGTAATT	CTGGAGCTAA	TACACGTG-C
Poikilolaimus oxycerca_SB200	-----ATCC	-----TACT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	190	200	210	220	230	240
Poikilolaimus regenfussi_SB199	-----ATCC	-----TACT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Distolabrellus veechi_DWF1604	-----	-----TGAG	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TACATGTT-C
Distolabrellus veechi_DF5024	-----	-----TGAG	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TACATGTT-C
Choriorhabditis dudichi	-----	-----CCCM	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Protorhabditis_sp	-----ATCC	-----TTTT	GGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACCTGCA-C
Protorhabditis_sp_DF5055	-----	-----CCTT	GGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACCATTA-A
Cruzema tripartitum_DF5015	-----	-----AAGA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Mesorhabditis_sp_PS1179	-----	-----ATTT	TGGATACT--	TCTAGTAATT	CTGGAGCTAA	TACACGTG-T
Mesorhabditis_spiculigera_SB15	-----	-----TCAA	TGGATACT--	TCTAGTAATT	CTGGAGCTAA	TACATGCA-A
Mesorhabditis_anisomorpha_SB12	-----	-----TCCC	CGGATACT--	TCTAGTAATT	CTGGAGCTAA	TACGGCGC-A
Teratorhabditis palmarum_DF501	-----	-----TTCT	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TAC--GTA-A
Teratorhabditis_synpapillata_S	-----	-----TTTC	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TAC--GTA-C
Caenorhabditis briggsae_PB102	-----	-----AATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis elegans_N2	-----GATCC	-----TCATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis_sp_CB5161	-----	-----TATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis_sp_PS1010	-----	-----TAATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis_japonica	-----	-----TGAAA	AGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCG-A
Caenorhabditis_sp_DF5170	-----	-----TATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-T
Caenorhabditis_sp_SB341	-----	-----CAAA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis_plicata	-----	-----CTTAAA	TGGATACT--	TGCGGAAATT	CTGGAGCTAA	TACATGCC-G
Caenorhabditis_vulgaris	-----	-----TATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis_sonorae	-----	-----TCTTA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis_drosophilae	-----	-----CATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-T
Phasmarhabditis_hermaphrodita	-----	-----CTAT	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-A
Phasmarhabditis_neopapillosa	-----	-----CTTT	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-A
Phasmarhabditis_neopapillosa	-----	-----CTGA	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Pellioditis_mediterranea_SB173	-----	-----CTAG	TGGATAAC--	TTCGGTAATT	CTGGAGCTAA	TACATGCA-A
Pellioditis_marina	-----	-----CTGA	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Prodontorhabditis_wirthi	-----	-----AACC	AGGATACT--	TGCGGAAATA	CTGGAGCTAA	TACTTGCTA
Crustorhabditis_scanica	-----	-----CTTT	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TACATGCC-C
Dolichorhabditis_sp_CEW1	-----	-----TTAG	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGTC-A
Oscheius_sp_DF5000	-----	-----TTAA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Oscheius_sp_BW282	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Oscheius_myriophila_EM435	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Oscheius_insectivora	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Oscheius_dolichuroides	-----	-----TTAA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Rhabditis_blumi_DF5010	-----	-----CCTT	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Rhabditis_sp_PS1191	-----	-----CATT	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Rhabditis_sp_PS1010	-----	-----AATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Rhabditis_myriophila_EM435	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Rhabditis_colombiana	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Rhabditella_axei_DF5006	-----	-----TATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Rhabditella_sp_DF5044	-----	-----TATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Cuticularia_sp_PS2083	-----ATCC	-----TACT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Necator_americanus	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Kaliocephalus_cristatus	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Ancylostoma_caninum	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Angiostrongylus_cantonensis	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-T
Angiostrongylus_costaricensis	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-T
Angiostrongylus_malaysiensis	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-T
Angiostrongylus_dujardini	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-G
Angiostrongylus_vasorum	-----	-----TTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Aulurostrongylus_abstrusus	-----	-----CACC	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-G
Didelphostrongylus_hayesi	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-G
Crenosoma_mephitidis	-----	-----CATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Crenosoma_vulpis	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Otostrongylus_circumlitus	-----	-----CATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Troglostrongylus_wilsoni	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Filaroides_martis	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Oslerus_osleri	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Parafilaroides_decorus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-A
Metastrongylus_salmi	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Metastrongylus_elongatus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Skrjabinogylus_chitwoodrum	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGTG-A
Parelaphostrongylus_odocoilei	-----	-----TTAA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGTG-C
Muelerius_capillaris	-----	-----ACAA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Protostrongylus_rufescens	-----	-----AATC	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Halocercus_invasivatus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-a
Pseudalius_inflexus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATTCT-A
Stenurus_minor	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-A
Torynurus_convolutus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-A
Syngamus_trachea	-----	-----GTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Stephanurus_dentatus	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Nematodirus_battus	-----	-----TTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-T
Strongylus_equinus	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Labiostrongylus_bipapillosus	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Petrovinema_poculatum	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Cylicocyclus_insignis	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
NUCLEOTIDES INCLUDED	-----	-----nnnnn	nnnnnnnnnn--	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn--

	190	200	210	220	230	240
Pratylenchoides_ritteri	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Pratylenchoides_magnicauda	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Hirschmanniella_sp._JH_2003	-----CACC	-----TACM	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Radopholus_similis	-----ACC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Pratylenchus_goodeyi_VF	-----CCCA	-----CACA	CGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACGTGCA-C
Boleodorus_thylactus_clone2	-----ATCC	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Philippine Sequence 1	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Philippine Sequence 2	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Philippine Sequence 3	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCG-A
Philippine Sequence 4	-----TAAA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-C
Philippine Sequence 5	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCG-A
Philippine Sequence 6	-----	-----TTTC	AGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Philippine Sequence 7	-----	-----TATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCT-A
Tahiti Sequence 1	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Tahiti Sequence 2	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Thailand Sequence 1	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Ogasawara Sequence 1	-----TAAA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-C
Ogasawara Sequence 2	-----	-----TATTT	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Ivory Coast Sequence 1	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 2	-----	-----AAAC	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 3	-----	-----TAATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-A
Ivory Coast Sequence 4	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 5	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Ivory Coast Sequence 6	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 7	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATTCT-A
Singapore Sequence 1	-----	-----ATCTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACACGTA-A
Singapore Sequence 2	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Singapore Sequence 3	-----	-----AATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Singapore Sequence 4	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Nigerian Sequence 1	-----	-----TAAC	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Nigerian Sequence 2	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCG-A
Nigerian Sequence 3	-----	-----ATAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-A
NUCLEOTIDES INCLUDED	-----	-----mmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm--

	250	260	270	280	290	300
Gordius_aquaticus	AAAAG-TCGA	ATC-----	-----	-----	GCGAGGTCGG	ACCGCTTTT-
Priapulid_sp._caudatus	TCAAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCTTTT-
Brachionus_plicatilis	AAAAGCTCCG	ACC-----	-----	-----	GTATGGGAAG	AGCGCTTTT-
Chordodes_morgani	AAACG-TCGA	ATC-----	-----	-----	GCGAGGTTTCG	ACTGCTTTT-
Paractinolaimus_macrolaimus	AAAAGCTCTG	CCC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Aporcelaimellus_obtusicaudatus	AAAAGCTCCG	CCC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Wilsonema_schuurmansstekhoveni	ACAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCATTT-
Mesodorylaimus_sp._cf._nigritul	AAAAGCTCCG	ACC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Mesodorylaimus_bastiani	AAAAGCTCTG	ACC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Mesodorylaimus_japonicus	AAAAGCTCTG	CCCT-----	-----	-----	TCACGGAAG	AGCGCATTT-
Pungentus_sp._PDL_2005	TAAAGCTCTG	CCC-----	-----	-----	GTAAGGAACG	AGCGCATTT-
Allocladylaimus_sp	TAAAGCTCCG	CCC-----	-----	-----	GAAAGGAACG	AGCGCATTT-
Eudorylaimus_carteri	TAAAGCTCTG	CCC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Microdorylaimus_sp	TAAAGCTCTG	ACC-----	-----	-----	GAAAGGAACG	AGCGCATTT-
Longidorus_elongatus	AAAAGCTCAG	ACT-----	-----	-----	GAAAGGAATG	AGCGCATTT-
Xiphinema_rivesi	AAAAGCTCGT	TTC-----	-----	-----	GCAAGATTCG	AGCGCATTT-
Tylencholaimus_sp	AAAAGCTCCG	CCC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Anoplostoma_sp._BHMM_2005	TTATATCCAG	ACC-----	-----	-----	TTACGGAATG	GATGCGATT-
Adoncholaimus_fuscus	CAAAACCCCG	AC-----	-----	-----	-TAACGAAGG	GGTGCGTTT-
Enoplus_meridionalis	TTATGCCCTG	ACT-----	-----	-----	TCACGGAAG	GGCGCGTTT-
Enoplus_brevis_U88336	TTAAGTCCAG	ACC-----	-----	-----	TCACGGAACG	GACGCGGTT-
Enoplus_communis	TTAAGTCCAG	ACCT-----	-----	-----	-CACGGAACG	GACGCGGTT-
Enoploides_brunettii	CAAAAGCCCTA	TTGC-----	-----	-----	-----AG	GGCGCATTT-
Syringolaimus_striatocaudatus	TTATGCTCAG	ACC-----	-----	-----	TTACGGAATG	AGCGCATTT-
Ironus_dentifurcatus	CAAAAGCCCTG	ACC-----	-----	-----	GTAAGGAAGG	GGTGCACTT-
Calyptronema_maxweberi	CAAAACCCCA	ACT-----	-----	-----	-TTACGGAGG	GGTGCGTTT-
Viscosia_sp._BHMM_2005	CAAAACCCAA	CCT-----	-----	-----	-TACGGTGG	GGTGCGTTT-
Viscosia_viscosa	CAAAACCCAA	CCT-----	-----	-----	-TACGGTGG	GGTGCGTTT-
Pontonema_vulgare	CAAAACACCC	GACT-----	-----	-----	TCGATGGAGG	GGTGCGATTT
Oncholaimus_sp._BHMM_2005	CAAAAGCCCTG	ACT-----	-----	-----	-TCGGAAGG	GGTGCGTTT-
Alaimus_sp._PDL_2005	TGAAGCCGAA	ACC-----	-----	-----	TTACGGAATC	GGCGCATTT-
Prismatolaimus_intermedius	GAAAGCTCTG	ACC-----	-----	-----	GTTTCGGGAGG	AGCGCATTT-
Tobrilus_gracilis	CAAAAGCTCGG	ACC-----	-----	-----	TCACG-AAAC	AGCGCATTT-
Tripyla_cf._filicaudata_JH_2004	CAAAAGCTTCG	ACC-----	-----	-----	TTACGGAAGG	AGCGCATTT-
Bathylaimus_sp	CAAA-CGGCA	CCC-----	-----	-----	TTATGGGAGC	CGCGCTTTT-
Bathylaimus_assimilis	CAAA-CGGCA	CCC-----	-----	-----	TTATGGGAGC	CGCGCTTTT-
Bathylaimus_sp._BHMM_2005	CAAA-CGGCG	ACC-----	-----	-----TTTC	ACGAGGGAGC	CGCGCTTTT-
Tripyloides_sp._BHMM_2005	CAAA-CTGTG	ACTTC-----	-----	-----GC	AAGAAGAAGC	AGTGCTTTT-
Trischistoma_monohystera	TCAA-CTCTG	ACC-----	-----	-----	TTACGGAAGG	AGCGCGTTT-
Mermis_nigrescens	GAAAGCTCGG	ACC-----	-----	-----	TTCCGGGAAG	AGCGCATTT-
Mylonchulus_arenicolus	GAAAGCTCCG	ACCC-----	-----	-----	TTGAGGAAG	AGCGCATTT-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	250	260	270	280	290	300					
Anatonchus tridentacus	GTAAGCTCGG	ACC-----	-----	-----	TTAGGGAAGG	AGCGCATTT-					
Mononchus truncatus	GAAAAGCTCGG	ACC-----	-----	-----	TTT-GGAAAG	AGCGCATTT-					
Prionchulus muscorum	GAAGGCTCGG	ACC-----	-----	-----	TTTTGGGAAG	AGCGCATTT-					
Clarkus sp	GCAGGCTCGG	ACC-----	-----	-----	TTTTGGGAAG	AGCGCATTT-					
Trichinella spiralis	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCTTTT-					
Trichinella papuae	AAAAACTTCA	ACTG-----	-----	-----	AAATGGTTGA	AGTGCTTTT-					
Trichinella britovi	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCTTTT-					
Trichinella murrelli	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCTTTT-					
Trichinella pseudospiralis	AAAAACTTCA	ACTA-----	-----	TA	CAATGGTTGA	AGTGCTTTT-					
Trichinella nativa	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCTTTT-					
Trichinella zimbabwensis	AAAAACTTCA	ACTG-----	-----	-----	AAATGGTTGA	AGTGCTTTT-					
Trichinella nelsoni	AAAAACTTCA	ACTG-----	-----	-----	AATGGTTGA	AGTGCTTTT-					
Trichuris suis	CGAAGCTCGG	TCGCG-----	-----	-CG	CAACGCTCGG	AGCGCATT-					
Trichuris trichiura	CGAAGCTCAG	TCGC-----	-----	-GCGC	TGCGCGTCGG	AGCGCGTTT-					
Trichuris muris	CAAAGCTTCG	GCGC-----	-----	-GC	AATGCGCGGA	GACGCGATT-					
Tyololaimophorus minor	AAAAAAGCTCAG	ACC-----	-----	-----	TCGGGGAATG	AGTGCTTTT-					
Paratrichodorus pachydermus	TAAA-CGACG	ACC-----	-----	-----	TTACGGAAGT	CGTGCAATT-					
Paratrichodorus anemones	TAAA-CGACA	ACC-----	-----	-----	TCACGGAAGT	CGTGCAATT-					
Trichodorus primitivus	AAATCGCAC	ACC-----	-----	-----	TCACGGGAATG	CGTGCAATT-					
Axonolaimus helgolandicus	AACAGCTCTG	ACC-----	-----	-----	GCAAGGGATG	AGCGCATTT-					
Ascolaimus elongatus	AACAGCTCTG	ACC-----	-----	-----	GCAAGGGATG	AGCGCATTT-					
Odontophora rectangula	C-AAGCTCTG	ACC-----	-----	-----	GCAAGGGATG	AGCGCATTT-					
Cylindrolaimus sp. 202149	CAAAGCTCTG	ACC-----	-----	-----	GTAAGGGAGG	AGTGCAATT-					
Tylocephalus auriculatus	AAAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCATTT-					
Plectus acuminatus BS9	TAAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCATTT-					
Plectus aquatilis	TAAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCATTT-					
Anaplectus sp	CCAAGCTCCG	ACC-----	-----	-----	GTAAGGGACG	AGCGCATTT-					
Anisakis sp_WKT	CAAAGCTCCG	AT-----	-----	-----	TA-TTTGACG	AGCGCATCT-					
Anisakis sp_Nadler	CAAAGCTCCG	AT-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Contracaecum multipapillatum	CAAAGCTCCG	AT-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Pseudoterranova decipiens	CAAAGCTCCG	AT-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Raphidascaris acus	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Terranova caballeri	CAAAGCTCCG	AA-----	-----	-----	TC-ATTGACG	AGCGCATCT-					
Ascaris suum	CAAAGCTCCG	AT-----	-----	-----	TT-TCTGACG	AGCGCATCT-					
Ascaris lumbricoides	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Baylisascaris procyonis	CAAAGCTCCG	AA-----	-----	-----	TT-TCTGACG	AGCGCATCT-					
Baylisascaris transfuga	CAAAGCTCCG	AT-----	-----	-----	TT-TCTGACG	AGCGCATCT-					
Parascaris equorum	CAAAGCTCCG	AT-----	-----	-----	TT-TCTGACG	AGCGCATCT-					
Porrocaecum depressum	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Toxascaris leonina	CAAAGCTCAG	AT-----	-----	-----	TT-TCTGACG	AGCGCATTT-					
Heterocheilus tunicatus	CAAAGCTCCG	AA-----	-----	-----	TC-GTCGACG	AGCGCATCT-					
Goezia pelagia	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Hysterothylacium fortalezae	CAAAGCTCCG	AA-----	-----	-----	TC-TTTGACG	AGCGCATCT-					
Hysterothylacium pelagicum	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Hysterothylacium reliquens	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Iheringascaris iniquies	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-					
Toxocara canis	CAAAGCTCCG	AT-----	-----	-----	TT-TGTGACG	AGCGCATCT-					
Nemhelix bakeri	CAAAGCTCCG	AT-----	-----	-----	TT-T-TGACG	AGCGCATCT-					
Raillietnema sp. V3060	CAAAGCCCCG	AT-----	-----	-----	TT-T-TGACG	GGCGCATCT-					
Cruzia americana	CAAAGCTCCG	AC-----	-----	-----	TC-GTTGACG	AGCGCATCT-					
Heterakis sp_14690	CAAAGCTCCG	AT-----	-----	-----	---AATGACG	AGCGCATCT-					
Heterakis gallinarum	CAAAGCTCCG	AT-----	-----	-----	---TTTGACG	AGCGCATCT-					
Paraspidodera sp_21303	CGAAGCTCTG	AT-----	-----	-----	-T-TTTGACG	AGCGCATCT-					
Chromodora nudicapitata	CGAAGGTCAA	GC-----	-----	-----	GC-AAGCTTG	ATCGCACTT-					
Chromadora sp. BHMM_2005	TCAAGTCCCG	AC-----	-----	-----	TT-CGGAAGG	GACGCAATT-					
Atrochromadora microlaima	TCAAGTCCCG	AC-----	-----	-----	TT-CGGAAGG	GACGCAATT-					
Chromadorina germanica	AACAG--CCA	AC-----	-----	-----	TT-CGGAAGG	CTGGCAATT-					
Chromadorita tentabundum	TCAAGCCCTA	ACCT-----	-----	-----	TT-TGGAAGG	GGCGCAATT-					
Dichromadora sp. BHMM_2005	TCAAGTCCCG	AC-----	-----	-----	TT-CGGAAGG	GACGCAATT-					
Neochromadora BHMM_2005	TCAAGCCCCA	ACC-----	-----	-----	TGACGGGCGG	GGCGCAATT-					
Spilophorella paradoxa	TTAAACCCCA	AAC-----	-----	-----	TTACGTCGGG	GGTGCAATT-					
Paracanthonchus caecus	CAAAAACCTG	AC-----	-----	-----	TT-CGGAAGG	GGTGCAATT-					
Paracyatholaimus intermedius	CAAAAACCTG	AC-----	-----	-----	TT-CGGAAGG	GGTGCAATT-					
Praeacanthonchus punctatus	GAAAGCCCCG	AC-----	-----	-----	TT-CGGGAGG	GGCGCAATT-					
Praeacanthonchus sp	GAAAGCCC-G	AC-----	-----	-----	TT-CGGGAGG	GGCGCAATT-					
Cyatholaimus sp. BHMM_2005	CAAAAACCTG	AC-----	-----	-----	TT-CGGAAGG	GGTGCAATT-					
Spirinia parasitifera	AAAAAGTCCA	ACC-----	-----	-----	TTACGGGAAG	AGTGCAATT-					
Acanthopharynx micans	CCAAGCTCTG	ACC-----	-----	-----	TTACGGGAAG	AGCGCAATT-					
Xyzzors sp	CCAAGCTCTG	ACC-----	-----	-----	TTGGGGGAAG	AGTGCAATT-					
Metachromadora sp	CCAAGCTCCG	ACC-----	-----	-----	TTGCGGAAGG	AGCGCAATT-					
Metachromadora remanei	CCAAGCTCCG	ACC-----	-----	-----	TTGCGGAAGG	AGCGCAATT-					
Catanema sp	ACAAGCTCCG	ACT-----	-----	-----	TCGCGGAAGG	AGCGCAATT-					
Eubostrichus diana	TCAGGCTGCG	ACC-----	-----	-----	TTTTGGAAGC	AGTGCAATT-					
Eubostrichus topiarus	CCAAGCTCCA	ACC-----	-----	-----	TTACGGGAAG	AGCGCAATT-					
Eubostrichus parasitiferus	CCAAGCTCCA	ACC-----	-----	-----	TTACGGGAAG	AGCGCAATT-					
Laxus oneistus	TCAGGCTCCG	ACC-----	-----	-----	TTACGGGAAG	AGTGCAATT-					
Laxus cosmopolitus	TCAGGCTCCG	ACC-----	-----	-----	TTGCGGAAGG	AGCGCAATT-					
Leptonemella sp	CTCAGCTTCG	ACC-----	-----	-----	TTACGGGAAG	AGCGCAATT-					
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----					

	250	260	270	280	290	300
Heterorhabditis hepialus	CTATGCCCCA	ACT-----	-----	-----	--TCGGAAGG	GGTGCAATT-
Heterorhabditis zelandica	CTATGCCCCA	ACT-----	-----	-----	TC--GGAAGG	GGTGCAATT-
Parasitorhabditis_sp_SB281	GCGCTTC---	-----	-----	-----	-----G	GCGTCATTT-
Rhabditoides inermiformis	CAAAAGCTCTG	ACC-----	-----	-----	GTAAGGAACG	AGCGCATTT-
Rhabditoides inermis_DF5001	CAAACCCAG	ATC-----	-----	-----	GTAAGAGGCG	GGTGCAATT-
Rhabditoides regina_DF5012	GACCTT----	-----	-----	-----	-----CG	GGTTCATTT-
Poikilolaimus oxycerca_SB200	CAAAAGCTCTG	ACA-----	-----	-----	---CCCTCGG	AGCGCATTT-
Poikilolaimus regenfussi_SB199	CAAAGCTCTG	ACA-----	-----	-----	---CCCTCGG	AGCGCATTT-
Distolabrellus veechi_DWF1604	GCACGGGCAA	CCG-----	-----	-----	-----	-TGTCATTT-
Distolabrellus veechi_DF5024	GCACGGGCAA	CCG-----	-----	-----	-----	-TGTCATTT-
Choriorhabditis dudichi	TATGCCCCGA	CTC-----	-----	-----	---ACGGAGG	GGTGCAATT-
Protorhabditis_sp	ACATAACCGG	ACG-----	-----	-----	---CAAGTACG	GGTGCAATT-
Protorhabditis_sp_DF5055	CAAAGCTTGG	AAG-----	-----	-----	---CAATTCCA	AGTGCAGTT-
Cruzanema tripartitum_DF5015	CGAAACCCCG	AC-----	-----	-----	-TTGCGAAGG	GGTGCAATT-
Mesorhabditis_sp_PS1179	GCACCTTC---	-----	-----	-----	-----G	GTGTCATTT-
Mesorhabditis spiculigera_SB15	GTGCTTC---	-----	-----	-----	-----G	GCACCATTT-
Mesorhabditis anisomorpha_SB12	CTTCGGY---	-----	-----	-----	-----	--GTATAC-
Teratorhabditis palmarum_DF501	GCGTGCTTCG	G-----	-----	-----	-----	-CACCACTT-
Teratorhabditis synpapillata_S	TCGTGCTTCG	G-----	-----	-----	-----	-CACCACTT-
Caenorhabditis briggsae_PB102	CTAAACCCCA	ACG-----	-----	-----	--TAAGGCGG	GGTGCAATT-
Caenorhabditis elegans_N2	CTATACCCCA	ACG-----	-----	-----	--CAAGGCGG	GGTGCAATT-
Caenorhabditis_sp_CB5161	CTATACCCCA	ACG-----	-----	-----	--CAAGGCGG	GGTGCAATT-
Caenorhabditis_sp_PS1010	TTAAACCCCTG	ACG-----	-----	-----	--CAAGGACG	GGTGCAATT-
Caenorhabditis japonica	CTATACTCCA	ACG-----	-----	-----	--CAAGGCGG	GGTGCAATT-
Caenorhabditis_sp_DF5170	TTAAACCTCG	ACG-----	-----	-----	--TAAGGAGA	GGTGCAATT-
Caenorhabditis_sp_SB341	CTAAGCCCTG	ACG-----	-----	-----	--TAAGGAGG	GGTGCAATT-
Caenorhabditis plicata	AAATTCCCGG	ACG-----	-----	-----	--CAAGGAGG	GGGGCAATA-
Caenorhabditis vulgaris	CTAAGCCCCA	ACG-----	-----	-----	--TAAGGCGG	GGTGCAATT-
Caenorhabditis sonoriae	CTAAGCCCTG	ACG-----	-----	-----	--CAAGGAGG	GGTGCAATT-
Caenorhabditis drosophilae	TTAAACCTCA	ACG-----	-----	-----	--TAAGGCGA	GGTGCAATT-
Phasmarhabditis hermaphrodita	CAATGCTCAG	TTT-----	-----	-----	---CGGCTG	AGTGCAGTT-
Phasmarhabditis neopapillosa	CAATGCTCAG	TTT-----	-----	-----	---CGGCTG	AGTGCAGTT-
Phasmarhabditis neopapillosa	TAAAGCCTCG	ACT-----	-----	-----	TC--GGAAGA	GGTGCAGAT-
Pellioditis mediterranea_SB173	TTAAGCCTCG	ACT-----	-----	-----	TC--GGAAGA	GGTGCAGAT-
Pellioditis marina	TAAAGCCTCG	ACT-----	-----	-----	TC--GGAAGA	GGTGCAGAT-
Prodontorhabditis wirthi	TGATGTCTCG	CCCGACG--	-----	-----	--TAAGTCAG	GCTGCGGTT-
Crustorhabditis scanica	--ACGTCACT	-----	-----	-----	---YTCTGA	CGT--CATTT-
Dolichorhabditis_sp_CEW1	CAAAGCTAG	GCTC--G--	-----	-----	---CTTA	GCTGCAGGT-
Oscheius_sp_DF5000	TTAAAGCTAG	G-TTC----	-----	-----	---GTCTA	GCTGCAGGT-
Oscheius_sp_BW282	CAAACCTCG	ACTTT----	-----	-----	---GAAGA	GGTGCAGAT-
Oscheius myriophila_EM435	CAAACCTCG	ACTTT----	-----	-----	---GAAGA	GGTGCAGAT-
Oscheius insectivora	TAAACCCCG	ACTTT----	-----	-----	---GAAGG	GGTGCAGAT-
Oscheius dolichuroides	AAAAAGCTAG	ATTC-----	-----	-----	---GTCTA	GCTGCAGGT-
Rhabditis blumi_DF5010	C-ATGCCGTG	TCITT--G--	-----	-----	---ACAC	GGTGCATTT-
Rhabditis_sp_PS1191	CGAAACCCCG	ACTTT----	-----	-----	---GAAGG	GGTGCAGAT-
Rhabditis_sp_PS1010	TTAAACCTCG	ACGCAAG--	-----	-----	---GACG	GGTGCATTT-
Rhabditis myriophila_EM435	CAAACCTCG	ACTTT----	-----	-----	---GAAGA	GGTGCAGAT-
Rhabditis colombiana	TAAAGCCCG	-----	-----	-----	TC--ACGAAG	GGTGCAGAT-
Rhabditella axei_DF5006	TAAAGCCCAT	CCTTACG--	-----	-----	---GGTG	GGTGCAGAT-
Rhabditella_sp_DF5044	TAAAGCCCAT	CCTTACG--	-----	-----	---GGTG	GGTGCAGAT-
Cuticularia_sp_PS2083	CAAAGCTCTG	ACACCCCT-	-----	-----	---CGG-	AGCGCATTT-
Necator americanus	ATAAACCTCG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Kalicephalus cristatus	TTATACCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Ancylostoma caninum	ATAAACCTCG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus cantonensis	ATAAACCTCG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus costaricensis	ATAAACCTCG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus malaysiensis	ATAAACCTCG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus dujardini	TTAAGCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus vasorum	TTAACCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Aulurostrongylus abstrusus	TAGGACCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Didelphostrongylus hayesi	ATAAACCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Crenosoma mephitis	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Crenosoma vulpis	TTAGACCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Otostrongylus circumlitus	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Troglostrogylus wilsoni	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Filaroides martis	TTAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCATTT-
Oslerus osleri	TAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Parafilaroides decorus	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Metastrongylus salmi	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Metastrongylus elongatus	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Skrjabinstrongylus chitwoodrum	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Parelaphostrongylus odocoilei	ATAAACCTG	ACT-----	-----	-----	CT--GTGAAAG	GGTGCATTT-
Muelerius capillaris	TAAATCCTG	ACT-----	-----	-----	TT--TGAAAG	GATGCAATT-
Protostrongylus rufescens	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Halocercus invaginatus	TTAGACCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Pseudalius inflexus	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Stenurus minor	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
Torynurus convolutus	TTAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCATTT-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	250	260	270	280	290	300
Syngamus trachea	GTAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Stephanurus dentatus	ATAAGCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Nematodirus battus	TTAGGCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Strongylus equinus	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Labiostrongylus bipapillosus	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Petrovinema poculatum	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Cylicocyclus insignis	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Chabartia ovina	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Cyclodontostomum purvisi	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Zoniolaimus mawsonae	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Hypodontus macropi	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Deletrocephalus dimidiatus	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Dictyocaulus eckerti_P7B8	TAAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Dictyocaulus capreolus_P3B2	TAAAACCCCTA	ACG-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Dictyocaulus sp._P6A1	TAAAACCCCG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Dictyocaulus filaria	AATAACCCCTG	ACG-----	-----	-----	TT--TTGAAAG	GGTGCAAGTT--
Dictyocaulus capreolus_P2C10	TAAAACCCCTA	ACG-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Dictyocaulus viviparus	TAGAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Haemonchus sp._V3091	ATAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Haemonchus contortus	ATAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Haemonchus placei	ATAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Haemonchus similis	ATAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Ostertagia ostertagi	TTAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Ostertagia leptospicularis	TTAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Nippostrongylus brasiliensis	TAAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Heligmosomoides polygyrus	GAAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Trichostrongylus colubriformis	ATAAACCCCTG	ACT-----	-----	-----	TT--CGAAAG	GGTGCAAGTT--
Tetrabothriostongylus mackerr	TTAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Herpestrostrongylus pythionis	AGAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Nicollina cameroni	AGAAACCCCTG	ACT-----	-----	-----	TT--TGAAAG	GGTGCAAGTT--
Filarimena flagrifer	TGAAACCCCTG	ACT-----	-----	-----	CT--TGAAAG	GGTGCAAGTT--
Amidostomum cygni	ATATACCCCTG	ACT-----	-----	-----	AT--TGAAAG	GGTGCAAGTT--
Teratocephalus lirellus	CAAAGCTCAG	ACC-----	-----	-----	TTTCGGGACG	AGCGCATCTT--
Brumptaemilius justini	CAAAGCTCGG	ATT-----	-----	-----	TT--TGAAAG	AGCGCATCTT--
Anguillicola crassus	CAACGCTCCG	ATT-----	-----	-----	TAC--TGACG	AGCGCATCTT--
Dracunculus medinensis	CAAAGCTCCG	GAT-----	-----	-----	TTTATTGACG	AGCGCATCTT--
Dracunculus oesophageus	CAAAGCTCTG	AGA-----	-----	-----	TATATCGAGG	AGCGCATCTT--
Dracunculus sp._V3104	CAAAGCTCCG	ATT-----	-----	-----	TTA--TTGACG	AGCGCATCTT--
Philonema sp_A	CAAAGCTCCA	AAT-----	-----	-----	TTT--TGACG	AGCGCATCTT--
Philometra obturans	CAAAGCTCGG	ACTAC-----	-----	-----	AAA--TTT--AGACG	AGCGCATCTT--
Camallanus oxycephalus	CAAAGCTCTG	AT-----	-----	-----	TTTT--TGACG	AGCGCATCTT--
Acanthocheilonema viteae	CAAAGCTCCG	AC-----	-----	-----	TTAT--GGACG	AGCGCATCTT--
Loa loa	CAAAGCTCCG	AC-----	-----	-----	TTTT--GAACG	AGCGCATCTT--
Onchocerca cervicalis	CAAAGCTCCG	AC-----	-----	-----	TTTT--GGACG	AGCGCATCTT--
Dirofilaria immitis	CAAAGCTCTG	AC-----	-----	-----	TTTTTTGAACG	AGCGCATCTT--
Brugia malayi	CAAAGCTCCG	AA-----	-----	-----	TTTT--AAACG	AGCGCATCTT--
Wuchereria bancrofti	CAAAGCTCCG	AT-----	-----	-----	TTTT--AGACG	AGCGCATCTT--
Litomosoides sigmodontis	CAAAGCTCCG	AC-----	-----	-----	TGTTTGAACG	AGCGCATCTT--
Setaria digitata	CAAAGCTCCG	AC-----	-----	-----	TTTT--GGACG	AGCGCATCTT--
Gnathostoma turgidum	CAAAGCTCCG	AC-----	-----	-----	TCTG--TGACG	AGCGCATCTT--
Gnathostoma neoprocyonis	CAAAGCTCCG	AC-----	-----	-----	TTTG--TGACG	AGCGCATCTT--
Gnathostoma binucleatum	CAAAGCTCCG	AC-----	-----	-----	TCTG--TGACG	AGCGCATCTT--
Physaloptera alata	CAAAGCTCTG	AC-----	-----	-----	TTC--GGATG	AGCGCATCTT--
Physaloptera turgida	CAAAGCTCTG	AC-----	-----	-----	TTT--GGACG	AGCGCATCTT--
Ascarophis arctica	CAAAGCTCCG	AC-----	-----	-----	TTACTGGACG	AGCGCATCTT--
Spinitectus carolini	CAAAGCTCCG	AC-----	-----	-----	TTT--TTAGTGGACG	AGCGCATCTT--
Thelazia lacrymalis	CAAAGCTCAA	AC-----	-----	-----	TAC--GGATG	AGCGCATCTT--
Aphelenchus avenae	TAAAGCTCTG	AT-----	-----	-----	C--TCGCGGGAAG	AGTGCATTTT--
Aphelenchoides fragariae	ATAAGTACTT	TTCA-----	-----	-----	G--CAATGAGAAT	TATGCACTTT--
Bursaphelenchus sp	CAAGCCTCGG	CC-----	-----	-----	T--TCGGGTTCCG	GGTGCAAGTT--
Deladenus sp	ACATGCTCGG	ACC-----	-----	-----	TTGCGGAAAG	AGTGCATTTT--
Criconema sp	CGATGCTGTC	CC-----	-----	-----	TCGCGGGGTC	AGTGCATTTT--
Hemicyclophora conida	CGAAG--CTG	TC-----	-----	-----	CCCCAAGGGC	AGTGCATTTT--
Paratylenchus dianthus	CGAAGTTGTT	CC-----	-----	-----	TCACGGAGC--	AATGCATTTT--
Pratylenchus thornei	CAAAGCTCCG	AC-----	-----	-----	CCGAGGGGAG	AGCGCATTTT--
Tylenchulus semipentrans	CGAAGCTACC	TC-----	-----	-----	TTGC--GGGGT	AGTGCATTTT--
Ditylenchus angustus	CTAAGCTCTT	ACC-----	-----	-----	TTACGGAAAG	AGTGCATTTT--
Subanguina radicola	CAAAGCTCTG	ACC-----	-----	-----	TTACGGAAAG	AGTGCATTTT--
Tylenchorhynchus maximus	CAAAGCTCCG	ACC-----	-----	-----	GTAAGGGAAG	AGCGCATTTT--
Geocenamus quadriifer	CAAAGCTCTG	ACC-----	-----	-----	TTACGGGGAAG	AGCGCATTTT--
Globodera pallida	CAAAGCTCCA	ATCTT-----	-----	-----	CCGAGAGCGG	AGTGCATTTT--
Scutellonema bradys	CAAAGCTTCG	ATCCCTT---	-----	-----	GA--GGAGAGG	AGTGCATTTT--
Helicotylenchus dihystrera	CAAAGCTCCG	ATCCCTC---	-----	-----	GC--GGAGAGG	AGCGCATTTT--
Rotylenchus robustus	CAAAGCTCCG	ATCCCTT---	-----	-----	GCTGGAGAGG	AGCGCATTTT--
Meloidogyne arenaria	TAAAGCTTTG	TCC-----	-----	-----	TTACGG--AAA	AGCGCATTTT--
Meloidogyne incognita_KT	TAAAGCTTTG	TCC-----	-----	-----	TTACGGAAAG	--GCGCATTTT--
Meloidogyne javanica	CAAAGCTTTG	TCC-----	-----	-----	TTACGGWWMA	--GCGCATTTT--
Meloidogyne artiellia	CAAAGCTTTG	ACCCTTT---	-----	-----	ACGGGGAAAA	AGCGCATTTT--
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	250	260	270	280	290	300
Meloidogyne_duytsi	CAAAGCTTTG	TCC-----	-----	-----	-----	TTCCGGAAA-	AGCGCATTT-
Meloidogyne_exigua	TAAAGCTTTG	TCC-----	-----	-----	-----	TTACGGAAA-	AGCGCATTT-
Meloidogyne_hapla	TAAAGCTTTG	TCC-----	-----	-----	-----	TTACGGAAA-	AGCGCATTT-
Meloidogyne_ichinohei	AAAAGCTCCG	ACCC-----	-----	-----	-----	CTCGGGGAG	AGCGTATTT-
Meloidogyne_maritima	TAAAGCTTTG	TCC-----	-----	-----	-----	TTACGGAAA-	AGCGCATTT-
Meloidogyne_microtyla	TAAAGCTTTG	TCC-----	-----	-----	-----	TTACGGAAA-	AGCGCATTT-
Nacobbus_ aberrans	AAAAGCTCCG	ACC-----	-----	-----	-----	TAAAGGAAA-	AGCGCATTT-
Pratylenchoides_ritteri	CAAAGCTCCG	ACC-----	-----	-----	-----	TTACGGGAA-	AGCGCATTT-
Pratylenchoides_magnicauda	CAAAGCTCCG	ACC-----	-----	-----	-----	TTACGGGAA-	AGCGCATTT-
Hirschmanniella_sp._JH_2003	CAAAGCTCCG	ACC-----	-----	-----	-----	GCGAGGGA-	GGCGCATTT-
Radopholus_similis	AAAAGCTCCC	AAC-----	-----	-----	-----	GCAAGGAGG	AGCGCATTT-
Pratylenchus_goodeyi_VF	CAAAGCTCTG	ATCCGCGCTT	-----	-----	-----	GCGTGGAAG	AGCGCATTT-
Boleodorus_thylactus_clone2	CAAAGCTCCG	ACC-----	-----	-----	-----	TCGCGGGAG	AGCGCATTT-
Philippine Sequence 1	ATAAACCTTG	ACT-----	-----	-----	-----	TT--CGAAAG	GGTGCAATT-
Philippine Sequence 2	ATAAACCTTG	ACT-----	-----	-----	-----	TT--CGAAAG	GGTGCAATT-
Philippine Sequence 3	TTTAGCCCCG	ACT-----	-----	-----	-----	CA--CGAAGG	GGTACGGTT-
Philippine Sequence 4	AAAAGTTGAA	TCT-----	-----	-----	-----	TA--TGGATT	AATGCGTAT-
Philippine Sequence 5	ATAAACCTTG	ACT-----	-----	-----	-----	TT--CGAAAG	GGTGCAATT-
Philippine Sequence 6	CAAAGCCTTG	ACT-----	-----	-----	-----	TC--GGAAAG	GGTGCAATT-
Philippine Sequence 7	TACATCCCCA	ACG-----	-----	-----	-----	--TAAGGCGG	GGTGCAATT-
Tahiti Sequence 1	ATAAACCTTG	ACT-----	-----	-----	-----	TT--CGAAAG	GGTGCAATT-
Tahiti Sequence 2	ATAAACCTTG	ACT-----	-----	-----	-----	TT--CGAAAG	GGTGCAATT-
Thailand Sequence 1	CAAAGCTCCG	AT-----	-----	-----	-----	--TTGAAG	AGCGCATCT-
Ogasawara Sequence 1	AAAAGTTGAA	ACT-----	-----	-----	-----	-TATGTTTC	AATGCGTAT-
Ogasawara Sequence 2	CAATGCTCAG	TTT-----	-----	-----	-----	----CGGCTG	AGTGCAGTT-
Ivory Coast Sequence 1	TTAAACCTTG	ACT-----	-----	-----	-----	TT--TGAAAG	GGTGCAATT-
Ivory Coast Sequence 2	TTAAACCTTG	ACT-----	-----	-----	-----	TT--TGAAAG	GGTGCAATT-
Ivory Coast Sequence 3	YTAAACCTTG	ACT-----	-----	-----	-----	TT--TGAAAG	GGTGCAATT-
Ivory Coast Sequence 4	TTAAACCTTG	ACT-----	-----	-----	-----	TT--TGAAAG	GGTGCAATT-
Ivory Coast Sequence 5	CAAAGCTCCG	AT-----	-----	-----	-----	---TTGACG	AGCGCATCT-
Ivory Coast Sequence 6	TTAAACCTTG	ACT-----	-----	-----	-----	TT--TGAAAG	GGTGCAATT-
Ivory Coast Sequence 7	TAAACCTTG	ACT-----	-----	-----	-----	TT--TGAAAG	GGTGCAATT-
Singapore Sequence 1	TTTAGCCCCG	ACT-----	-----	-----	-----	CA--CGAAGG	GGTACGGTT-
Singapore Sequence 2	CAAAGCTCCG	AT-----	-----	-----	-----	---TTGAAG	AGCGCATCT-
Singapore Sequence 3	CTAAACCTTG	ACT-----	-----	-----	-----	--TAAGGCGG	GGTGCAATT-
Singapore Sequence 4	TAAACCTTG	ACT-----	-----	-----	-----	TT--TGAAAG	GGTGCAATT-
Nigerian Sequence 1	CTAAACCTTG	ACT-----	-----	-----	-----	TT--TGAAAG	GGTGCAATT-
Nigerian Sequence 2	ATAACCTTG	ACT-----	-----	-----	-----	TT--CGAAAG	GGTGCAATT-
Nigerian Sequence 3	GTAAACCTTG	ACT-----	-----	-----	-----	TA--TGAAAG	GGTGCAATT-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	-----

	310	320	330	340	350	360
Gordius_aquaticus	A-TTAGA---	ACAAAACTA	ACG-----	-----	-----	-----	-----
Priapulius_caudatus	A-TTAGA---	CC-AAACCAA	TCG-----	-----	-----	GG TTT----	CG
Brachionus_plicatilis	A-TTAGA---	CCAAAAACCAA	TGG-----	-----	-----	---GG T----	CG
Chordodes_morgani	A-TTAGA---	ACAAAACTA	ACA-----	-----	-----	-----	-----
Paractinolaimus_macrolaimus	A-TTAGAA--	TAAAAACCAA	TCA-----	-----	-----	---GG TTT----	CG
Aporcelaimellus_obtusicaudatus	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---GG TTT----	CG
Wilsonema_schuurmansstekhoveni	A-TTAGA---	CCAAAAACCAA	TCG-----	-----	-----	---GG CTT----	CG
Mesodorylaimus_sp_cf_nigritul	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---GG CCT----	CG
Mesodorylaimus_bastiani	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---GG TTT----	CG
Mesodorylaimus_japonicus	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---GG CTT----	CG
Pungentus_sp._PDL_2005	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---GG TTT----	CG
Allodorylaimus_sp	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---AG CTT----	CG
Eudorylaimus_carteri	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---AG CTT----	CG
Microdorylaimus_sp	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---AG CTT----	CG
Longidorus_elongatus	A-TTAKAA--	TAAAAACCAA	TCG-----	-----	-----	---GG TCT-A--	AAA
Xiphinema_rivesi	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---GG CCCAT--	AAA
Tylencholaimus_sp	A-TTAGAA--	TAAAAACCAA	TCG-----	-----	-----	---GG CCT---	CGT
Anoplostoma_sp._BHMM_2005	A-TTAGAA--	TAAA-ACCAA	GCA-----	-----	-----	---GG CTT---	GCC
Adoncholaimus_fuscus	G-TTACT---	TCAA-ACCAA	TCG-----	-----	-----	---GG CTT----	CG
Enoplus_meridionalis	A-TTAGA---	CCAAAAACCAA	TCG-----	-----	-----	---GG CTT----	GT
Enoplus_brevis_U88336	A-TTAGA---	CCAAAAACCAA	TCG-----	-----	-----	---GG CTT----	G
Enoplus_communis	A-TTAGA---	CCAAAAACCAA	TCG-----	-----	-----	---GG CTT----	GT
Enoploides_brunettii	A-TTAGG---	ACAAAAACCAA	TCG-----	-----	-----	---GG CTT----	GCC
Syngnathoides_striatocaudatus	A-TTAGA---	ACAAAAACCAA	TCG-----	-----	-----	---GA CTT----	CGG
Ironus_dentifurcatus	A-TTAGA---	ACAAAAACCAA	TCG-----	-----	-----	---AG CCT----	CTG
Calyptronema_maxweberi	G-TTACT---	TCAA-ATCAA	TCG-----	-----	-----	---GA CTT----	CG
Viscosia_sp._BHMM_2005	G-TTACA---	ACAA-ACCGA	TCA-----	-----	-----	---GA CCT----	CG
Viscosia_viscosa	G-TTACT---	TCAA-ACCAA	TCG-----	-----	-----	---GA CTT----	CG
Pontonema_vulgare	GATTACT---	TCAA-ATCAA	TCG-----	-----	-----	---GA CTT----	CG
Oncholaimus_sp._BHMM_2005	G-TTACT---	TCAA-ACTGA	TCG-----	-----	-----	---GT CTT----	CG
Alaimus_sp_PDL_2005	A-TTAGA---	TCAAAAACCAA	TCG-----	-----	-----	---GA CTT----	CG
Prismatolaimus_intermedius	A-TTAGA---	ACAAAAACCAA	TCG-----	-----	-----	---GG CCTT----	CG
Tobrilus_gracilis	A-TTAGA---	GCAAAAAACCAA	TCG-----	-----	-----	---GG CTT----	CG
Tripyla_cf_filicaudata_JH_2004	A-TTAGA---	CTAAAGCCAA	TCG-----	-----	-----	---GA CCT----	TCGG
NUCLEOTIDES INCLUDED	m-mmmmm---	mmmm-mmmmm	mmmm-----	-----	-----	-----	-----

	310	320	330	340	350	360
Bathylaimus_sp	A-TTAGA---	ACAAAACCAA	TCG-CTTCA-	-----	-----	GTG CTT----	CG
Bathylaimus_assimilis	A-TTAGA---	ACAAAACCAA	TCG-CTTCA-	-----	-----	GTG CTT----	CG
Bathylaimus_sp._BHMM_2005	A-TTAGA---	ACAAAACCAA	TCG-CGCTG-	-----	-----	TGG TTT----	CG
Tripyloides_sp._BHMM_2005	A-TTAGA---	ACAAAACCAA	TCG----CG-	-----	-----	CTA CTT----	CG
Trischistoma_monohystera	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GG CTT----	CG
Mermis_nigrescens	A-TTAGA---	TCAAAACCAA	TCG-----	-----	-----	GG CTT----	TT
Mylonchulus_arenicolus	A-TTAGA---	CCAAAACCAA	TCG----A-	-----	-----	GTA CTT----	CG
Anatonchus_tridentacus	A-TTAGA---	CCAAAACCAA	TCA-----	-----	-----	AG CTT----	CG
Mononchus_truncatus	A-TTAGA---	CCAAAACCAA	TCG-----	-----	-----	AG TTT----	CG
Prionchulus_muscorum	A-TTAGA---	CCAAAACCAA	TCG-----	-----	-----	AG CTT----	CG
Clarkus_sp	A-TTAGA---	CCAAAACCAA	TCG-----	-----	-----	GG CTT----	CG
Trichinella_spiralis	A-TTAGAA--	CAAAAACCGA	ACG-----	-----	-----	GCG TTT----	GA
Trichinella_papuae	A-TTAGAA--	CAAAAACCGA	ACG-----	-----	-----	GCG TCT----	GG
Trichinella_britovi	A-TTAGAA--	CAAAAACCGA	ACG-----	-----	-----	GCG TTT----	GG
Trichinella_murrelli	A-TTAGAA--	CAAAAACCGA	ACG-----	-----	-----	GCG TTT----	GG
Trichinella_pseudospiralis	A-TTAGAA--	CAAAAACCGA	ACG-----	-----	-----	GCG TTC----	GA
Trichinella_nativa	A-TTAGAA--	CAAAAACCGA	ACG-----	-----	-----	GCG TTT----	GA
Trichinella_zimbabwensis	A-TTAGAA--	CAAAAACCGA	ACG-----	-----	-----	GCG TCT----	GG
Trichinella_nelsoni	A-TTAGAA--	CAAAAACCGA	ACG-----	-----	-----	GCG TTT----	GA
Trichuris_suis	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GAC GCA----	G
Trichuris_trichiura	A-TTTGA---	ACAAAACCAA	TCG-----	-----	-----	GAC GCA----	G
Trichuris_muris	A-TTGGT---	ACAAAACCAA	TCG-----	-----	-----	GAC TTT----	CG
Tyololaimophorus_minor	A-TTAGT---	-CTAAACCAA	TCGG-----	-----	-----	GCC GCA----	AG
Paratrichodorus_pachydermus	G-TTAGA---	CC-AAACCAA	TCG-----	-----	-----	GTC TT-----	CG
Paratrichodorus_anemones	G-TTAGA---	-AAAAACCAA	TCG-----	-----	-----	GTC TT-----	CG
Trichodorus_primitivus	A-TTAGA---	-TTAAACCAA	TCG-----	-----	-----	GTC CTT-----	CGG
Axonolaimus_helgolandicus	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT-----	GCC
Ascolaimus_elongatus	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT-----	GCC
Odontophora_rectangula	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GG- TT-----	CGT
Cylindrolaimus_sp._202149	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT-----	CGG
Tylocephalus_auriculatus	A-TTAGA---	CCAAAACCAA	TCG-----	-----	-----	GGC TT-----	CGG
Plectus_acuminatus_BS9	A-TTAGA---	CCAAAACCAA	TCG-----	-----	-----	GGC TT-----	CGG
Plectus_aquaticus	A-TTAGA---	CCAAAACCAA	TCG-----	-----	-----	GGC TT-----	CGG
Anaplectus_sp	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT-----	CGG
Anisakis_sp_WKT	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Anisakis_sp_Nadler	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Contracaecum_multipapillatum	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TA-----	ACG
Pseudoterranova_decipiens	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Raphidascaris_acus	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Terranova_caballeroi	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT CT-----	CGG
Ascaris_suum	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Ascaris_lumbricoides	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Baylisascaris_procyonis	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Baylisascaris_transfuga	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Parascaris_equorum	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Porrocaecum_depressum	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Toxascaris_leonina	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Heterocheilus_tunicatus	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT CT-----	CGG
Goezia_pelagia	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Hysterothylacium_fortalezae	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Hysterothylacium_pelagicum	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GAC CT-----	CGG
Hysterothylacium_reliquens	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Iheringascaris_inquies	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CCG
Toxocara_canis	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Nemhelix_bakeri	A-TTAGA---	TTAAAGCCAA	TCG-----	-----	-----	GGT TT-----	CGG
Raillietnema_sp._V3060	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GAT TT-----	CGG
Cruzia_americana	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Heterakis_sp_14690	A-TTAGA---	TTAAACCAA	TCA-----	-----	-----	GGT TT-TGT-	CGA
Heterakis_gallinarum	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGT TT-----	CGG
Paraspidodera_sp_21303	A-TTAGA---	TTAAACCAA	TCG-----	-----	-----	GGC YT-----	CGG
Chromodora_nudicapitata	C-TTAGA---	-AAAGACCAA	TTG-----	-----	-----	GC TT-----	CGG
Chromadora_sp._BHMM_2005	G-TTAGA---	ACAAGACCAA	TTG-----	-----	-----	GC TT-----	CGG
Atrochromadora_microlaima	G-TTAGA---	ACAAGACCAA	TTG-----	-----	-----	GC TT-----	CGG
Chromadorina_germanica	A-TTAGT---	ACAAGACCAA	TTG-----	-----	-----	GC TT-----	CGG
Chromadorita_tentabundum	A-TTAGA---	ACAAAACCAA	TTG-----	-----	-----	GC TT-----	CGG
Dichromadora_sp._BHMM_2005	G-TTAGA---	ACAAGACCAA	TTG-----	-----	-----	GC TT-----	CGG
Neochromadora_BHMM_2005	A-TTAGA---	ACAAAACCAA	TTG-----	-----	-----	GC TT-----	CGG
Spilophorella_paradoxa	A-TTAGA---	ACAAAACCAA	TTG-----	-----	-----	GC TT-----	CGG
Paracanthonchus_caecus	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GAC TT-----	G
Paracyatholaimus_intermedius	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT-----	G
Praeacanthonchus_punctatus	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT-----	G
Praeacanthonchus_sp	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT-----	G
Cyatholaimus_sp._BHMM_2005	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GAC TT-----	G
Spirinia_parasitifera	A-TTAGA---	CCAAGACCAA	TCA-----	-----	-----	GGC TT-----	TGC
Acanthopharynx_micans	A-TTAGA---	CCAAGACCAA	TTA-----	-----	-----	GGC TC-----	CGG
Xyzzors_sp	G-TTAGA---	CCAAGACCAA	TCA-----	-----	-----	GGC TT-----	TGC
Metachromadora_sp	A-TTAGA---	CCAAGACCAA	TCG-----	-----	-----	GGC TC-----	TGC
Metachromadora_remanei	A-TTAGA---	CCAAGACCAA	TCG-----	-----	-----	GGC TC-----	TGC
NUCLEOTIDES INCLUDED	m-mmmmm---	mmmm-mmmmm	mmmm-----	-----	-----	-----	-----

	310	320	330	340	350	360
Parelaphostrongylus_odocoilei	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT----	CGG
Muelerius_capillaris	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Protostrongylus_rufescens	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Halocercus_invaginatus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT TT----	CGG
Pseudalius_inflexus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT TT----	CGG
Stenurus_minor	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT TT----	CGG
Torynurus_convolutus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT TT----	CGG
Syngamus_trachea	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Stephanurus_dentatus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Nematodirus_battus	A-TTAGA---	GCAA-ATCAA	TCT-----	-----	-----	CC TT----	CGG
Strongylus_equinus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Labiostrongylus_bipapillosus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Petrovinema_poculatum	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Cylicocyclus_insignis	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Chabartia_ovina	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Cyclodontostomum_purvisi	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Zoniolaimus_mawsonae	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Hypodontus_macropi	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Deletrocephalus_dimidiatus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Dictyocaulus_eckerti_P7B8	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----	CGG
Dictyocaulus_capreolus_P3B2	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----	CGG
Dictyocaulus_sp._P6A1	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----	CGG
Dictyocaulus_filaria	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TCCT TA----	CGG
Dictyocaulus_capreolus_P2C10	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----	CGG
Dictyocaulus_viviparus	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----	CGG
Haemonchus_sp._V3091	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CT TT----	CGG
Haemonchus_contortus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CT TT----	CGG
Haemonchus_placei	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CT TT----	CGG
Haemonchus_similis	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CT TT----	CGG
Ostertagia_ostertagi	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Ostertagia_leptospicularis	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Nippostrongylus_brasileinsis	A-TTAGT---	GCAA-ACCAA	TCA-----	-----	-----	TC TT----	CGG
Heligmosomoides_polygyrus	A-TTAGA---	GCAA-ACCAA	TCA-----	-----	-----	CC TT----	CGG
Trichostrongylus_colubriformis	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CC TT----	CGG
Tetrabothriostongylus_mackerr	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----	CGG
Herpestrongylus_pythonis	A-TTAGA---	GCAA-ACCAA	TCA-----	-----	-----	CC TT----	CGG
Nicollina_cameroni	A-TTAGA---	GCAA-ACCAA	TCA-----	-----	-----	CC TT----	CGG
Filarimena_flagrifer	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CC CT----	CGG
Amidostomum_cygni	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CC TT----	CGG
Teratocephalus_lirellus	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT----	CGG
Brumptaemilius_justini	A-TTAGA---	TTAAAACCAA	TCG-----	-----	-----	GGT CT----	CGG
Anguillicola_crassus	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	AGC TG----	CGG
Dracunculus_medinensis	A-TAAGGA--	TTAAAACCAA	GTC-----	-----	-----	GTAGTCT TTAAA--	CGG
Dracunculus_oesophageus	A-TTAGA---	CTAAAACCAA	TCG-----	-----	-----	AGCGGT TTTAAAAC	CTG
Dracunculus_sp._V3104	A-TTAGA---	CTAAAACCAA	TCG-----	-----	-----	AGCCAA TTAAA--	CGG
Philonema_sp._A	A-TTAGA---	TTAAAACCAA	TCGGGCATAT	AACTTCATTC	GTGGAGTGAT	A-----	GG
Philometra_obturans	A-TTAGAC--	TAAAACCAA	TCGAGATGGT	GTCTCTCCGC	GCCATGCGGG	-TGGCATAAC	
Camallanus_oxyccephalus	A-TTAGAC--	CAAAAACCAA	TCG-----	-----	-----	AGA TTATTTCGT	CG
Acanthocheilonema_viteae	A-TTAGA---	TTAAAACCAA	TCG-----	-----	-----	GGT TATTGGCTTA	
Loa_loa	A-TTAGA---	TTAAAACCAA	TCG-----	-----	-----	GGT TATT----	AG
Onchocerca_cervicalis	A-TTAGA---	TTAAAACCAA	TCG-----	-----	-----	GGT TATT----	GG
Dirofilaria_immitis	A-TTAGA---	TTAAAACCAA	TCGGAACAAA	-----	-----	TTT GATCT--	GA
Brugia_malayi	A-TTAGA---	TTAAAACCAA	TCG-----	-----	-----	GATT ATTAATTTTA	
Wuchereria_bancrofti	A-TTAGA---	TTAAAACCAA	TCG-----	-----	-----	GATT ATTAATTAAC	
Litomosoides_sigmodontis	A-TTAGA---	TTAAAACCAA	TGG-----	GGTGGTC	ATCAGCTGCT	GCTGCTGGCC	
Setaria_digitata	A-TTAGA---	TTAAAACCAA	TCG-----	GGTGGTC	-----	GGT TATTT--	GA
Gnathostoma_turgidum	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	AGC TTC-----	GG
Gnathostoma_neoprocyonis	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	AGC TTC-----	GG
Gnathostoma_binucleatum	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	AGC TTC-----	GG
Physaloptera_alata	A-TTAGA---	TTAAAACCAA	TCG-----	-----	-----	GGC TTC-----	CG
Physaloptera_turgida	A-TTAGA---	TTAAAACCAA	TCG-----	-----	-----	GGC TTC-----	AA
Ascarophis_arctica	A-TTAGG---	TTAAAACCAA	TCG-----	-----	-----	GGC TT-----	TG
Spinitectus_carolini	A-TTAGA---	TTAAATCCAA	TCG-----	-----	-----	GTG AAC-----	TT
Thelazia_lacrymalis	A-TTAGAATG	AAAAAACCAA	TGG-----	-----	-----	GGT TT-----	ATG
Aphelenchus_avenae	A-TTAGA---	TCAAAACCAA	GCG-----	-----	-----	G-C TT-----	CG
Aphelenchoides_fragariae	A-TCGGA---	GCAAAACTAA	TCT-----	-----	-----	-----	T----
Bursaphelenchus_sp	A-TTGGA---	GCAAAACCAA	TTT-----	-----	-----	G- TA-----	
Deladenus_sp	A-TTAGA---	ACAAAACCAA	GCG-----	-----	-----	G-C TT-----	CGG
Criconema_sp	G-TTGGA---	GCAAAACCAA	GCG-----	-----	-----	G-C TT-----	CGG
Hemicycliophora_conida	A-TTGGA---	GCAAAACCAA	GCG-----	-----	-----	AC TT-----	CGG
Paratylenchus_dianthus	A-TTGA---	GCAAAACCAA	GCG-----	-----	-----	G-C TT-----	CGG
Pratylenchus_thornei	A-TTAGA---	ACAAAACCAA	GCG-----	-----	-----	G-C TT-----	CGG
Tylenchulus_semipetrans	A-TTGGA---	GCAAAACCAA	GCG-----	-----	-----	G-C TT-----	CGG
Ditylenchus_angustus	A-TTAGA---	ACAAAACCAA	GCG-----	-----	-----	G-C TT-----	G
Subanguina_radicola	A-TTAGA---	ACAAAACCAA	GCG-----	-----	-----	AC TT-----	CGG
Tylenchorhynchus_maximus	A-TTAGA---	ACAAAACCAA	GCG-----	-----	-----	G-C TT-----	CGG
Geocnamus_quadriker	A-TTCGA---	ACAAAACCAA	GCG-----	-----	-----	G-C TT-----	CGG
Globodera_pallida	G-TTCGCC--	ACAAAACCAA	GCG-----	-----	-----	C-C TT-----	CGG
NUCLEOTIDES INCLUDED	m-mmmmm--	mmmm-mmmmm	mmmm-----	-----	-----	-----	-----

	370	380	390	400	410	420	
Viscospia_viscosa	G--TCTG---	-----	-----	AAATCAAGTG	A--TATTGAA	CAATTTA---	
Pontonema_vulgare	G--TCTG---	-----	-----	ATGTCAAATG	A--TATTGAA	CAATTTA---	
Oncholaimus_sp._BHMM_2005	G--ACTG---	-----	-----	TAATCAAGTG	A--GATTGAA	CTATTTA---	
Alaimus_sp._PDL_2005	G--TCCG---	-----	-----	CTCTTTGGTG	A--ATCTGAA	TAACTCA---	
Prismatolaimus_intermedius	G--GCCCG---	-----	-----	CCAGTTGGTG	A--ATCTGAA	TAACCTT---	
Tobrilus_gracilis	G--CCCG---	-----	-----	TTCTTTGGTG	A--ATCTGAA	TAACCTCT---	
Tripyla_cf._filicaudata_JH_2004	G--TCCG---	-----	-----	TCCATTGGTG	A--TTCTAAA	TAACCTCT---	
Bathylaimus_sp.	G--CATTGTT	GCG-----	-----	TAATTTGGTG	A--GTCTGAA	TAACCTTG---	
Bathylaimus_assimilis	G--CATTGTT	GCG-----	-----	TAATTTGGTG	A--GTCTGAA	TAACCTTG---	
Bathylaimus_sp._BHMM_2005	G--CTGCAGT	GCG-----	-----	TCATTTGGTG	A--GTCTGAA	TAACCTTG---	
Tripyloides_sp._BHMM_2005	G--TGGTGCG	-----	-----	TTATTTGGTG	A--ATCTGAA	TAACCTTG---	
Trischistoma_monohystera	G--CCCG---	-----	-----	TCATTTGGTG	A--ATCTGAA	TAACCTTT---	
Mermis_nigrescens	G--CTCG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACCTCA---	
Mylonchulus_arenicolus	G--TGCTTG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACCTTA---	
Anatonchus_tridentacus	G--CTTG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACCTAA---	
Mononchus_truncatus	G--CTTG---	-----	-----	ATTGTTGGTG	A--CTCTGAA	TAACCTAA---	
Prionchulus_muscorum	G--CTTG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACCTAA---	
Clarkus_sp.	G--CCTG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACCTAA---	
Trichinella_spiralis	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACCTATT---	
Trichinella_papuae	C--TGATGCT	GAACGTCGAA	A-----	CTCTTTGGTG	A--ATCTGAA	TAACCTATT---	
Trichinella_britovi	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACCTATT---	
Trichinella_murrelli	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACCTATT---	
Trichinella_pseudospiralis	C--TTGTGTT	GAACGTCGAA	A-----	CTCTTTGGTG	A--ATCTGAA	TAACCTATT---	
Trichinella_nativa	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACCTATT---	
Trichinella_zimbabwensis	C--TGATGCT	GAACGTCGAA	A-----	CTCTTTGGTG	A--ATCTGAA	TAACCTATT---	
Trichinella_nelsoni	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACCTATT---	
Trichuris_suis	G--CTATCTA	TTGGTCTGGG	TCCGCGA---	AAGGTTGGTG	A--ATCGGAA	TAACCTAT---	
Trichuris_trichiura	G--CTAGCTA	TTGGTCTGAG	TCCGCGA---	AATGTTGGTG	A--ATCGGAA	TAACCTAT---	
Trichuris_muris	C--TCTTTGC	GTCCGTCCGT	CA-----	ACGTTTGGTG	A--ATCGGAA	TAACCTAT---	
Tyololaimophorus_minor	G--TCCG---	-----	-----	CTCTTTGGTG	A--ATCTGAA	TAACCTTA---	
Paratrachodorus_pachydermus	G--ACCG---	-----	-----	CCATTTGGTG	A--ACTCTGAA	TAACCTTT---	
Paratrachodorus_anemones	G--ACCG---	-----	-----	CCATTTGGTG	A--CTCTGAA	TAATTTCC---	
Trichodorus_primitivus	G--ACCG---	-----	-----	CCATTTGGTG	A--ACTCTGAA	TAACCTTA---	
Axonolaimus_helgolandicus	---T-----	-----	-----	GTCTTTGGTG	G--ATCTGAA	TAACCTCA---	
Ascolaimus_elongatus	---T-----	-----	-----	GTCTTTGGTG	G--ATCTGAA	TAACCTCA---	
Odontophora_rectangula	---CCC-----	-----	-----	GTCTTTGGTG	G--ATCTGAA	TAACCTCA---	
Cylindrolaimus_sp._202149	---CCT-----	-----	-----	GTGTTTGGTG	G--ATCTGAA	TAACCTCA---	
Tylocephalus_auriculatus	---CCTG---	-----	-----	AACGTTGGTG	A--CTCTGAA	TAACCTGA---	
Plectus_acuminatus_BS9	---CCTG---	-----	-----	AACGTTGGTG	A--CTCTGAA	TAACCTGA---	
Plectus_aquatilis	---CCTG---	-----	-----	AACGTTGGTG	A--CTCTGAA	TAACCTGA---	
Anaplectus_sp.	---CCTG---	-----	-----	AGGTTTGGTG	A--ATCTGAA	TAACCTGA---	
Anisakis_sp._WKT	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATA---	
Anisakis_sp._Nadler	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATG---	
Contracecum_multipapillatum	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATA---	
Pseudoterranova_decipiens	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATA---	
Raphidascaris_acus	---CCCG---	-----	-----	TTTGTTGGTG	A--CTCTGAA	TAACCTATA---	
Terranova_caballeroi	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTAAG---	
Ascaris_suum	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATA---	
Ascaris_lumbricoides	---CCCG---	-----	-----	TAAATTGGTG	A--CTCTGAA	TAACCTATA---	
Baylisascaris_procyonis	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATA---	
Baylisascaris_transfuga	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATA---	
Parascaris_equorum	---CCCG---	-----	-----	TTAATTGGTG	A--CTCTGAA	TAACCTATA---	
Porrocaecum_depressum	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATA---	
Toxascaris_leonina	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATT---	
Heterocheilus_tunicatus	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATG---	
Goezia_pelagia	---CCCG---	-----	-----	TTTGTTGGTG	A--CTCTGAA	TAACCTATG---	
Hysterothylacium_fortalezae	---CCCG---	-----	-----	TTTGTTGGTG	A--CTCTGAA	TAACCTATG---	
Hysterothylacium_pelagicum	---CCCG---	-----	-----	TTTGTTGGTG	A--CTCTGAA	TAACCTATA---	
Hysterothylacium_reliquens	---CCCG---	-----	-----	TTTGTTGGTG	A--CTCTGAA	TAACCTATG---	
Iheringascaris_inquies	---CCCG---	-----	-----	TTTGTTGGTG	A--CTCTGAA	TAACCTATG---	
Toxocara_canis	---CCCG---	-----	-----	TAAATTGGTG	A--CTCTGAA	TAACCTGTA---	
Nemhelix_bakeri	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAGCTATG---	
Raillietnema_sp._V3060	---TCCG---	-----	-----	TCATTTGGTG	A--CTCTGAA	TAGCTATC---	
Cruzia_americana	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACCTATG---	
Heterakis_sp._14690	---CCTG---	-----	-----	TTCATTGGTG	A--CTCTGAA	TAGCTCA---	
Heterakis_gallinarum	---CCTG---	-----	-----	TTTATTGGTG	A--CTCTGAA	TAGCTTA---	
Paraspidodera_sp._21303	---CCCG---	-----	-----	TCAACTGGTG	A--CTCTGAA	TAGCTATA---	
Chromodora_nudicapitata	C---CA---	-----	-----	TCCATTGGTG	A--ATCTCC	GAAGAAA---	
Chromadora_sp._BHMM_2005	C---CA---	-----	-----	TCTATTGGTG	A--ATCTGAG	CAACTTA---	
Atrochromadora_microlaima	C---CA---	-----	-----	TCTATTGGTG	A--ATCTGAG	CAACTTA---	
Chromadorina_germanica	C---CA---	-----	-----	TCTATTGGTG	A--ATCTGAA	TAACCTAC---	
Chromadorita_tentabundum	C---CA---	-----	-----	TGAATTGGTG	A--ATCTGAA	TAACCTAC---	
Dichromadora_sp._BHMM_2005	C---CA---	-----	-----	TCTATTGGTG	A--ATCTGAG	CAACTTA---	
Neochromadora_BHMM_2005	C---CA---	-----	-----	TTAATTGGTG	A--ATCTGAA	TAACCTAC---	
Spilophorella_paradoxa	C---CA---	-----	-----	TTCTTTGGTG	A--ATCTGAA	TAACCTAC---	
Paracanthochus_caecus	T---CCG---	-----	-----	TAGTTTGGTG	A--ATCTGAA	TAACCTCA---	
Paracyatholaimus_intermedius	C---CCG---	-----	-----	TCGTTTGGTG	A--ATCTGAA	TAACCTCA---	
Praeacanthochus_punctatus	C---CCG---	-----	-----	TCGTTTGGTG	A--ATCTGAA	TAACCTCA---	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

[illegible]

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370      380      390      400      410      420

Panagrolaimus_subelongatus  --AATG----- TTTTGTGTTG A--AKCTGAA TATATCAT--
Panagrolaimus_davidi        G-ATG----- TTTTGTGTTG A--AACTGAA TATATCAT--
Panagrolaimus_cf_rigidus_AF40 G-ATG----- TTTTGTGTTG A--ACCTGAA TATATCT--
Panagrolaimus sp. Sourhope ED2 G-ATTG----- TTTAGTGTG A--ATCTGAA TATATCT--
Panagrolaimus sp. Sourhope ED2 G-ATTG----- TTTAGTGTG A--ATCTGAA TATATCT--
Panagrolaimus sp. Sourhope ED2 G-AATG----- TTTAGTGTG A--ACCTGAA TATATCT--
Panagrolaimus sp. Sourhope ED2 G-AATG----- TTTAGTGTG A--ACCTGAA TATATCT--
Steinernema_carpocapsae     -TCG----- TT TATGTTGGTG A--CTCTGAG TAACCTT--
Strongyloides_stercoralis   -----G CTCCTTGGTG A--TTCATAA TAACCTC--
Strongyloides_ratti        -----TTG A--CTCAAAA TATCCTC--
Diploscapter_sp_PS1897     -GACG----- TATTGTGGCG A--CTCCAAA TAGCCCA--
Diploscapter_sp_PS2017     -GACG----- TATTGTGGCG A--CTCCAAA TAGCCCA--
Heterorhabditis_bacteriophora -ACG----- TAGTTTGTG A--CTCTGAA TAACGCA--
Heterorhabditis_hepialus   -ACG----- TAGTTTGTG A--CTCTGAA TAACGCA--
Heterorhabditis_zelandica  -ACG----- TAGTTTGTG A--CTCTGAA TAACGCA--
Parasitorhabditis_sp_SB281 -CG----- TAAAGTGATG A--ATCTGAA TATCTT--
Rhabditoides_inermiformis  -TCG----- TTGTTTGTG A--CTCTGAA TAACCTA--
Rhabditoides_inermis_DF5001 -GAT----- ---ACTGGTG A--CTCTGAA TAACCCA--
Rhabditoides_regina_DF5012 ----- TTGAGTGATG A--CTCTGAT TATCTTG--
Poikilolaimus_oxycerca_SB200 -CCG----- TTTGTTGTTG A--CTCTGAA TAACCTA--
Poikilolaimus_regenfussi_SB199 -CCG----- TTTGTTGTTG A--CTCTGAA TAACCTA--
Distolabrellus_veechi_DWF1604 -C-G----- TG TAGTGATG A--CTCTGAA TATCTC--
Distolabrellus_veechi_DF5024 -C-G----- TATAGTGATG A--CTCTGAA TATCTC--
Choriorhabditis_dudichi    -GAG----- G ACGTTTGTG A--CTCTGAA TAACCTA--
Protorhabditis_sp         -ACG----- TATTGTGGCG A--CTCCAAA TAGCGCA--
Protorhabditis_sp_DF5055   -AGG----- TTTTGTGTTG A--ATCTGAG TAAAGTTC--
Cruzanema_tripartitum_DF5015 -ACG----- TAGTTTGTG A--CTCTGAG TAATGCA--
Mesorhabditis_sp_PS1179    -CG----- TCAAGTGATG A--TTCTGAA TATCTT--
Mesorhabditis_spiculigera_SB15 -CAACG---- -GTTGT TTCGGTGATG A--TTCTGAA TAACCTC--
Mesorhabditis_anisomorpha_SB12 -CG----- TTGAGTGATG A--TTCTGAA TGTCTT--
Teratorhabditis_palmarum_DF501 -CG----- TTTTGTGTTG A--CTCTGAA TATTTT--
Teratorhabditis_synpapillata_S -CG----- TTTTGTGTTG A--CTCTGAA TATTTT--
Caenorhabditis_briggsae_PB102 -ACG----- TTGTTTGTG A--CTCTGAA TAAAGCA--
Caenorhabditis_elegans_N2   -ACG----- TTGTTTGTG A--CTCTGAA TAAAGCA--
Caenorhabditis_sp_CB5161    -ACG----- TTGTTTGTG A--CTCTGAA TAAAGCA--
Caenorhabditis_sp_PS1010    -ACG----- TAGTTTGTG A--CTCTGAA TAAAGCA--
Caenorhabditis_japonica     -ACG----- TTGTTTGTG A--CTCTGAA TACCCTC--
Caenorhabditis_sp_DF5170    -ACG----- TAATTTGTG A--CTCTGAA TAAAGCA--
Caenorhabditis_sp_SB341     -ACG----- TTGACTGTTG A--CTCTGAA TAATGCA--
Caenorhabditis_plicata      -ACG----- TTTTGTGTTG A--CTCTGAA CAATCCC--
Caenorhabditis_vulgaris     -ACG----- TTGTTTGTG A--CTCTGAA TAAAGCA--
Caenorhabditis_sonorae      -ACG----- TTGACTGTTG A--CTCTGAA TAAAGCA--
Caenorhabditis_drosophilae  -ACG----- TAATTTGTG A--CTCTGAA TAAAGCA--
Phasmarhabditis_hermaphrodita -GCG----- TAGTTTGTG A--CTCTGAA TACCTAG--
Phasmarhabditis_neopapillosa -GCG----- TAGTTTGTG A--ATCTAAA TACCTAG--
Phasmarhabditis_neopapillosa -ACG----- TAGTTTGTG A--CTCTGAA TACCCTA--
Pellioditis_mediterranea_SB173 -ACG----- TAGTTTGTG A--CTCTGAA TACCCTA--
Pedlioditis_marina         -ACG----- TAGTTTGTG A--CTCTGAA TACCCTA--
Prodontorhabditis_wirthi    -CGG----- ACTGTG-TTG A--CACTGAA TAACGCTC--
Crustorhabditis_scanica     -GTW----- AAGTGATGAC T--CTGGATA TCCTGYA--
Dolichorhabditis_sp_CEW1    -ACG----- TAGTTTGCTG A--CTCTGAA TACC-CA--
Oscheius_sp_DF5000         -ACG----- TAGTTTGTG A--CTCTGAA TACCAG--
Oscheius_sp_BW282          -ACG----- TTGTTTGTG A--CTCTGAA TATCGCA--
Oscheius_myriophila_EM435   -ACG----- TTGTTTGTG A--CTCTGAA TATCGCA--
Oscheius_insectivora       -ACG----- TTGTTTGTG A--CTCTGAA TAACGTA--
Oscheius_dolichuroides     -ACG----- TAGTTTGTG A--CTCTGAA TACCCA--
Rhabditis_blumi_DF5010     -GCG----- TTGTTTGTG A--CTCTGAA TAACGTA--
Rhabditis_sp_PS1191        -ACG----- TAGTTTGTG A--CTCTGAA TAACCTC--
Rhabditis_sp_PS1010        -ACG----- TAGTTTGTG A--CTCTGAA TAAAGCA--
Rhabditis_myriophila_EM435  -ACG----- TTGTTTGTG A--CTCTGAA TATCGCA--
Rhabditis_colombiana        -ACG----- TAGTTTGTG A--CTCTGAA TAACGCT--
Rhabditella_axei_DF5006    -ACG----- AAGTTTGTG A--CTCTGAA TATCGTA--
Rhabditella_sp_DF5044      -ACG----- AAGTTTGTG A--CTCTGAA TATCGTA--
Cuticularia_sp_PS2083      -CCG----- TTTGTTGTTG A--CTCTGAA TAACGTA--
Necator_americanus         -ATG----- TAGTTTGCTG A--CTCTAAA TAACGCT--
Kalicephalus_cristatus     -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCT--
Ancylostoma_caninum        -GTG----- TAGTTTGCTG A--CTCTAAA TAACGCT--
Angiostrongylus_cantonensis -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCT--
Angiostrongylus_costaricensis -ATG----- CAGTTTGCTG A--CTCTGAA TAACGCA--
Angiostrongylus_malaysiensis -ATG----- CAGTTTGCTG A--CTCTGAA TAACGCA--
Angiostrongylus_dujardini   -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCA--
Angiostrongylus_vasorum     -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCA--
Aulurostrongylus_abstrusus  -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCA--
Didelphostrongylus_hayesi   -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCA--
Crenosoma_mephitidis       -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCA--
Crenosoma_vulpis           -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCA--
Otostrongylus_circumlitus   -ATG----- TAGTTTGCTG A--CTCTGAA TAACGCA--
NUCLEOTIDES INCLUDED      ----- -mmmmmmmm m--mmmmmm mmmmmmm--

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	370	380	390	400	410	420
Pratylenchus_thornei	--CCG-----				TCCATTGTTG	A--CTCAGAA	TAACTAA---
Tylenchulus_semipentrans	--CCG-----				-TTTTTGCTG	A--CTCAGAA	TAACTAA---
Ditylenchus_angustus	--CCG-----				TGTTTTGTTG	A--CTCAAA	TAACTCA---
Subanguina_radicola	--TCG-----				TTTTTTGTTG	A--CTCAGAA	TAACTAA---
Tylenchorhynchus_maximus	--CCG-----				TCCAATGTTG	A--CTCAGAA	TAACTAA---
Geocenamus_quadrifer	--CCG-----				TCCGCAGTTG	A--CTCAGAA	TAACTAA---
Globodera_pallida	--GCG-----				TCCAGTGTG	A--CTCAGAA	CAACTAA---
Scutellonema_bradys	--TCG-----				TTCAGTGTG	A--ACCAGAA	CAACTAA---
Helicotylenchus_dihystera	--GCG-----				TCCATTGCTG	A--ACCAGAA	CAACTCA---
Rotylenchus_robustus	--GCG-----				TTCAGTGTG	A--CTCAGAA	TAACTAA---
Meloidogyne_arenaria	--CTG-----				CTTCTTGTG	A--CTCAGAA	TAACTTA---
Meloidogyne_incognita_KT	--CTG-----				CTTCTTGTG	A--CTCAGAA	TAACTTA---
Meloidogyne_javanica	--CTG-----				CTTYTTGTTG	A--CTCAGAA	TAACTWA---
Meloidogyne_artiellia	--CTGTCAA-				ACGTTTGTTG	A--CTCAGAA	TAACTAA---
Meloidogyne_duytsi	--CTG-----				CTTCTTGTG	A--CTCAGAA	TAACTAA---
Meloidogyne_exigua	--CTG-----				CCTTTTGTG	A--CTCAGAA	TAACTAA---
Meloidogyne_hapla	--CTG-----				CTTCTTGTG	A--CTCAGAA	TAACTAA---
Meloidogyne_ichinohei	--CTG-----				TTCATTGTG	A--CTCAGAA	TAACTCA---
Meloidogyne_maritima	--CTG-----				CTTCTTGTG	A--CTCAGAA	TAACTAA---
Meloidogyne_microtyla	--CTG-----				CTTCTTGTG	A--CTCAGAA	TAACTAA---
Nacobbus_ aberrans	--CCG-----				TCCACTGTG	A--ACCAGAA	TAACTAA---
Pratylenchoides_ritteri	--CCG-----				TTCGCAGTTG	A--CTCAGAA	TAACTAA---
Pratylenchoides_magnicauda	--CCG-----				TTCGCAGTTG	A--CTCAGAA	TAACTAA---
Hirschmanniella_sp._JH_2003	--CCG-----				CCATGAGTTG	A--CTCAGAA	TAACTAA---
Radopholus_similis	--GGG-----				TTGAGTGTG	A--ATCAGAG	CAACTAA---
Pratylenchus_goodeyi_VF	--CCG-----				TCCATTGTG	A--CTCAGAA	TAACTAA---
Boleodorus_thylactus_clone2	A-CGG-TGCT CG-----				TGATTGTG	A--CTCAGAA	TAACTAA---
Philippine Sequence 1	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Philippine Sequence 2	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Philippine Sequence 3	--ACG-----				-CGGCTGTG	A--CTCTGAA	TAACGCT---
Philippine Sequence 4	--ATG-----				TTTTGTGTTG	A--AACTGAA	TATATCT---
Philippine Sequence 5	--GTG-----				TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Philippine Sequence 6	--ACG-----				TTGTTTGTG	A--CTCTGAA	TATCGTA---
Philippine Sequence 7	--ACG-----				TTGTTTGTG	A--CTCTGAA	TAAAGCA---
Tahiti Sequence 1	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Tahiti Sequence 2	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Thailand Sequence 1	--CCTG----				TTCATTGGTG	A--CTCTGAA	TAGCTTA---
Ogasawara Sequence 1	--ATG-----				TTTAGTGTG	A--AACTGAA	TATATCT---
Ogasawara Sequence 2	--GCG-----				TTGTTTGTG	A--CTCTGAA	TACCAAG---
Ivory Coast Sequence 1	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Ivory Coast Sequence 2	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Ivory Coast Sequence 3	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Ivory Coast Sequence 4	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Ivory Coast Sequence 5	--CCTG----				TTCATTGGTG	A--CTCTGAA	TAGCTTA---
Ivory Coast Sequence 6	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Ivory Coast Sequence 7	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Singapore Sequence 1	--ACG-----				-CGGCTGTG	A--CTCTGAA	TAACGCT---
Singapore Sequence 2	--CCTG----				TTCATTGGTG	A--CTCTGAA	TAGCTTA---
Singapore Sequence 3	--ACG-----				TTGTTTGTG	A--CTCTGAA	TAAAGCA---
Singapore Sequence 4	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Nigerian Sequence 1	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Nigerian Sequence 2	--GTG-----				TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Nigerian Sequence 3	--ATG-----				TAGTTTGCTG	A--CTCTGAA	TAACGCA---
NUCLEOTIDES INCLUDED	-----				-mmmmmmmm	m--mmmmmm	mmmmmm----

	430	440	450	460	470	480
Gordius_aquaticus	GCCGATCGCA --CGGTCTC-	-GT-ACCGGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA		
Priapulus_caudatus	GCTGATCGCA --CGGTCTC-	-GC-ACCGGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA		
Brachionus_plicatilis	GCTGATCGCA --TGGCCTA-	-GA-GCCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CCTA		
Chordodes_morgani	GCCGATCGCA --TGGTCTC-	-GT-ACCGGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA		
Practinolaimus_macrolaimus	GCCAATCGCA --CAGTCCTA	-GT-ACTGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Aporcelaimellus_obtusicaudatus	GCCAATCGCA --CAGTCCTA	-GT-ACTGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Wilsonema_schuermansstekhoveni	GCTGATCGCA --TGGTCTTT	-GT-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Mesodorylaimus_sp_cf_nigritul	GCCAATCGCA --CAGTCCTA	-GC-ACTGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Mesodorylaimus_bastianii	GCCAATCGCA --CAGTCCTA	-GT-ACTGGC	GATGTATCT-	TTC--AAATG	TCTGC-CTTA		
Mesodorylaimus_japonicus	GCCAATCGCA --CAGTCCTA	-GT-ACTGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Pungentus_sp._PDL_2005	GCCAATCGCA --CAGTCCTA	-GT-ACTGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Allodorylaimus_sp	GCCAATCGCA --CAGTCCTA	-GT-ACTGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Eudorylaimus_carteri	GCCAATCGCA --CAGTCCTA	-GT-ACTGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Microdorylaimus_sp	GCCAATCGCA --CAGTCCTA	-GT-ACTGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Longidorus_elongatus	GCTGATCGCA --CGGTC-TA	-GT-ACCGGC	GAMGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Xiphinema_rivesi	GCCGATCGCA --CGGTCCTA	-GT-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Tylencholaimus_sp	GCCAATCGCA --CGGTCCTA	-GT-ACCGGC	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA		
Anoplostoma_sp._BHMM_2005	G-TGATCGCA --CGGTCTTT	-GT-ACCGGC	GACATATCC-	TTC--AAATG	TCTGC-CTTA		
Adoncholaimus_fuscus	GCTGATCGCA --CGGTCTAT	-GC-ACCGGC	GACATGTCT-	TTC--AAACG	TCTGC-CTTA		
Enoplus_meridionalis	GTAGATCGCA --CGGTCTCT	-GA-ACCGGC	GACTCGTCA-	TTC--AAATG	TCTGC-CTTA		
NUCLEOTIDES INCLUDED	mmmmmmmm--	-----	-----	-----	-----	mmmmmm	mmmmmm-mmmmm

	430	440	450	460	470	480	
Enoplus_brevis_U88336	GCAGATCGCA	--CGGTCTCT	-GC-ACCGGC	GACATGTCA-	TTC--AAATG	TCTGC-CTTA	
Enoplus_communis	GCAGATCGCA	--CGGTCTCT	-GC-ACCGGC	GACATGTCA-	TTC--AAATG	TCTGC-CTTA	
Enoplodes_brunettii	GCTGATCGCA	--CAGTCTCT	-GT-ACCTGC	GATGTATCT-	TTC--AAATA	TCTGC-CTTA	
Syringolaimus_striatocaudatus	GCAGATCGCA	--TGGTCTTT	-GC-ACCGGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA	
Ironus_dentifurcatus	GCCGATCGAA	--CGGTCTTT	-GA-ACCGTC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA	
Calyptronema_maxweberi	GCTGAACGAA	--CGGTCTAA	-GA-ACCGAC	GTCATATCC-	TTC--AAACG	TCTGC-CTTA	
Viscosia_sp._BHMM_2005	GCTGATCGCA	--CGGTCTCA	-GA-ACCGGC	GACATATCC-	TTC--AAATG	TCTGC-CTTA	
Viscosia_viscosa	GCTGATCGCA	--CGGTCTGA	-GA-ACCGGC	GACATATCC-	TTC--AAATG	TCTGC-CTTA	
Pontonema_vulgare	NNNNAACGCA	--CGGTCCGA	-GT-ATTGGC	GACATATCC-	TTC--AAATG	TCTGC-CTTA	
Oncholaimus_sp._BHMM_2005	GCTGATCGCA	--CGGTCTAA	-GC-ACCGGC	GACATATCC-	TTC--AAATG	TCTGC-CTTA	
Alaimus_sp._PDL_2005	GCCGATCGCA	--GGTCTCTC	-GA-ACCGGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA	
Prismatolaimus_intermedius	GCAGATCGCA	--CGGTCTCT	-GT-ACCGGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA	
Tobrilus_gracilis	GCAGATCGCA	--CGGTCTCT	-GC-ACCGGC	GACGCATCT-	TTC--AAATG	TCTGC-CTTA	
Tripyla_cf._filicaudata_JH_2004	GCCGATCGCA	--TGGTCTCT	-GC-ACCGGC	GACGCACCT-	TTC--AAATG	TCTGT-CTCA	
Bathylaimus_sp.	GCAGATCGCA	--CGGTCTA-	-GC-ACCGGC	GACGAATCT-	TTC--AAATG	TCTGC-CTTA	
Bathylaimus_assimilis	GCAGATCGCA	--CGGTCTA-	-GC-ACCGGC	GACGAATCT-	TTC--AAATG	TCTGC-CTTA	
Bathylaimus_sp._BHMM_2005	GCAGATCGCA	--CGGTCTA-	-GT-ACCGGC	GACGCATCT-	TTC--AAATG	TCTGC-CTTA	
Tripylodes_sp._BHMM_2005	GCAGATCGCA	--TGGTCTA-	-GT-ACCGGC	GACGCATCT-	TTC--AAATG	TCTGC-CTTA	
Trischistoma_monohystera	GCCGATCGCA	--TGGTCTAA	-GT-ACCGGC	GACGCATCT-	TTC--AAATG	TCTGC-CTTA	
Mermis_nigrescens	GTTGATCGCA	--CAGTCTCT	-GC-ACCTGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA	
Mylonchulus_arenicolus	GTTGATCGCA	--CAGTCTTA	-GT-ACCTGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA	
Anatonchus_tridentatus	GTTGATCGCA	--CAGTCTTA	-GT-ACCTGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA	
Mononchus_truncatus	GTTGATCGCA	--CAGTCTT-	-GT-ACCTGC	GACGTATCT-	TTC--AAATG	TCTGC-CTTA	
Prionchulus_muscorum	GTCGATCGCA	--CAGTCTTA	-GT-ACCTGC	GACGTATTT-	TTC--AAATG	TCTGC-CTTA	
Clarkus_sp.	GTCGATCGCA	--CAGTCTTA	-GT-ACCTGC	GACGTATTT-	TTC--AAATG	TCTGC-CTTA	
Trichinella_spiralis	TCTGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATCACT-	TTC--AAGTG	TCTGC-CTTA	
Trichinella_papuae	TCTGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATCACT-	TTC--AAGTG	TCTGC-CTTA	
Trichinella_britovi	TCTGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATCACT-	TTC--AAGTG	TCTGC-CTTA	
Trichinella_murrelli	TCTGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATCACT-	TTC--AAGTG	TCTGC-CTTA	
Trichinella_pseudoispiralis	TCTGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATCACT-	TTC--AAGTG	TCTGC-CTTA	
Trichinella_nativa	TCTGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATCACT-	TTC--AAGTG	TCTGC-CTTA	
Trichinella_zimbabwensis	TCTGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATCACT-	TTC--AAGTG	TCTGC-CTTA	
Trichinella_nelsoni	TCTGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATCACT-	TTC--AAGTG	TCTGC-CTTA	
Trichuris_suis	GCTGATCGCA	--CGGTCTCA	-GC-ACCGGC	GACGAACT--	TTG--AAATG	ACTTG-CTCA	
Trichuris_trichiura	GCTGATCGCA	--CGGTCTCA	-GT-ACCGGC	GACGATGCT-	TTG--AAACG	ACTTG-CTCA	
Trichuris_trichiura	GCTGATCGCA	--CGGTCTCA	-GC-ACCGGC	GACGAATCT-	TTG--AAATG	ACTTG-CTCA	
Tylolaimophorus_minor	GCTGATCGCA	--CGGTCTCT	-GT-ACCGGC	GACAAATCT-	TTC--AAATG	TCTGT-TTTA	
Paratrichodorus_pachydermus	GCTGATCGCA	--CAGTCTCT	-GT-ACCTGC	GACAAATCT-	TTC--AAGTG	TCTGC-CTTA	
Paratrichodorus_anemones	GCTGATCGCA	--CAGTCTCT	-GT-ACCTGC	GACAAATCT-	TTC--AAGTG	TCTGA-CTTA	
Trichodorus_primitivus	GCTGATCGCA	--CAGTCTCT	-GT-ACCTGC	GACAAATCT-	TTC--AAGTG	TCTGC-CTTA	
Axonolaimus_helgolandicus	GTTGATCGCA	--TGGTCTCT	-GC-ACCGGC	GACGTATCT-	TCC--AAGTG	TCTGC-CTTA	
Ascolaimus_elongatus	GTTGATCGCA	--TGGTCTCT	-GC-ACCGGC	GACGTATCT-	TCC--AAGTG	TCTGC-CTTA	
Odontophora_rectangula	GCTGATCGCA	--TGGTCTCT	-GC-ACCGGC	GACGTATCT-	TCC--AAGTG	TCTGC-CTTA	
Cylindrolaimus_sp._202149	GCTGATCGCA	--TGGTCTCT	-GT-ACCGGC	GACGTATCT-	TCC--AAGTG	TCTGC-CTTA	
Tylocephalus_auriculatus	GCTGATCGCA	--TGGTCTCT	-GT-ACCGGC	GACGTATCA-	TTC--AAGTG	TCTGC-CTTA	
Plectus_acuminatus_BS9	GCTGATCGCA	--TGGTCTTT	-GT-ACCGGC	GACGCATCT-	TTC--AAGTG	TCTGC-CTTA	
Plectus_aquatilis	GCTGATCGCA	--TGGTCTTT	-GT-ACCGGC	GACGCATCT-	TTC--AAGTG	TCTGC-CTTA	
Anaplectus_sp.	GCTGATCGCA	--TGGTCTTT	-GT-ACCGGC	GACGCATCT-	TTC--AAGTG	TCTGC-CTTA	
Anisakis_sp._WKT	GCTGATCGCA	--TGGTCTT-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Anisakis_sp._Nadler	GCTGATCGCA	--TGGTCTT-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Contracaecum_multipapillatum	GCTGATCGCA	--TGGTCTT-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Pseudoterranova_decipiens	GCTGATCGCA	--TGGTCTT-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Raphidascaris_acus	GCTGATCGCA	--TGGTCTA-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Terranova_caballeri	GCTGATCGCA	--TGGTCTT-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Ascaris_suum	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Ascaris_lumbricoides	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Baylisascaris_procyonis	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Baylisascaris_transfuga	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACATGTCT-	ATC--AAGTG	TCTGC-CTTA	
Parascaris_equorum	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Porrocaecum_depressum	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Toxascaris_leonina	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Heterocheilus_tunicatus	GCTGATCGCA	--TGGTCTCT	-GC-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Goezia_pelagia	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Hysterothylacium_fortalezae	GCTGATCGCA	--TGGTCTCT	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Hysterothylacium_pelagicum	GCTGATCGCA	--TGGTCTA-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Hysterothylacium_reliquens	GCTGATCGCA	--TGGTCTA-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Iheringascaris_inquies	GCTGATCGCA	--TGGTCTA-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Toxocara_canis	GCTGATCGCA	--TGGTCTA-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Nemhelix_bakeri	GCTGATCGCA	--TGGTCTA-	-GT-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Raillietnema_sp._V3060	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Cruzia_americanana	GCTGATTGCA	--TGGCTCT-	-GC-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Heterakis_sp._14690	GCTGATCGCA	--CGGTCTT-	-GC-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Heterakis_gallinarum	GCTGATCGCA	--TGGTCTT-	-GA-ACCGGC	GACGTGTCT-	ATC--AAGTG	TCTGC-CTTA	
Paraspidodera_sp._21303	GCTGATCGCA	--TGGTCTCT	-GC-ACCGGC	GACGYGTCT-	ATC--AAGTG	TCTGC-CTTA	
Chromodora_nudicapitata	GCAGATCGCA	--CGGTCTA-	-GT-ACCGGC	GACATATCC-	TTC--ATGTG	TCTGC-CTTA	
Chromadora_sp._BHM_2005	GCAGATCGCA	--TGGGCTCT	-GT-CCCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA	
Atrochromadora_microlaima	GCAGATCGCA	--TGGGCTCT	-GT-CCCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA	
Chromadorina_germanica	GCTGATCGCA	--CGAGCTCT	-GT-CTCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA	
NUCLEOTIDES INCLUDED	mmmmmmmm--	-----	-----	-----	-----mmmmmm	mmmmmm--mmmm	

	430	440	450	460	470	480	
Chromadorita_tentabundum	GCTGATCGCA	--CGGTCTC-	-GT-ACCGGC	GACGTATCC-	TTC--AAGTG	TCTGC-CTTA	
Dichromadora_sp._BHMM_2005	GCAGATCGCA	--TGGGCTC-	-GT-CCCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA	
Neochromadora_BHMM_2005	GCTGATCGCA	--CGGTCTC-	-GT-ACCGGC	GACGTATCC-	TTC--AAGTG	TCTGC-CTTA	
Spilophorella_paradoxa	GCTGATCGCA	--TGGGCTC-	-GT-CCCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA	
Paracanthorchus_caecus	GTTGATCGCA	--CAGTCCTC	-GC-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Paracyatholaimus_intermedius	GTGGATCGCA	--CAGTCTC-	-GT-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Praeacanthorchus_punctatus	GTTGATCGCA	--CAGTCCTC	-GA-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Praeacanthorchus_sp	GTTGATCGCA	--CAGTCCTC	-GA-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Cyatholaimus_sp._BHMM_2005	GTTGATCGCA	--CAGTCCTC	-GC-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Spirinia_parasitifera	GCTGATCACA	--CGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Acanthopharynx_micans	GCTGATCACA	--CGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Xyzzors_sp	GCTGATCACA	--CGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Metachromadora_sp	GCTGATCACA	--CGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Metachromadora_remanei	GCTGATCACA	--CGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Catanema_sp	GCAGATCACA	--CGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Eubostriechus_dianae	GCTGATCACA	--TGGTCTC-	-GT-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Eubostriechus_topiarus	GCTGATCACA	--TGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Eubostriechus_parasitiferus	GCTGATCACA	--TGGTCTTA	-GT-ACCGAT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Laxus_oneistus	GCTGATCACA	--TGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Laxus_cosmopolitus	GCTGATCACA	--CGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Leptonemella_sp	GCTGATCACA	--CAGTCCTA	-GC-ACTGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Robbea_hypermnestra	GCTGATCGCA	--TGGTCTTA	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Stilbonema_majum	GCTGATCACA	--CGGTCTC-	-GC-ACTGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Monoposthia_costata	ACGGATCGCA	--CGGCCTTC	-GT-GCCGGC	GACGCATCA-	TTC--AAATT	TCTGC-CCTA	
Nudora_bipapillata	GCAGATCGCA	--TGGTCCA-	-GT-ACCGGC	GACACATTC-	CAG--GAGGG	AGTGC-CTTA	
Calomicrolaimus_parahonestus	GCAGATCGCA	--TGGTCTTA	-GC-ACCGGC	GACATATCT-	TTC--AAGTG	TCTGC-CTTA	
Calomicrolaimus_sp._BHMM_2005	GCAGATCGCA	--TGGTCTTA	-GC-ACCGGC	GACATATCT-	TTC--AAGTG	TCTGC-CTTA	
Molgolaimus_demani	GCAGATCGCA	--TGGTCTTA	-GT-ACCGGC	GACATATCT-	TTC--AAGTG	TCTGC-CTTA	
Diplogaster_lethieri	GCGGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGTACTG-	GTC--GAGCG	GGTGC-CCTA	
Aduncospiculum_halicti	GCTGATCGCA	--CGGTCTT-	-GCAACCGGC	GACGTATCG-	TTC--GAGTG	TCTGC-CTTA	
Pristionchus_lheritieri	GCGGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGTACTG-	GTC--GAGCG	GGTGC-CCTA	
Pristionchus_pacificus	GCGGATCGTA	--TGGTCTT-	-GT-ACCGAC	GACGTACTG-	GTC--GAGTG	GGTGC-CCTA	
Pristionchus_pacificus	GCGGATCGTA	--TGGTCTT-	-GT-ACCGAC	GACGTACTG-	GTC--GAGTG	GGTGC-CCTA	
Sabatieria_punctata_STRAIN_343	GCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Sabatieria_sp._355_BHMM_2005	GCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Sabatieria_celtica	TCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGCATCT-	TTC--AAGTG	TCTGC-CTTA	
Sabatieria_punctata_STRAIN_200	GCCGATCGCA	--CGGTCTC-	-GT-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Sabatieria_punctata_STRAIN_223	GCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGCATCT-	TTC--AAGTG	TCTGC-CTTA	
Sabatieria_sp._210_BHM_2005	GCCGATCGCA	--CGGTCTC-	-GT-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Setosabatieria_hilarula	TCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGCATCT-	TTC--AAGTG	TCTGC-CTTA	
Desmolaimus_zeelandicus	GCCGATCGCA	--CAGTCTT-	-GT-ACTGGC	GACGGACCA-	TTC--GAGTG	TCTGC-CTTA	
Terschellingia_longicaudata	GCTGATCGCA	--CGGTCTTT	-GC-ACCGGC	GACGCATCT-	TTC--AAGTG	TCTGC-CTTA	
Cyartoneura_elegans	GCCGATCGCA	--CGGTCTTT	-GC-ACCGGC	GACGCATCT-	TTC--AAGTG	TCTGC-CTTA	
Tridentulus_sp	GCAAAATCACA	--TGGTCTC-	-GT-ACCGGT	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA	
Diplolaimellodes_meyli	GCAGATCGCA	--TGGGCTC-	-GT-CCCGGC	GACAAATCT-	TTC--AAGTG	TCTGC-CTTA	
Diplolaimella_diavengatensis	GCAGATCGCA	--TGGGCTC-	-GT-CCCGGT	GCCAAATCT-	TTC--AAGTG	TCTGC-CTTA	
Geomonhystera_disjuncta	GCTGATCGCA	--CGGTCTA-	-GT-GCCGGC	GACATATCT-	TTC--AAGTG	TCTAC-CTTA	
Sphaerolaimus_hirsutus	GCGGATCGCA	--TGGTCTTA	-GT-ACCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA	
Theristus_acer	GCTGATCGCT	T-CGGTCTC-	-GT-ACCGGC	GACATGTCT-	TTC--AAGTG	TCTGC-CTTA	
Daptonema_procerus	GCAGATCGCT	--TGGTTTA-	-AT-ACCGGC	GACGTATCC-	TTC--AAGTG	TCTGC-CTTA	
Daptonema_hirsutum	GCAGATCGCT	T-CGGTTTC-	-AC-ACCGGC	GACATATCC-	TTC--AAGTG	TCTGC-TTTA	
Daptonema_normandicum	GCAGATCGCT	T-CGGTCTC-	-GT-ACCGGC	GATGTATCC-	TTC--AAGTG	TCTGC-CTTA	
Daptonema_oxycerca	GCAGATCGCT	T-CGGTTTA-	-AT-ACCGGC	GACGTATCC-	TTC--AAGTG	TCTGC-CTTA	
Daptonema_setosum	GCAGATCGCT	T-CGGTTTC-	-AC-ACCGGC	GACATATCC-	TTC--AAGTG	TCTGC-TTTA	
Desmodora_communis	GCTGATCACA	--CGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Desmodora_ovigera	GCTGATCACA	--CAGTCTTA	-GC-ACTGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA	
Metadesmolaimus_sp	GCAGATCGCT	T-CGGTTTC-	-AC-ACCGGC	GACATATCC-	TTC--AAGTG	TCTGC-TTTA	
Dentostomella_sp	GCTGATCCCA	--CGGTAA-	-GC-ACCGGC	GGCGTTTCT-	ATC--AAGTA	TCTGC-CTTA	
Bunonema_franzi	TCCGATCGCA	--TGGTCTC-	-GT-ACCGGC	GACGTATCAT	TTC--AAGAC	TCTGC-CCTA	
Bunonema_sp	GCCGATCGCA	--TGGGCTC-	-GG-GCCGGC	GACGCATCAT	ATC--AAGTC	TCTGC-CCTA	
Seleborca_complexa	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Acrobeloides_nanus	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Acrobeloides_bodenheimeri	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Acrobeloides_sp_PS1146	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Acrobeles_ciliatus	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--GAGTT	TCTGC-CGTA	
Acrobeles_sp_PS1156	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--GAGTT	TCTGC-CGTA	
Acrobeles_complexus_WCUG2	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--GAGTT	TCTGC-CGTA	
Cephaloboides_sp_SB227	GATAATCGAA	--GGTT--	-AA-TCCATC	GATTAAAGC-	ATT--AGGTG	TCTGC-CCTA	
Cephalobus_cubaensis	GCTGATCGCA	--CGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Cephalobus_sp_PS1143	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Cephalobus_sp_PS1196	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Cephalobus_oryzae_PS1165	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Cervidellus_alutus	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Pseudacrobelus_variabilis	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Triligulla_aluta	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Zeldia_punctata	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	-ACGTGTCT-	TTC--AAGTA	TCTGC-CGTA	
Myolaimus_sp_U81585	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACAGTTCT-	TTC--AAGTT	TCTGC-CTTA	
Rhabditophanes_sp._KR3021	GCTGAAC---	-----TA-	-CT-CACGTA	GTCATGCCG-	TAT--AAGTA	TCTGC-TTTA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	-----	-----	-----	-----	mmmmmmmmmm	

	430	440	450	460	470	480
Angiostrongylus_dujardini	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Angiostrongylus_vasorum	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Aulurostrongylus_abstrusus	GCATATCG-G	--CGGCTT--	-GA--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Didelphostrongylus_hayesi	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Crenosoma_mephitidis	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Crenosoma_vulpis	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Otostrongylus_circumlatus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Troglostrongylus_wilsoni	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Filaroides_martis	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Oslerus_osleri	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Parafilaroides_decorus	GCATATCG-A	--CGGCTT--	-GT--TCGTC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Metastrongylus_salini	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Metastrongylus_elongatus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Skrjabingylus_chitwoodrum	GCATATCG-G	--CGGCTT--	-GA--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Parelaphostrongylus_odocoeilei	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Muelerius_capillaris	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Protostrongylus_rufescens	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Halocercus_invaginatatus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AGA--AAGTG	TCTGC-CCTA
Pseudalius_inflexus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Stenurus_minor	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Torynurus_convolutus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Syngamus_trachea	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AGGTG	TCTGC-CCTA
Stephanurus_dentatus	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Nematodirus_battus	GCATATCG-G	--CGGCTT--	-GA--CCGCT	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Strongylus_equinus	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Labiostrongylus_bipapillosus	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Petrovinema_poculatum	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Cylicocyclus_insignis	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Chabartia_ovina	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Cyclodontostomum_purvisi	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Zoniolaimus_mawsonae	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Hypodontus_macropi	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Deletrocephalus_dimidiatus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_eckerti_P7B8	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_capreolus_P3B2	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_sp._P6A1	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_filaria	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_capreolus_P2C10	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_viviparus	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Haemonchus_sp._V3091	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Haemonchus_contortus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Haemonchus_placei	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Haemonchus_similis	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Ostertagia_ostertagi	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Ostertagia_leptospicularis	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Nippostrongylus_brasileinsis	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Heligmosomoides_polygyrus	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Trichostrongylus_colubriformis	GCATATCG-G	--CGGCTT--	-GT--YCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Tetrabothriostongylus_mackerr	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Herpestrostrongylus_pythonis	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Nicollina_cameroni	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Filarimena_flagrifer	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Amidostomum_cygni	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Teratocephalus_lirellus	GCTGATCGCA	--TGGTCTT--	-GC--ACCGGC	GACTCATCT-	TTC--AAGTG	TCTGC-CTTA
Brumptaemilius_justini	GCTGATCGCA	--TGGTCTT--	-GA--ACCGGC	GACGTGTCA-	ATC--AAGTG	TCTGC-CTTA
Anguillicola_crassus	GCTGATCGCA	--TGGTCTT--	-GA--ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CTTA
Dracunculus_medinensis	GCTGATCGCA	--TGGTCTA-	-GT--ACCGGC	GACATATCT-	ATC--AAGTA	TCTGC-CTTA
Dracunculus_oesophageus	GCCGATCGTA	--TGGTCTA-	-GT--ACCGAC	GACATATCT-	ATC--AAGTA	TCTGC-CTTA
Dracunculus_sp._V3104	GCTGATCGCA	--TGGTCTA-	-GT--ACCGGC	GACATATCT-	ATC--AAGTA	TCTGC-CTTA
Philonema_sp._A	GCTGATCGCA	--CGGTCTT--	-GC--ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CTTA
Philometra_obturator	GCTGATCGCA	--TGGTCTC-	-GT--ACCGGC	GACTTATCT-	ATC--AAGTA	TCTGC-CTTA
Camallanus_oxyccephalus	GCTGATCGCA	--TGGTCTC-	-GT--ACCGGC	GACGTATCT-	ATC--AAGTG	TCTGC-CTTA
Acanthocheilonema_viteae	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CTTA
Loa_loa	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACGTATCT-	ATC--AAGTG	TCTGC-CTTA
Onchocerca_cervicalis	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACATATCT-	ATCTCAAGTG	TCTGC-CTTA
Dirofilaria_immitis	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACGTATCT-	ATC--AAGTG	TCTGC-CTTA
Brugia_malayi	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACGTATCT-	ATC--AAGTG	TCTGC-CTTA
Wuchereria_bancrofti	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACGTATCT-	ATC--AAGTG	TCTGC-CTTA
Litomosoides_sigmodontis	GCTGATCGCA	--TGGTCTT--	-GC--ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CTTA
Setaria_digitata	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACGTATCT-	ATC--AAGTG	TCTGC-CTTA
Gnathostoma_turgidum	GCTGATCGCA	--TGGTCTC-	-GT--ACCGGC	GACATGTCA-	ATC--AAGTA	TCTGC-CTTA
Gnathostoma_neoprocyonis	GCTGATCGCA	--CGGTCTC-	-GC--ACCGGC	GACATGTCA-	ATC--AAGTA	TCTGC-CTTA
Gnathostoma_binucleatum	GCTGATCGCA	--CGGTCTC-	-GT--ACCGGC	GACATGTCA-	ATC--AAGTA	TCTGC-CTTA
Physaloptera_alata	GCTGATCGCA	--TGGTCTC-	-GT--ACCGGC	GACGTATCT-	ATC--AAGTG	TCTGC-CTTA
Physaloptera_turgida	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACGTATCT-	ATC--AAGTG	TCTGC-CTTA
Ascarophis_arctica	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACATGTCT-	ATC--AAGTG	TCTGC-CTTA
Spinitectus_carolini	GCTGATCGCA	--TGGTCTT--	-GC--ACCGGC	GACGTGTCT-	ATC--AAGTA	TCTGC-CTTA
Thelazia_lacrymalis	GCTGATCGCA	--TGGTCTT--	-GT--ACCGGC	GACGTATCT-	CAC--AAGTG	TCTGC-CTTA
NUCLEOTIDES INCLUDED	mmmmmmmm--	-----	-----	-----	-----mmmmmm	mmmmmm--mmmm

	490	500	510	520	530	540
Heterakis_sp_14690	TCA-AC	TTC	GATGG	TAGTT	TAAATGC	CTT
Heterakis_gallinarum	TCA-AC	TTC	GATGG	TAGTT	TAAATGC	CTT
Paraspidodera_sp_21303	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Chromodora_nudicapitata	TCA-AC	TGTC	GATGG	TAGTT	TATTGGA	CTT
Chromadora_sp._BHMM_2005	TCA-AC	TGTC	GATGG	TAGTT	TATTGGA	CTT
Atrochromadora_microlaima	TCA-AC	TGTC	GATGG	TAGTT	TATTGGA	CTT
Chromadorina_germanica	TCA-AC	TGTC	GATGG	TAGTT	TATTGGA	CTT
Chromadorita_tentabundum	TCA-AC	TTC	GATGG	TAGTC	TATAAGC	CTT
Dichromadora_sp._BHMM_2005	TCA-AC	TGTC	GATGG	TAGTT	TATTGGA	CTT
Neochromadora_BHMM_2005	TCA-AC	TTC	GATGG	TAGTT	TACATGA	CTT
Spilophorella_paradoxa	TCA-AC	TTC	GATGG	TAGTT	TACATGA	CTT
Paracanthonchus_caecus	TCA-AC	TTC	GATGG	TAGTT	TATATGA	CTT
Paracyatholaimus_intermedius	TCA-AC	TGTC	GATGG	TAGTT	TATATGA	CTT
Praeacanthonchus_punctatus	TCA-AC	TTC	GATGG	TAGTT	TACATGA	CTT
Praeacanthonchus_sp	TCA-AC	TTC	GATGG	TAGTT	TACATGA	CTT
Cyatholaimus_sp._BHMM_2005	TCA-AC	TTC	GATGG	TAGTT	TATATGA	CTT
Spirinia_parasitifera	TCA-AC	TTC	GATGG	TAGTT	TACGTGC	CTT
Acanthopharynx_micans	TCA-AC	TGTC	GATGG	TAGTT	TATGTGC	CTT
Xyzzors_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Metachromadora_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Metachromadora_remanei	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Catanema_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Eubostrichus_dianae	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Eubostrichus_topiarus	TCA-AC	TTC	GATGG	TAGTT	TATATGC	CTT
Eubostrichus_parasitiferus	TCA-AC	TTC	GATGG	TAGTT	TATATGC	CTT
Laxus_ohneistus	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Laxus_cosmopolitus	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Leptonemella_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Robbea_hypermnestra	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Stilbonema_majum	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Monoposthia_costata	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Nudora_bipapillata	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Calomicrolaimus_parahonestus	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Calomicrolaimus_sp._BHMM_2005	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Molgolaimus_demani	TCA-AC	TTC	GATGG	TAGTT	TATATGA	CTT
Diplogaster_lethieri	TCA-AC	TAT	GATGG	TAGTT	TATGTGC	CTT
Aduncospiculum_halicti	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Pristionchus_lheritieri	TCA-AC	TAT	GATGG	TAGTT	TATGTGC	CTT
Pristionchus_pacificus	TCA-AC	TAT	GATGG	TAGTT	TATGTGC	CTT
Pristionchus_pacificus	TCA-AC	TAT	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_punctata_STRAIN_343	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_sp._355_BHMM_2005	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_celtica	TCA-AC	TGTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_punctata_STRAIN_200	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_punctata_STRAIN_223	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_sp._210_BHM_2005	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Setosabatieria_hilarula	TCA-AC	TGTC	GATGG	TAGTT	TATGTGC	CTT
Desmolaimus_zeelandicus	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Terschellingia_longicaudata	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Cyarttonema_elegans	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Tridentulus_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Diplolaimelloides_meyli	TCA-AC	TTC	GATGG	TAGTT	TCTGTGC	CTT
Diplolaimella_diavengatensis	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Geomonhystera_disjuncta	TCA-AC	TTC	GATGG	TAGTT	TATATGC	CTT
Sphaerolaimus_hirsutus	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Theristus_acer	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Daptonema_procerus	TCA-AC	TTC	GATGG	TAGTT	TCTACGC	CTT
Daptonema_hirsutum	TCA-AC	TTC	GTTGG	TAGTT	TATACGC	CTT
Daptonema_normandicum	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Daptonema_oxycerca	TCA-AC	TTC	GATGG	TAGTT	TCTACGC	CTT
Daptonema_setosum	TCA-AC	TTC	GTTGG	TAGTT	TATACGC	CTT
Desmodora_communis	TCA-AC	TGTC	GATGG	TAGTT	TATGTGC	CTT
Desmodora_ovigera	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Metadesmolaimus_sp	TCA-AC	TTC	GTTGG	TAGTT	TATACGC	CTT
Dentostomella_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Bunonema_franzi	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Bunonema_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Seleborca_complexa	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
Acrobeloides_nanus	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
Acrobeloides_bodenheimeri	TCA-AC	TTC	GATGG	TAGTT	TTTCTGA	CTT
Acrobeloides_sp_PS1146	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
Acrobeles_ciliatus	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
Acrobeles_sp_PS1156	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
Acrobeles_complexus_WCUG2	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
Cephaloboides_sp_SB227	TCA-AC	---	GATGG	TAGTT	TATTAGA	CTT
Cephalobus_cubaensis	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
Cephalobus_sp_PS1143	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
Cephalobus_sp_PS1196	TCA-AC	TTC	GATGG	TAGTT	TATCTGA	CTT
NUCLEOTIDES INCLUDED	mmmm	mmmm	mmmm	mmmm	mmmm	mmmm

	490	500	510	520	530	540	
Cuticularia_sp_PS2083	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	CAACGGG-TA	ACGGAAAATA	
Necator_americanus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Kalicephalus_cristatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Ancylostoma_caninum	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Angiostrongylus_cantonensis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Angiostrongylus_costaricensis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Angiostrongylus_malaysiensis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Angiostrongylus_dujardini	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Angiostrongylus_vasorum	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Aulurostrongylus_abstrusus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Didelphostrongylus_hayesi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Crenosoma_mephitidis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Crenosoma_vulpis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Otostrongylus_circumlitus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Troglostrongylus_wilsoni	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Filaroides_martisi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Oslerus_osleri	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Parafilaroides_decorus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Metastrongylus_salini	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Metastrongylus_elongatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Skrjabinstrongylus_wilsoni	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Parelaphostrongylus_odocoilei	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Muelerius_capillaris	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Protostrongylus_rufescens	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Halocercus_invaginatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Pseudalius_inflexus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Stenurus_minor	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Torynurus_convolutus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Syngamus_trachea	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Stephanurus_dentatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Nematodirus_battus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Strongylus_equinus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Labiostrongylus_bipapillosus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Petrovinema_poculatum	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Cylicocycylus_insignis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Chabertia_ovina	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Cyclodontostomum_purvisi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Zoniolaimus_mawsonae	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Hypodontus_macropi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Deletrocephalus_dimidiatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Dictyocaulus_eckerti_P7B8	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Dictyocaulus_capreolus_P3B2	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Dictyocaulus_sp._P6A1	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Dictyocaulus_filaria	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Dictyocaulus_capreolus_P2C10	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Dictyocaulus_viviparus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Haemonchus_sp._V3091	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Haemonchus_contortus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Haemonchus_placei	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Haemonchus_similis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Ostertagia_ostertagi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Ostertagia_leptospicularis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Nippostrongylus_brasileinsis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Heligmosomoides_polygyrus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Trichostrongylus_colubriformis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Tetrabothriostongylus_mackerr	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Herpestrongylus_pythonis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Nicollina_cameroni	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Filarimena_flagrifer	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Amidostomum_cygni	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA	
Teratocephalus_lirellus	TCA-ACCTTC	GATGGTAGAT	TATGTGTCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Brumptaemilius_justini	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Anguillicola_crassus	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Dracunculus_medinensis	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Dracunculus_oesophageus	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Dracunculus_sp._V3104	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Philonema_sp_A	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Philometra_obturus	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Camallanus_oxyccephalus	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Acanthocheilonema_viteae	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Loa_loa	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Onchocerca_cervicalis	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Dirofilaria_immitis	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Brugia_malayi	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Wuchereria_bancrofti	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Litomosoides_sigmodontis	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Setaria_digitata	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
Gnathostoma_turgidum	TCA-ACCTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	490 500 510 520 530 540
Gnathostoma_neoprocyonis	TCA-AC TTTC GATGGTAGGT TATGTGCCT- ACCATGGTTG TAACGGG-TA ACGGAGAATA
Gnathostoma_binucleatum	TCA-AC TTTC GATGGTAGGT TATGTGCCT- ACCATGGTTG TAACGGG-TA ACGGAGAATA
Physaloptera_alata	TCA-AC TTTC GATGGTAGTT TATGTGCCT- ACCATGGTTG TAACGGG-TA ACGGAGAGTA
Physaloptera_turgida	TCA-AC TTTC GATGGTAGTT TATGTGCCT- ACCATGGTTG TAACGGG-TA ACGGAGAATA
Ascarophis_arctica	TCA-AC TTTC GATGGTAGTT TATGTGCCT- ACCATGGTTG TAACGGG-TA ACGGAGAATA
Spinitectus_carolini	TCA-AC TTTC GATGGTAGTT TATGTGCCT- ACCATGGTTG TAACGGG-TA ACGGAGAATA
Thelazia_lacrymalis	TCA-AC TTTC GATGGTAGTT TATGTGCCT- ACCATGGTTG TAACGGG-TA ACGGAGAATA
Aphelenchus_avenae	TCA-AC TTTC GATGGTAGTG TAGTGGACT- ACCATGGTTG TGACGGG-TA ACGGAGGATA
Aphelenchoides_fragariae	TCA-AC TTTC GATGGTAGTG TATTGGACT- GCCATGGTGT TGACGGG-TA ACGGAGAATC
Bursaphelelenchus_sp	TCA-AC TATC GTTGGTAGTT TATTGGACT- ACCATGGTGT TGACGGG-TA ACGGAGAATC
Deladenus_sp	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTGG TGACGGG-TA ACGGGGGATA
Criconema_sp	TCA-AC TTTC GATGGTAGTG TACGTGACT- ACCATGGTTG TGACGGG-TA ACGGAGGATA
Hemicycliophora_conida	TCA-AC TTTC GATGGTAGTG TACGTGACT- ACCATGGTTG TGACGGG-TA ACGGAGGATA
Paratylenchus_dianthus	TCA-AC TTTC GATGGTAGTG TACGTGACT- ACCATGGTGT TGACGGG-TA ACGGAGGATA
Pratylenchus_thornei	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTGG TGACGGG-TA ACGGAGGATA
Tylenchulus_semipetrans	TCA-AC TTTC GATGGTAGTG TACGTGACT- ACCATGGTGT TGACGGG-TA ACGGAGGATA
Ditylenchus_angustus	TCA-AC TTTC GACGGTAGTG TATTGGACT- ACCGTGGTGG TGACGGG-TA ACGGAGGATA
Subanguina_radicola	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTGG TGACGGG-TA ACGGAGGATA
Tylenchorhynchus_maximus	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTGA TGACGGG-TA ACGGAGGATA
Geocenamus_quadriifer	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTGG TGACGGG-TA ACGGAGGATA
Globodera_pallida	TCA-AC TTTC GATGGTAGTG TACCTGACT- ACCATGGTGA TGACGGG-TA ACGGAGGATA
Scutellonema_bradys	TCA-AC TTTC GATGGTAGTG TACCTGACT- ACCATGGTTG TGACGGG-TA ACGGAGGATC
Helicotylenchus_dihystera	TCA-AC TTTC GATGGTAGTG TACCTGACT- ACCATGGTTG TGACGGG-TA ACGGAGGATA
Rotylenchus_robustus	TCA-AC TTTC GATGGTAGTG TACCTGACT- ACCATGGTGA TGACGGG-TA ACGGAGGATA
Meloidogyne_arenaria	TCA-AC TT-- GACGGGAGCA TAATCGACT- CCCGTGGTGG TGACGGA-TA ACGGAGGATC
Meloidogyne_incognita_KT	TCA-AC TT-- GACGGGAGCA TAATCGACT- CCCGTGGTGG TGACGGA-TA ACGGAGGATC
Meloidogyne_javanica	TCA-AC MT-- GACGGGAGCA TAATCGACT- CCCGTGGTGG TGACGGA-TA ACGGAGGATC
Meloidogyne_artiellia	TCA-AC TT-- GACGGGAGTA TAATCGACT- CCCGTGGTGG TGACGGA-TA ACGGAGAATC
Meloidogyne_duytsi	TCA-AC TT-- GACGGGAGCA TAACCGACT- CCCGTGGTGG TGACGGA-TA ACGGAGGATC
Meloidogyne_exigua	TCA-AC TT-- GACGGGAGCA TAATCGACT- CCCGTGGTGG TGACGGA-TA ACGGAGGATC
Meloidogyne_hapla	TCA-AC TT-- GACGGGAGCA TAATCGACT- CCCGTGGTGG TGACGGA-TA ACGGAGGATC
Meloidogyne_ichinohei	TCATACTT-- GATGTTAGTA TCAGTGGCT- AACATGGTGG TGACGGA-TA ACGGAGGATC
Meloidogyne_maritima	TCA-AC TT-- GACGGGAGCA TAATCGACT- CCCGTGGTGG TGACGGA-TA ACGGAGGATC
Meloidogyne_microtyla	TCA-AC TT-- GACGGGAGCA TAATTGACT- CCCGWGGTGA TGACGGA-TA ACGGAGGATC
Nacobbus_ aberrans	TCA-AC TTTC GATGGTAGCG TATCTGCCT- ACCATGGTGA TGACGGG-TA ACGGAGGATA
Pratylenchoides_ritteri	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTGG TGACGGG-TA ACGGAGGATA
Pratylenchoides_magnicauda	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTGG TGACGGG-TA ACGGAGGATA
Hirschmanniella_sp._JH_2003	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTGG TGACGGG-TA ACGGAGGATA
Radopholus_similis	TCA-AC TTTC GACGGTAGTG TCTCTGGCT- ACCGTGGTGG TGACGGG-TA ACGGAGGATC
Pratylenchus_goodeyi_VF	TCA-AC TTTC GATGGTAGTG TCAGTGACT- ACCATGGTGG TGACGGT-TA ACGGAGGATC
Boleodorus_thylactus_clone2	TCA-AC TTTC GATGGTAGTG TATCTGACT- ACCATGGTTG TGACGGG-TA ACGGAGGATA
Philippine Sequence 1	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Philippine Sequence 2	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Philippine Sequence 3	TCA-ACC--A GATGGTAGCC TATTAGACT- ACCATGGTTA TAACGGG-TA ACGGAGAATC
Philippine Sequence 4	TCA-AC TTTC TATATAAGTA TATAGTACT- TATATGGTTT TGACGGA-TA ACGGAGTATT
Philippine Sequence 5	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Philippine Sequence 6	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TCACGGG-TA ACGGAGAATA
Philippine Sequence 7	TCA-AC T--A GATGGTAGTT TATTGGACT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Tahiti Sequence 1	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Tahiti Sequence 2	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Thailand Sequence 1	TCA-AC TTTC GATGGTAGTT TAAATGCCT- ACCATGGTTG TTACGGG-TA ACGGAGAATA
Ogasawara Sequence 1	TCA-AC TTTC TATATAAGTA TATAGTACT- TATATGGTTT TGACGGA-TA ACGGAGTATT
Ogasawara Sequence 2	TCA-AC T--A GATGTTAGTC TATTAGTCT- AACATGGTTG TAACGGG-TA ACGGAGAATC
Ivory Coast Sequence 1	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Ivory Coast Sequence 2	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Ivory Coast Sequence 3	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Ivory Coast Sequence 4	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Ivory Coast Sequence 5	TCA-AC TTTC GATGGTAGTT TAAATGCCT- ACCATGGTTG TTACGGG-TA ACGGAGAATA
Ivory Coast Sequence 6	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Ivory Coast Sequence 7	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Singapore Sequence 1	TCA-ACC--A GATGGTAGCC TATTAGACT- ACCATGGTTA TAACGGG-TA ACGGAGAATC
Singapore Sequence 2	TCA-AC TTTC GATGGTAGTT TAAATGCCT- ACCATGGTTG TTACGGG-TA ACGGAGAATA
Singapore Sequence 3	TCA-AC T--A GATGGTAGTT TATTGGACT- ACCATGGTTG TTACGGG-TA ACGGAGAATA
Singapore Sequence 4	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Nigerian Sequence 1	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TG ACGGAGAATA
Nigerian Sequence 2	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
Nigerian Sequence 3	TCA-ACC--T GATGGTAGTC TATTAGTCT- ACCATGGTTA TTACGGG-TA ACGGAGAATA
NUCLEOTIDES INCLUDED	mmmmmm--mm- mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmm--mm mmmmmmmmmmm

	550 560 570 580 590 600
Gordius_aquaticus	AGGGTT-CG- ATTCC--GGA GAGGGAGCCT GAGAAACGGC TACCACATCC AAGGAAGGCA
Priapulius_caudatus	AGGGTT-CG- ATTCC--GGA GAGGGAGCAT GAGAAACGGC TACCACATCC AAGGAAGGCA
Brachionus_plicatilis	AGGGTT-CG- ATTCC--GGA GAGGGAGCAT GAGAAACGGC TACCACATCT ACGGAAGGCA
Chordodes_morgani	AGGGTT-CG- ATTCC--GGA GAGGGAGCCT GAGAAACGGC TACCACATCC AAGGAAGGCA
Paractinolaimus_macrolaimus	AGGGTT-CG- ACTCC--GGA GAGGGAGCCT GAGAAACGGC TACCACATCC AAGGAAGGCA
Aporcelaimellus_obtusicaudatus	AGGGTT-CG- ACTCC--GGA GAGGGAGCCT GAGAAACGGC TACCACATCC AAGGAAGGCA
NUCLEOTIDES INCLUDED	mmmmmm--mm- mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	550	560	570	580	590	600
Wilsonema_schuermansstekhoveni	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Mesodorylaimus_sp_cf_nigritul	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Mesodorylaimus_bastianii	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Mesodorylaimus_japonicus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pungentus_sp._PDL_2005	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Allodorylaimus_sp	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Eudorylaimus_carteri	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Microdorylaimus_sp	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Longidorus_elongatus	AGGGTT-YG-	ACTCC--GGA	GAGGGAGCCT	GWGAAACGGC	TACCACATCC	AAGGAAGGCA
Xiphinema_rivesi	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tylencholaimus_sp	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Anoplostoma_sp._BHMM_2005	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Adoncholaimus_fuscus	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Enoplus_meridionalis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Enoplus_brevius_U88336	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Enoplus_communis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Enoploides_brunettii	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Syringolaimus_striatocaudatus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ironus_dentifurcatus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Calyptronema_maxweberi	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Viscosia_sp._BHMM_2005	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Viscosia_viscosa	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pontonema_vulgare	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Oncholaimus_sp._BHMM_2005	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Alaimus_sp._PDL_2005	AGGGTT-CG-	ATACC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Prismatolaimus_intermedius	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Tobrilus_gracilis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tripyla_cf_filicaudata_JH_2004	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAKCGGC	TACCACATCC	AAGGAAGGCA
Bathylaimus_sp	AGGGTT-TG-	ACTCT--GGA	GAGGACGCCT	GAGAGACGGC	TACCACATCC	AAGGAAGGCA
Bathylaimus_assimilis	AGGGTT-TG-	ACTCT--GGA	GAGGACGCCT	GAGAGACGGC	TACCACATCC	AAGGAAGGCA
Bathylaimus_sp._BHMM_2005	AGGGTT-TG-	ACTCT--GGA	GAGGACGCCT	GAGAGACGGC	TACCACATCC	AAGGAAGGCA
Tripylodes_sp._BHMM_2005	AGGGTT-TG-	ACTCT--GGA	GAGGACGCCT	GAGAGACGGC	TACCACATCC	AAGGAAGGCA
Trischistoma_monohystera	AGGGTT-CG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Mermis_nigrescens	AGG-TT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Mylonchulus_arenicolus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Anatonchus_tridentacus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Mononchus_truncatus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Prionchulus_muscorum	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Clarkus_sp	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Trichinella_spiralis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichinella_papuae	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichinella_britovi	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichinella_murrelli	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichinella_pseudospiralis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichinella_nativa	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichinella_zimbabwensis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichinella_nelsoni	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichuris_suis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichuris_trichiura	AGGGTT-CG-	GCTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichuris_muris	AGGGTTTCG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tylolaimophorus_minor	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Paratrichodorus_pachydermus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Paratrichodorus_anemones	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichodorus_primitivus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Axonolaimus_helgolandicus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Ascolaimus_elongatus	AGGGTT-CG-	ACTCC--GGA	RAGGGAGCCT	GAAAAATGGC	TACCACATCC	AAGGAAGGCA
Odontophora_rectangula	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Cylindrolaimus_sp._202149	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tylocephalus_auriculatus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Plectus_acuminatus_BS9	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Plectus_aquatilis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Anaplectus_sp	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Anisakis_sp_WKT	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Anisakis_sp_Nadler	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Contracaecum_multipapillatum	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pseudoterranova_decipiens	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Raphidascaris_acus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Terranova_caballeri	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ascaris_suum	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ascaris_lumbricoides	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Baylisascaris_procyonis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Baylisascaris_transfuga	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Parascaris_equorum	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA
Porrocaecum_depressum	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Toxascaris_leonina	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Heterocheilus_tunicatus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Goezia_pelagia	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Hysterothylacium_fortalezae	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
NUCLEOTIDES INCLUDED	mmmmmm-mm-	mmmmmm--mm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	550	560	570	580	590	600
Acrobeles_ciliatus	AGGGTT-CG-	ACTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Acrobeles_sp_PS1156	AGGGTT-CG-	ACTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Acrobeles_complexus_WCUG2	AGGGTT-CG-	ACTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Cephaloboides_sp_SB227	AGGGTT-CG-	TCTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Cephalobus_cubaensis	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Cephalobus_sp_PS1143	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Cephalobus_sp_PS1196	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Cephalobus_oryzae_PS1165	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Cervidellus_alutus	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Pseudacrobeles_variabilis	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Triligulla_aluta	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Zeldia_punctada	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
Myolaimus_sp_U81585	AGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditophanes_sp_KR3021	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Brevibuca_sp_SB261	AGGGTT-CG-	ACTCC--GGA	GAGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Halicephalobus_gingivalis	AGGGTT-CG-	TCTCC--GGA	GAGAAATGCCT	TAAAAACGGC	TTTACATCC	AAGGAAGGCA
Panagrobolus_stammeri	AGGGTT-CG-	ACTCC--GGA	GAAATCGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Plectonchus_sp_PDL0025	AGGGTT-CG-	ACTCC--GGA	GAAATCGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Turbatrix_aceti	AGGGTT-CG-	ACTCC--GGA	GAAAACGCCT	GAGAAACGGC	GGTTACATCC	AAGGAAGGCA
Panagrellus_redivivus	ATGGTT-CG-	ACTCC--GGA	GAAATAGCCT	GAGAAACGGC	TTTTACATCC	AAGGAAGGCA
Panagrellus_redivivus_PS1163	AGGGTT-CG-	ACTCC--GGA	GAAATAGCCT	GAGAAACGGC	TTTTACATCC	AAGGAAGGCA
Panagrolaimus_subelongatus	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Panagrolaimus_davidi	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Panagrolaimus_cf_rigidus_AF40	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Panagrolaimus_sp_Sourhope_ED2	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Panagrolaimus_sp_Sourhope_ED2	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Panagrolaimus_sp_Sourhope_ED2	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Panagrolaimus_sp_Sourhope_ED2	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Panagrolaimus_sp_Sourhope_ED2	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
Steinernema_carpcapsae	AGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Strongyloides_stercoralis	AGGGTT-CG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Strongyloides_ratti	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Diploscapter_sp_PS1897	GGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Diploscapter_sp_PS2017	GGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Heterorhabditis_bacteriophora	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Heterorhabditis_hepialus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Heterorhabditis_zelandica	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Parasitorhabditis_sp_SB281	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	TAGAAACGGC	AACCACATCT	AAGGAAGGCA
Rhabditoides_inermiformis	AGGGTT-CG-	ACTCC--GTA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditoides_inermis_DF5001	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditoides_regina_DF5012	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	TAGAAACGGC	AACTACATCT	AAGGAAGGCA
Poikilolaimus_oxycerca_SB200	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Poikilolaimus_regenfussi_SB199	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCT	AAGGAAGGCA
Distolabrellus_veechi_DWF1604	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	TAGAAACGGC	AACCACATCT	AAGGAAGGCA
Distolabrellus_veechi_DF5024	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	TAGAAACGGC	AACCACATCT	AAGGAAGGCA
Choriorhabditis_dudichi	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Protorhabditis_sp	GGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Protorhabditis_sp_DF5055	GGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Cruzema_tripartitum_DF5015	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Mesorhabditis_sp_PS1179	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	GAGAAACGGC	AACCACATCT	AAGGAAGGCA
Mesorhabditis_spiculigera_SB15	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	GAGAAACGGC	AACCACATCT	AAGGAAGGCA
Mesorhabditis_anisomorpha_SB12	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	GAGACACGGC	AACCACATCT	AAGGAAGGCA
Teratorhabditis_palmarum_DF501	AGGGTT-TG-	ACTCC--GGA	GA-GTATCCT	GAGAGACGGC	AAACACATCT	AAGGAAGGCA
Teratorhabditis_synapillata_S	AGGGTT-TG-	ACTCC--GGA	GAGTATGCCT	GAGAGACGGC	AAACACATCT	AAGGAAGGCA
Caenorhabditis_briggsae_PB102	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_elegans_N2	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_sp_CB5161	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_sp_PS1010	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_japonica	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_sp_DF5170	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_sp_SB341	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_plicata	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_vulgaris	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_sonorae	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Caenorhabditis_drosophilae	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Phasmarhabditis_hermaphrodita	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Phasmarhabditis_neopapillosa	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Phasmarhabditis_neopapillosa	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pellioiditis_mediterranea_SB173	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pellioiditis_marina	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Prodontorhabditis_wirthi	ATG-TT-CG-	TCTCC--GGA	GAGCCGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Crustorhabditis_scanica	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	KAGAAACGGC	AGCCACATCT	AAGGAAGGCA
Dolichorhabditis_sp_CEW1	AGGGTT-CG-	GCTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Oscheius_sp_DF5000	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Oscheius_sp_BW282	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Oscheius_myriophila_EM435	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Oscheius_insectivora	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Oscheius_dolichuroides	AGGGTT-CG-	GCTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
NUCLEOTIDES INCLUDED	mmmmmm-mm-	mmmmmm--mm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	550	560	570	580	590	600
Onchocerca_cervicalis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dirofilaria_immitis	AGGGTT-CG-	ACNCA--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Brugia_malay	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Wuchereria_bancrofti	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Litosomoides_sigmodontis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Setaria_digitata	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Gnathostoma_turgidum	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Gnathostoma_neoprocyonis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Gnathostoma_binucleatum	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Physaloptera_alata	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Physaloptera_turgida	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ascarophis_arctica	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Spinitectus_carolini	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Thelazia_lacrymalis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Aphelenchus_avenae	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Aphelenchoides_fragariae	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Bursaphelenchus_sp	AGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Deladenus_sp	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Criconema_sp	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Hemicycliophora_conida	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Paratylenchus_dianthus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pratylenchus_thornei	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tylenchulus_sempentrans	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ditylenchus_angustus	AGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Subanguina_radicola	AGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tylenchorhynchus_maximus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Geocenamus_quadriker	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Globodera_pallida	AGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Scutellonema_bradys	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Helicotylenchus_dihystera	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rotylenchus_robustus	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_arenaria	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_incognita_KT	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_javanica	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_artiellia	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_duytsi	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_exigua	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_hapla	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_ichinohei	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_maritima	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Meloidogynae_microtyla	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Nacobbus_aberrans	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pratylenchoides_ritteri	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pratylenchoides_magnicauda	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Hirschmanniella_sp._JH_2003	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Radopholus_similis	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pratylenchus_goodeyi_VF	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Boleodorus_thylactus_clone2	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philippine Sequence 1	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philippine Sequence 2	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philippine Sequence 3	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philippine Sequence 4	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philippine Sequence 5	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philippine Sequence 6	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philippine Sequence 7	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tahiti Sequence 1	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tahiti Sequence 2	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Thailand Sequence 1	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ogasawara Sequence 1	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ogasawara Sequence 2	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ivory Coast Sequence 1	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ivory Coast Sequence 2	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ivory Coast Sequence 3	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ivory Coast Sequence 4	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ivory Coast Sequence 5	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ivory Coast Sequence 6	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ivory Coast Sequence 7	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Singapore Sequence 1	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Singapore Sequence 2	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Singapore Sequence 3	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Singapore Sequence 4	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Nigerian Sequence 1	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Nigerian Sequence 2	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Nigerian Sequence 3	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
NUCLEOTIDES INCLUDED						

	610	620	630	640	650	660
Gordius_aquaticus	GCAG-GCGCG	CAAA-TTACC	CA---CTCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Priapulius_caudatus	GCAG-GCAGC	CAAA-TTACC	CA---TTCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Brachionus_plicatilis	GCAG-GCGCG	CAAA-TTACC	CAC---TCCT	AGA---ACG-	--GGGAGG--	-TAGTG-ACG
Chordodes_morgani	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	GGC---TCG-	--GGGAGG--	-TAGTG-ACG
Paractinolaimus_macrolaimus	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Aporcelaimellus_obtusicaudatus	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Wilsonema_schuurmansstekhoveni	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Mesodorylaimus_sp_cf_nigritul	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Mesodorylaimus_bastiani	GCAG-GCGCG	CAAA-TTACC	CAC---TTTC	AGA---ACG-	--AAGAGG--	-TAGTG-ACG
Mesodorylaimus_japonicus	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Pungentus_sp_PDL_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Allodorylaimus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Eudorylaimus_carteri	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Microdorylaimus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Longidorus_elongatus	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Xiphinema_rivesi	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Tylencholaimus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Anoplostoma_sp_BHMM_2005	GCAG-GCAGC	CAAA-TTACC	CAC---TCTT	GGT---ACG-	--AGGAGG--	-TAGTG-ACG
Adoncholaimus_fuscus	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Enoplus_meridionalis	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	AAC---ACG-	--GGGAGG--	-TAGTG-ACG
Enoplus_brevis_U88336	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	AAT---ACG-	--GGGAGG--	-TAGTG-ACG
Enoplus_communis	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	AAT---ACG-	--GGGAGG--	-TAGTG-ACG
Enoploides_brunettii	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGA---AAG-	--GGGAGG--	-TAGTG-ACG
Syringolaimus_striatocaudatus	GCAG-GCAGC	CAAA-TTACC	CAA---TCCC	CGA---ACG-	--GGGAGG--	-TAGTG-ACG
Ironus_dentifurcatus	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---AAG-	--GGGAGG--	-TAGTG-ACG
Calyptronema_maxweberi	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Viscosia_sp_BHMM_2005	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Viscosia_viscosa	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Pontonema_vulgare	GCAG-GCAGC	CAAA-TTACC	CAC---TCCG	GCA---CGG-	--GGAGG--	-TAGTG-ACG
Oncholaimus_sp_BHMM_2005	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Alaimus_sp_PDL_2005	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GTGAGG--	-TAGTG-ACG
Prismatolaimus_intermedius	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Tobrilus_gracilis	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Tripyla_cf_filicaudata_JH_2004	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Bathylaimus_sp	GCAG-GCAGC	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Bathylaimus_assimilis	GCAG-GCAGC	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Bathylaimus_sp_BHMM_2005	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---TCG-	--GGGAGG--	-TAGTG-ACG
Tripyloides_sp_BHMM_2005	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Trischistoma_monohystera	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Mermis_nigrescens	GCACTGCGCG	CAAAATTACC	CAA---TCCC	AGT---ACG-	--GGGAG--	-TAGTG-ACG
Mylonchulus_arenicolus	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Anatonchus_tridentatus	GCAG-GCGCG	CAAA-TTACC	CAA---TCCC	AGA---ACG-	--GGGAGG--	-TAGTG-ACG
Mononchus_truncatus	GCAG-GCGCG	CAAA-TTACC	CAA---TCCC	AGA---ATG-	--GGGAGG--	-TAGTG-ACG
Prionchulus_muscorum	GCAG-GCGCG	CAAA-TTACC	CAA---TCCC	AGA---ACG-	--GGGAGG--	-TAGTG-ACG
Clarkus_sp	GCAG-GCGCG	CAAA-TTACC	CAA---TCCC	AGA---ACG-	--GGGAGG--	-TAGTG-ACG
Trichinella_spiralis	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_papuae	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_britovi	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_murrelli	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_pseudospiralis	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_nativa	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_zimbabwensis	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_nelsoni	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichuris_suis	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	AGA---TCG-	--GGGAGG--	-TAGTG-ACG
Trichuris_trichiura	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	AGA---TCG-	--GGGAGG--	-TAGTG-ACG
Trichuris_muris	GCAG-GCAGC	CAAA-TTACC	CAC---TCCC	AGA---TCG-	--GGGAGG--	-TAGTG-ACG
Tyololaimophorus_minor	GCAG-GCGCG	TAAA-TTACC	CAT---TCCC	GGT---TCG-	--GGGAGG--	-TAGTG-ACG
Paratrachodorus_pachydermus	GCAG-GCGCG	CAAA-TTACC	CAC---T-TC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Paratrachodorus_anemones	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Trichodorus_primitivus	GCAG-GCGCG	CAAA-TTACC	CAC---TCGC	AGA---ACG-	--CGGAGG--	-TAGTG-ACG
Axonolaimus_helgolandicus	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	GGA---ACG-	--AGGAGG--	-TAGTG-ACG
Ascolaimus_elongatus	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	GGA---ACG-	--AGGAGG--	-TAGTG-ACG
Odontophora_rectangula	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	GGA---ACG-	--AGGAGG--	-TAGTG-ACG
Cylindrolaimus_sp_202149	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Tylocephalus_auriculatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Plectus_acuminatus_BS9	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Plectus_aquatilis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Anaplectus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Anisakis_sp_WKT	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Anisakis_sp_Nadler	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Contracaecum_multipapillatum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Pseudoterranova_decipiens	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Raphidascaris_acus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Terranova_caballeroi	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ascaris_suum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ascaris_lumbricoides	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Baylisascaris_procyonis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Baylisascaris_transfuga	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
NUCLEOTIDES INCLUDED	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm

	610	620	630	640	650	660	
Parascaris equorum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Porrocaecum depressum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Toxascaris leonina	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Heterocheilus tunicatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Goezia pelagia	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Hysterothylacium fortalezae	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Hysterothylacium pelagicum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Hysterothylacium reliquens	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Itheringascaris iniquies	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Toxocara canis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Nemhelix bakeri	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Raillietnema sp. V3060	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Cruzia americana	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Heterakis sp. 14690	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Heterakis gallinarum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Paraspidodera sp. 21303	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Chromodora nudicapitata	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---GCG-	--AGGAGG--	-TAGTG-ACG	
Chromadora sp. BHMM 2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Atrochromadora microlaima	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG	
Chromadorina germanica	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG	
Chromadorita tentabundum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG	
Dichromadora sp. BHMM 2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG	
Neochromadora BHMM 2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG	
Spilophorella paradoxa	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG	
Paracanthonchus caecus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Paracatholaimus intermedius	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Praeacanthonchus punctatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG	
Praeacanthonchus sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG	
Cyatholaimus sp. BHMM 2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Spirinia parasitifera	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG	
Acanthopharynx micans	GCTG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Xyzzors sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG	
Metachromadora sp	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	AGT---TTG-	--AGGAGG--	-TAGTG-ACG	
Metachromadora remanei	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	AGT---TTG-	--AGGAGG--	-TAGTG-ACG	
Catanema sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---CCG-	--AGGAGG--	-TAGTG-ACG	
Eubostrichus dianae	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Eubostrichus topiarus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Eubostrichus parasitiferus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Laxus oneistus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG	
Laxus cosmopolitus	GCAG-GCAGC	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG	
Leptonemella sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---TCG-	--AGGAGG--	-TAGTG-ACG	
Robbea hypermnestra	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG	
Stilbonema majum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Monoposthia costata	GCAG-GCGCG	CAAA-TTACA	CAA---TGAC	AGCAAATTG-	--TTGATG--	-TAGTG-ACA	
Nudora bipapillata	GCAG-GCGCG	CAAA-TTACA	CAA---TGAC	AGCAAAATG-	--TTGATG--	-TAGTG-ACA	
Calomicrolaimus parahonestus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Calomicrolaimus sp. BHMM 2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG	
Molgolaimus demani	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Diplogaster lethieri	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACT	
Aduncospiculum halicti	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---GCG-	--AGGAGG--	-TAGTA-ACT	
Pristionchus lheritieri	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACT	
Pristionchus pacificus	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACT	
Pristionchus pacificus	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACT	
Sabatieria punctata STRAIN 343	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Sabatieria sp. 355 BHMM 2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Sabatieria celtica	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG	
Sabatieria punctata STRAIN 200	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Sabatieria punctata STRAIN 223	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Sabatieria sp. 210 BHM 2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Setosabatieria hilarula	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG	
Desmolaimus zeelandicus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Terschellingia longicaudata	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Cyartonema elegans	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Tridentulus sp	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	GGT---ATG-	--AGGAGG--	-TAGTG-ACG	
Diplolaimelloides meyli	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-AAG	
Diplolaimella diavengatensis	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG	
Geomonhystera disjuncta	GCAG-GCGCG	CAAA-TTACA	CAA---TCTC	AGC---ACG-	--AGGATG--	-TAGTG-ACG	
Sphaerolaimus hirsutus	GCAG-GCGCG	TAAC-TTACC	CAC---TCTC	AGA---TTG-	--AGGAGG--	-TAGTG-ACG	
Theristus acer	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG	
Daptonema procerus	GCAG-GCAGC	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Daptonema hirsutum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Daptonema normandicum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Daptonema oxyerca	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Daptonema setosum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Desmodora communis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG	
Desmodora ovigera	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAC---TCG-	--AGGAGG--	-TAGTG-ACG	
Metadesmolaimus sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG	
Dentostomella sp	GCAG-GCGCG	CAAA-TTACC	CAC---TNTC	GGC---ATG-	--AGGAGT--	-TAGTG-ACG	
NUCLEOTIDES INCLUDED	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	

	610	620	630	640	650	660
Bunonema_franzi	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-AAT
Bunonema_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-AAT
Seleborca_complexa	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeloides_nanus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeloides_bodenheimeri	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeloides_sp_PS1146	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeles_ciliatus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeles_sp_PS1156	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeles_complexus_WCUG2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cephaloboides_sp_SB227	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TTA-----	--GTGAGA--	-TAGTG-ACT
Cephalobus_cubaensis	GCAG-GCGCG	NAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cephalobus_sp._PS1143	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cephalobus_sp._PS1196	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cephalobus_oryzae_PS1165	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cervidellus_alutus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Pseudacrobeles_variabilis	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Triligulla_aluta	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Zeldia_punctada	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Myolaimus_sp_U81585	GCAG-GCGCG	CAAA-TTATC	CAC---TCTC	GGC---ACG-	--AGGAGA--	-TAGTG-ACG
Rhabditophanes_sp._KR3021	GCAG-GCGCG	AAAA-TTACC	CAA---TTTT	AGT---TAA-	--AAGAGG--	-TAGTG-ACG
Brevibucca_sp._SB261	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Halicephalobus_gingivalis	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---GCG-	--AGGAGG--	-TAGTG-ACG
Panagrobelus_stammeri	GCAGG-GCGG	AAAA-TTACC	CAC---TCTT	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Plectonchus_sp._PDL0025	GCAG-GCGCG	AAAA-TTACC	CAC---TCTT	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Turbatrix_aceti	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	GGT---TCG-	--AGGAGG--	-TAGTG-ACG
Panagrellus_redivivus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTT	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrellus_redivivus_PS1163	GCAG-GCGCG	AAAA-TTACC	CAC---TCTT	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus_subelongatus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus_davidi	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus_cf_rigidus_AF40	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus_sp._Sourhope_ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus_sp._Sourhope_ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus_sp._Sourhope_ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus_sp._Sourhope_ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus_sp._Sourhope_ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Steinernema_carpocapsae	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Strongyloides_stercoralis	GCAG-GCGCG	AAAA-TTACC	CAA---TTTT	AGT---TAA-	--AAGAGG--	-TAGTG-ACG
Strongyloides_ratti	GCAG-GCGCG	AAAA-TTACC	CAA---TTTT	AGT---TAA-	--AAGAGG--	-TAGTG-ACG
Diploscapter_sp_PS1897	GCAG-GCGCG	AAAC-TTATC	CAC---TGTC	AAA---A---	--ATGAGA--	-TAGTG-ACT
Diploscapter_sp_PS2017	GCAG-GCGCG	AAAC-TTATC	CAC---TGTC	AAA---A---	--ATGAGA--	-TAGTG-ACT
Heterorhabditis_bacteriophora	GCAG-GCGCG	TAAC-TTATC	CAC---TC-C	TAA---TCG-	--GTGAGA--	-TAGTG-ACT
Heterorhabditis_hepialus	GCAG-GCGCG	TAAC-TTATC	CAC---TC-C	TAA---TCG-	--GTGAGA--	-TAGTG-ACT
Heterorhabditis_zelandica	GCAG-GCGCG	TAAC-TTATC	CAC---TC-C	TAA---TCG-	--GTGAGA--	-TAGTG-ACT
Parasitorhabditis_sp_SB281	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---KCG-	--AGGAGG--	-TAGTG-AAA
Rhabditoides_inermiformis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTA-ACA
Rhabditoides_inermis_DF5001	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---ACG-	--AGGAGG--	-TAGTG-ACT
Rhabditoides_regina_DF5012	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACA
Poikilolaimus_oxycerca_SB200	GCAG-GCGCG	CAAA-TTACC	CAC---TGGA	GGT---GCT-	--CCGAGG--	-TAGTG-ACT
Poikilolaimus_regenfussi_SB199	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACT
Distolabrellus_veechi_DWF1604	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Distolabrellus_veechi_DF5024	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Choriorhabditis_dudichi	GCAG-GCGTG	AAAC-TTATC	CAC---TGCA	TAT---AG-	---TGAGA--	-TAGTG-ACT
Protorhabditis_sp	GCAG-GCGCG	AAAC-TTATC	CAC---TGTA	TCA---AA-	---TGAGA--	-TAGTG-ACT
Protorhabditis_sp_DF5055	gcag-gcgcg	aaac-ttatc	cac---tgtt	gaa---aa-	---tgaga--	-tagtg-act
Cruznema_tripartitum_DF5015	GCAG-GCGCG	TAAC-TTATC	CAT---TATA	AAT---T---	--ATGAGA--	-TAGTG-ACT
Mesorhabditis_sp_PS1179	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Mesorhabditis_spiculigera_SB15	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---GCG-	--AGGAGG--	-TAGTG-ACA
Mesorhabditis_anisomorpha_SB12	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---GCG-	--AGGAGG--	-TAGTA-ACA
Teratorhabditis_palmarum_DF501	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Teratorhabditis_synpapillata_S	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Caenorhabditis_briggsae_PB102	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_elegans_N2	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_sp_CB5161	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_sp_PS1010	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAT---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_japonica	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_sp_DF5170	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GTG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_sp_SB341	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAC---A---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_plicata	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GGG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_vulgaris	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_sonorae	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAC---A---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis_drosophilae	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GTG---T---	--ATGAGA--	-TAGTG-ACT
Phasmarhabditis_hermaphrodita	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	CTT---T---	--GTGAGA--	-TAGTG-ACT
Phasmarhabditis_neopapillosa	ACAG-GCGCG	TAAC-TTATC	CAC---TACT	CTT---T---	--GTGAGA--	-TAGCA-ACT
Phasmarhabditis_neopapillosa	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Pellioditis_mediterranea_SB173	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	ACA-----	--GTGAGA--	-TAGTG-ACT
Pellioditis_marina	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Prodontorhabditis_wirthi	GCAG-GCGCG	AAAC-TTACT	CAC---TATN	ATA---G---	--ATGAAG--	-TAGTA-ACT
Crustorhabditis_scanica	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
NUCLEOTIDES INCLUDED	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm

	610	620	630	640	650	660
Dracunculus_sp._V3104	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---AAG-	--AGGAGG--	-TAGTG-ACG
Philonema_sp_A	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---GTG-	--AGGAGG--	-TAGTG-ACG
Philometra_obturans	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACAG-	--AGGAGG--	-TAGTG-ACG
Camallanus_oxycapahalus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acanthocheilonema_viteae	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Loa_loa	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Onchocerca_cervicalis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Dirofilaria_immitis	GCAG-GCGCG	CAAA-TTACC	CA---TCTC	AGA---ATG-	--AGGATG--	-TAGTG-ACG
Brugia_malayi	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Wuchereria_bancrofti	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Litomosoides_sigmodontis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Setaria_digitata	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGA---ATG-	--AGGAGG--	-TAGTG-ACG
Gnathostoma_turgidum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Gnathostoma_neoprocyonis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Gnathostoma_binucleatum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Physaloptera_alata	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Physaloptera_turgida	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ascarophis_arctica	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Spinitectus_carolini	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Thelazia_lacrymalis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Aphelenchus_avenae	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Aphelenchoides_fragariae	GCAG-GCGCG	CAAC-TTATC	CAC---TGTT	CAT---TGA-	--ACGAGA--	-TAGTG-ACG
Bursaphelenchus_sp	GCAG-GCGCG	AAAA-TTACC	CAA---TATC	AAA---ACG-	--ATGAGG--	-TAGTG-ACG
Deladenus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGA---ACG-	--AGGAGG--	-TAGTG-AAG
Criconema_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGT---TCG-	--AGGAGG--	-TAGTG-ACG
Hemicycliophora_conida	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGT---TCG-	--AGGAGG--	-TAGTG-ACG
Paratylenchus_dianthus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Pratylenchus_thornei	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---GCG-	--AGGAGG--	-TAGTG-ACG
Tylenchulus_semipetrans	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Ditylenchus_angustus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Subanguina_radicola	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Tylenchorhynchus_maximus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Geocenamus_quadriker	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Globodera_pallida	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAC---ATG-	--AGGAGG--	-TAGTG-ACG
Scutellonema_bradys	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAC---ACGC	GGAGGAGG--	-TAGTG-ACG
Helicotylenchus_dihystera	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAC---GCG-	--AGGAGG--	-TAGTG-ACG
Rotylenchus_robustus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_arenaria	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_incognita_KT	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_javanica	GCAG-GCGCG	CAAA-TTACC	MAC---TTTC	GGC---TCC-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_artiellia	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_duytsi	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_exigua	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_hapla	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_ichinohei	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_maritima	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_microtyla	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Nacobbus_aberrans	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Pratylenchoides_ritteri	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Pratylenchoides_magnicauda	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Hirschmanniella_sp._JH_2003	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--CGGAGGAG	GTAGTG-ACG
Radopholus_similis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ATG-	--AGGAGG--	-TAGTG-ACG
Pratylenchus_goodeyi_VF	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACG
Boleodorus_thylactus_clone2	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACAC	TGAGGAGG--	-TAGTG-ACG
Philippine Sequence 1	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Philippine Sequence 2	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Philippine Sequence 3	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TTT-----	--GTGAGA--	-TAGAG-ACG
Philippine Sequence 4	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Philippine Sequence 5	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Philippine Sequence 6	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTA-ACT
Philippine Sequence 7	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Tahiti Sequence 1	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Tahiti Sequence 2	GCAG-GCGCG	AAAC-TTATC	TAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Thailand Sequence 1	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ogasawara Sequence 1	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Ogasawara Sequence 2	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	CCT---T---	--GTGAGA--	-TAGGG-ACA
Ivory Coast Sequence 1	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 2	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 3	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 4	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 5	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ivory Coast Sequence 6	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 7	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Singapore Sequence 1	GCAG-GCGCG	TAAC-TTATC	CAC---TACA	TTT-----	--GTGAGA--	-TAGAG-ACG
Singapore Sequence 2	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ATG-	--AGGAGG--	-TAGTG-ACG
Singapore Sequence 3	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Singapore Sequence 4	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Nigerian Sequence 1	GCAG-GCGCG	AAAC-TTATC	CAA---TATT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
NUCLEOTIDES INCLUDED	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm

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610 620 630 640 650 660
GCAG-GCGCG TAAC-TTATC CAC---TCTT GAA---GAG- --ATGAGA-- -TAGTG-ACT
GCAG-GCGCG AAAC-TTATC CAA---TCTT GAA---TAG- --ATGAGA-- -TAGTG-ACT
mmmm-mmmmm mmmm-mmmmm mmm---mmmm mm----- -mmmmmm-----mmmmm-mm

[illegible]

	670	680	690	700	710	720
Pseudoterranova_decipiens	AAAAATAACG	AG-ACCGTTC	TCTCT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Raphidascaris_acus	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Terranova_caballeroi	AAAAATAACG	AG-ACCGTTC	TCTCT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Ascaris_suum	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Ascaris_lumbricoides	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Baylisascaris_procyonis	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Baylisascaris_transfuga	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Parascaris_equorum	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Porrocaecum_depressum	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Toxascaris_leonina	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Heterocheilus_tunicatus	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Goezia_pelagia	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Hysterothylacium_fortalezae	AAAAATAACG	AG-ACCGTTC	TCTTT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Hysterothylacium_pelagicum	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Hysterothylacium_reliquens	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Itheringascaris_inquies	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Toxocara_canis	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Nemhelix_bakeri	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Raillietnema_sp._V3060	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Cruzia_americana	AAAAATAACG	AG-GCGTTC	TCTAT---GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Heterakis_sp._14690	AAAAATAACA	AG-ACCGTTC	TCTTT---AGA	GGCCG-GTTA	TT-GGAATGA	GTA-CAGTTT
Heterakis_gallinarum	AAAAATAACA	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TT-GGAATGA	GTA-CAATTT
Paraspidodera_sp._21303	AAAAATAACG	AG-GCGTTC	TCTAT---GA	GGCCG-GCCA	TC-GGAATGG	GTA-CAATTT
Chromodora_nudicapitata	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACTT
Chromadora_sp._BHMM_2005	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATTT
Atrochromadora_microlaima	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATTT
Chromadorina_germanica	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATTT
Chromadorita_tentabundum	AAAAATAACG	AG-ACAGTTC	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATCT
Dichromadora_sp._BHMM_2005	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATTT
Neochromadora_BHMM_2005	AAAAATAACG	AG-ACAGTTC	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATCT
Spilophorella_paradoxa	AAAAATAACG	AG-ACAGTTC	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATCT
Paracanthonchus_caecus	AAAAATAACA	AG-ATGGTTC	TCTAT---GA	GGTCC-ATCA	TT-GGAATGG	GTA-CAATTT
Paracantholaimus_intermedius	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGTCC-GTCA	TC-GGAATGG	GTA-CAATTT
Praeacanthonchus_punctatus	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGTCC-GTCA	TC-GGAATGG	GTA-CAATTT
Praeacanthonchus_sp	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGTCC-GTCA	TC-GGAATGG	GTA-CAATTT
Cyatholaimus_sp._BHMM_2005	AAAAATAACA	AG-ATGGTTC	TCTAT---GA	GGTCC-GTCA	TT-GGAATGG	GTA-CAATTT
Spirinia_parasitifera	AAAAATAACG	AG-ATGGTTC	TCTAT---GA	GGCCC-ATCA	TC-GGAATGG	GTA-CAATTT
Acanthopharynx_micans	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATCT
Xyzzors_sp	AAAAATAACG	AG-ATGGTTC	TCTAT---GA	GGCCC-ATCA	TC-GGAATGG	GTA-CAATTT
Metachromadora_sp	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGCCC-GTCA	TC-GGAATGA	GTA-CAATTT
Metachromadora_remanei	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGCCC-GTCA	TC-GGAATGA	GTA-CAATTT
Catanema_sp	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
Eubostrichus_dianae	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
Eubostrichus_topiarus	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
Eubostrichus_parasitiferus	AAAAATAACA	AG-ATGGTTC	TCTTT---GA	GGCCC-GTCA	TT-GGAATGG	GTA-CAATTT
Laxus_oneistus	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
Laxus_cosmopolitus	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
Leptonemella_sp	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TT-GGAATGG	GTA-CAATTT
Robbea_hypermnestra	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
Stilbonema_majum	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGTCC-GTCA	TC-GGAATGA	GTA-CAATTT
Monoposthia_costata	AGAGGTAACG	AG-GCGGTTC	TCTGT---	GGTCC-GTCA	TC-GTATCGA	GTA-CAACTC
Nudora_bipapillata	AGAAATAACG	AG-ACGGTTC	TCTAT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAACTT
Calomicrolaimus_parahonestus	AAAAATAACG	AG-ACAGTTC	TCTTT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACCC
Calomicrolaimus_sp._BHMM_2005	AAAAATAACG	AG-ACAGTTC	TCTTT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACCC
Molgolaimus_demani	AAAAATAACG	AG-ACAGTTC	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACCC
Diplogaster_lethieri	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGTCT-GTCA	TT-GAAATGA	GCA-CAACTT
Aduncospiculum_halicti	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGCCT-GTCA	TC-GTAATGG	GTG-TATATT
Pristionchus_lheritieri	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGTCT-GTCA	TT-GAAATGA	GCA-CAACTT
Pristionchus_pacificus	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGTCT-GTCA	TT-GCAATGA	GCA-CAACTT
Pristionchus_pacificus	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGTCT-GTCA	TT-GCAATGA	GCA-CAACTT
Sabatieria_punctata_STRAIN_343	AAAAATAACA	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TT-GGAATGA	GAA-CAATTC
Sabatieria_sp._355_BHMM_2005	AAAAATAACA	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TT-GGAATGA	GAA-CAATTC
Sabatieria_celtica	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
Sabatieria_punctata_STRAIN_200	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
Sabatieria_punctata_STRAIN_223	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
Sabatieria_sp._210_BHM_2005	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
Setosabatieria_hilarula	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
Desmolaimus_zeelandicus	AGAAATAACG	AG-GCGTCTC	TCTTT---GA	GGGTC-GCCA	TC-GGAATGA	GGA-CAATGC
Terschellingia_longicaudata	AAAAATAACG	AG-GCGGTTC	TCTAT---GA	GGCCC-GCTA	TC-GGAATGA	GAA-CAATAT
Cyartonema_elegans	AAAAATAACG	AG-GCGGTTC	TCTTT---GA	GGCCC-GCTA	TC-GGAATGA	GAA-CAACAC
Tridentulus_sp	AAAAATACCA	AG-CCGAGGC	TCATT---GA	GTTTC-GGCA	TT-GGAATGA	GAA-CAATCT
Diplolaimelloides_meyli	AAAAATACCA	AG-AGGAGGC	TCATT---GA	GTTTC-GTCA	TT-GGAATGA	GTA-CAATCT
Diplolaimella_diavengatensis	AAAAATACCA	AG-GTGAGGC	CCTAT---GG	GTTTC-GCCA	TT-GGAATGA	GAA-CAATAT
Geomonhystera_disjuncta	AAACATACCG	AG-GCGATGC	CCTTTC---GG	GTGTC-GTCA	TC-GGAATGA	GTC-GAATCT
Sphaerolaimus_hirsutus	AAAAATATCG	AG-CTGGTAG	TCTTTC---GA	TTATC-AGCA	TC-GGAATGA	GAA-CAATAT
Theristus_acer	AAAAATAACG	AG-ATGGGAC	TCTATC---GA	GTACC-ATCA	TC-GGAATGA	GAA-CAATCT
Daptonema_procerus	AAAAATAACG	AG-ATAACCC	GCTAT---GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATTT
Daptonema_hirsutum	AAAAATAACG	AG-ATAACCC	GCTCTA---GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATCT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mm-mmmmmmm	mmmm-----m	mmmmmm-mmmmm	mm-mmmmmmm	mmmm-mmmmmmm

	670	680	690	700	710	720
Daptonema_normandicum	AAAAATAACG	AG-ACGATTC	TCTTTT--GA	GTATC-GTCA	TC-GGAATGA	GAA-CAATCC
Daptonema_oxycerca	AAAAATAACG	AG-ATAACCC	GCTATC--GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATTT
Daptonema_setosum	AAAAATAACG	AG-ATAACCC	GCTCTA--GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATCT
Desmodora_communis	AAAAATAACG	AG-ACGGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATCT
Desmodora_ovigera	AAAAATAACG	AG-ACGGTTC	TCTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAACTT
Metadesmolaimus_sp	AAAAATAACG	AG-ATAACCC	GCTCTA--GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATCT
Dentostomella_sp	AAAAATAACG	AG-ACCGTTC	TCAAT--GA	GGCCG-GTTA	TC-GGAATGA	ATA-CAATTT
Bunonema_franzi	AGAAATAACA	AA-GTAGATG	TCTTT--GA	TGTCT-ACTA	TT-GGAATGG	GTA-CAATTT
Bunonema_sp	AGAAATAACA	AA-GCAGATG	TCTCT--GA	TGTCT-GCTA	TT-GGAATGG	GTA-CAATTC
Seleborca_complexa	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeloides_nanus	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeloides_bodenheimeri	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeloides_sp_PS1146	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeles_ciliatus	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeles_sp_PS1156	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeles_complexus_WCUG2	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Cephaloboides_sp_SB227	AAATATAAAG	AG-ACCAATC	CTCT----G	GATTG-GTTA	TT-TCAATGA	GAT-GAGCTT
Cephalobus_cubaensis	AAAAATAACG	AG-ACCATTC	TCTCAT--GA	GGCTG-GTCA	TC-GGAATGG	GTA-CAATTT
Cephalobus_sp_PS1143	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Cephalobus_sp_PS1196	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Cephalobus_oryzae_PS1165	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Cervidellus_alutus	AAAAATAACG	AG-ACCGTTC	TCTT-T--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Pseudacrobeles_variabilis	AAAAATAACG	AG-ACCGTTC	TCTTCT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Triligulla_aluta	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Zeldia_punctada	AAAAATAAC-	AG-ACCGTTC	TCTTCT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Myolaimus_sp_U81585	AAAAATAACG	AG-GATGTCT	TCTTT--GA	GGCCA-TCTA	TT-GTAATGA	GTA-CAATTT
Rhabditophanes_sp_KR3021	AAAAATGACA	AG-GCTACAA	ATTTT--GT	TTGTA-GTCA	TT-GGAAATC	TTT-GAGTCT
Brevibucca_sp_SB261	AAAAATAACA	AG-GGCGTAC	TCTTT--GA	GGCCG-CCTA	TT-GGAATGG	GTA-CAATTT
Halicephalobus_gingivalis	TGAAATGACN	AGATANGTIN	TGTAT--AG	AATNT-ANCA	TT-GGAATGG	TTT-AAATAC
Panagrolaimus_stammeri	AGAAATGACA	AGGCTTACTC	CTTAT--GG	GGTGA-GTCA	TT-GGAATGG	GTA-CAATTT
Plectonchus_sp_PDL0025	TGAAATGACA	AGGCTTATTC	CTTAT--GG	AATGA-GTCA	TT-GGAATGG	GTC-CAATTT
Turbatrix_aceti	TGAAATGGCA	GAACTCTTTT	CTTTA--TGG	AACGG-ATTA	TT-GCAATGA	TGT-GAGTTT
Panagrellus_redivivus	AGAAAGTGACA	AGATCTGTCTM	CTTCG--GG	GGCGG-GTCA	TT-GGAATGG	TTT-GAATTT
Panagrellus_redivivus_PS1163	AGAAAGTGACA	AGATCTGTCC	CTTCG--GG	GGCGG-GTCA	TT-GGAATGG	TTT-GAATTT
Panagrolaimus_subelongatus	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_davidi	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_cf_rigidus_AF40	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_sp_Sourhope_ED2	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_sp_Sourhope_ED2	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_sp_Sourhope_ED2	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_sp_Sourhope_ED2	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Steinernema_carpocapsae	AAAAATAACA	AG-ACTGGTC	GCTTT--GC	GAACA-GTTA	TT-GGAATGG	GTA-CAATTT
Strongyloides_stercoralis	AAAAATGACA	ACCAAAATATT	ATTAT--TA	ATATT-TGGA	TT-GAAATC	TTT-AAGTTT
Strongyloides_ratti	AAAAATGACA	ACCAAAATATT	ATTTT--TA	GTATT-TGGA	TT-GAAATC	TTT-AAGTTT
Diploscapter_sp_PS1897	AAAAATATAA	AA-TCCCTCC	CTCAT--G	GGAGG-GATA	TT-TAAATGA	GTA-GAGGTT
Diploscapter_sp_PS2017	AAAAATATAA	AA-TCCCTCC	CTCAT--G	GGAGG-GATA	TT-TAAATGA	GTA-GAGGTT
Heterorhabditis_bacteriophora	AAAAATAAAA	AG-ACCATTC	CT-AT--G	GAACG-GTTA	TT-TCAATGA	GTA-GATCTT
Heterorhabditis_hepialus	AAAAATAAAA	AG-ACCATTC	CT-AT--G	GAACG-GTTA	TT-TCAATGA	GTA-GATCTT
Heterorhabditis_zelandica	AAAAATAAAA	AG-ACCATTC	CTAT--G	GAACG-GTTA	TT-TCAATGA	GTA-GATCTT
Parasitorhabditis_sp_SB281	ACAACTAACG	AG-ACTGCTC	CCTAC--GG	GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Rhabditoides_inermiformis	ACAACTAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Rhabditoides_inermis_DF5001	AACAATAACG	AG-ACAGCTC	TCTTT--GA	GGGTT-GTTA	TC-GGAATGG	GTT-CAATCT
Rhabditoides_regina_DF5012	ACCAATAACG	AG-ACTCAGA	CCTTT--GG	TTTGA-GTCA	TC-GGAATGG	GTC-CAAGT
Poikilolaimus_oxycerca_SB200	AACAATAACG	AG-ACCGTCT	TCTAT--GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Poikilolaimus_regenfussi_SB199	AACAATAACG	AG-ACCGTCT	TCTTT--GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Distolabrellus_veechi_DWF1604	ACAACTAACG	AG-ACTGCGT	CCTAT--GG	GCGCA-GTCA	TC-GAAATGG	GTA-CAATTT
Distolabrellus_veechi_DF5024	ACAACTAACG	AG-ACTGCGT	CCTAT--GG	GCGCA-GTCA	TC-GAAATGG	GTA-CAATTT
Choriorhabditis_dudichi	AAACATAAAA	AG-ACCATTC	CGTTC--G	GAGCG-GTTT	TT-TCAATGA	ATA-GAGTTT
Protorhabditis_sp	AAACATATAA	AA-TCCCTCC	CAAAT--G	GGAGG-GATA	TT-TAAATGA	GTA-GAGGTT
Protorhabditis_sp_DF5055	AAGAATAAAA	AA-CCCTACC	CTTAG--G	GGTGG-GGTA	TT-TGAATGA	GTC-AAGTTC
Cruzanema_tripartitum_DF5015	ANACATAAAA	AG-ACCATTC	CTAAC--G	GAACG-GTTA	TT-TCAATGA	TAC-GAGTGT
Mesorhabditis_sp_PS1179	ACAACTAACG	AG-ACTGCTC	CCTAT--GG	GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Mesorhabditis_spiculigera_SB15	ACAACTAACG	AG-ACTGCTC	CCTAT--GG	GAGCA-GTCA	TC-GAAATGG	GTA-AAATTC
Mesorhabditis_anisomorpha_SB12	ACGAATAACG	AG-ACTGTTC	CCTCT--GG	GAACA-GTCA	TC-GAAATGG	GTC-TAATAC
Teratorhabditis_palmarum_DF501	ACAACTAACG	AG-ACTGCTC	CTACG--G	GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Teratorhabditis_synpapillata_S	ACAACTAACG	AG-ACTGCTC	CCTAT--GG	GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Caenorhabditis_briggsae_PB102	AAAAATATAA	AG-ACTCATC	CTTTT--G	GATGA-GTTA	TT-TCAATGA	GTT-GAATAC
Caenorhabditis_elegans_N2	AAAAATATAA	AG-ACTCATC	CTTTT--G	GATGA-GTTA	TT-TCAATGA	GTT-GAATAC
Caenorhabditis_sp_CB5161	AAAAATATAA	AG-ACTCATC	CTTTT--G	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_sp_PS1010	AAAAATATAA	AG-ACTCATC	CTTTT--G	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_japonica	AAAAATATAA	AG-ACTCATC	CTTTT--G	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_sp_DF5170	AAAAATATAA	AG-ACTCATC	CTTTT--G	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_sp_SB341	AAAAATAAAA	AG-ACCCCTC	CCAAC--G	GAGGG-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_plicata	AAAAATAGAA	AG-AGTCATT	CTTTAT--G	GATGA-CTCA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_vulgaris	AAAAATATAA	AG-ACTCATC	CTTTT--G	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_sonorae	AAAAATAAAA	AG-ACCCCTC	CTTT--G	GAGGG-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_drosophilae	AAAAATATAA	AG-ACTCATC	CTTC--G	GGATGAGTTA	TT-TCAATGA	GTT-GAATAT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mm-mmmmmmm	mmmm-----m	mmmmmm-mmmmm	mm-mmmmmmm	mmmm-mmmmmmm

	670	680	690	700	710	720
Phasmarhabditis_hermaphrodita	GAACATAAAAA	AG-ACTCATC	TTTCT----	A GATGG-GTTA	TT-TCAATGA	GTT-GAAGCTT
Phasmarhabditis_neopapillosa	GAACATAAAAA	AG-ACTCATC	TTTCT----	A GATGG-GTTA	TT-TCAATGA	GTT-GAAGCTT
Phasmarhabditis_neopapillosa	AAAAATAAAA	AG-ACCGATC	CTAT-----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Pellioditis_mediterranea_SB173	AAAAATAAAA	AG-ATCAATC	CTAT-----	G GATTG-ATTA	TT-TCAATGA	GTT-GAGCTTT
Pellioditis_marina	AAAAATAAAA	AG-ACCGATC	CTAT-----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Prodontorhabditis_wirthi	GGAATAACAA	AA-CCTTTGC	CCTAT----	GG GTAGA-GGTA	TT-TGAATGA	GAA-GATATT
Crustorhabditis_scanica	ACAACTAACG	AG-ACGTGTC	TCTAT----	GA GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Dolichorhabditis_sp_CEW1	AAAAATAAAA	AG-TCCAATC	CTAAC----	G GATCG-GATA	TT-TCAATGA	ATT-GAGCTTT
Oscheius_sp_DF5000	AAAAATAAAA	AG-ACCATTC	-TAAC----	G GAACG-GTTA	TT-TCAATGA	ATT-GAGCTTT
Oscheius_sp_BW282	AAAAATAAAA	AG-ACCAATC	CTCAC----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Oscheius_myriophila_EM435	AAAAATAAAA	AG-ACCAATC	CTCAC----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Oscheius_insectivora	AAAAATAAAA	AG-ACCAATC	CTTAC----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Oscheius_dolichuroides	AAAAATAAAA	AG-ACCATTC	CTAAC----	G GAACG-GTTA	TT-TCAATGA	ATT-GAGCTTT
Rhabditis_blumi_DF5010	GAAAATAAAA	AG-ACCAATC	CTCT-----	G GATTG-GTTA	TT-TCAATGA	ATT-GAGCTTT
Rhabditis_sp_PS1191	GAAAATAAAA	AG-ACCAATC	CAC-----	G GATTG-GTTA	TT-TCAATGA	ATT-GAGCTTT
Rhabditis_sp_PS1010	AAAAATATAA	AG-ACTCATC	CTTTT----	G GATGA-GTTA	TT-TCAATGA	GTT-GAATTTT
Rhabditis_myriophila_EM435	AAAAATAAAA	AG-ACCAATC	CTCAC----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Rhabditis_colombiana	AAAAATAAAA	AG-ACCAATC	CTTAC----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Rhabditella_axei_DF5006	AAAAATAAAG	AG-ACCAATC	CTTT-----	G GATTG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Rhabditella_sp_DF5044	AAAAATAAAG	AG-ACCAATC	CTTT-----	G GATTG-GTTA	TT-TCAATGA	GTT-GAGCTTT
Cuticularia_sp_PS2083	AACAATAACG	AG-ACCGTCT	TCTAT----	GA AGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Necator_americanus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Kalicephalus_cristatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Ancylostoma_caninum	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_cantonensis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_costaricensis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_malaysiensis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_dujardini	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_vasorum	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Aulurostrongylus_abstrusus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Didelphostrongylus_hayesi	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Crenosoma_mephitidis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Crenosoma_vulpis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Otostrongylus_circumlatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Troglostrongylus_wilsoni	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Filaroides_martis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Oslerus_osleri	AAAAATAAAA	AG-ACCGTTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Parafilaroides_decorus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Metastrongylus_salmoi	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Metastrongylus_elongatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Skrjabinstrongylus_chitwoodrum	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Parelaphostrongylus_odocoilei	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Muelerius_capillaris	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Protostrongylus_rufescens	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Halocercus_invaginatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Pseudalius_inflexus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Stenurus_minor	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Torynurus_convolutus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Syngamus_trachea	AAAAATAAAA	AG-ACCGTTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Stephanurus_dentatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Nematodirus_battus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GAGCTTT
Strongylus_equinus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Labiostrongylus_bipapillosus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Petrovinema_poculatum	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Cylicocyclus_insignis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Chabartia_ovina	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Cyclodontostomum_purvisi	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Zoniolaimus_mawsonae	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Hypodontus_macropi	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Deletrocephalus_dimidiatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_eckerti_P7B8	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_capreolus_P3B2	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_sp_P6A1	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_filaria	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_capreolus_P2C10	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_viviparus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Haemonchus_sp_V3091	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Haemonchus_contortus	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Haemonchus_placei	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Haemonchus_similis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Ostertagia_ostertagi	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Ostertagia_leptospicularis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Nippostrongylus_brasileinsis	AAAAATAAGA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Heligmosomoides_polygyrus	AAAAATAAGA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Trichostrongylus_colubriformis	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Tetrabothriostongylus_mackerr	AAAAATAAAA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Herpestrongylus_pythonis	AAAAATAAGA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Nicollina_cameroni	AAAAATAAGA	AG-ACCATTC	CTAT-----	G GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mm-mmmmmmm	mmmm-----m	mmmmmm-mmmmm	mm-mmmmmmm	mmmm-mmmmmmm

	670	680	690	700	710	720
Filarimena flagrifer	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA GTT-GATCAT
Amidostomum cygni	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTTA	TT-TCAATGA GTT-GATCAT
Teratocephalus lirellus	AAAAATAACG	AG-ACCGTTC	TCTAT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Brumptaemilius justini	AAAAATAACG	AG-ACCGTTC	TCTWT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Anguillicola crassus	AAAAATAACG	GG-GCCGTTC	TCTTT---	GA	GGCCG-GTTA	CC-GGAATGG GTA-CAATCT
Dracunculus medinensis	AAAAATAACG	AG-ACCGTTC	TCATT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Dracunculus oesophageus	AAAAATAACG	GG-ACCGTTC	TCATT---	GA	GGCCG-GTTA	TT-GGAATGG GTA-CAATTT
Dracunculus sp. V3104	AAAAATAACG	AG-ACCGTTC	TCATT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Philonema sp. A	AAAAATAACG	AG-ACCGTTC	TCAAT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Philometra obturans	AAAAATAACG	AG-ACCGTTC	TCGAT---	GA	GGCCG-GTTA	TC-GGAATGG GCA-CAATCT
Camallanus oxycephalus	AAAAATAACG	AG-ACCGTTC	TCTTC---	GA	GGCCG-GTTA	TC-GGAATGA GTA-CAATTT
Acanthocheilonema viteae	AAAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Loa loa	AAAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Onchocerca cervicalis	AAAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Dirofilaria immitis	AAAAATAACG	AG-ACCGTTC	TCTAT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Brugia malayi	AAAAATAACG	AG-ACCGTTC	TCTTT---GR	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Wuchereria bancrofti	AAAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Litomosoides sigmodontis	AAAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTTA	TC-GGAATGN NTA-CAATTT
Setaria digitata	AAAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Gnathostoma turgidum	AAAAATAACG	AA-ACCGATC	TCAAT---	GA	GGCCG-GTTA	TC-GGAATGA GTC-ACGCTT
Gnathostoma neoprocyonis	AAAAATAACG	AA-ACCGATC	TCAAT---	GA	GGCCG-GTTA	TC-GGAATGA GTC-ACGCTT
Gnathostoma binucleatum	AAAAATAACG	AA-ACCGATC	TCAAT---	GA	GGCCG-GTTA	TC-GGAATGA GTC-ACGCTT
Physaloptera alata	AAAAATAACG	AG-ACCGTTC	TCTTT---	GG	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Physaloptera turgida	AAAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Ascarophis arctica	AAAAATAACG	AG-ACCGTTC	TCTTC---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Spinitectus carolini	AAAAATAACG	AG-ACCGTTC	TCTTC---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Thelazia lacrymalis	AAAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTTA	TC-GGAATGG GTA-CAATTT
Aphelenchus avenae	AAAAATAACG	AG-ACCGTTC	TCTAT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATCT
Aphelenchoides fragariae	AGAAATAACA	AG-TCCGTTC	TCTTT---	GA	GGTCG-GACA	TT-GGAATGG ATG-AAATCT
Bursaphelenchus sp	AGAAATAACG	AG-ATCGTTC	TCTTT---	GA	GGTCG-GTTA	TC-GGAATGG GTA-CAATCC
Deladenus sp	AGAAATAACG	AG-ACCGTTC	TCTAC---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATCC
Criconema sp	AAAAATAACA	AG-GCTGTTC	TCTAC---	GA	GGCCA-GCCA	TT-GGAATGG GTA-CAACAC
Hemicycliophora conida	AAAAATAACA	AG-GCCGTTC	TCTTAT---	GA	GGCCG-GCCA	TT-GGAATGG GTA-CAACTC
Paratylenchus dianthus	AAAAATAACA	AG-GCCGTTC	TCTTT---	GA	GGCCG-GCCA	TT-GGAATGG GTA-CAACTT
Pratylenchus thornei	AGAAATAACG	AG-GCCGTTC	TCTTT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Tylenchulus semipetrans	AAAAATAACG	AG-ACCGTTC	TCTAC---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAACTC
Ditylenchus angustus	AGAAATAACG	AG-ACCGTTC	TTTTT---	AA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Subanguina radicola	AAAAATAACG	AG-ACCGTTC	TCTAT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATCT
Tylenchorhynchus maximus	AGAAATATCG	AG-ACCGTTC	TCTTAA---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATCT
Geocenamus quadrifer	AGAAATAACG	AG-ACCGTTC	TCTAC---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Globodera pallida	AGAAATAACG	AG-ACCGATC	TCTTAT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Scutellonema bradys	AGAAATAACG	AG-GCCGTTC	TCTTAC---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Helicotylenchus dihystrera	AGAAATAACA	AG-ACCGTTC	TCTTAC---	GA	GGCCG-GTTA	TT-GGAATGG GTA-CAATTT
Rotylenchus robustus	AGAAATAACG	AG-ACCGTTC	TCACAA---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATCT
Meloidogyne arenaria	AGAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Meloidogyne incognita KT	AGAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Meloidogyne javanica	AGAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Meloidogyne artiellia	AGAAATAACG	AG-ATCGTTC	TCTTCA---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAACTT
Meloidogyne duytsi	AGAAATAACG	AG-ATCGTTC	TCTTAT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Meloidogyne exigua	AGAAATAACG	AG-ACCGTTC	TCTTT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Meloidogyne hapla	AGAAATAACG	AG-ATCGTTC	TCAAAT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Meloidogyne ichinohei	AGAAATAACG	AG-GCCGTTC	TCTTCT---	GA	GGCCG-GTCA	TC-GGAATGG GAA-AAATTT
Meloidogyne maritima	AGAAATAACG	AG-ATCGTTC	TCTTAT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Meloidogyne microtyla	AGAAATAACG	AG-ATCGTTC	TCATAT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Nacobbus aberrans	AGAAATAACG	AG-ACAGTTC	TCTTAT---	GA	GGCCT-GTCA	TC-GGAATGG GTA-CAATCT
Pratylenchoides ritteri	AGAAATAACG	AG-ACCGTTC	TCTAC---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Pratylenchoides magnicauda	AGAAATAACG	AG-ACCGTTC	TCTAC---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATTT
Hirschmanniella sp. JH_2003	AGAAATAACG	AG-ACCGTTC	TCTTCA---	GA	GGCCA-GTCA	TC-GGAATGG GTA-CAATTT
Radopholus similis	AGAAATAACG	AG-GCCGTTC	TCATAC---	GA	GGCCG-GCCA	TT-GGAATGG AAG-AGGTTT
Pratylenchus goodeyi VF	AGAAATAACG	AG-ACCGTTC	TCTTCA---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATCC
Boleodorus thylactus clone2	AGAAATAACG	AG-GCCGTTC	TCTAT---	GA	GGCCG-GTCA	TC-GGAATGG GTA-CAATCT
Philippine Sequence 1	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTTA	TT-TCAATGA GTT-GATCAT
Philippine Sequence 2	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTTA	TT-TCAATGA GTT-GATCAT
Philippine Sequence 3	AAAAATAGAA	AG-GGTAATC	CTCT----	G	GATTT-CCTA	TT-TCAATGA GTT-GAGCCC
Philippine Sequence 4	AGAAATGACA	AG-ATCTATT	CCTTCG---	GG	GATGG-ATCA	TT-GGAATGG TTT-GAATTT
Philippine Sequence 5	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA GTT-GATCAT
Philippine Sequence 6	AAAAATAAAA	AG-GCCAATC	CTAT----	G	GATCG-GTTA	TT-TCAATGA GCT-GAGCTT
Philippine Sequence 7	AAAAATATAA	AG-ACTCATC	CTTTT---	G	GATGA-GTTA	TT-TCAATGA GTT-GAATTT
Tahiti Sequence 1	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTTA	TT-TCAATGA GTT-GATCAT
Tahiti Sequence 2	AAAAATAAAA	AG-ACCATT	CTAT----	G	GATCG-GTTA	TT-TCAATGA GTC-GWTCAT
Thailand Sequence 1	AAAAATAACA	AG-ACCGTTC	TCTT---	AGA	GGCCG-GTTA	TT-TCAATGA GTA-CAATTT
Ogasawara Sequence 1	AGAAATGACA	AGATCTATT	CTTCG---	GG	GATGG-ATCA	TT-GGAATGG TTT-GAATTT
Ogasawara Sequence 2	GTACATAAAA	AG-ACTCATC	TTTC----	TA	GATGG-GTTA	TT-TCAATGA GTT-GAATTT
Ivory Coast Sequence 1	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA GTT-GATCAT
Ivory Coast Sequence 2	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA GTT-GATCAT
Ivory Coast Sequence 3	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAATCGGTCA	TT-TCAATGA GTT-GATCAT
Ivory Coast Sequence 4	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA GTT-GATCAT
Ivory Coast Sequence 5	AAAAATAACA	AG-ACCGTTC	TCTAT---	GA	GGCCG-GTTA	TT-GGAATGA GTA-CAATTT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mm-mmmmmmm	mmmm-----m	mmmmmm-mmmmm	mm-mmmmmmm	mmmm-mmmmmmm

	670 680 690 700 710 720
Ivory Coast Sequence 6	AAAAATAAAA AG-ACCATTCT AT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Ivory Coast Sequence 7	AAAAATAAAA AG-ACCATTCT AT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Singapore Sequence 1	AAAAATAGAA AG-GGTAATC CTCT-----G GATTTCCTA TT-TCAATGA TTT-GAGCCC
Singapore Sequence 2	AAAAATAACA AG-ACCGTTC TCTT---AGA GGCCG-GTTA TT-GGAATGA GTA-CAATAT
Singapore Sequence 3	AAAAATATAA AG-ACTCATC CTTTT---G GATGA-GTTA TT-TCAATGA GTT-GAATAC
Singapore Sequence 4	AAAAATAAAA AG-ACCATTCT AT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Nigerian Sequence 1	AAAAATAAAA AG-ACCATTCT AT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Nigerian Sequence 2	AAAAATAAAA AG-ACCATTCT AT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Nigerian Sequence 3	AAAAATAAAA AG-ACCATTCT AT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mm-mmmmmmm mmmmm-----m mmmmm-mmmmm mm-mmmmmmm mmm-mmmmmmm

	730 740
Gordius_aquaticus	A-AATCCTTT AACGAGGATC TATTGGAG
Priapulius_caudatus	A-AATCCTTT AACGAGGATC TATTGGAG
Brachionus_plicatilis	A-AAACCCTT AACGAGGATC TATTGGAG
Chordodes_morgani	A-AATCCTTT AACGAGGATC TATTGGAG
Paractinolaimus_macrolaimus	A-AATCCTTT AACGAGGATC TATTGGAG
Aporcelaimellus_obtusicaudatus	A-AATCCTTT AACGAGGATC TATTGGAG
Wilsonema_schuurmansstekhoveni	A-AACCCTTT AACGAGGACC TATGAGAG
Mesodorylaimus_sp_cf_nigritul	A-AATCCTTT AACGAGGATC TATTGGAG
Mesodorylaimus_bastiani	A-AATCCTTT AACGAGGATC TATTGGAG
Mesodorylaimus_japonicus	A-AATCCTTT AACGAGGATC TATTGGAG
Pungentus_sp._PDL_2005	A-AATCCTTT AACGAGGATC TATTGGAG
Allodorylaimus_sp	A-AATCCTTT AACGAGGATC TATTGGAG
Eudorylaimus_carteri	A-AATCCTTT AACGAGGATC TATTGGAG
Microdorylaimus_sp	A-AATCCTTT AACGAGGATC TATTGGAG
Longidorus_elongatus	A-AATCCTTT AACGAGGATC TATTGGAG
Xiphinema_rivesi	A-AATCCTTT AACGAGGATC TATTGGAG
Tylencholaimus_sp	A-AATCCTTT AACGAGGATC TATTGGAG
Anoplostoma_sp._BHMM_2005	A-AATCCTTT AACGAGGATC TATTGGAG
Adoncholaimus_fuscus	A-AATCCTTT ATCGAGGATC TATTGGAG
Enoplus_meridionalis	A-AATCCTTT AACGAGGATC TATTGGAG
Enoplus_brevis_U88336	A-AATCCTTT AACGAGGATC TATTGGAG
Enoplus_communis	A-AATCCTTT AACGAGGATC TATTGGAG
Enoploides_brunettii	A-AATCCTTT AACGAGGATC TATTGGAG
Syringolaimus_striatocaudatus	A-AATCCTTT AACGAGGATC CATTGGAG
Ironus_dentifurcatus	A-AATCCTTT AATGAGGATC TATTGGAG
Calyptronema_maxweberi	A-AATCCTTT ATCGAGGATC TATTGGAG
Viscosia_sp._BHMM_2005	A-AATCATTT ATCGAGGATC TATTGGAG
Viscosia_viscosa	A-AATCATTT ATCGAGGATC TATTGGAG
Pontonema_vulgare	A-AATCATTT ATCGAGGATC TATTGGAG
Oncholaimus_sp._BHMM_2005	A-AATCATTT ATCGAGGATC TATTGGAG
Alaimus_sp_PDL_2005	A-AATCCTTT AACGAGGATC AATTGGAG
Prismatolaimus_intermedius	A-AATCCTTT AACGAGGAT- TATTGGAG
Tobrilus_gracilis	A-AATCCTTT AACGAGGATC TATTGGAG
Tripyla_cf_filicaudata_JH_2004	A-AATCCTTT AACGAGGAAC TATTGGAG
Bathylaimus_sp	A-AATCCTTT AACGAGGATC TATTGGAG
Bathylaimus_assimilis	A-AATCCTTT AACGAGGATC TATTGGAG
Bathylaimus_sp._BHMM_2005	A-AATCCTTT AACGAGGATC TATTGGAG
Tripyloides_sp._BHMM_2005	A-AATCCTTT AACGAGGATC TATTGGAG
Trischiostoma_monohystera	A-AATCCTTT AACGAGGATC TATTGGAG
Mermis_nigrescens	A-AATCCTTT AACGAGGATC TATTAGAG
Mylonchulus_arenicolus	A-AATCCTTT AACGAGGATC TATTAGAG
Anatonchus_tridentatus	A-AAAACCTTT AACGAGGATC TATTAGAG
Mononchus_truncatus	A-AATCCTTT AACGAGGATC TATTAGAG
Prionchulus_muscorum	A-AATCCTTT AACGAGGATC TATTAGAG
Clarkus_sp	A-AATCCTTT AACGAGGATC TATTAGAG
Trichinella_spiralis	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_papuae	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_britovi	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_murrelli	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_pseudospiralis	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_nativa	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_zimbabwensis	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_nelsoni	A-CACACGCT AACGAGTAGC AATTGGAG
Trichuris_suis	A-CACAAGCT CGGCTAAATC TATTGGAG
Trichuris_trichiura	A-CATAAGTT CGGCTAAATC TATTGGAG
Trichuris_muris	A-CATAAGCT CGGCTAATTC TATTGGAG
Tylolaimophorus_minor	A-AATCCTTT AACGAGGATC TATTGGAG
Paratrichodorus_pachydermus	A-AATCCTTT AACGAGGATC TATTGGAG
Paratrichodorus_anemones	A-AATCCTTT AATGAGGATC TATTGGAG
Trichodorus_primitivus	A-AATCCTTT AACGAGGATC TATTGGAG
Axonolaimus_helgolandicus	A-AACCTTTT AACGAGGATC TACGAGAG
Ascolaimus_elongatus	A-AACCTTTT AACGAGGATC TACGAGAG
Odontophora_rectangula	A-AACCTTTT AACGAGGATC TACGAGAG
Cylindrolaimus_sp._202149	A-AACCTTTT AACGAGGATC TACGAGAG
NUCLEOTIDES INCLUDED	m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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....|....| ....|....| ....|...
730      740
Tylocephalus_auriculatus A-AACCCCTTT AACGAGGACC TATGAGAG
Plectus_acuminatus_BS9 A-AACCCCTTT AACGAGGACC TATGAGAG
Plectus_aquatilis A-AACCCCTTT AACGAGGACC TATGAGAG
Anaplectus_sp A-AACCCCTTT AACGAGGACC TATGAGAG
Anisakis_sp_WKT A-AACCCCGTT AACGAGGATC TATGAGAG
Anisakis_sp_Nadler A-AACCCCGTT AACGAGGATC TATGAGAG
Contracaecum_multipapillatum A-AACCCCGTT AACGAGGATC TATGAGAG
Pseudoterranova_decipiens A-AACCCCGTT AACGAGGATC TATGAGAG
Raphidascaris_acus A-AACCCCGTT AACGAGGATC TATGAGAG
Terranova_caballeroi A-AACCCCGTT AACGAGGATC TATGAGAG
Ascaris_suum A-AACCCCGTT AACGAGGATC TATGAGAG
Ascaris_lumbricoides A-AACCCCGTT AACGAGGATC TATGAGAG
Baylisascaris_procyonis A-AACCCCGTT AACGAGGATC TATGAGAG
Baylisascaris_transfuga A-AACCCCGTT AACGAGGATC TATGAGAG
Parascaris_equorum A-AACCCCGTT AACGAGGAAC TATGAGAG
Porrocaecum_depressum A-AACCCCGTT AACGAGGATC TATGAGAG
Toxascaris_leonina A-AACCCCGTT AACGAGGATC TATGAGAG
Heterocheilus_tunicatus A-AACCCCGTT AACGAGGATC TATGAGAG
Goezia_pelagia A-AACCCCGTT AACGAGGATC TATGAGAG
Hysterothylacium_fortalezae A-AACCCCGTT AACGAGGATC TATGAGAG
Hysterothylacium_pelagicum A-AACCCCGTT AACGAGGATC TATGAGAG
Hysterothylacium_reliquens A-AACCCCGTT AACGAGGATC TATGAGAG
Iheringascaris_inquies A-AACCCCGTT AACGAGGATC TATGAGAG
Toxocara_canis A-AACCCCGTT AACGAGGATC TATGAGAG
Nemhelix_bakeri A-AACCCCGTT AACGAGGATC TATGAGAG
Raillietnema_sp._V3060 A-AACCCCGTT AACGAGGATC TATAAGAG
Cruzia_americana A-AACCCCGTT AACGAGGATC TATGAGAG
Heterakis_sp_14690 A-AATCCGTT AACGAGGATC TATGAGAG
Heterakis_gallinarum A-AATCCGTT AACGAGGATC TATGAGAG
Paraspidodera_sp_21303 A-AACCCCGTT AACGAGGATC TATGAGAG
Chromodora_nudicapitata A-AACCCCTTT AACGAGGATC TATTGGAG
Chromadora_sp._BHMM_2005 A-AACCCCTTT AACGAGGATC TATTGGAG
Atrochromadora_microlaima A-AACCCCTTT AACGAGGATC TATTGGAG
Chromadorina_germanica A-AACCCCTTT AACGAGGATC TATTGGAG
Chromadorita_tentabundum A-AACCCCTTT AACGAGGATC TATTGGAG
Dichromadora_sp._BHMM_2005 A-AACCCCTTT AACGAGGATC TATTGGAG
Neochromadora_BHMM_2005 A-AACCCCTTT AACGAGGATC TATTGGAG
Spilophorella_paradoxa A-AACCCCTTT AACGAGGATC TATTGGAG
Paracanthonchus_caecus A-AACCCCTTT AACGAGGATC TATTGGAG
Paracyatholaimus_intermedius A-AACCCGTT AACGAGGATC TATTGGAG
Praeacanthonchus_punctatus A-AACCCCTTT AACGAGGATC TATTGGAG
Praeacanthonchus_sp A-AACCCCTTT AACGAGGATC TATTGGAG
Cyatholaimus_sp._BHMM_2005 A-AACCCCTTT AACGAGGATC TATTGGAG
Spirinia_parasitifera A-AACCCCTTT AACGAGGATC TATTGGAG
Acanthopharynx_micans A-AACCCCTTT AACGAGGATC TATTGGAG
Xyzzors_sp A-AACCCCTTT AACGAGGATC TATTGAAG
Metachromadora_sp A-AACCCCTTT AATGAGGATC TATTGGAG
Metachromadora_remanei A-AACCCCTTT AATGAGGATC TATTGGAG
Catanema_sp A-AACCCCTTT AACGAGGATC TATTGGAG
Eubostrichus_dianae A-AACCCCTTT AACGAGGATC TATTGGAG
Eubostrichus_topiarus A-AACCCCTTT AACGAGGATC TATTGGAG
Eubostrichus_parasitiferus A-AACCCCTTT AACGAGGATC TATTGGAG
Laxus_oneistus A-AACCCCTTT AACGAGGATC TATTGGAG
Laxus_cosmopolitus A-AACCCCTTT AACGAGGATC TATTGGAG
Leptonemella_sp A-AACCCCTTT AACGAGGATC TATTGGAG
Robbea_hypermnestra A-AACCCCTTT AACGAGGATC TACTGGAG
Stilbonema_majum A-AACCCCTTT AACGAGGATC TATTGGAG
Monoposthia_costata A-AAAAATTT AACGATA-TC AATTGGAG
Nudora_bipapillata A-AACCCCTTT AACGAGGATC AATTGGAG
Calomicrolaimus_parahonestus A-AACCCCTTT AACGAGGATC TATTGGAG
Calomicrolaimus_sp._BHMM_2005 A-AACCCCTTT AACGAGGATC TATTGGAG
Molgolaimus_demani A-AACCCCTTT AACGAGGATC TATTGGAG
Diplogaster_lethieri A-AAGACTTT AACGAAG-TC TATGGGAG
Aduncospiculum_halicti A-AACCCCTTT AACGAGGATC TATGAGAG
Pristionchus_lheritieri A-AAGACTTT AACGAAG-TC TATGGGAG
Pristionchus_pacificus A-AAGACTTT AACGAAG-TC TATGGGAG
Pristionchus_pacificus A-AAGACTTT AACGAAG-TC TATGGGAG
Sabatieria_punctata_STRAIN_343 A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_sp._355_BHMM_2005 A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_celtica A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_punctata_STRAIN_200 A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_punctata_STRAIN_223 A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_sp._210_BHM_2005 A-AATCCTTT AACGAGGATC TAGCGGAG
Setosabatieria_hilarula A-AATCCTTT AACGAGGATC TAGCGGAG
Desmolaimus_zeelandicus A-AACGACTT ACCGAATT-C TATCAGAG
Terschellingia_longicaudata A-AATCCTTT AACGAGGATC TAGCGGAG
Cyartonema_elegans A-AATCCTTT AACGAGGATC TAGCGGAG
Tridentulus_sp A-AATCTTTT AACGAGGATC TAGTGGAG
NUCLEOTIDES INCLUDED m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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730      740
Diplolaimelloides_meyli      A-AATCTTTT AACGAGGATC TAGTGGAG
Diplolaimella_diavengatensis A-AATCTTTT AACGAGTAAC AAGTGGAG
Geomonhystera_disjuncta      A-AAAAATTC GGCAAGGATC TAGTGGAG
Sphaerolaimus_hirsutus       A-AATGTGTT ATCGAGTAAC TAGCAGAG
Theristus_acer                A-AACCCTTT AACGAGGACC TAGCGGAG
Daptonema_procerus           A-AATCCTTT AACGAGGACC TAGCGGAG
Daptonema_hirsutum           A-AATCCTTT AACGAGGACC TAGCGGAG
Daptonema_normandicum        A-AATCCTTT AACGAGGACC TAGCGGAG
Daptonema_oxycerca           A-AATCCTTT AACGAGGACC TAGCGGAG
Daptonema_setosum            A-AATCCTTT AACGAGGACC TAGCGGAG
Desmodora_communis           A-AACCCTTT AACGAGGATC TATTGGAG
Desmodora_ovigera            A-AACCCTTT AACGAGGATC TATTGGAG
Metadesmolaimus_sp           A-AATCCTTT AACGAGGACC TAGCGGAG
Dentostomella_sp            A-AACCCTTT AATGAGGATC TATGAGAG
Bunonema_franzi              A-AAACCCAT AACGAGGACC TATGAGAG
Bunonema_sp                  A-AAACCCAT AACGAGGATC TATGAGAG
Seleborca_complexa           A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeloides_nanus           A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeloides_bodenheimeri    A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeloides_sp_PS1146       A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeles_ciliatus           A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeles_sp_PS1156          A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeles_complexus_WCUG2    A-AACCCTTT AACGAGTATC TATGAGAG
Cephaloboides_sp_SB227       A-AATAGCTC TTCGAGGACC CAGTGGAG
Cephalobus_cubaensis         A-AACCCTTT AACGAGTATC TATGAGAG
Cephalobus_sp_PS1143         A-AACCCTTT AACGAGTATC TATGAGAG
Cephalobus_sp_PS1196         A-AACCCTTT AACGAGTATC TATGAGAG
Cephalobus_oryzae_PS1165     A-AACCCTTT AACGAGTATC TATGAGAG
Cervidellus_alutus           A-AACCCTTT AACGAGTATC TATGAGAG
Pseudacrobeles_variabilis    A-AACCCTTT AACGAGTATC TATGAGAG
Triligulla_aluta             A-AACCCTTT AACGAGTATC TATGAGAG
Zeldia_punctada              A-AACCCTTT AACGAGTATC TATGAGAG
Myolaimus_sp_U81585          A-AATCCTTT AACGAGGATC TATGAGAG
Rhabditophanes_sp_KR3021     A-AATAAC-T CGTT-GG-TA AAGGAAAG
Brevibucca_sp_SB261          A-AACCCTTT AACGAGGATC TATGAGAG
Halicephalobus_gingivalis    A-AATTCATT AAAGATTATC AATGAGAG
Panagrobelus_stammeri        A-AACCCTTT AACGATTATC TATGAGAG
Plectonchus_sp_PDL0025       A-AACCCTTG AACGAACG-C AAAAAGAG
Turbatrix_aceti              ATAAACCTTA CAAATTG-C TATGAGAG
Panagrellus_redivivus        A-CAACCTTC CTAGATTATC TATGAGAG
Panagrellus_redivivus_PS1163 A-CAACCTTC CTAGATTATC TATGAGAG
Panagrolaimus_subelongatus   A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus_davidi         A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus_cf_rigidus_AF40 A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus_sp_Sourhope_ED2 A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus_sp_Sourhope_ED2 A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus_sp_Sourhope_ED2 A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus_sp_Sourhope_ED2 A-AAATCATT AAAGAGTATC AATGAGAG
Steinernema_carpocapsae      A-AACTCTTT AACGAGGACC TATGAGAG
Strongyloides_stercoralis    A-AATMACCT TGTGTGTAAG GGAAGGGG
Strongyloides_ratti          A-AATAAC-T TGTGTGTAAG GGAAGGGG
Diploscapter_sp_PS1897       A-AAAAGCTC TTCGAGAAAC TAAGAGAG
Diploscapter_sp_PS2017       A-AAAAGCTC TTCGAGAAAC TAAGAGAG
Heterorhabditis_bacteriophora A-AATCTATC TTCGAGTATC TAGTGGAG
Heterorhabditis_hepialus     A-AATCTATC TTCGAGTATC TAGTGGAG
Heterorhabditis_zelandica    A-AATCTATC TTCGAGTACC TAGTGGAG
Parasitorhabditis_sp_SB281   A-AACCCTTT AACGAGGATC TATGAGAG
Rhabditoides_inermiformis    A-AACCCTTT AACGAGGATC TATGAGAG
Rhabditoides_inermis_DF5001  A-AACCCTTT ATCGAGGATC TATGAGAG
Rhabditoides_regina_DF5012   A-AACCCTTT AACGATGATC AATGAGAG
Poikilolaimus_oxycerca_SB200 A-AACCCTTT AACGAGGATC TATGAGAG
Poikilolaimus_regenfussi_SB199 A-AACCCTTT AACGAGGATC TATGAGAG
Distolabrellus_veechi_DWF1604 A-AACCCTTT AACGAGGATC TATGAGAG
Distolabrellus_veechi_DF5024 A-AACCCTTT AACGAGGATC TATGAGAG
Choriorhabditis_dudichi      A-AAACACTT TATGAGGATC AAATGGAG
Protorhabditis_sp            A-AAAACTCT TTCGAGAAAC TAAGAGAG
Protorhabditis_sp_DF5055     A-AATAACTT CTCGAGAAAC TAAGAGAG
Cruzanema_tripartitum_DF5015 A-AATATCTC GTAAAGGAAC TAGTGGAG
Mesorhabditis_sp_PS1179      A-AACCCTTT AACGAGGATC AATGAGAG
Mesorhabditis_spiculigera_SB15 A-AACCCTTT AACGAGGATC AATGAGAG
Mesorhabditis_anisomorpha_SB12 A-AACCCTTG AACGAGGATC AATGAGAG
Teratorhabditis_palmarum_DF501 A-AACCCTTT AACGAGGATC TATGAGAG
Teratorhabditis_synpapillata_S A-AACCCTTT AACGAGGATC TATGAGAG
Caenorhabditis_briggsae_PB102 A-AATGATTC TTCGAGTATC TAGTGGAG
Caenorhabditis_elegans_N2     A-AATGATTC TTCGAGTAGC AAGGAGAG
Caenorhabditis_sp_CB5161      A-AATGATTC TTCGAGTAGC AAGGAGAG
Caenorhabditis_sp_PS1010     A-AACAATTC TTCGAGTAAC AAGGAGAG
NUCLEOTIDES INCLUDED         m-mmmmmmmmm mmmmmmmmmmm mmm.....

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730      740
Caenorhabditis_japonica      A-AATTATTC TTCGAGTAGC AAGGAGAG
Caenorhabditis_sp_DF5170     A-AATCGTTC TTCAGGTAAC AAGGAGAG
Caenorhabditis_sp_SB341      A-AATGATTC TTCGAGGACC CAGGAGAG
Caenorhabditis_plicata       A-AACTAATC TTCGAGTAAC AAGGAGAG
Caenorhabditis_vulgaris      A-AATGATTC TTCGAGTAGC AAGGAGAG
Caenorhabditis_sonorae       A-AATGATTC TTCGAGGACC CAGGAGAG
Caenorhabditis_drosophilae   A-AATCGTTC TTCAGGTAAC AAGGAGAG
Phasmarhabditis_hermaphrodita A-AATAGTTC TTCGAGGACC CAGTGGAG
Phasmarhabditis_neopapillosa A-AATAGTTC TTCGAGGACC CAGTGGAG
Phasmarhabditis_neopapillosa A-AATAGCTC TTCGAGGACC CAGTGGAG
Pellioiditis_mediterranea_SB173 A-AATAGCTC TTCGAGGACC CAGTGGAG
Pellioiditis_marina          A-AATAGCTC TTCGAGGACC CAGTGGAG
Prodontorhabditis_wirthi     A-AAAATATC TCCGAGAAAC TAAGAGAG
Crustorhabditis_scanica      A-AACCCCTTT AACGAGGATC TATGAGAG
Dolichorhabditis_sp_CEW1     A-AATAGCTC TATGAGGACC TAATGGAG
Oscheius_sp_DF5000           A-AATAGCTC TATGATGATC TAGTGGAG
Oscheius_sp_BW282            A-AATAGCTC TTCGAGGATC TAGTGGAG
Oscheius_myriophila_EM435     A-AATAGCTC TTCGAGGATC TAGTGGAG
Oscheius_insectivora         A-AATAGCTG TTCGAGGACC TAGTGGAG
Oscheius_dolichuroides       A-AATAGCTC TATGATGATC TAGTGGAG
Rhabditis_blumi_DF5010       A-AACAGCTC TTTGAGTAAC AAGTGGAG
Rhabditis_sp_PS1191          A-AATAGCTC TTTGAGTAAC TAGTGGAG
Rhabditis_sp_PS1010          A-AACAATTC TTCGAGTAAC AAGGAGAG
Rhabditis_myriophila_EM435    A-AATAGCTC TTCGAGGATC TAGTGGAG
Rhabditis_colombiana          A-AATAGCTC TTCGAGGACC TAGTGGAG
Rhabditella_axei_DF5006       A-AATAGCTC TTCGAGGACC CAGTGGAG
Rhabditella_sp_DF5044         A-AATAGCTC TTCGAGGACC CAGTGGAG
Cuticularia_sp_PS2083         A-AACCCCTTT AACGAGGATC TATGAGAG
Necator_americanus           A-AACCTTTT TTCGAGGATC AAGTGGAG
Kalicephalus_cristatus       A-AACCTTTT TTCGAGGATC AAGTGGAG
Ancylostoma_caninum          A-AACCTTTT TTCGAGGATC AAGTGGAG
Angiostrongylus_cantonensis  A-AACCTTTT TTCGAGTATC CAGTGGAG
Angiostrongylus_costaricensis A-AACCTTTT TTCGAGTATC AAGTGGAG
Angiostrongylus_malaysiensis  A-AACCTTTT TTCGAGTATC AAGTGGAG
Angiostrongylus_dujardini     A-AACCTTTT TTCGAGTATC AAGTGGAG
Angiostrongylus_vasorum       A-AACCTTTT TTCGAGTACC AAGTGGAG
Aulurostrongylus_abstrusus    A-AACCTCTT TTCGAGTATC AAGTGGAG
Didelphostrongylus_hayesi     A-AACATATT TTCGAGTATC AAGTGGAG
Crenosoma_mephitidis         A-AACCTTTT TTCGAGGATC AAGTGGAG
Crenosoma_vulpis             A-AACCTTTT TTCGAGGATC AAGTGGAG
Otostrongylus_circumlitus     A-AACCTTTT TTCGAGGATC AAGTGGAG
Troglostrongylus_wilsoni      A-AACCTTTT TTCGAGGATC AAGTGGAG
Filaroides_martis             a-AACCTTTT TTCGAGTATC AAGTGGAG
Oslerus_osleri               A-AACATTTT TTCAAGTATC AAGTGGAG
Parafilaroides_decorus        A-AACCTTTT TTCGAGTATC AAGTGGAG
Metastrongylus_salmi          A-AACCTTTT TTCGAGTATC AAGTGGAG
Metastrongylus_elongatus      A-AACCTTTT TTCGAGTATC AAGTGGAG
Skrjabinstrongylus_chitwoodrum A-AACATTTT TTCGAGGATC AAGTGGAG
Parelaphostrongylus_odocoilei A-AACCTTTT TTCGAGTATC AAGTGGAG
Muellerius_capillaris         A-AACGTTTT TTCGAGTATC AAGTGGAG
Protostrongylus_rufescens     A-AACCTTTT TTCGAGTATC AAGTGGAG
Halocercus_invaginatus       A-AACTATTT TTCGAGTATC GAGTGGAG
Pseudalius_inflexus          A-AACTTTTT TTCGAGTATC AAGTGGAG
Stenurus_minor               A-AACCTTTT TTCGAGTATC AAGTGGAG
Torynurus_convolutus         A-AACCTTTT TTCGAGTATC AAGTGGAG
Syngamus_trachea             A-AACCTTTT TTCGAGGATC AAGTGGAG
Stephanurus_dentatus         A-AACTTTTT TTCGAGGATC AAGTGGAG
Nematodirus_battus           A-AACCTTTT TTCGAGGATC AAGTGGAG
Strongylus_equinus           A-AACCTCTT TTCGAGGATC AAGTGGAG
Labiostongylus_bipapillosus  A-AACCTTTT TTCGAGGATC AAGTGGAG
Petrovinema_poculatum        A-AACCTCTT TTCGAGGATC AAGTGGAG
Cylicocyclus_insignis        A-AACCTCTT TTCGAGGATC AAGTGGAG
Chabertia_ovina              A-AACCTTTT TTCGAGGATC AAGTGGAG
Cyclodontostomum_purvisi     A-AACCTTTT TTCGAGGATC AAGTGGAG
Zoniolaimus_mawsonae         A-AACCTTTT TTCGAGGATC AAGTGGAG
Hypodontus_macropi           A-AACCTTTT TTCGAGGATC AAGTGGAG
Deletrocephalus_dimidiatus   A-AACCTTTT TTCGAGGATC AAGTGGAG
Dictyocaulus_eckerti_P7B8     A-AACGTTTT TTCGAGTATC GAGTGGAG
Dictyocaulus_capreolus_P3B2   A-AACGTTTT TTCGAGTATC GAGTGGAG
Dictyocaulus_sp._P6A1         A-AACGTTTT TTCGAGTATC GAGTGGAG
Dictyocaulus_filaria         A-AACCATTT TTCGAGTATC AAGTGGAG
Dictyocaulus_capreolus_P2C10 A-AACGTTTT TTCGAGTATC GAGTGGAG
Dictyocaulus_viviparus       A-AACGTTTT TTCGAGTATC GAGTGGAG
Haemonchus_sp._V3091         A-AACCTTTT TTCGAGGATC AAGTGGAG
Haemonchus_contortus         A-AACCTTTT TTCGAGGATC AAGTGGAG
Haemonchus_placei            A-AACCTTTT TTCGAGGATC AAGTGGAG
Haemonchus_similis           A-AACCTTTT TTCGAGGATC AAGTGGAG
Ostertagia_ostertagi         A-AACCTTTT TTCGAGGATC AAGTGGAG
NUCLEOTIDES INCLUDED         m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmm

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730      740
Ostertagia_leptospicularis A-AACCTTTT TTCGAGGATC AAGTGGAG
Nippostrongylus_brasileinsis A-AACCTTTT TTCGAGGAAC AAGTGGAG
Heligmosomoides_polygyrus A-AACCTTTT TTCGAGGATC CAGTGGAG
Trichostrongylus_colubriformis A-AACCTTTT TTCGAGGATC AAGTGGAG
Tetrabothriostrongylus_mackerr A-AACCTTTT TTCGAGGATC AAGTGGAG
Herpestrostrongylus_pythonis A-AACCTTTT TTCGAGGATC AAGTGGAG
Nicollina_cameroni A-AACCTTTT TTCGAGGATC AAGTGGAG
Filarimena_flagrifer A-AACCTTTT TTCGAGGATC AAGTGGAG
Amidostomum_cygni A-AACTTTT TTCGAGGATC AAGTGGAG
Teratocephalus_lirellus A-AACCTTTT AACGAGGATC TATGAGAG
Brumptaemilius_justini A-AACCCGTT AACGAGGATC TATGAGAG
Anguillicola_crassus A-AACCTTTT AACGAGGATC TATGAGAG
Dracunculus_medinensis A-AACCTGTT AACGAGGACC TATGAGAG
Dracunculus_oesophageus A-AACCTGTT AACGAGGATC TATGAGAG
Dracunculus_sp._V3104 A-AACCTGTT AACGAGGATC TATGAGAG
Philonema_sp_A A-AACCCGTT AACGAGGATC TATGAGAG
Philometra_obturans A-AACCCGTT AACGAGGATC AATGAGAG
Camallanus_oxyccephalus A-AAGCCGTT AACAGGATC TATGAGAG
Acanthocheilonema_viteae A-AACCTGTT AACGAGGATC TATGAGAG
Loa_loa A-AACCCGTT AACGAGGATC TATGAGAG
Onchocerca_cervicalis A-AACCCGTT AACGAGGATC TATGAGAG
Dirofilaria_immitis A-AACCTGTT AACGAGGATC TATGAGAG
Brugia_malayi A-AACCTGTT AACGAGGATC TATGAGAG
Wuchereria_bancrofti A-AACCCGTT AACGAGGATC TATGAGAG
Litomosoides_sigmodontis A-AACCTGTT AACGAGGATC TATGAGAG
Setaria_digitata A-AACCCGTT AACGAGGATC TATGAGAG
Gnathostoma_turgidum A-AACCTCT AACGAGGATC TATGAGAG
Gnathostoma_neoprocyonis A-AACCTCT AACGAGGATC TATGAGAG
Gnathostoma_binucleatum A-AACCTCT AACGAGGATC TATGAGAG
Physaloptera_alata A-AACCCGTT AACGAGGATC TATGAGAG
Physaloptera_turgida A-AACCCGTT AACGAGGATC TATGAGAG
Ascarophis_arctica A-AACCCGTT AACGAGGATC TATGAGAG
Spinitectus_carolini A-AACCCGTT AACGAGGATC TATGAGAG
Thelazia_lacrymalis A-AACCTGTT AACGAGGATC TATGAGAG
Aphelenchus_avenae A-AACCTTT AACGAGTATC TATGAGAG
Aphelenchoides_fragariae A-AACAATTT CATGAGTACC TACGAGAG
Bursaphelenchus_sp A-GACATTTT AACGAGTATC TACGAGAG
Deladenus_sp A-AACCTTTT AACGAGTATC TATGAGAG
Criconema_sp A-AACCTTTT AACGAGTATC AATGAGAG
Hemicycliophora_conida A-AACCTTTT AACGAGTATC AATGAGAG
Paratylenchus_dianthus A-AACCTTTT AACGAGTATC AATGAGAG
Pratylenchus_thornei A-AACCTTTT AACGAGTATC TATGAGAG
Tylenchulus_semipetrans A-AACCTTTT AACGAGTATC AATGAGAG
Ditylenchus_angustus A-AACCTTTT GACGAGTATC TATGAGAG
Subanguina_radicola A-AACCTTTT AACGAGTATC TATGAGAG
Tylenchorhynchus_maximus A-AACCTTTT AACGAGTATC TATGAGAG
Geocenamus_quadriifer A-AACCTTTT AACGAGTATC TATGAGAG
Globodera_pallida A-AACCTTTT AACGAGTATC TATGAGAG
Scutellonema_bradys A-AACCTTTT AACGAGTATC TATGAGAG
Helicotylenchus_dihystera A-AACCTTTT AACGAGTATC TATGAGAG
Rotylenchus_robustus A-AACCTTTT AACGAGTATC TATGAGAG
Meloidogyne_arenaria A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne_incognita_KT A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne_javanica A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne_artiellia A-AACCTTTT AACGAGTATC TACGAGAG
Meloidogyne_duytsi A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne_exigua A-AACCTTTT AACGAGTATC AAGGAGAG
Meloidogyne_hapla A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne_ichinohei A-AATCCTTT ATCGAGTATC TACGAGAG
Meloidogyne_maritima A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne_microtyla A-AACCTTTT AACGAGTATC AAGCAGAG
Nacobbus_aberrans A-AACCTTTT AACGAGTATC TATGAGAG
Pratylenchoides_ritteri A-AACCTTTT AACGAGTATC TATGAGAG
Pratylenchoides_magnicauda A-AACCTTTT AACGAGTATC TATGAGAG
Hirschmanniella_sp._JH_2003 A-AACCTTTT AACGAGTATC TATGAGAG
Radopholus_similis A-AAACCCCC ATTGAGTATC TACGAGAG
Pratylenchus_goodeyi_VF A-AACCTTTT AACGAGTATC TATGAGAG
Boleodorus_thylactus_clone2 A-AACCTTTT AACGAGTATC TATGAGAG
Philippine Sequence 1 A-AACCTTTT TTCGAGTATC CAGTGGAG
Philippine Sequence 2 A-AACCTTTT TTCGAGTATC CAGTGGAG
Philippine Sequence 3 A-AATAGCTC TTGGATGACC CAG....
Philippine Sequence 4 A-AAACCATT AAAGAGTATC AATGAGAG
Philippine Sequence 5 A-AACCTTTT TTCGAGGATC AAG....
Philippine Sequence 6 A-AATAGCTC TACGAGGACC CAGTGGAG
Philippine Sequence 7 A-AATGATTC TTCGAGTAGC AAG....
Tahiti Sequence 1 A-AACCTTTT TTCGAGTATC CAGTGGAG
Tahiti Sequence 2 A-AATCTTTT CTTGAGTATC CAG....
Thailand Sequence 1 A-AATCCGTT AACGAGGATC TATGAGAG
NUCLEOTIDES INCLUDED m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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	730 740
Ogasawara Sequence 1	A-AAACCATT AAAGAGTATC AATGAGAG
Ogasawara Sequence 2	A-AATAGTTC TTCGAGGACC CAGTGAGG
Ivory Coast Sequence 1	A-AACCTTTT TTCGAGTATC AAGTGGAG
Ivory Coast Sequence 2	A-AACCTTTT TTCGAGTATC AAGTGGAG
Ivory Coast Sequence 3	A-AACCTTTT TTCGAGTATC GAATGGAG
Ivory Coast Sequence 4	A-AACCTTTT TTCGAGTATC AAGTGGAG
Ivory Coast Sequence 5	A-AATCCGTT AACGAGGATC TATGAGAG
Ivory Coast Sequence 6	A-AACCTTTT TTCGAGTATC AAGTGGAG
Ivory Coast Sequence 7	A-AACCTTTT TTCGAGTATC AAGTGGAG
Singapore Sequence 1	A-AATAGCTC TTGAT-GACC CAGTGGAG
Singapore Sequence 2	A-AATCCGTT AACGAGGATC
Singapore Sequence 3	A-AATGATTG TTCGAGTAGC AAGGAGAG
Singapore Sequence 4	A-AACCTTTT TTCGAGTATC AAGTGGAG
Nigerian Sequence 1	A-AACCTTTT TTCGAGTATC AAGTGGAG
Nigerian Sequence 2	A-AACCTTTT TTCGAGGATC AAGTGGAG
Nigerian Sequence 3	A-AACCTTTT TTCGAGTATC AAGTGGAG
NUCLEOTIDES INCLUDED	m-m-m-m-m-m-m-m m-m-m-m-m-m-m-m m-m-m-m-m-m-m-m

Appendix 6.3: Log likelihood scores of the different models used for comparison for the (A) the Nematoda and (B) the Rhabditida within Nematoda using the SSU rRNA gene to determine the phylogenetic positions of the nematode sequences from a trial population of *Achatina fulica* and *Laevicaulis alte* in the Philippines. A total of 376 unambiguously aligned nucleotide sites of the 5' end of the SSU rRNA gene were used. Log Likelihood Ratio Tests for both the Nematoda and the Rhabditida revealed GTR+ Γ as the optimal model. Note that the K2P and F81 models are non-nested and therefore cannot be compared using LRT.

(A) Nematoda

(B) Rhabditida

Model	Log-Likelihood Score	Model	Log-Likelihood Score
JC	21909.35435	JC	14354.91018
JC+ Γ	19844.13051	JC+ Γ	13100.12574
F81	21.855.63587	F81	14325.96300
F81+ Γ	19747.80473	F81+ Γ	13031.31768
K2P	21623.78574	K2P	14234.74221
K2P+ Γ	19518.62434	K2P+ Γ	12957.97008
HKY85	21549.39123	HKY	14192.26735
HKY85+ Γ	19361.80971	HKY+ Γ	12844.49245
TN93	21435.08416	TrN93	14129.30390
TN93+ Γ	19353.15979	TN93+ Γ	12846.67437
GTR	21370.61122	GTR	13930.31599
GTR+Γ	19339.54695	GTR+Γ	12725.79552

[illegible]605

	130	140	150	160	170	180
Angiostrongylus cantonensis	ATCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGTATAAA	CCCTGACTTT
Angiostrongylus costaricensis	ATCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGTATAAA	CCCTGACTTT
Angiostrongylus malaysiensis	ATCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGTATAAA	CCCTGACTTT
Angiostrongylus dujardini	ATCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGGTTAAG	CCCTAACTTT
Angiostrongylus vasorum	ATCT-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT
Aelurostrongylus abstrusus	GTTC-ACCTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCAGTAGGA	CCCTGACTTT
Didelphostrongylus hayesi	TTTG-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCAGATAAA	CTCTGACTTT
Crenosoma mephitiditis	ATCC-ATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT
Otostrongylus circumlitus	ATCC-ATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT
Troglostrongylus wilsoni	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT
Filaroides martis	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT
Oslerus osleri	GACG-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATAAAA	CCCTGACTTT
Parafilaroides decorus	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCTATTAAA	CCCTGACTTT
Metastrongylus salmi	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT
Skrjabinogylus chitwoodorum	TTCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGTGATTAAA	CCCTGACTTT
Parelaphostrongylus odocoilei	ATCCTTAATG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TATGCATAAA	CCCTGACTCT
Muellerius capillaris	ATGA-CAATG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCAATAAAA	TCCTGACTTT
Halocercus invaginatus	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCTATTAGA	CCCTGACTTT
Pseudalius inflexus	ATCTCAATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TTCTATTAAA	CCCTGACTTT
Stenurus minor	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCTATTAAA	CCCTGACTTT
Torynurus convolutus	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCTATTAAA	CCCTGACTTT
Nematodirus battus	ATCCTTTTTG	GATAACTGCG	GCAATTCTGG	AGCTAATACA	TGCGTTTAGG	CCCTGACTTT
Nippostrongylus brasiliensis	GTCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCAATAAAA	CCCTGACTTT
NUCELOTIDES INCLUDED	mmmm-mmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	190	200	210	220	230	240
Angiostrongylus cantonensis	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Angiostrongylus costaricensis	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGCAGTTT	GCTGACTCTG
Angiostrongylus malaysiensis	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGCAGTTT	GCTGACTCTG
Angiostrongylus dujardini	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Angiostrongylus vasorum	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Aelurostrongylus abstrusus	-CGGAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Didelphostrongylus hayesi	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATCTTCG	GATGTAGTTT	GCTGACTCTG
Crenosoma mephitiditis	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Otostrongylus circumlitus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Troglostrongylus wilsoni	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Filaroides martis	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Oslerus osleri	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATCTACG	GATGTAGTTT	GCTGACTCTG
Parafilaroides decorus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Metastrongylus salmi	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTACG	GATGTAGTTT	GCTGACTCTG
Skrjabinogylus chitwoodorum	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Parelaphostrongylus odocoilei	GTGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Muellerius capillaris	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATCTTCG	GATGTAGTTT	GCTGACTCTG
Halocercus invaginatus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Pseudalius inflexus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Stenurus minor	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Torynurus convolutus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG
Nematodirus battus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCTCCTTCG	GGTGCTGTTT	GCTGACTCTG
Nippostrongylus brasiliensis	-TGAAAGGGT	GCAATTATTA	GTGCAAAACCA	ATCATCTTCG	GATGTTGTTT	GCTGACTCTG
NUCELOTIDES INCLUDED	-mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	250	260	270	280	290	300
Angiostrongylus cantonensis	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Angiostrongylus costaricensis	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	TGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Angiostrongylus malaysiensis	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	TGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Angiostrongylus dujardini	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Angiostrongylus vasorum	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Aelurostrongylus abstrusus	AATAACGCAG	CATATCGGCG	GCTTGATTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Didelphostrongylus hayesi	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Crenosoma mephitiditis	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Otostrongylus circumlitus	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Troglostrongylus wilsoni	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Filaroides martis	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Oslerus osleri	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Parafilaroides decorus	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Metastrongylus salmi	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Skrjabinogylus chitwoodorum	AATAACGCAG	CATATCGGCG	GCTTGATTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Parelaphostrongylus odocoilei	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Muellerius capillaris	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Halocercus invaginatus	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Pseudalius inflexus	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Stenurus minor	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Torynurus convolutus	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Nematodirus battus	AATAATGCAG	CATATCGGCG	GCTTGACCGC	TGATAATCCG	AAAAAGTGTC	TGCCCTATCA
Nippostrongylus brasiliensis	AATAACGCAG	CATATCGGCG	GCTTGTTTCG	TGATAATCCG	AAAAAGTGTC	TGCCCTATCA
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	310	320	330	340	350	360
Angiostrongylus cantonensis	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Angiostrongylus costaricensis	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Angiostrongylus malaysiensis	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Angiostrongylus dujardini	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Angiostrongylus vasorum	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Aelurostrongylus abstrusus	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Didelphostrongylus hayesi	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Crenosoma mephitiditis	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Otostrongylus circumlitus	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Troglostrongylus wilsoni	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Filaroides martis	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Oslerus osleri	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Parafilaroides decorus	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Metastrongylus salmi	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Skrjabinogylus chitwoodorum	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Parelaphostrongylus odocoilei	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Muellerius capillaris	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Halocercus invaginatus	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Pseudalius inflexus	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Stenurus minor	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Torynurus convolutus	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Nematodirus battus	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
Nippostrongylus brasiliensis	ACCTGATGGT AGTCTATTAG TCTACCATGG TTATTACGGG TAACGGAGAA TAAGGGTTTCG						
NUCELOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm						
	370	380	390	400	410	420
Angiostrongylus cantonensis	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Angiostrongylus costaricensis	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Angiostrongylus malaysiensis	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Angiostrongylus dujardini	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Angiostrongylus vasorum	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Aelurostrongylus abstrusus	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Didelphostrongylus hayesi	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Crenosoma mephitiditis	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Otostrongylus circumlitus	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Troglostrongylus wilsoni	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Filaroides martis	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Oslerus osleri	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Parafilaroides decorus	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Metastrongylus salmi	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Skrjabinogylus chitwoodorum	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Parelaphostrongylus odocoilei	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Muellerius capillaris	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Halocercus invaginatus	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Pseudalius inflexus	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Stenurus minor	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Torynurus convolutus	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Nematodirus battus	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
Nippostrongylus brasiliensis	ACTCCGGAGA GGGAGCCTTA GAAACGGCTA CCACATCCAA GGAAGGCAGC AGGC CGCGAAA						
NUCELOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm						
	430	440	450	460	470	480
Angiostrongylus cantonensis	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Angiostrongylus costaricensis	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Angiostrongylus malaysiensis	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Angiostrongylus dujardini	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Angiostrongylus vasorum	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Aelurostrongylus abstrusus	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Didelphostrongylus hayesi	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Crenosoma mephitiditis	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Otostrongylus circumlitus	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Troglostrongylus wilsoni	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Filaroides martis	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Oslerus osleri	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Parafilaroides decorus	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Metastrongylus salmi	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Skrjabinogylus chitwoodorum	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Parelaphostrongylus odocoilei	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Muellerius capillaris	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Halocercus invaginatus	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Pseudalius inflexus	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Stenurus minor	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Torynurus convolutus	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Nematodirus battus	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
Nippostrongylus brasiliensis	CTTATCCAAT CTTGAATAGA TGAGATAGTG ACTAAAAATA AAAAGACCAT TCCTATGGAA						
NUCELOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm						

	490	500	510	520	530	540	
Angiostrongylus cantonensis	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAGTGGA	GGGCAAGTCT	
Angiostrongylus costaricensis	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Angiostrongylus malaysiensis	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Angiostrongylus dujardini	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Angiostrongylus vasorum	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ACCAAGTGGA	GGGCAAGTCT	
Aelurostrongylus abstrusus	CGGTCATTTC	AATGAGTTGA	TCATAAACTT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Didelphostrongylus hayesi	CGGTTATTTC	AATGAGTTGA	TCATAAACAT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Crenosoma mephitiditis	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	ATCAAGTGGA	GGGCAAGTCT	
Otostrongylus circumlitus	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	ATCAAGTGGA	GGGCAAGTCT	
Troglostrongylus wilsoni	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	ATCAAGTGGA	GGGCAAGTCT	
Filaroides martis	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Oslerus osleri	CGGTCATTTC	AATGAGTTGA	TCATAAACAT	TTTTTCAAGT	ATCAAGTGGA	GGGCAAGTCT	
Parafilaroides decorus	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Metastrongylus salmi	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Skrjabinstrongylus chitwoodorum	CGGTCATTTC	AATGAGTTGA	TCATAAACAT	TTTTTCGAGG	ATCAAGTGGA	GGGCAAGTCT	
Parelaphostrongylus odocoilei	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Muellerius capillaris	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Halocercus invaginatus	CGGTTATTTC	AATGAGTTGA	TCATAAACTA	TTTTTCGAGT	ATCGAGTGGA	GGGCAAGTCT	
Pseudalius inflexus	CGGTTATTTC	AATGAGTTGA	TCATAAACTT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Stenurus minor	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Torynurus convolutus	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCAAGTGGA	GGGCAAGTCT	
Nematodirus battus	CGGTCATTTC	AATGAGTTGA	GCTTAAACCT	TTTTTCGAGG	ATCAAGTGGA	GGGCAAGTCT	
Nippostrongylus brasiliensis	CGGTCATTCC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	AACAAGTGGA	GGGCAAGTCT	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	550	560	570	580	590	600	
Angiostrongylus cantonensis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Angiostrongylus costaricensis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Angiostrongylus malaysiensis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Angiostrongylus dujardini	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Angiostrongylus vasorum	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Aelurostrongylus abstrusus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Didelphostrongylus hayesi	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Crenosoma mephitiditis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Otostrongylus circumlitus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Troglostrongylus wilsoni	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Filaroides martis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Oslerus osleri	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Parafilaroides decorus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Metastrongylus salmi	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Skrjabinstrongylus chitwoodorum	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Parelaphostrongylus odocoilei	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Muellerius capillaris	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Halocercus invaginatus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Pseudalius inflexus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Stenurus minor	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Torynurus convolutus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Nematodirus battus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Nippostrongylus brasiliensis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	610	620	630	640	650	660	
Angiostrongylus cantonensis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Angiostrongylus costaricensis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Angiostrongylus malaysiensis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Angiostrongylus dujardini	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Angiostrongylus vasorum	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Aelurostrongylus abstrusus	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Didelphostrongylus hayesi	AAAGCTCGTA	GTTGGATCTG	AGTCATATGC	AATGATTTCG	TATTAGCGTT	AATTATTGTT	
Crenosoma mephitiditis	AAAGCTCGTA	GTTGGATCTG	AGTCGCATGC	AGTAGTTCGC	CTTTGGCGTT	AATTACTGTT	
Otostrongylus circumlitus	AAAGCTCGTA	GTTGGATCTG	AGTCGCATGC	AATGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Troglostrongylus wilsoni	AAAGCTCGTA	GTTGGATCTG	AGTCACATGC	AGTGATTTCG	CTTTGGCGTT	AATCATTGTT	
Filaroides martis	AAAGCTCGTA	GTTGGATCTG	AGTCGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Oslerus osleri	AAAGCTCGTA	GTTGGATCTG	AGTCGCATGC	AGTGGTTCGC	CCTTGGCGTT	AATCACTGTT	
Parafilaroides decorus	AAAGCTCGTA	GTTGGATCTG	AGTCACATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Metastrongylus salmi	AAAGCTCGTA	GTTGGATCTG	AGTCGCATGC	AGTGATTTCG	CTTTGGCGTT	AATCACTGTT	
Skrjabinstrongylus chitwoodorum	AAAGCTCGTA	GTTGGATCTG	AGTCACATGC	AATGATTTCG	CTTTGGCGTT	AATTATTGTT	
Parelaphostrongylus odocoilei	AAAGCTCGTA	GTTGGATCTG	AGTCGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Muellerius capillaris	AAAGCTCGTA	GTTGGATCTG	AGTCGCATGC	AATGATTTCG	CTTTGGCGTT	AATTATTGTT	
Halocercus invaginatus	AAAGCTCGTA	GTTGGATCTG	AGTCACATGC	AATTGGTTGC	CTTTGGCGTT	AATCATTGTT	
Pseudalius inflexus	AAAGCTCGTA	GTTGGATCTG	AGTCACATGC	AATAATTAC	CTTTGGTTGT	AATTATTGTT	
Stenurus minor	AAAGCTCGTA	GTTGGATCTG	AGTCACACGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Torynurus convolutus	AAAGCTCGTA	GTTGGATCTG	AGTCACACGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Nematodirus battus	AAAGCTCGTA	GTTGGATCTG	AGTCACATGC	AGTGGTTCG	CTTTGGCGTT	AATCGCTGTT	
Nippostrongylus brasiliensis	AAAGCTCGTA	GTTGGATCTG	AGTTACATGC	AG-TGGTTAC	CTTTGGTTGT	AATCGCTGTT	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	670	680	690	700	710	720
Angiostrongylus cantonensis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Angiostrongylus costaricensis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATCTCGATT	TCTTTAGTGG	CTAGCGAGTT
Angiostrongylus malaysiensis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTCTAGTGA	CTGGCGAGTT
Angiostrongylus dujardini	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATCTCGATT	TCTTTAGTGG	CTAGCGAGTT
Angiostrongylus vasorum	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Aelurostrongylus abstrusus	GCGACTATTT	----GCTGGT	TTTCTGCAGA	GATTTTCGATT	TCTGTAGTGG	CTAGCGAGTT
Didelphostrongylus hayesi	GTGACTAGCA	--TTGCTGGT	TTTCTATTGA	GATTTTCGATT	TCATTAGTGA	CTAGCGAGTT
Crenosoma mephitiditis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Otostrongylus circumlitus	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Troglostrongylus wilsoni	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Filaroides martis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATCTCGATT	TCTTTAGTGG	CTAGCGAGTT
Oslerus osleri	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGA	CTAGCGAGTT
Parafilaroides decorus	GTGACTATTT	----GCTGGT	TTTCTATTGA	GATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Metastrongylus salmi	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Skrjabinstrongylus chitwoodorum	GTGACTATTT	----GCTGGT	TTTCTATTGA	GATTTTCGATT	TCATAGTGG	CTAGCGAGTT
Parelaphostrongylus odocoilei	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTATAGTGG	CTAGCGAGTT
Muellerius capillaris	GTGACTATTT	----GCTGGT	TTTCTATAGT	AATTTTCGATT	TCTATAGTGG	CTAGCGAGTT
Halocercus invaginatus	GTGACTATCT	RT--GCTGGT	TTTCTATTGA	GATTTTCGATT	TCATAGTGG	CTAGCGAGTT
Pseudalius inflexus	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTATAGTGG	CTAGCGAGTT
Stenurus minor	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Torynurus convolutus	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Nematodirus battus	GTGACTATTT	----GCTGGT	TTTCTATTGA	GGTTTCGATT	TCTTTAGTGG	CTAGCGAGTT
Nippostrongylus brasiliensis	GTGACTATTT	----GCTGGT	TTTCTATTAA	AGCTTCGGCT	TTTTTAGTGG	CTAGCGAGTT
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	730	740	750	760	770	780
Angiostrongylus cantonensis	TACTTTGAAT	AAATTAAAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Angiostrongylus costaricensis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Angiostrongylus malaysiensis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Angiostrongylus dujardini	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGA	TCGATCATGG
Angiostrongylus vasorum	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Aelurostrongylus abstrusus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGA	TCGATCATGG
Didelphostrongylus hayesi	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Crenosoma mephitiditis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Otostrongylus circumlitus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Troglostrongylus wilsoni	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Filaroides martis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Oslerus osleri	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Parafilaroides decorus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Metastrongylus salmi	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Skrjabinstrongylus chitwoodorum	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Parelaphostrongylus odocoilei	TACTTTGAAT	AAATTAAAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Muellerius capillaris	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTTA-CA	GCTTGAATGT	GTGATCATGG
Halocercus invaginatus	TACTTTGAAT	AAATTAGAGT	GCTCAAAACA	AGCGTTACGT	GCTTGAATGT	GTGATCATGG
Pseudalius inflexus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Stenurus minor	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Torynurus convolutus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Nematodirus battus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
Nippostrongylus brasiliensis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT---T	GCTTGAATGG	TCGATCATGG
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	790	800	810	820	830	840
Angiostrongylus cantonensis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Angiostrongylus costaricensis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Angiostrongylus malaysiensis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Angiostrongylus dujardini	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Angiostrongylus vasorum	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Aelurostrongylus abstrusus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Didelphostrongylus hayesi	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Crenosoma mephitiditis	AATAATAGAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Otostrongylus circumlitus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Troglostrongylus wilsoni	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Filaroides martis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Oslerus osleri	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Parafilaroides decorus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Metastrongylus salmi	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Skrjabinstrongylus chitwoodorum	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Parelaphostrongylus odocoilei	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Muellerius capillaris	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Halocercus invaginatus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Pseudalius inflexus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Stenurus minor	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Torynurus convolutus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Nematodirus battus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
Nippostrongylus brasiliensis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGGAAGTA	ATGATTAAGA
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	850	860	870	880	890	900
Angiostrongylus cantonensis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Angiostrongylus costaricensis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Angiostrongylus malaysiensis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Angiostrongylus dujardini	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Angiostrongylus vasorum	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Aelurostrongylus abstrusus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Didelphostrongylus hayesi	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Crenosoma mephitiditis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Otostrongylus circumlitus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Troglostrongylus wilsoni	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Filaroides martis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Oslerus osleri	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Parafilaroides decorus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Metastrongylus salmi	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Skrjabinngylus chitwoodorum	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Parelaphostrongylus odocoilei	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Muellerius capillaris	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Halocercus invaginatus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Pseudalius inflexus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Stenurus minor	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Torynurus convolutus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Nematodirus battus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Nippostrongylus brasiliensis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	
	910	920	930	940	950	960
Angiostrongylus cantonensis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Angiostrongylus costaricensis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Angiostrongylus malaysiensis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Angiostrongylus dujardini	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Angiostrongylus vasorum	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Aelurostrongylus abstrusus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Didelphostrongylus hayesi	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Crenosoma mephitiditis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Otostrongylus circumlitus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Troglostrongylus wilsoni	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Filaroides martis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Oslerus osleri	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Parafilaroides decorus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Metastrongylus salmi	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Skrjabinngylus chitwoodorum	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Parelaphostrongylus odocoilei	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Muellerius capillaris	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Halocercus invaginatus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Pseudalius inflexus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Stenurus minor	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Torynurus convolutus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Nematodirus battus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
Nippostrongylus brasiliensis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCTTCAT	TAATCAAGAA	CGAAAGTCAG	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	
	970	980	990	1000	1010	1020
Angiostrongylus cantonensis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Angiostrongylus costaricensis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Angiostrongylus malaysiensis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Angiostrongylus dujardini	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Angiostrongylus vasorum	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Aelurostrongylus abstrusus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Didelphostrongylus hayesi	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Crenosoma mephitiditis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Otostrongylus circumlitus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Troglostrongylus wilsoni	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Filaroides martis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Oslerus osleri	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Parafilaroides decorus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Metastrongylus salmi	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Skrjabinngylus chitwoodorum	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Parelaphostrongylus odocoilei	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Muellerius capillaris	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Halocercus invaginatus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Pseudalius inflexus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Stenurus minor	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Torynurus convolutus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Nematodirus battus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Nippostrongylus brasiliensis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1030	1040	1050	1060	1070	1080	
Angiostrongylus cantonensis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Angiostrongylus costaricensis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Angiostrongylus malaysiensis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Angiostrongylus dujardini	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Angiostrongylus vasorum	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Aelurostrongylus abstrusus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Didelphostrongylus hayesi	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Crenosoma mephitis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Otostrongylus circumlitus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Troglostrongylus wilsoni	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Filaroides martis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Oslerus osleri	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Parafilaroides decorus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Metastrongylus salmi	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Skrjabinogylus chitwoodorum	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Parelaphostrongylus odocoilei	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Muellerius capillaris	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Halocercus invaginatus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Pseudalius inflexus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Stenurus minor	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Torynurus convolutus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Nematodirus battus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
Nippostrongylus brasiliensis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCCT	
NUCELOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1090	1100	1110	1120	1130	1140	
Angiostrongylus cantonensis	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Angiostrongylus costaricensis	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Angiostrongylus malaysiensis	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Angiostrongylus dujardini	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Angiostrongylus vasorum	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Aelurostrongylus abstrusus	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Didelphostrongylus hayesi	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Crenosoma mephitis	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Otostrongylus circumlitus	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Troglostrongylus wilsoni	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Filaroides martis	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Oslerus osleri	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Parafilaroides decorus	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Metastrongylus salmi	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Skrjabinogylus chitwoodorum	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Parelaphostrongylus odocoilei	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Muellerius capillaris	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Halocercus invaginatus	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Pseudalius inflexus	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Stenurus minor	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Torynurus convolutus	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Nematodirus battus	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
Nippostrongylus brasiliensis	GGGGTAGTAT	GGTTGCAAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG	
NUCELOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1150	1160	1170	1180	1190	1200	
Angiostrongylus cantonensis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Angiostrongylus costaricensis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Angiostrongylus malaysiensis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Angiostrongylus dujardini	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Angiostrongylus vasorum	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Aelurostrongylus abstrusus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Didelphostrongylus hayesi	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Crenosoma mephitis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Otostrongylus circumlitus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Troglostrongylus wilsoni	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Filaroides martis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Oslerus osleri	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Parafilaroides decorus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Metastrongylus salmi	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Skrjabinogylus chitwoodorum	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Parelaphostrongylus odocoilei	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Muellerius capillaris	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Halocercus invaginatus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Pseudalius inflexus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Stenurus minor	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Torynurus convolutus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Nematodirus battus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
Nippostrongylus brasiliensis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCCG	GACACCGTAA	
NUCELOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	

	1210 1220 1230 1240 1250 1260
Angiostrongylus cantonensis	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Angiostrongylus costaricensis	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Angiostrongylus malaysiensis	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Angiostrongylus dujardini	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Angiostrongylus vasorum	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Aelurostrongylus abstrusus	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Didelphostrongylus hayesi	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Crenosoma mephitiditis	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Otostrongylus circumlitus	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Troglostrongylus wilsoni	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Filaroides martis	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Oslerus osleri	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Parafilaroides decorus	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Metastrongylus salmi	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Skrjabinngylus chitwoodorum	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Parelaphostrongylus odocoilei	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Muellerius capillaris	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Halocercus invaginatus	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Pseudalius inflexus	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Stenurus minor	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Torynurus convolutus	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Nematodirus battus	G-ATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Nippostrongylus brasiliensis	GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
NUCELOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	1270 1280 1290 1300 1310 1320
Angiostrongylus cantonensis	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Angiostrongylus costaricensis	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Angiostrongylus malaysiensis	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Angiostrongylus dujardini	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Angiostrongylus vasorum	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Aelurostrongylus abstrusus	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Didelphostrongylus hayesi	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Crenosoma mephitiditis	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Otostrongylus circumlitus	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Troglostrongylus wilsoni	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Filaroides martis	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Oslerus osleri	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Parafilaroides decorus	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Metastrongylus salmi	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Skrjabinngylus chitwoodorum	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Parelaphostrongylus odocoilei	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Muellerius capillaris	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Halocercus invaginatus	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Pseudalius inflexus	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Stenurus minor	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Torynurus convolutus	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Nematodirus battus	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Nippostrongylus brasiliensis	GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
NUCELOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	1330 1340 1350 1360 1370 1380
Angiostrongylus cantonensis	AGTGACTAGA TTATTGA--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Angiostrongylus costaricensis	AGTGACTAGA TTAAGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Angiostrongylus malaysiensis	AGTGACTAGA TTAAGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Angiostrongylus dujardini	AGTGACTAGA TTAAGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Angiostrongylus vasorum	AGTGACTAGA TTAAGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Aelurostrongylus abstrusus	AGTGACTAGA TTAAGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Didelphostrongylus hayesi	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Crenosoma mephitiditis	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Otostrongylus circumlitus	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Troglostrongylus wilsoni	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Filaroides martis	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Oslerus osleri	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Parafilaroides decorus	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Metastrongylus salmi	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Skrjabinngylus chitwoodorum	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Parelaphostrongylus odocoilei	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Muellerius capillaris	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Halocercus invaginatus	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Pseudalius inflexus	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Stenurus minor	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Torynurus convolutus	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Nematodirus battus	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
Nippostrongylus brasiliensis	AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCG GTGTTTA-GC
NUCELOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	1390	1400	1410	1420	1430	1440	
Angiostrongylus cantonensis	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Angiostrongylus costaricensis	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Angiostrongylus malaysiensis	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Angiostrongylus dujardini	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Angiostrongylus vasorum	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Aelurostrongylus abstrusus	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Didelphostrongylus hayesi	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Crenosoma mephitiditis	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Otostrongylus circumlitus	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Troglostrongylus wilsoni	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Filaroides martis	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Oslerus osleri	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Parafilaroides decorus	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Metastrongylus salmi	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Skrjabinogylus chitwoodorum	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Parelaphostrongylus odocoilei	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Muellerius capillaris	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Halocercus invaginatus	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Pseudalius inflexus	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Stenurus minor	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Torynurus convolutus	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Nematodirus battus	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
Nippostrongylus brasiliensis	CGCACGAGAT	TGAGCGATAA	CAGGTCTGTG	ATGCCCTTAG	ATGTCCGGGG	CTGCACGGGC	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1450	1460	1470	1480	1490	1500	
Angiostrongylus cantonensis	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Angiostrongylus costaricensis	GCTACAATGG	AAGAATCAAC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Angiostrongylus malaysiensis	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Angiostrongylus dujardini	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Angiostrongylus vasorum	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Aelurostrongylus abstrusus	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Didelphostrongylus hayesi	GCTACAATGG	AAGAATCAAC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Crenosoma mephitiditis	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Otostrongylus circumlitus	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Troglostrongylus wilsoni	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Filaroides martis	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Oslerus osleri	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Parafilaroides decorus	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Metastrongylus salmi	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Skrjabinogylus chitwoodorum	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Parelaphostrongylus odocoilei	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Muellerius capillaris	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Halocercus invaginatus	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Pseudalius inflexus	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Stenurus minor	GCTACAATGG	AAGATGCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Torynurus convolutus	GCTACAATGG	AAGATGCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Nematodirus battus	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
Nippostrongylus brasiliensis	GCTACAATGG	AAGAATCAGC	TGGCCTATCC	ATTGCCGAAA	GGTATTGGTA	AACCGTTGAA	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1510	1520	1530	1540	1550	1560	
Angiostrongylus cantonensis	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Angiostrongylus costaricensis	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Angiostrongylus malaysiensis	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Angiostrongylus dujardini	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Angiostrongylus vasorum	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Aelurostrongylus abstrusus	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Didelphostrongylus hayesi	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Crenosoma mephitiditis	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Otostrongylus circumlitus	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Troglostrongylus wilsoni	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Filaroides martis	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Oslerus osleri	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Parafilaroides decorus	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Metastrongylus salmi	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Skrjabinogylus chitwoodorum	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Parelaphostrongylus odocoilei	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Muellerius capillaris	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Halocercus invaginatus	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Pseudalius inflexus	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Stenurus minor	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Torynurus convolutus	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Nematodirus battus	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
Nippostrongylus brasiliensis	ACTCTTCCGT	GACCCGGGATA	GGGAATTGTA	ATTATTTCCT	TTGAACGAGG	AATTCCTAGT	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1570	1580	1590	1600	1610	1620
Angiostrongylus cantonensis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Angiostrongylus costaricensis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Angiostrongylus malaysiensis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Angiostrongylus dujardini	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Angiostrongylus vasorum	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Aelurostrongylus abstrusus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Didelphostrongylus hayesi	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Crenosoma mephitiditis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Otostrongylus circumlitus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Troglostrongylus wilsoni	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Filaroides martis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Oslerus osleri	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Parafilaroides decorus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Metastrongylus salmi	AAGTGTGAGT	CATCAGCTCA	CGTGTATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Skrjabinigylus chitwoodorum	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Parelaphostrongylus odocoilei	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Muellerius capillaris	AAGTGTGAGT	CATTAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Halocercus invaginatus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Pseudalius inflexus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Stenurus minor	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Torynurus convolutus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Nematodirus battus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
Nippostrongylus brasiliensis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1630	1640	1650	1660	1670	1680
Angiostrongylus cantonensis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Angiostrongylus costaricensis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Angiostrongylus malaysiensis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Angiostrongylus dujardini	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Angiostrongylus vasorum	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCAGGTC
Aelurostrongylus abstrusus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Didelphostrongylus hayesi	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TATATTGAGG	CCTTCGGGTC
Crenosoma mephitiditis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATTGAGG	CCTTCGGGTC
Otostrongylus circumlitus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATTGAGG	CCTTCGGGTC
Troglostrongylus wilsoni	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATTGAGG	CCTTCGGGTC
Filaroides martis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Oslerus osleri	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Parafilaroides decorus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	CGTATTGAGG	CCTTCGGGTC
Metastrongylus salmi	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Skrjabinigylus chitwoodorum	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATTGAGG	CCTTCGGGTC
Parelaphostrongylus odocoilei	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Muellerius capillaris	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Halocercus invaginatus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Pseudalius inflexus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Stenurus minor	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Torynurus convolutus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC
Nematodirus battus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATCGAGG	CCTTCGGGTC
Nippostrongylus brasiliensis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATCGAGG	CCTTCGGGTC
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1690	1700	1710	1720	1730	1740
Angiostrongylus cantonensis	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Angiostrongylus costaricensis	GCGATATAGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Angiostrongylus malaysiensis	GCAATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Angiostrongylus dujardini	GCGATATAGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Angiostrongylus vasorum	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	G.....
Aelurostrongylus abstrusus	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Didelphostrongylus hayesi	GTGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Crenosoma mephitiditis	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Otostrongylus circumlitus	GTGGTATAGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Troglostrongylus wilsoni	GCGGTATAGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Filaroides martis	GCGGTATGGC	GGGAAAACAG	TTCAATCGCA	ATGGCT-GAA	CCGGG-AAAA	GTCGT-AACA
Oslerus osleri	GTGATATGGT	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Parafilaroides decorus	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCG.....
Metastrongylus salmi	GCGGTATGGT	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Skrjabinigylus chitwoodorum	GCGATATAGT	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Parelaphostrongylus odocoilei	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-A...
Muellerius capillaris	GCGATATGGC	GGGAAA-CAG	TT.....
Halocercus invaginatus	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	G.....
Pseudalius inflexus	GTGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Stenurus minor	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Torynurus convolutus	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Nematodirus battus	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AACA
Nippostrongylus brasiliensis	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	-TCGT-AACA
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmm-mm	mm.....

	1750 1760 1770
Angiostrongylus cantonensis	AGGTATCTG.
Angiostrongylus costaricensis	AGGTATCTG.
Angiostrongylus malaysiensis	AGGTATCTG.
Angiostrongylus dujardini	AGGTATCTG.
Angiostrongylus vasorum
Aelurostrongylus ABSTRUSUS	AGGTATCTGT AGGTGAACCT GCAGATGGAT CATCG
Didelphostrongylus hayesi	AGGTATCT..
Crenosoma mephitiditis	AGGTATCT..
Otostrongylus circumlitus	AGGTATCTGT AGGTGAACCT GG.....
Troglostrongylus wilsoni	AGG.....
Filaroides martis	AG.....
Oslerus osleri	AGGTATCTGT AGGT.....
Parafilaroides decorus
Metastrongylus salmi	AGGTAT....
Skrjabingylus chitwoodorum	AGGTATCTGT AGG.....
Parelaphostrongylus odocoilei
Muellerius capillaris
Halocercus invaginatus
Pseudalius inflexus	AG.....
Stenurus minor	AGGTAT....
Torynurus convolutus	AGG.....
Nematodirus battus	AGGTATCTGT AGGTGAACCT GCAGATGGAT CATCG
Nippostrongylus brasiliensis	AGGTATCTGT AGGTGAACTG
NUCELOTIDES INCLUDED

Appendix 6.5: Log likelihood scores of the different models used for comparison for the Sub-order Metastrongylina using 1628 unambiguously aligned nucleotide sites from the near full-length SSU rRNA gene. Log Likelihood Ratio Tests revealed GTR+ Γ as the optimal model. Note that the K2P and F81 models are non-nested and therefore cannot be compared using LRT.

Model	Log-Likelihood Score
JC	4572.59040
JC+ Γ	4273.02522
F81	4555.55232
F81+ Γ	4256.28503
K2P	4495.17230
K2P+ Γ	4193.80011
HKY	4477.60518
HKY+ Γ	4172.09473
TN93	4475.64206
TN93+ Γ	4172.09461
GTR	4455.11204
GTR+Γ	4149.04297

Appendix 7.1: Log likelihood scores of the different models used for comparison for the (A) the Nematoda and (B) the Rhabditida within Nematoda using SSU rRNA gene to determine the phylogenetic positions of the nematode type sequences from global populations of *Achatina fulica*, African populations of *Achatina* sp. and Philippine populations of *Laevicaulis alte*. A total of 376 unambiguously aligned nucleotide sites of the 5' end of the SSU rRNA gene were used. Log Likelihood Ratio Tests for both the Nematoda and the Rhabditida reveal GTR+ Γ as the optimal model. Note that the K2P and F81 models are non-nested and therefore cannot be compared using LRT.

(A) Nematoda

(B) Rhabditida

Model	Log-Likelihood Score	Model	Log-Likelihood Score
JC	22626.94349	JC	14988.30034
JC+ Γ	20485.39787	JC+ Γ	13664.52274
F81	22551.59800	F81	14962.62351
F81+ Γ	20366.41757	F81+ Γ	13565.99042
K2P	22346.17796	K2P	14843.88334
K2P+ Γ	20156.78913	K2P+ Γ	13508.78184
HKY85	22261.34758	HKY	14795.57154
HKY85+ Γ	19983.22991	HKY+ Γ	13374.22988
TN93	22163.39719	TN93	14760.22240
TN93+ Γ	19957.42496	TN93+ Γ	13388.72096
GTR	22099.17630	GTR	14645.08708
GTR+Γ	19963.29509	GTR+Γ	13341.76120